

1 **Supplementary Information**

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3 **Bacteria associated with spores of arbuscular mycorrhizal fungi: a review.**

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5 Biology and Fertility of Soils

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7 Monica Agnolucci¹, Arianna Grassi^{1*}, Manuela Giovannetti¹, Olga A. Lastovetsky^{2*}

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9 ¹ Department of Agriculture, Food and Environment, University of Pisa, Via del Borghetto 80, 56124 Pisa, Italy

10 ² School of Biology and Environmental Science, University College Dublin, Belfield, Dublin 4, Ireland

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12 *Corresponding authors: Olga A. Lastovetsky, School of Biology and Environmental Science, University College Dublin,

13 Belfield, Dublin 4, Ireland.: Tel.: +35 37162392. Email address: olga.lastovetsky@ucd.ie

14 Arianna Grassi, Department of Agriculture, Food and Environment, University of Pisa, Via del Borghetto 80, 56124 Pisa,

15 Italy. Tel.: +39 050 2216640. Email address: arianna.grassi@phd.unipi.it

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17 **Supplementary Table:**

18 *Table S1*

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32 **Table S1.** Occurrence and functional properties of bacteria (excluding known endobacteria *CaGg* and *CaMg*) associated
33 with AMF spores, as detected by transmission electron microscopy (TEM), culture-dependent (CD) and culture-
34 independent (CI) methods. The names of AMF taxa refer to the current names of each species (see INVAM 20024 and
35 amf-phylogeny.com).

AMF species	Bacterial taxa (genus/species/strain)	Functional characterization	Decontamination procedure and molecular method	References
<i>Funneliformis caledonius</i>	Unidentified bacteria embedded in spore wall layers	-	TEM Surface- decontaminated spores (20 min with 2% chloramine T + 0.02% streptomycin sulphate + trace of Teepol by the "filter paper sandwich method")	Macdonald and Chandler 1981
<i>Diversispora epigaea</i>	<i>Pseudomonas</i> sp. <i>Corynebacterium</i> spp. Lactobacillaceae	Spore germination increases	CD Surface- decontaminated spores (5 min with sodium hypochlorite 0.05%)	Mayo et al. 1986
<i>Rhizophagus clarus</i>	Unidentified bacteria embedded in spore wall layers	-	TEM Surface- decontaminated spores (SDS wash, multiple 20–30 min chloramine-T exposures)	Walley and Germida 1995
<i>Funneliformis mosseae</i>	Unidentified bacteria embedded in spore wall layers	-	TEM Spore without surface decontamination	Filippi et al. 1998
<i>R. clarus</i> NT4	<i>Arthrobacter ilicis</i> WA2 <i>Bacillus alvei</i> TA8 <i>B. brevis</i> WA4 <i>B. chitinosporus</i> LA6a <i>B. circulans</i> WA3 <i>B. firmus</i> YA5 <i>B. laterosporus</i> LA6b <i>B. longisporus</i> LA4 <i>B. megaterium</i> LA1 <i>B. pabuli</i> LA3	Variable effects on spore germination, depending on the strain	CD Surface- decontaminated spores (60 min with chloramine-T and streptomycin)	Xavier and Germida 2003
<i>R. clarus</i> NT4	<i>B. pabuli</i> LA3	Increase in plant shoot N and P content	CD Surface- decontaminated spores	Id.

			(60 min with chloramine-T + streptomycin)	
<i>Funneliformis geosporus</i> and <i>Septoglomus constrictum</i>	<i>Cellvibrio</i> , <i>Chondromyces</i> , <i>Flexibacter</i> , <i>Lysobacter</i> , <i>Pseudomonas</i>	-	CI (PCR-DGGE) (Spores rinsed 3 times in sterile water)	Roesti et al. 2005
<i>Funneliformis mosseae</i>	<i>Arthrobacter atrocyneus</i> <i>A. globiformis</i> <i>A. ilicis</i> <i>A. oxydans</i> <i>A. ramosus</i> <i>Micrococcus halobius</i> <i>M. kristinae</i> <i>M. roseus</i> <i>Aureobacterium saperdae</i> <i>Curtobacterium citreum</i> <i>Cellulomonas flavigena</i> <i>Bacillus brevis</i> <i>B. megaterium</i> GC subgroup A <i>B. megaterium</i> GC subgroup B <i>Agrobacterium radiobacter</i> <i>Burkholderia pickettii</i> <i>Acidovorax delafieldii</i> <i>Hydrogenophaga pseudoflava</i> <i>Pseudomonas alcaligenes</i> <i>P. chlororaphis</i> <i>P. pseudoalcaligenes</i> <i>P. putida</i> biotype B <i>P. stutzeri</i> <i>Stenotrophomonas maltophilia</i>	10% of the isolates showed <i>in vitro</i> growth inhibition of the pathogen <i>Rhizoctonia solani</i>	CD Surface-decontaminated spores (30 washes with sterile potassium phosphate)	Bharadwaj et al. 2008
<i>Rhizophagus intraradices</i>	<i>A. globiformis</i> <i>A. ilicis</i> <i>A. oxydans</i> <i>A. ramosus</i> <i>Micrococcus kristinae</i> <i>M. luteus</i> GC subgroup C <i>M. roseus</i> <i>Clavibacter michiganense insidiosum</i> <i>Cellulomonas flavigena</i> <i>Corynebacterium bovis</i> <i>Bacillus brevis</i> <i>B. chitinosporus</i> <i>B. subtilis</i> <i>Agrobacterium radiobacter</i> <i>Achromobacter piechaudii</i>	40% of the isolates showed <i>in vitro</i> growth inhibition of the pathogen <i>Rhizoctonia solani</i>	CD Surface-decontaminated spores (30 washes with sterile potassium phosphate)	Id.

