

New insights into the mechanisms underlying 5-fluorouracil-induced intestinal toxicity based on transcriptomic and metabolomic responses in human intestinal organoids

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Journal: *Archives of Toxicology*

Supplementary Information

Supplementary Table 1. Input parameters for 5-fluorouracil (5-FU) human physiologically-based pharmacokinetic (PBPK) modelling in Simcyp and *in vitro* distribution modelling in SIVA (module 3). The virtual *in vitro* intracellular distribution (VIVD) model was developed to describe *in vitro* distribution in 2D cell culture systems (Fisher et al. 2019). Therefore, in this work, the model was used as an initial guide to select nominal concentrations for *in vitro* organoid studies.

Input parameters for 5-FU kinetic modelling		
Physicochemical	Molecular weight (Kim et al. 2019)	130.08 g/mol
	Lipophilicity (Kim et al. 2019) (Log $P_{o:w}$)	-0.89
	Compound type (Kim et al. 2019) pKa	Monoprotic acid 8.02
	Henry's law constant (25 °C)*	1.68×10^{-5} Pa m ³ /mol
Blood Binding	Blood to plasma ratio (Schaaf et al. 1987)	1.09
	Fraction unbound in plasma (Celio et al. 1983; Garrett et al. 1977)	0.9
Absorption	M-ADAM and MechPeff model (Pade et al. 2017) <i>Passive permeability ($P_{trans, 0}$) predicted from Log $P_{o:w}$</i>	10.3×10^6 cm/s
Distribution	Rodgers & Rowland model (Rodgers et al. 2005; Rodgers and Rowland 2006) <i>K_p scalar adjusted to replicate volume of distribution observed⁶</i>	0.2 L/kg
Elimination	Liver cytosol metabolism (V_{max} and K_m) (Niwa et al. 2005)	493 pmol/min/mg protein 3.9 μ M
	Renal clearance (Heggie et al. 1987)	8 L/hr
	Additional systemic clearance <i>Optimised to recover observed intravenous clearance (Heggie et al. 1987)</i>	37 L/hr

* Estimated from EPI Suite v4.11 using HENRYWIN v3.20 bond method

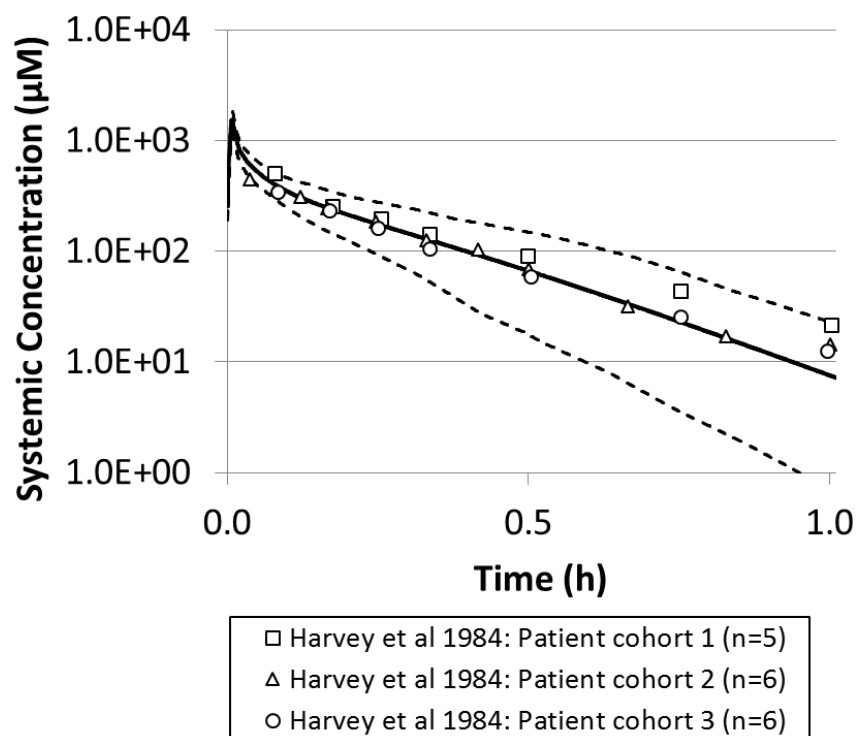
Supplementary Table 2. Human gut cell composition and culture conditions considered for 5-FU *in vitro* distribution modelling in SIVA.

