

Supporting information

Assessment of *Hedera helix* response in polluted environment through cross-advanced investigation techniques

Gabriela Lisa¹, Igor Cretescu¹, Catalin Tanase², Costel Mardari³, Nita Tudorachi⁴, Andra-Cristina Enache⁴, Petrisor Samoila⁴, Gabriela Soreanu^{1*}

¹“Cristofor Simionescu” Faculty of Chemical Engineering and Environmental Protection, “Gheorghe Asachi” Technical University of Iasi, 73 D. Mangeron Blvd., 700050 Iasi, Romania

²“Alexandru Ioan Cuza” University, Faculty of Biology, Carol I 20A, 700505 Iasi, Romania

³“Anastasie Fatu” Botanical Garden of “Alexandru Ioan Cuza” University, 7-9 Dumbrava Rosie, 700487 Iasi, Romania

⁴“Petru Poni” Institute of Macromolecular Chemistry, 41A Grigore Ghica Voda Alley, 700487 Iasi, Romania

*corresponding author: gsor@tuiasi.ro

Biofiltration system

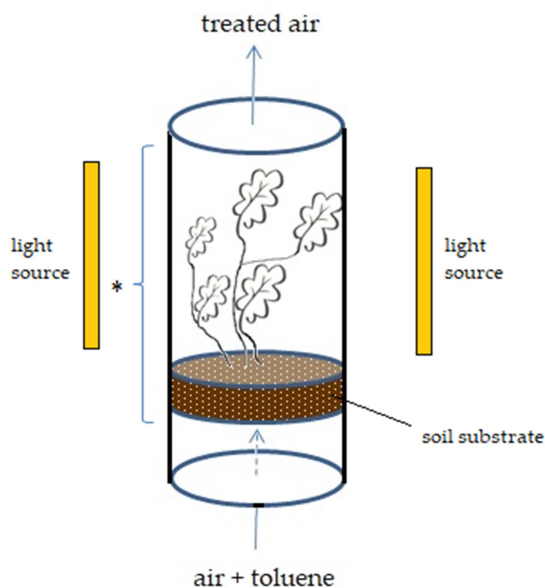


Figure S0. Schematic diagram of the botanical biofiltration system equipped with the investigated plant (*H. helix*) (*working packing volume). Air containing toluene was continuously fed at the bottom of the bioreactor, while the treated air was evacuated at the top, with different removal efficiencies (usually, 40-90%), depending on the inputs (polluted air loading rates). Leaf samples collected prior and after one month of bioreactor operation have been investigated by using FTIR, LP, SEM, EDX and TG/MS/FTIR techniques, revealing hidden plant-related aspects involved in toluene removal, with important benefits for a better process understanding and further development of such kind of biotechnology

