

Supplementary Material

Lamin B1 attrition promotes early human papillomavirus infection through increased nuclear access and blunted autophagic capacity.

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24 **Supplementary Table 1** Overview p values of all statistical comparisons shown in the figures.

Fig. S1c	x	y	p value	Sign.		x	y	p value	Sign.
Nuclear circ.	CTRLko	LMNAko	<1 ^e -10	***	Mean lamin A/C int.	CTRLko	LMNAko	<1 ^e -10	***
	CTRLko	LMNB1ko	<1 ^e -10	***		CTRLko	LMNB1ko	<1 ^e -10	***
	CTRLko	LMNB2ko	<1 ^e -10	***		CTRLko	LMNB2ko	<1 ^e -10	***
	LMNAko	LMNB1ko	<1 ^e -10	***		LMNAko	LMNB1ko	<1 ^e -10	***
	LMNAko	LMNB2ko	<1 ^e -10	***		LMNAko	LMNB2ko	<1 ^e -10	***
	LMNB1ko	LMNB2ko	<1 ^e -10	***		LMNB1ko	LMNB2ko	<1 ^e -10	***
Fig. 1a	x	y	p value	Sign.		x	y	p value	Sign.
Infection rate	CTRLkd	LMNAkd	0.99913	n.s.	Mean EGFP int.	CTRLkd	LMNAkd	0.98887	n.s.
	CTRLkd	LMNB1kd	<0.001	***		CTRLkd	LMNB1kd	<0.001	***
	CTRLkd	LMNB2kd	0.99801	n.s.		CTRLkd	LMNB2kd	0.99070	n.s.
Infection rate	CTRLko	LMNAko	0.166	n.s.	Mean EGFP int.	CTRLko	LMNAko	0.1182	n.s.
	CTRLko	LMNB1ko	<0.001	***		CTRLko	LMNB1ko	<0.001	***
	CTRLko	LMNB2ko	0.989	n.s.		CTRLko	LMNB2ko	1.000	n.s.
Fig. 1c	x	y	p value	Sign.		x	y	p value	Sign.
Infection rate	CTRLkd	LMNB1ko	0.00645	**	EGFP intensity	CTRLko	LMNB1ko	0.00322	**
Fig. 2a	x	y	p value	Sign.		x	y	p value	Sign.
Doubling time kd	CTRLkd	LMNAkd	0.94	n.s.	Doubling time ko	CTRLko	LMNAko	0.88	n.s.
	CTRLkd	LMNB1kd	0.94	n.s.		CTRLko	LMNB1ko	0.88	n.s.
	CTRLkd	LMNB2kd	0.94	n.s.		CTRLko	LMNB2ko	0.88	n.s.
Fig. 2c	x	y	p value	Sign.	Fig. 2d	x	y	p value	Sign.
Mitotic window	CTRLko	LMNB1ko	9.271 ^e -07	***	EdU spots	CTRLko	LMNB1ko	0.04677	*
Fig. 2f	x	y	p value	Sign.		x	y	p value	Sign.
Fraction cells with ruptures	CTRLko	LMNAko	0.193	n.s.	Rate rerupture	CTRLko	LMNAko	0.921	n.s.
	CTRLko	LMNB1ko	<0.001	***		CTRLko	LMNB1ko	<1 ^e -05	***
	CTRLko	LMNB2ko	1.000	n.s.		CTRLko	LMNB2ko	1.000	n.s.