VIVD input parameters

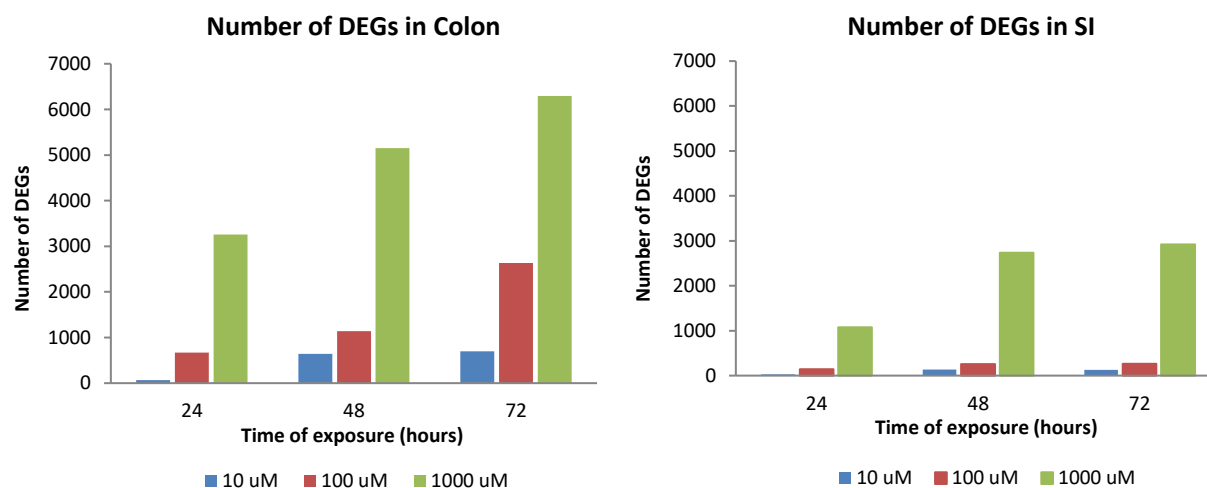
Cell composition¹	Fraction of cell volume comprising intracellular water	0.451
	Fraction of cell volume comprising neutral lipids	0.0487
	Fraction of cell volume comprising neutral phospholipids	0.0163
	Fraction of cell volume comprising lysosomes	0.01
	Fraction of cell volume comprising mitochondria	0.1
	pH of intracellular water	7
	pH inside lysosomes	4
	pH inside mitochondria	8
	Cell membrane potential	-41 mV
	Lysosome membrane potential	10 mV
	Mitochondria membrane potential	120 mV
	Intracellular concentration of acidic phospholipids	2.84 mg/g
Culture conditions	Fraction unbound in the culture medium ²	Assumed to be 1
	pH of culture medium	7.4
	Volume of culture medium per well	100 μ L
	Well diameter	6.4 mm
	Well volume	360 μ L
	Culture temperature	37 °C
	Number of cells per well	3000
	Cell diameter ³	5.3 μ m

1- Human gut tissue cell composition in the Simcyp Simulator (v18r2); 2 - No albumin or lipids present in the medium;

3 - Calculated based on average organoid area (4000 μ m²) and number of cells per organoid (180 cells).



Supplementary Figure 1. Verification of the 5-FU human PBPK model: predicted (black solid line) and observed (data points) systemic plasma concentration following a 15 mg/kg (609 mg/m²) IV bolus. The observed data was not used in the development or optimisation of the model, only in the verification of its performance. The markers show the mean concentration obtained in different studies with cancer patients (Harvey et al. 1984). The dashed lines refer to the 5th and 95th percentiles of the virtual population simulated (10 trials with 10 cancer patients each, 50% female).