ATR-FTIR spectroscopy

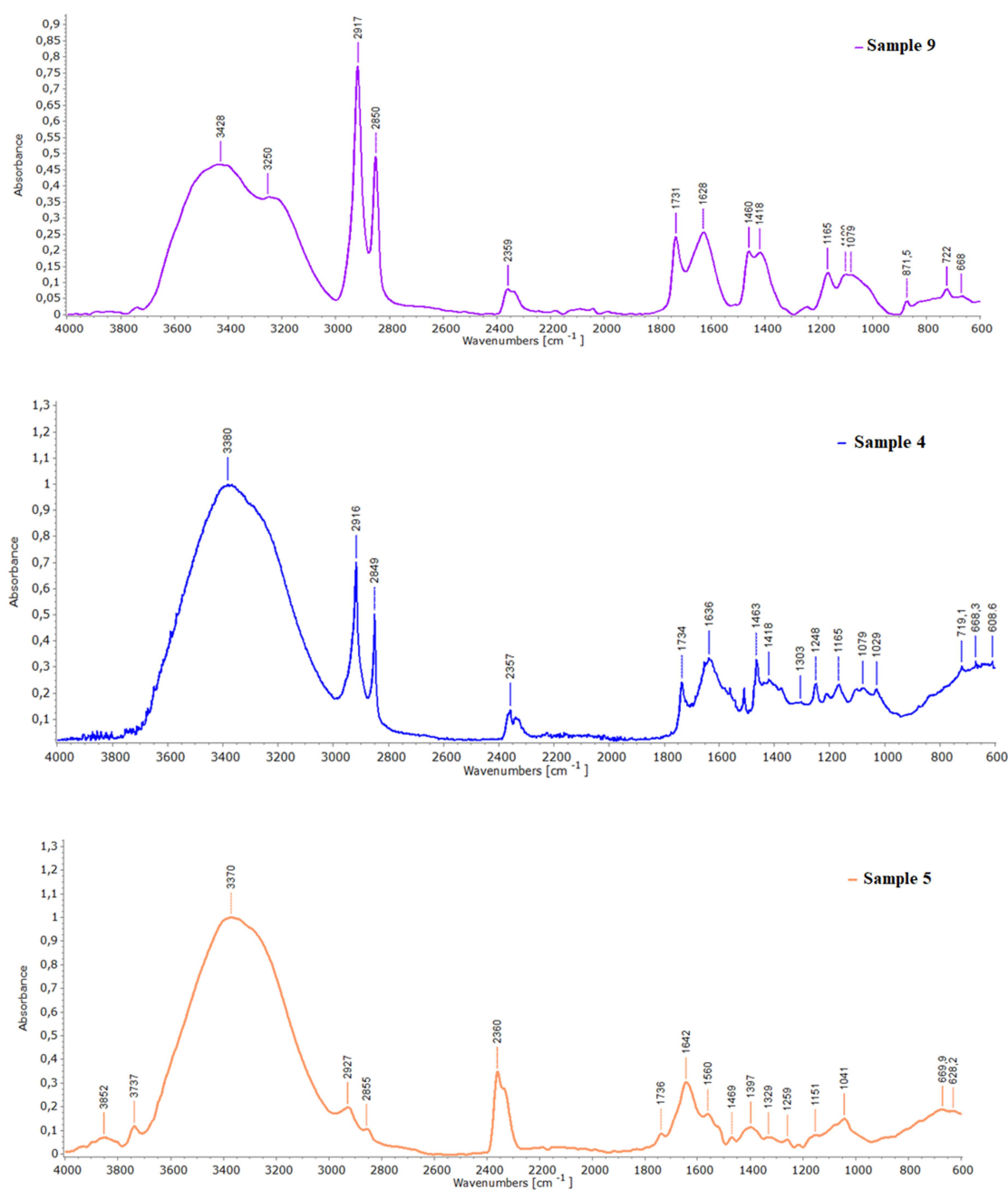
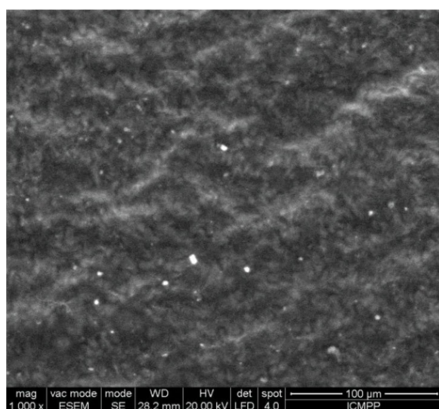


Figure S1. FTIR spectra

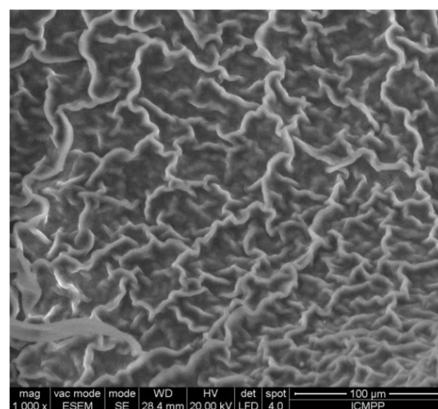
Table S1. The characteristic IR absorption bands in (cm^{-1}) of the Hedera Helix samples

