

Supplementary table 1. The table summarises the literature reviewed in order to design the synthetic aquaculture wastewater.

Source	Location	Fish species	Fish density	TP (mg·l ⁻¹)	P (phosphate, mg·l ⁻¹)	TN (mg·l ⁻¹)	N-Nitrate (mg·l ⁻¹)	N-Nitrite (mg·l ⁻¹)	N-ammonia (mg·l ⁻¹)	pH
1	Canada	Rainbow trout		8.7		50.2				
2	Germany	Rainbow trout	high	range: 0.42-0.49		range: 7.5-9.9	range: 3.2-4.6	range: 0.022-0.091	range: 1.1-2.6	
	Norway	Salmonid smolt	high	range: 0.05-0.3		range: 0.5-2				
3	Canada	Rainbow trout		7.15		41				
4	Canada	Brown trouta		6.2		49.2				
4	Canada	Lake trouta		6.8		65.3				
5	Several European countries	Artic char	range: 85-130 (kg/m³)				<10	<0.5		range: 6.5-8.5
		Atlantic salmon smolt	range: 45-50 (kg/m³)				≤90	<0.2		range: 6.8-7.3
		European eel	range: 50-120 (kg/m³)				range: 50-10	range: 0-1.5		range: 5-7.5
		European lobster								range: 7.8-8.2
		Pike perch	range: 15-60 (kg/m³)				≤56	range: 0-1.5		range: 6.5-7.5
		Rainbow trout	range: 50-80 (kg/m³)				<200	<1		range: 6.5-8
		Sturgeon	range: 80-100 (kg/m³)				<25	<0.5		range: 7-8
		Tilapia	range: 85-120 (kg/m³)				range: 100-200	range: 0.05-1		range: 6.5-8.5
6	Canada	Rainbow trout			0.06		2.2		0.5	
7	Israel	Tilapia	20 kg/m³	10					range: 10-23	range: 6.5-6.7
8	Canada				2.69		0.39		1.39	
9		synthetic aquaculture wastewater			19		20		20	
10	Ireland	Rainbow trout and perch	20		2.5		10.5		3.5	
11	Denmark	Rainbow trout		15		71				
12	USA (Alabama)	Catfish		0.56			2.36		2.35	
13	USA (Florida)	Catfish fingerlings	0.81 (fish/m³)			2.5±0.74 (SE) (range: 0.4-14.3)	0.058±0.007 (SE) (range: 0-0.13)	0.002±0.001 (SE)(range: 0-0.01)	0.439±0.107 (SE) (range: 0.05-1.8)	8.3±0.14 (SE) (range: 7.4-9.1)
	USA (Florida)	Catfish fingerlings	1.09 (fish/m³)	0.395±0.035 (SE) (range: 0.168-0.809)						
	USA (Florida)	Catfish fingerlings	0.83 (fish/m³)							
	USA (Florida)	Catfish fingerlings	0.89 (fish/m³)	0.33±0.039 (SE)(range: 0.17-0.454)		2.8±1.63 (SE)(range: 0.05-10.8)	0.048±0.015 (SE)(range: 0.01-0.1)	0.002±0.001 (SE)(range: 0-0.005)	0.33±0.083 (SE)(range: 0.05-0.64)	8.9±0.26 (SE)(range: 7.9-9.6)
	USA (Florida)	Catfish fingerlings	2.15 (fish/m³)	0.354±0.034 (SE) (range: 0.129-0.526)		1.5±0.31 (SE)(range: 0.3-3.7)	0.061±0.006 (SE)(range: 0.04-0.1)	0.003±0.001 (SE)(range: 0-0.01)	0.505±0.155 (SE)(range: 0.06-1.5)	8.5±0.19 (SE)(range: 7.4-9.3)
	USA (Florida)	Catfish fingerlings	1.37 (fish/m³)							
	UK	Catfish fingerlings	1.34 (fish/m³)	0.198±0.009 (SE) (range: 0.148-0.238)		2.6±0.36 (SE) (range: 0.8-4.9)	0.114±0.025 (SE) (range: 0.030-0.28)	0.03±0.001 (SE) 0-0.007	0.177±0.032 (SE)(range: 0.05-0.35)	8.3±0.21 (SE)(range: 7.4-9.3)
	USA (Florida)	Catfish fingerlings	0.83 (fish/m³)							
	USA (Florida)	Catfish fry	41.66 (fish/m³)	0.244±0.063 (SE)(range: 0.08-0.841)		1.4±0.14 (SE)(range: 0.7-12.4)	0.113±0.026 (SE)(range: 0.01-0.31)	0.008±0.004 (SE)(range: 0-0.048)	0.218±0.04 (SE)(range: 0.05-0.47)	8±0.16 (SE)(range: 7.3-9)
	USA (Florida)	Catfish fry	41.66 (fish/m³)							
	USA (Florida)	Buegill, reader sunfish, grass carp fry	8.47 (fish/m³)	0.104±0.009 (SE)(range: 0.069-0.151)		1.4±0.13 (SE)(range: 0.8-2.2)	0.088±0.014 (SE)(range: 0.05-0.17)	0.006±0.002 (SE)(range: 0-0.017)	0.128±0.025 (SE)(range: 0.05-0.28)	8±0.18 (SE)(range: 6.9-9.6)
	USA (Florida)	Buegill, reader sunfish, grass carp fry	8.47 (fish/m³)							
	USA (Florida)	Lasrgemouth bass, bluegill, reader sunfish fry	22 (fish/m³)	0.022±0.002 (SE)(range: 0.01-0.39)		0.8±0.06 (SE)(range: 0.5-1.1)	0.084±0.011 (SE)(range: 0.05-0.15)	0.003±0.001 (SE)(range: 0-0.005)	0.232±0.052 (SE)(range: 0.05-0.5)	9±0.21 (SE)(range: 7.8-9.9)
	USA (Florida)	Lasrgemouth bass, bluegill, grass carp fry	21.5 (fish/m³)							
14		Rainbow trout		0.49			60		10.6	
15	USA (Wisconsin)	Perch				0.5				
	USA (Wisconsin)	Perch				0.5				
	USA (Minnesota)	Perch				0.3				
	USA (Minnesota)	Perch				0.35				
	USA (Iowa)	Catfish				1				
	USA (Iowa)	Catfish				1				
	USA (Iowa)	Catfish				1				
	USA (Iowa)	Catfish				7				
	USA (Iowa)	Catfish				1.1				
	USA (Wisconsin)	Perch				1				
	USA (Wisconsin)	Perch				0.7				
	USA (Wisconsin)	Perch				0.8				
	USA (Wisconsin)	Perch				1				
16	Brazil								20	