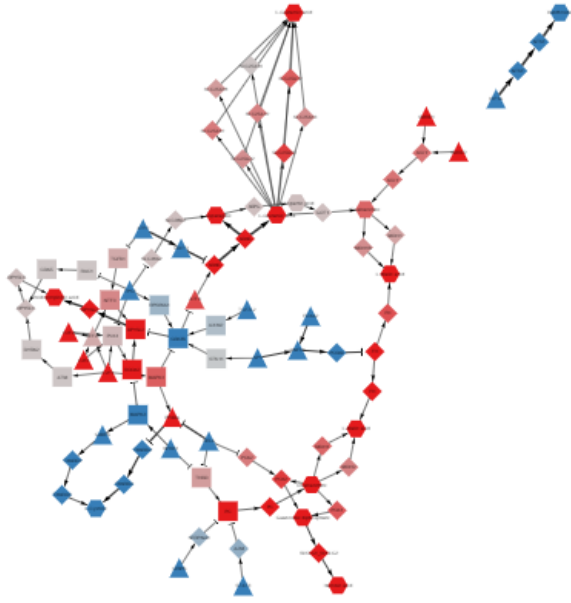
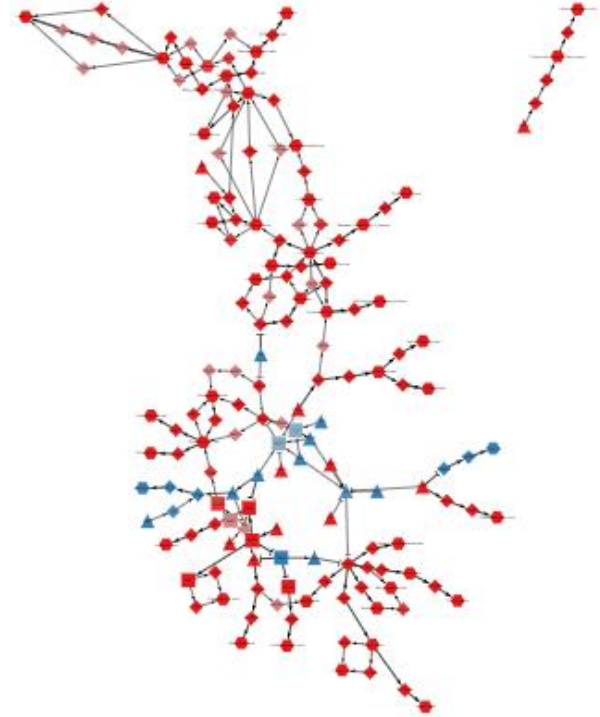


Supplementary Figure 2. Number of DEGs detected at each time point and concentration of 5-FU for colon and SI: blue corresponds to 10 μM , red corresponds to 100 μM and green corresponds to 1000 μM . Number of DEGs was corrected using the Bonferroni method.

A complex network graph visualization. The graph consists of numerous nodes, colored red and blue, connected by a dense web of edges. The nodes are distributed across the frame, with a large, dense central cluster and several smaller, more isolated clusters. The edges are thin, dark lines connecting the nodes, creating a complex, interconnected structure. The overall shape is roughly circular, with many branches extending outwards from the center. The red nodes are more numerous than the blue nodes, and they are more concentrated in the central area. The blue nodes are more spread out, particularly in the lower right and upper right regions. The edges are most dense in the central area, where many nodes are connected to multiple other nodes, and become sparser as they move towards the periphery. The background is plain white, making the network structure stand out clearly.

A complex network graph visualization. The nodes are colored red, blue, and grey, and are connected by edges. The graph shows a dense, branching structure with many interconnected nodes and edges, suggesting a highly complex network.

The figure displays a network graph with a large central cluster and several smaller, disconnected components. The nodes are colored red, blue, and black, and shaped as circles, squares, and triangles. The edges are black lines connecting the nodes. The central cluster is the most dense, with many nodes and edges. There are several smaller components, some of which are isolated from the main cluster. The nodes are distributed across the image, with a higher concentration in the center and some isolated nodes on the right side.

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Supplementary Figure 4. COSMOS networks obtained after multi-omics integration considering the high concentration of 5-FU and all time points: A, B and C are respective to colon organoids at 24h, 48h and 72h, respectively; D, E and F are respective to SI organoids at 24h, 48h and 72h, respectively. These networks can be further explored elsewhere.