	<i>A. xylosoxydans</i> <i>denitrificans</i> <i>Janthinobacterium lividum</i> <i>Pseudomonas alcaligenes</i> <i>P. fluorescens</i> biotype F <i>P. pseudoalcaligenes</i> <i>P. putida</i> biotype A <i>P. putida</i> biotype B <i>P. syringae</i> tagetes <i>Stenotrophomonas maltophilia</i>			
<i>R. intraradices</i>	<i>Stenotrophomonas maltophilia</i> FWC14 <i>Pseudomonas putida</i> biotype B FWC16 <i>P. fluorescens</i> biotype F FWC30 <i>Bacillus subtilis</i> FWC42 <i>P. putida</i> biotype A FWC70 <i>Arthrobacter ilicis</i> <i>S. maltophilia</i> LWC2	All the isolates showed <i>in vitro</i> growth inhibition of <i>Erwinia carotovora</i> var <i>carotovora</i> (Ecc), <i>P. infestans</i> , <i>Verticillium dahliae</i> and <i>Rhizoctonia solani</i> and production of siderophores	CD Surface-decontaminated spores (30 washes with sterile potassium phosphate)	Id.
<i>R. intraradices</i>	<i>P. putida</i> biotype A	Increased mycorrhizal colonization, number of primary and lateral roots, root and shoot length, and number of leaves in potato plants	CD Surface-decontaminated spores (30 washes with sterile potassium phosphate)	Id.
<i>Gigaspora margarita</i>	<i>Janthinobacterium lividum</i> <i>Paenibacillus polymyxa</i>	P solubilisation and <i>in vitro</i> growth inhibition of the phytopathogens <i>Pythium ultimum</i> , <i>Fusarium oxysporum</i> , <i>Rhizoctonia solani</i>	CD Surface-decontaminated spores (30 min with chloramine-T + streptomycin + chloramphenicol)	Cruz et al. 2008
<i>G. margarita</i>	<i>Amycolatopsis</i> sp. <i>Azospirillum</i> sp. <i>Flexibacter</i> sp. <i>Polyangium</i> sp. <i>Pseudonocardia</i> sp. <i>Ramlibacter</i> sp. <i>Rhizobium</i> sp. <i>Rubrivirax</i> spp. <i>Sphingomonas</i> sp. <i>Streptomyces</i> sp.	-	CI (PCR-DGGE) Bacteria colonizing spore surfaces (3 washes in water, followed by vortexing and analysis of the supernatant)	Long et al. 2008
<i>G. margarita</i>	<i>Bacillus</i> sp. BD-107 <i>Bacillus megaterium</i> EG24 <i>Bacillus subtilis</i> DF61 <i>Bacillus flexus</i> KSC_SF9c	<i>B. megaterium</i> increased AMF colonization. <i>B. megaterium</i> and <i>Bacillus</i> BD-107	CD Surface-decontaminated spores	Budi et al. 2012