Band assignment	Sample 9	Sample 4	Sample 5
	Band frequency/ cm^{-1}		
O–H stretching [1], N–H stretching alkyl or aryl amine [8]	3428, 3250	3380	3370, 3852, 3737
asymmetric CH_2 and CH_3 stretching vibration[2]	2917	2916	2927
symmetric CH_2 stretching vibration [2]	2850	2849	2855
C=O stretching vibration in unconjugated ketone, carbonyl and ester groups [2]	1731	1734	1736
C=O stretching vibration of Ph-(C=O)-groups in lignin [2], [3]	1628	1636	1642
asymmetric CH_3 stretching vibration, CH_2 scissoring vibration lignin and carbohydrates	1460, 1418	1463, 1418	1469
syringyl and condensed guaiacyl [5]	-	-	1329
C–O–C vibration [6]		1303	-
C–O stretching vibration [2]	1260		1259
C–OH stretch and C–H in-plane deformation in syringyl [1]		1248	
asymmetric C–O–C stretching vibration [2]	1165	1165	1151
C-C, C–OH and O– CH_3 stretching cellulose [1], [3], [7]	1079	1079, 1029	1041
C-H deformation in cellulose [3]	871	-	-
aromatic C–H out of plane bending vibration [2], [4]	722	719	-

Scanning Electron Microscopy (SEM)



Sample 4



Sample 5

Figure S2. SEM images for the adaxial surface of samples 4 and 5

Energy dispersive X-ray spectroscopy (EDX)

Table S2. Composition of ivy samples

Sample / Chemical element	Sample 9 Weight/ %	Sample 5 Weight/ %
C K	30.9	47.4
N K	4.1	1.0
O K	28.5	32.2
MgK	0.6	0.7
AlK	1.0	2.9
SiK	11.4	6.4
K K	0.7	1.1
CaK	11.0	1.7
FeL	0.0	0.9
PtM	11.8	5.6