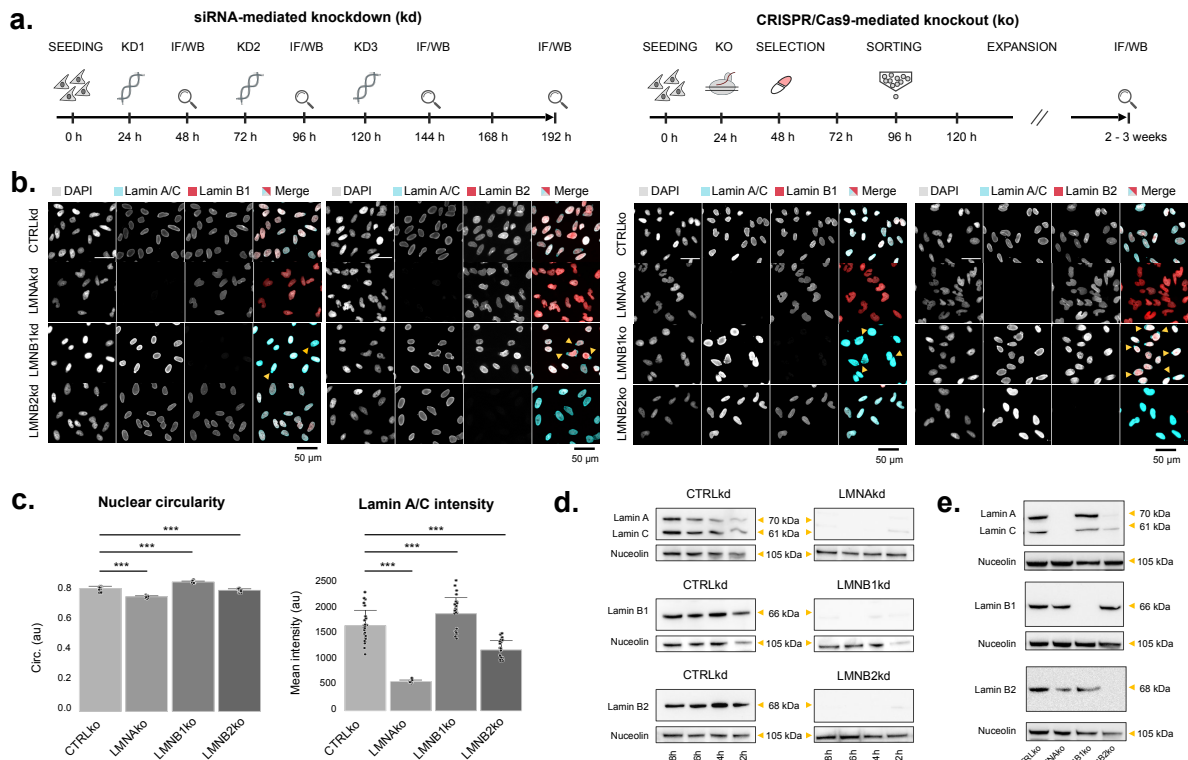
Fig. 2f	x	y	p value	Sign.				
EdU spots	CTRLko	LMNB1ko	0.002381	**				
Fig. S3	x	y	p value	Sign.		x	y	p value Sign.
+HPV PsV	CTRLko	LMNB1ko	0.688	n.s.	EGFP+	CTRLko	LMNB1ko	0.454 n.s.
Fig. 3a	x	y	p value	Sign.		x	y	p value Sign.
-HPV/	CTRLko	LMNB1ko	0.00479	**	EGFP-	CTRLko	LMNB1ko	0.234 n.s.
+HPV PsV	- HPV	+HPV	0.4271	n.s.	/EGFP+	EGFP-	EGFP+	0.645 n.s.
Fig. 3c	x	y	p value	Sign.		x	y	p value Sign.
-HPV/	CTRLko	LMNB1ko	<2 ^e -16	***	EGFP-	CTRLko	LMNB1ko	2.6 ^e -12 ***
+HPV PsV	- HPV	+HPV	0.15	n.s.	/EGFP+	EGFP-	EGFP+	2 ^e -16 ***
Fig. 3d	x	y	p value	Sign.		x	y	p value Sign.
24h	CTRLko	CTRLko	1.000	n.s.	48h	CTRLko	CTRLko	0.31976 n.s.
	CTRLkd	PMLkd				CTRLkd	PMLkd	
	CTRLko	LMNB1ko	0.01240	*		CTRLko	LMNB1ko	<0.001 ***
	CTRLkd	CTRLkd				CTRLkd	CTRLkd	
	CTRLko	LMNB1ko	0.99638	n.s.		CTRLko	LMNB1ko	0.94222 n.s.
	CTRLkd	PMLkd				CTRLkd	PMLkd	
	CTRLko	LMNB1ko	0.00666	**		CTRLko	LMNB1ko	<0.001 ***
	PMLkd	CTRLkd				PMLkd	CTRLkd	
	CTRLko	LMNB1ko	0.98656	n.s.		CTRLko	LMNB1ko	0.01488 *
	PMLkd	PMLkd				PMLkd	PMLkd	
	LMNB1ko	LMNB1ko	0.11160	n.s.		LMNB1ko	LMNB1ko	<0.001 ***
	CTRLkd	PMLkd				CTRLkd	PMLkd	
Fig. 4a	x	y	p value	Sign.				
Prot. degr.	DMSO	MG132	<2 ^e -16	***				
Fig. 4b	x	y	p value	Sign.		x	y	p value Sign.
DMSO	CTRLko	CTRLko	0.5669	n.s.	RM/CQ	CTRLko	CTRLko	0.869 n.s.
	HPV PsV-	HPV PsV+				HPV PsV-	HPV PsV+	
	LMNB1ko	LMNB1ko	0.0147	*		LMNB1ko	LMNB1ko	0.960 n.s.
	HPV PsV-	HPV PsV+				HPV PsV-	HPV PsV+	