1 Azevedo, P. A., Cho, C. Y., Leeson, S., & Bureau, D. P. (1998). Effects of feeding level and water temperature on growth, nutrient and energy utilization and waste outputs of rainbow trout (*Oncorhynchus mykiss*). *Aquatic Living Resources*, 11(4), 227-238.

2 Berghheim, A., & Brinker, A. (2003). Effluent treatment for flow through systems and European environmental regulations. *Aquacultural Engineering*, 27(1), 61-77.

3 Bureau, D. P., Gunther, S. J., & Cho, C. Y. (2003). Chemical composition and preliminary theoretical estimates of waste outputs of rainbow trout reared in commercial cage culture operations in Ontario. *North American Journal of Aquaculture*, 65(1), 33-38.

4 Cho, C. Y., Hynes, J. D., Wood, K. R., & Yoshida, H. K. (1994). Development of high-nutrient-dense, low-pollution diets and prediction of aquaculture wastes using biological approaches. *Aquaculture*, 124(1-4), 293-305.

5 Dalsgaard, J., Lund, I., Thorarinsdottir, R., Drenstvig, A., Arvonen, K., & Pedersen, P. B. (2013). Farming different species in RAS in Nordic countries: Current status and future perspectives. *Aquacultural engineering*, 53, 2-13.

6 Dumas, A., Laliberte, G., Lessard, P., & De La Noüe, J. (1998). Biotreatment of fish farm effluents using the cyanobacterium *Phormidium bohneri*. *Aquacultural engineering*, 17(1), 57-68.

7 Gendel, Y., & Lahav, O. (2013). A novel approach for ammonia removal from fresh-water recirculated aquaculture systems, comprising ion exchange and electrochemical regeneration. *Aquacultural engineering*, 52, 27-38.

8 Naylor, S., Brisson, J., Labelle, M. A., Drizo, A., & Comeau, Y. (2003). Treatment of freshwater fish farm effluent using constructed wetlands: the role of plants and substrate. *Water Science and Technology*, 48(5), 215-222.

9 Ng, Y. S., & Chan, D. J. C. (2018). Phytoremediation capabilities of *Spirodela polyrhiza*, *Salvinia molesta* and *Lemna* sp. in synthetic wastewater: a comparative study. *International journal of phytoremediation*, 20(12), 1179-1186.

10 O'Neill, E. A., Stejskal, V., Clifford, E., & Rowan, N. J. (2020). Novel use of peatlands as future locations for the sustainable intensification of freshwater aquaculture production—A case study from the Republic of Ireland. *Science of the Total Environment*, 706, 136044.

11 Roque d'orcastel, E., Blancheton, J. P., & Bellaud, A. (2009). Water quality and rainbow trout performance in a Danish Model Farm recirculating system: Comparison with a flow through system. *Aquacultural engineering*, 40(3), 135-143.

12 Schwartz, M. F., & Boyd, C. E. (1994). Channel catfish pond effluents. *The Progressive Fish-Culturist*, 56(4), 273-281.

13 Shireman, J. V., & Cichra, C. E. (1994). Evaluation of aquaculture effluents. *Aquaculture*, 123(1-2), 55-68.

14 Sindilariu, P. D., Brinker, A., & Reiter, R. (2009). Factors influencing the efficiency of constructed wetlands used for the treatment of intensive trout farm effluent. *Ecological Engineering*, 35(5), 711-722.

15 Yeo, S. E., Binkowski, F. P., & Morris, J. E. (2004). Aquaculture effluents and waste by-products characteristics, potential recovery, and beneficial reuse.

16 Zadineio, I. V., Alves, H. J., Moesch, A., Colpini, L. M. S., da Silva, L. C. R., & dos Santos, L. D. (2015). Influence of the chemical composition of smectites on the removal of ammonium ions from aquaculture effluents. *Journal of materials science*, 50(4), 1865-1875.