TG/MS/FTIR technique

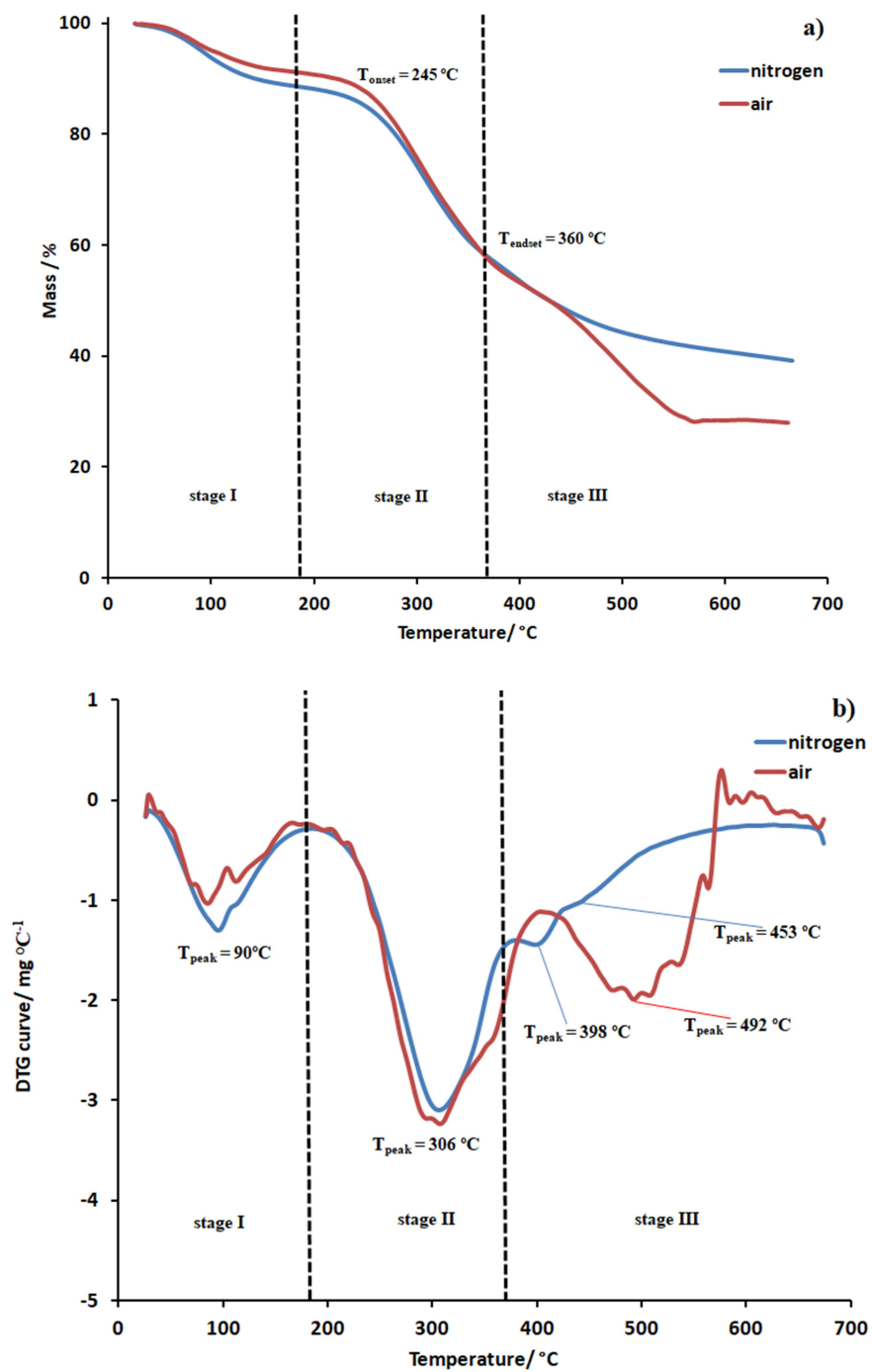


Figure S3. TG (a) and DTG (b) curves in air and nitrogen for sample 9

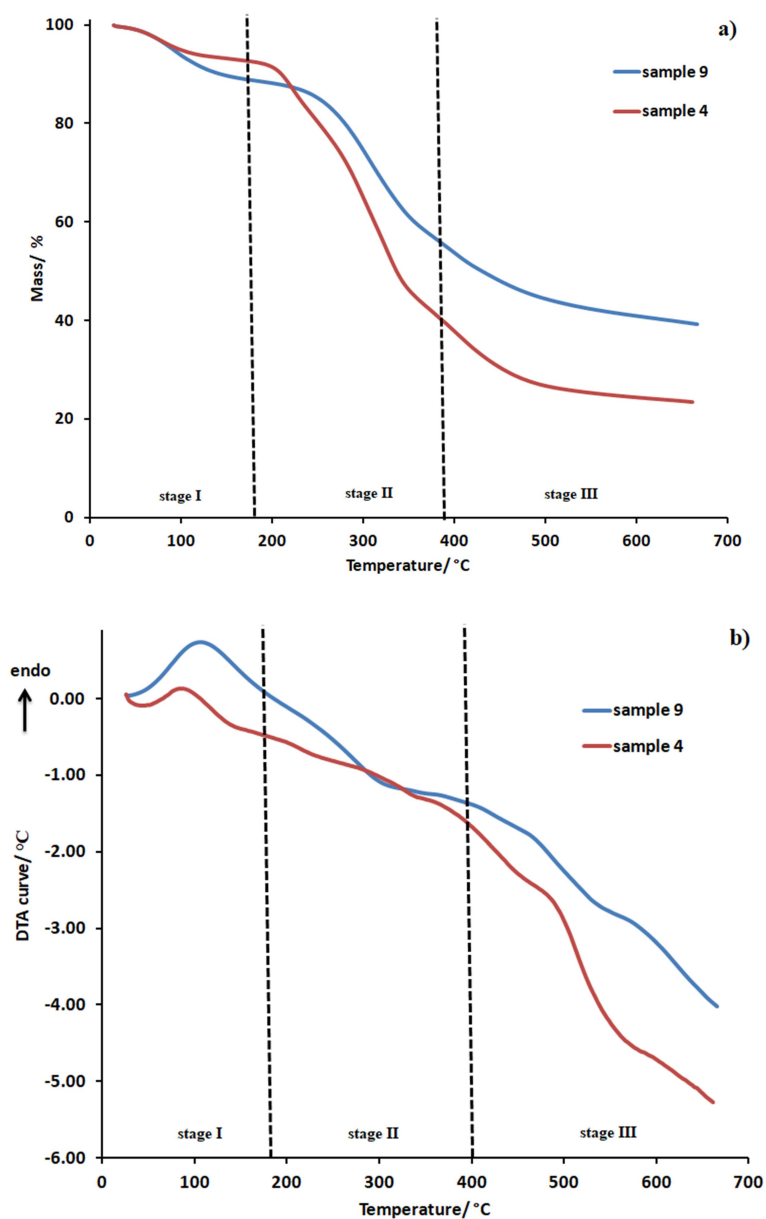