	CTRLko	LMNB1ko	0.9982	n.s.		CTRLko	LMNB1ko	<1 ^e -05	***
	HPV-	HPV PsV-				HPV-	HPV PsV-		
	CTRLko	LMNB1ko	0.4210	n.s.		CTRLko	LMNB1ko	<1 ^e -05	***
	HPV PsV+	HPV PsV+				HPV PsV+	HPV PsV+		
Fig. S5	x	y	p value	Sign.		x	y	p value	Sign.
DMSO	CTRLko	LMNB1ko	0.000653	***	RM/CQ	CTRLko	LMNB1ko	0.00628	**
Fig. 5c	x	y	p value	Sign.		x	y	p value	Sign.
-HPV/	CTRLko	LMNB1ko	0.516	n.s.	EGFP-	CTRLko	LMNB1ko	0.847	n.s.
+HPV PsV	-HPV	+HPV	0.124	n.s.	/EGFP+	EGFP-	EGFP+	<2 ^e -16	***
Fig. 5d	x	y	p value	Sign.		x	y	p value	Sign.
-HPV/	CTRLko	LMNB1ko	0.0238	*	EGFP-	CTRLko	LMNB1ko	0.0231	*
+HPV PsV	-HPV	+HPV	0.00814	**	/EGFP+	EGFP-	EGFP+	<2 ^e -16	***

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Supplementary Figures



Supplementary Figure S1. Loss of individual lamins provokes distinct morphological

changes. **a)** Workflow for establishment of lamin-depleted cells. Repetitive siRNA-mediated

genome silencing was used to establish a sustained knockdown (kd). HeLa cells were seeded

at time point 0 h and three subsequent knockdowns were performed on 24 h, 72 h and 120 h.