		increased shoot and dry weights of Neem plants	(ethanol, chloramine-T, + streptomycin + gentamycin)	
<i>G. margarita</i>	<i>Bacillus</i> sp. <i>Bacillus thuringiensis</i> <i>Paenibacillus rhizosphaerae</i>	Promotion of hyphal growth P solubilisation Variable antagonistic activity against phytopathogens	CD Surface-decontaminated spores (ethanol chloramine-T + streptomycin + chloramphenicol)	Cruz and Ishii 2011
<i>Funneliformis coronatum</i> IMA3	<i>Agrobacterium</i> , <i>Artrobacter</i> , <i>Sinorhizobium</i>	-	CI (PCR-DGGE) Surface-decontaminated spores (15 washes by 1500 rpm vortexing in sterile physiological solution)	Agnolucci et al. 2015
<i>F. mosseae</i> AZ225C	<i>Acidobacteria</i> , <i>Agrobacterium</i> , <i>Artrobacter</i> , <i>Bacteroidetes</i> , <i>Duganella</i> , <i>Herbaspirillum</i> , <i>Ideonella</i> , <i>Rhizobium</i> , <i>Sinorhizobium</i> , <i>Streptomyces</i> , <i>Paenibacillus</i>	-	Id.	Id.
<i>F. mosseae</i> IMA1	<i>Amycolatopsis</i> , <i>Bacteroidetes</i> , <i>Deltaproteobacteria</i>	-	Id.	Id.
<i>F. mosseae</i> IN101C	<i>Propionibacterium</i> , <i>Methylibium</i>	-	Id.	Id.
<i>R. intraradices</i> IMA5	<i>Sinorhizobium</i> , <i>Pseudomonas</i> , <i>Streptomyces</i> , <i>Deltaproteobacteria</i>	-	Id.	Id.
<i>R. intraradices</i> IMA6	<i>Artrobacter</i> , <i>Bacillus</i> , <i>Herbaspirillum</i> , <i>Massilia</i> , <i>Pseudomonas</i> , <i>Rhizobium</i> , <i>Streptomyces</i>	-	Id.	Id.
<i>R. intraradices</i> IMA6	<i>Arthrobacter phenanthrenivorans</i> N17 <i>Bacillus pumilus</i> CH10 <i>Fictibacillus barbaricus</i> TSA50 <i>Lysinibacillus fusiformis</i> CH19 <i>Nocardioideis albus</i> N13 <i>Sinorhizobium meliloti</i> CH5, CH8, CH17, N23, N28, N29, TSA3, TSA26, TSA41	19 out of 374 isolates showed high PGP activities: production of siderophores and IAA, solubilisation of mineral and organic P	CD Surface-decontaminated spores (15 washings by 1500 rpm vortexing in sterile physiological solution)	Battini et al. 2016

	<i>Streptomyces</i> W43N, W64, W77, W94, W115			
<i>F. caledonius</i> <i>Racocetra</i> <i>alborosea</i> <i>F. mosseae</i>	<i>Bacillus anthracis</i> <i>B. aryabhatai</i> <i>Lysinibacillus fusiformis</i>		CD Surface- decontaminated spores (<i>chloramine-T</i> + <i>streptomycin</i>)	Selvakumar et al. 2016
<i>G. margarita</i>	<i>Achromobacter</i> <i>Aquitalea</i> <i>Bosea</i> <i>Burkholderia</i> <i>Cupriavidus</i> <i>Ensifer</i> <i>Lysobacter</i> <i>Mitsuaria</i> <i>Proteus</i> <i>Pseudomonas</i> <i>Ralstonia</i> <i>Rhizobium</i> <i>Amycolatopsis</i> <i>Arthrobacter</i> <i>Curtobacterium</i> <i>Gordonia</i> <i>Leifsonia</i> <i>Mycobacterium</i> <i>Nocardia</i> <i>Streptomyces</i> <i>Bacillus</i> <i>Brevibacillus</i> <i>Paenibacillus</i>	P solubilisation, chitin degradation, stimulation of spore germination and hyphal growth, varying across the isolates	CD Surface- decontaminated spores (<i>washed 3 times in</i> <i>sterile NaCl</i>)	Long et al. 2017
<u><i>Gigaspora</i></u> <u><i>Racocentra</i></u> <u><i>Scutellospora</i></u> <u><i>Acaulospora</i></u> <u><i>Dentiscutata</i></u>	<i>Burkholderia</i> species		CI (Sanger sequencing) Surface- decontaminated spores (<i>chloramine-T</i> + <i>H₂O₂</i>)	Lastovetsky et al. 2018
<i>Acaulospora</i> <i>Ambispora</i> <i>Diversispora</i> <i>Funneliformis</i> <i>Glomus</i> <i>Paraglomus</i> <i>Scutellospora</i>	<i>Acidobacteria</i> , <i>Proteobacteria</i> <i>Verrucomicrobia</i> <i>Alphaproteobacteria</i> (<i>Rhizobiales</i> , <i>Rhodospirales</i>) <i>Gammaproteobacteria</i> (<i>Escherichia</i> , <i>Aquicella</i> , <i>Pseudomonas</i>) <i>Actinobacteria</i>	-	CI (Illumina MiSeq) Surface- decontaminated spores (<i>chloramine-T</i> + <i>H₂O₂</i>)	Lastovetsky et al. 2024
Glomerales Paraglomerales Archaeosporales Diversisporales	<i>Burkholderiales</i> <i>Micromonosporales</i> <i>Bacillales</i> <i>Pseudomonadales</i> <i>Flavobacteriales</i> <i>Aeromonadales</i>		CI (Illumina MiSeq) Surface- decontaminated (<i>chloramine-T</i> + <i>H₂O₂</i> + <i>chlorphenamine</i>)	Chávez- González et al. 2024

	<i>Lactobacilliales</i> <i>Streptomycetales</i> <i>pasteurellales</i> <i>Fusobacteriales</i> <i>Corynebacteriales</i> <i>Pseudonocardiales</i> <i>Neisseriales</i> <i>Rhizobiales</i> <i>Xanthomonadales</i> <i>Clostridiales</i> <i>Bacteroidales</i> <i>Caulobacterales</i>			
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