Figure S4. TG and DTA curves for samples 9 and 4 recorded in nitrogen

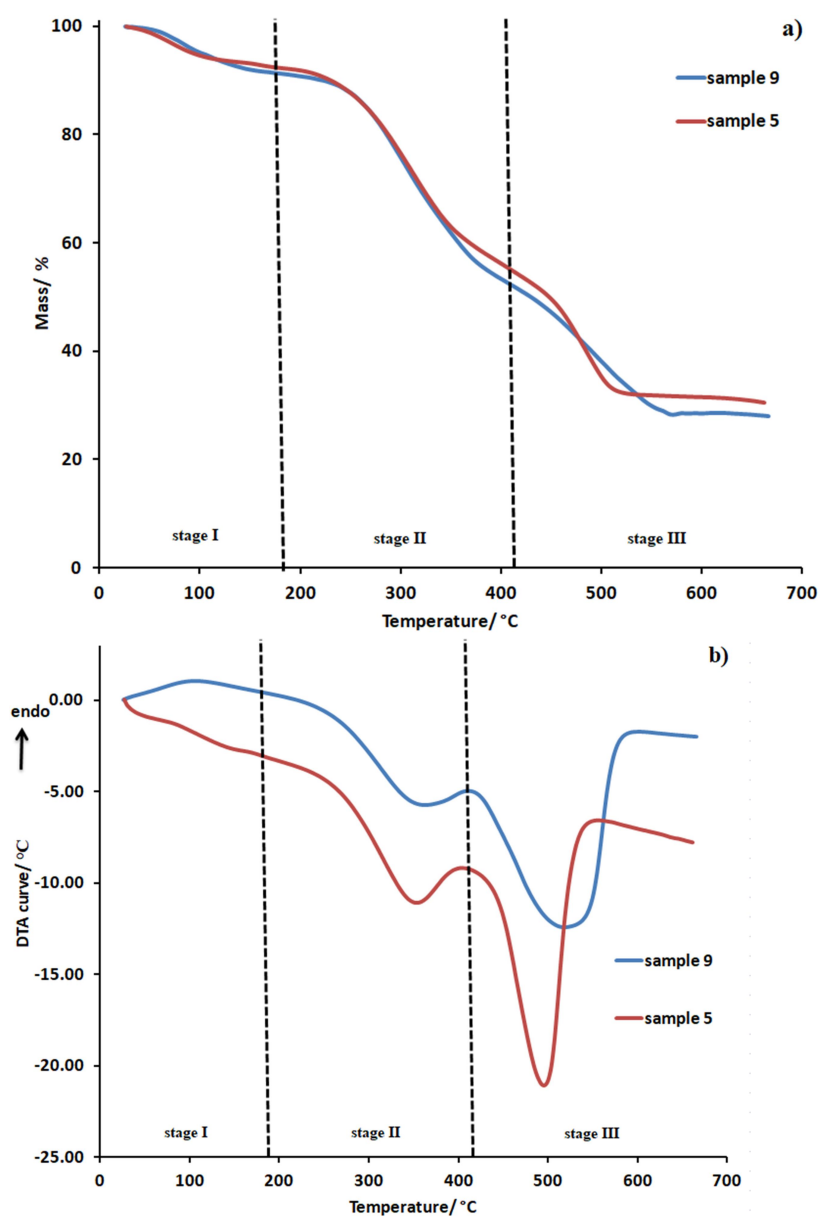


Figure S5. TG and DTA curves for samples 9 and 5 recorded in air