At different timepoints after cell seeding (*i.e.*, 48 h, 96 h, 144 h and 192 h), readouts were

obtained by immunofluorescent staining (IF) and western blot (WB). CRISPR/Cas9 genome

editing was used to obtain stable knockout (ko) cells. HeLa cells were seeded and CRISPR/Cas9

constructs were transfected 24 h later. Puromycin was used to enrich transfected cells prior to

single cell selection with FACS. Clonal expansion was followed by quantification of these

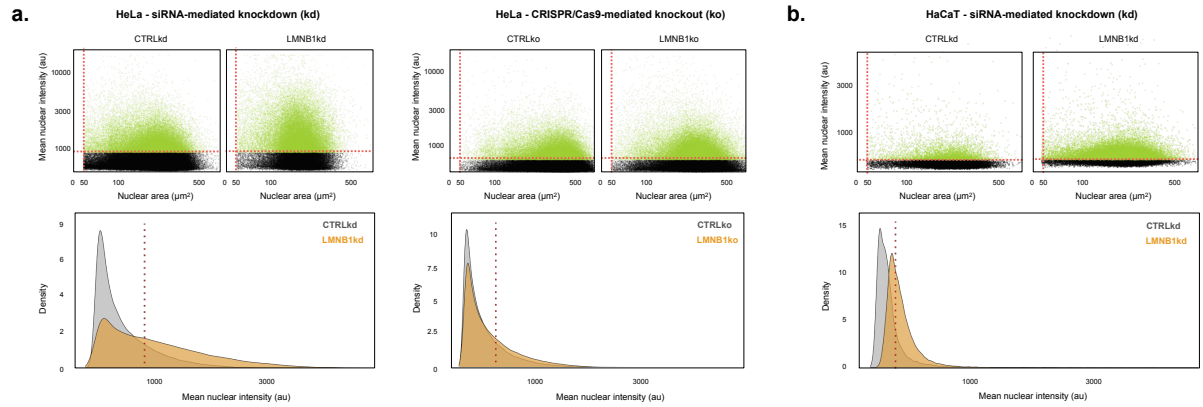
different colonies by IF and WB; **b)** IF reveals the absence of the protein of the respective target

genes and reveals significant morphological nuclear changes (insets). While CTRL cells mainly

display oval-shaped nuclei, *LMNA* depleted cells show dysmorphic nuclei with local loss of B-

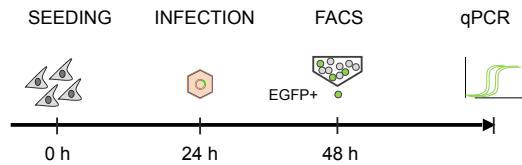
type lamins. Depletion of *LMNB1* provokes ample nuclear blebs and yields an increased lamin

42 A/C signal. Depletion of *LMNB2* does not result in overt changes; **c)** Nuclear circularity and
43 lamin A/C levels of lamin-depleted cells differ significantly from CTRL cells ($n_{\text{bio}} = 3$, $n_{\text{tech}} =$
44 5, $p < 0.05$, linear mixed effects model); **d)** WB confirms the virtual absence of the target
45 proteins in sustained kd as well as in **e)** stable ko cells.

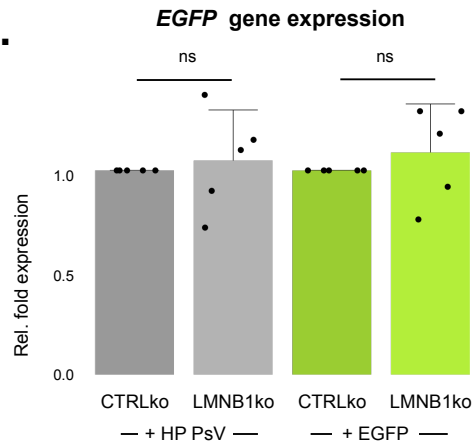


Supplementary Figure S2. Scatterplots and corresponding density plots illustrating the distribution of EGFP intensity as a function of the nuclear area of HP-PsV infected HeLa (a) and HaCaT (b) cells with and without depletion of lamin B1. The red dotted lines indicate fixed cutoffs for cell detection (min. area) or positively infected cells (min. EGFP intensity) ($n_{\text{bio}} = 2$).

a.



b.