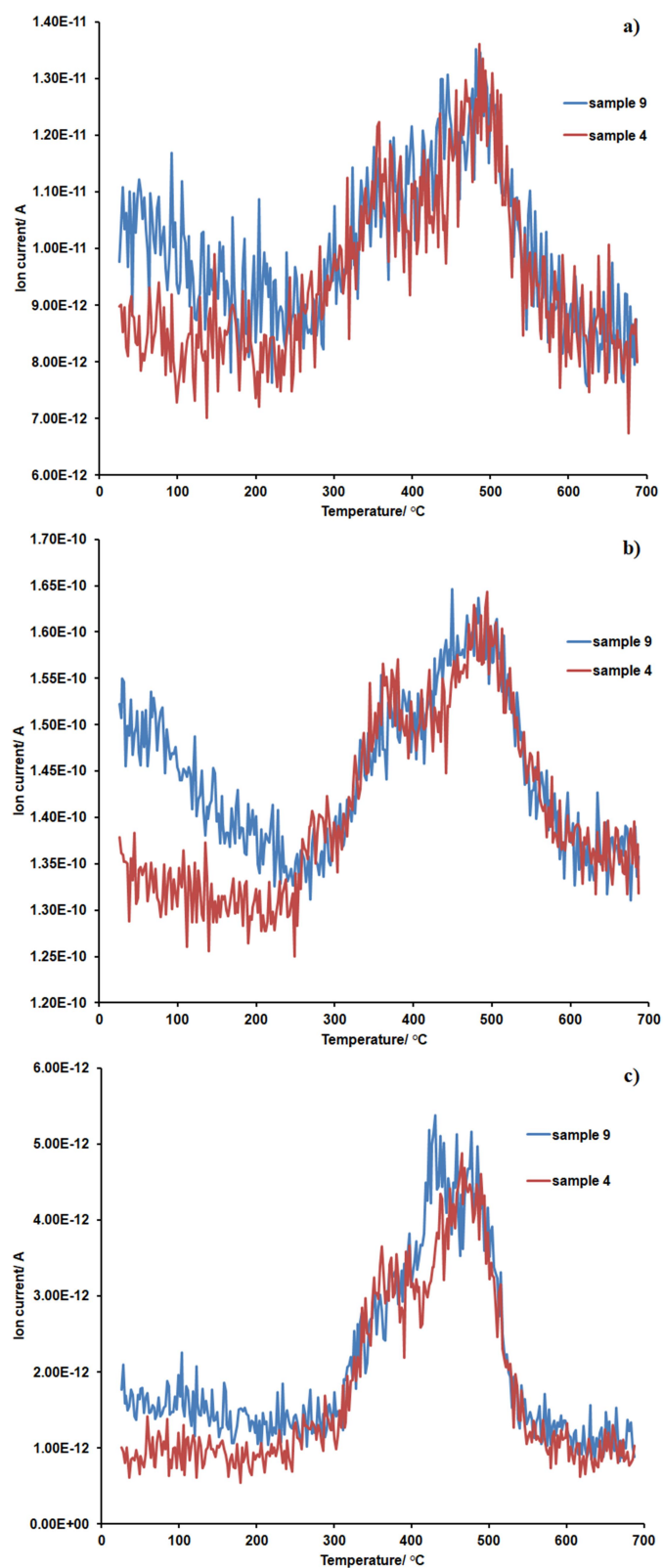


Figure S6. Ionic current variation with temperature for the fragments $m/z=26$ a), $m/z=27$ b) and $m/z=39$ c) for sample 9 and 4 (in nitrogen thermal analysis)