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54 **Supplementary Figure S3.** HP-PsV-infected *LMNB1* depleted cells do not show higher EGFP

55 gene expression levels. **a)** Workflow to determine the number of EGFP transcripts. CTRLko

56 and LMNB1ko cells were infected with HP-PsV for 24 h before cells were sorted with FACS

57 based on their EGFP-signal. Stringent selection criteria were used to sort true EGFP-positive

58 (EGFP+) and -negative (EGFP-) cells. RNA was collected from the selected cell populations

59 as well as from pooled, non-sorted HP-PsV infected CTRLko and LMNB1ko cells and used for

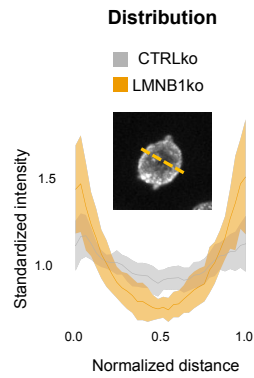
60 qPCR; **b)** The number of EGFP transcripts in LMNB1ko cells (expressed as fold change with

61 respect to the CTRL and normalized to the reference genes, $2^{-\Delta\Delta Ct}$) is not significantly

62 increased, not in pooled conditions, nor in flow-sorted EGFP-positive cells ($n = 5$, one sample

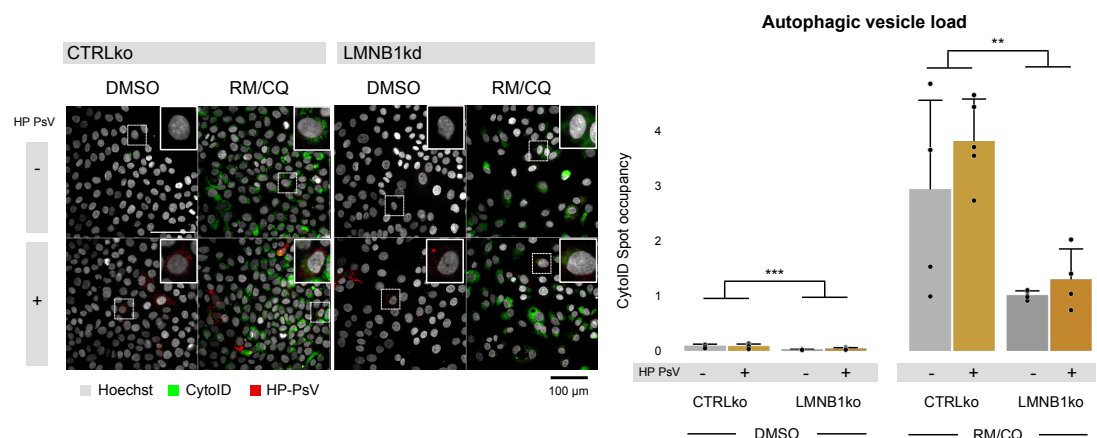
63 T-test, with Shapiro-Wilk normality test).

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66 **Supplementary Figure S4.** LMNB1ko cells show a higher accumulation of perinuclear
 67 methylated chromatin marker. The intensity of H3K9me23 is measured along the distance-
 68 normalized cross section of the nucleus and is standardized by the mean intensity per nucleus
 69 (n=20 cells).



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72 **Supplementary Figure S5.** Autophagic capacity is blunted in *LMNB1* depleted HaCaT cells.

73 Fluorescent images of cells stained with CytoID show strong enrichment of autophagic

74 vacuoles after treatment with a combination of Rapamycin (RM) (*i.e.*, inducer of autophagy)

75 and Chloroquine (CQ) (*i.e.*, inhibitor of lysosomal degradation) in CTRLko cells, but much less

76 prominent in LMNB1ko cells. This significant difference in autophagic capacity is also

77 confirmed in the quantification of the cellular CytoID spot occupancy ($n_{\text{bio}} = 3$, $n_{\text{tech}} = 5$, two-

78 way Anova with Tukey post hoc).