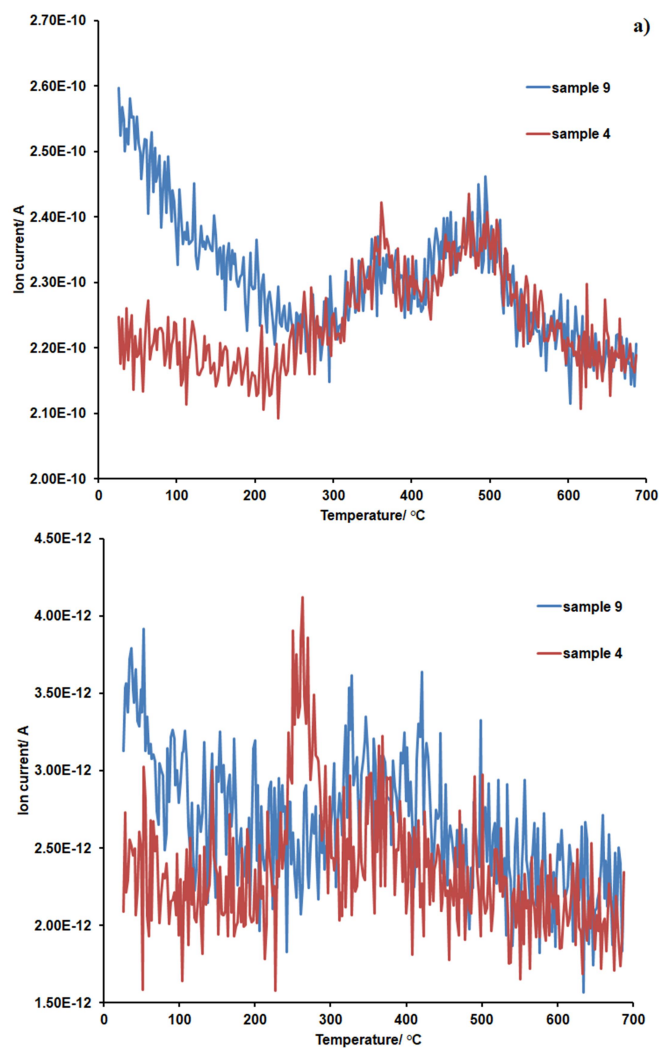


Figure S7. Ionic current variation with temperature for the fragments $m/z=29$ a) and $m/z=31$ b) for sample 9 and 4 (in nitrogen thermal analysis)

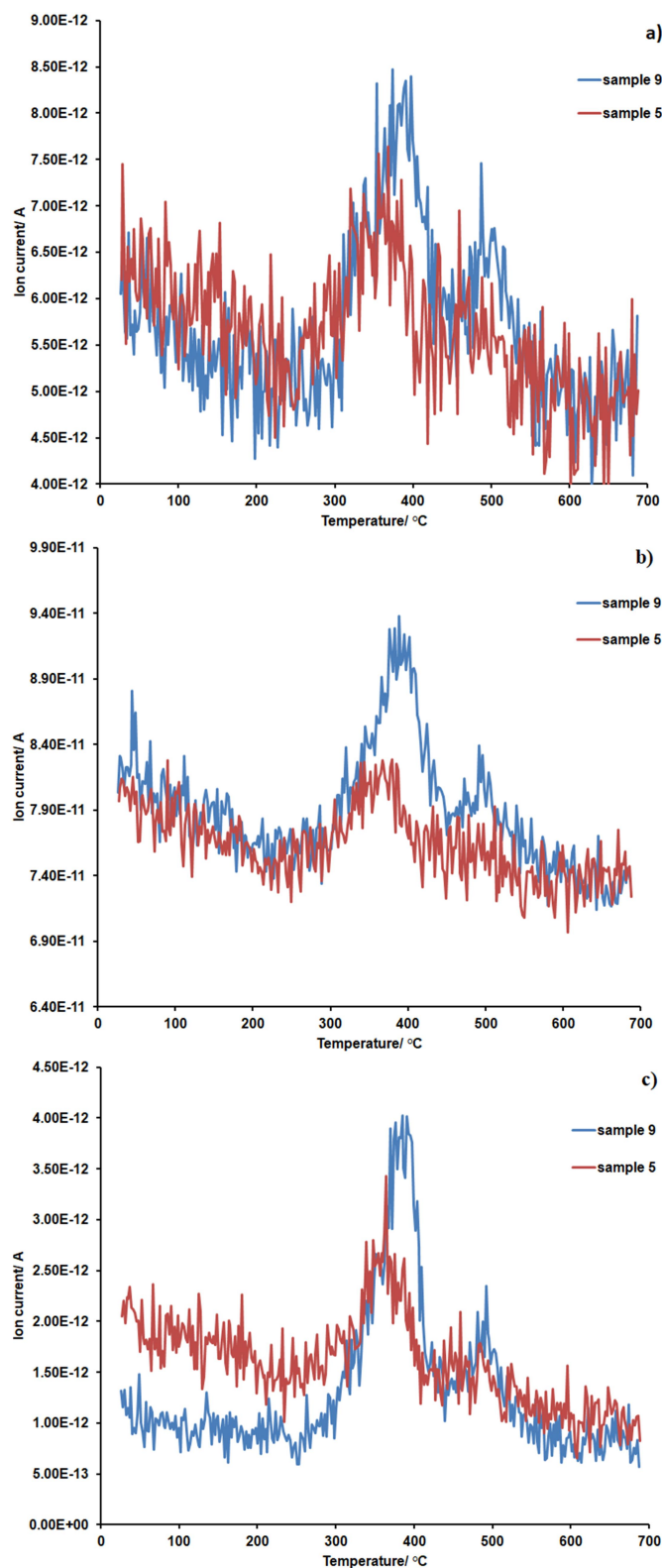


Figure S8. Ionic current variation with temperature for the fragments $m/z=26$ a), $m/z=27$ b) and $m/z=39$ c) for sample 9 and 5 (in air thermal analysis)

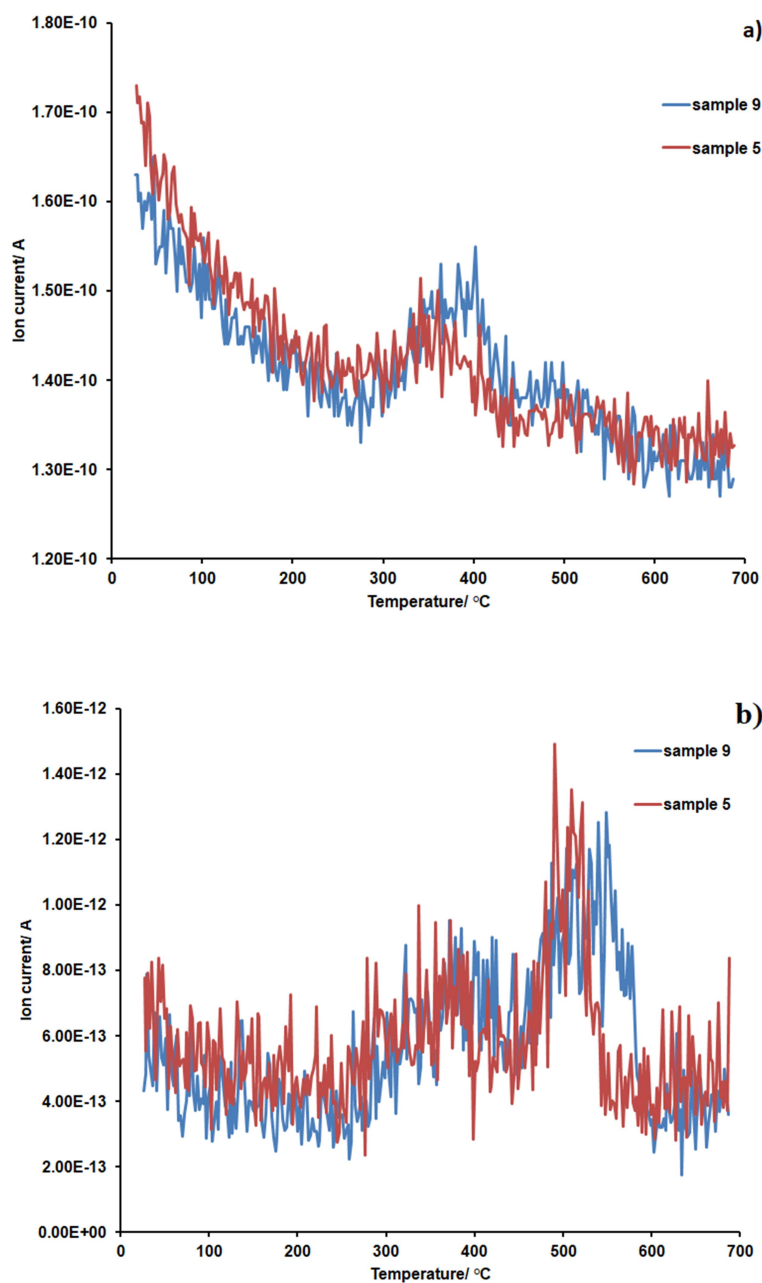


Figure S9. Ionic current variation with temperature for the fragments $m/z=29$ a) and $m/z=46$ b) for sample 9 and 5 (in air thermal analysis)

References

1. Shah MA, Hayder G, Kumar R, Kumar V, Ahamad T, Kalam MA, Soudagar MEM, Shamshuddin SZM, Mubarak NM (2023) Development of sustainable biomass residues for biofuels applications. Sci Rep 13:14248. <https://doi.org/10.1038/s41598-023-41446-1>

2. Părpăriță E, Brebu M, Uddin MA, Yanik J, Vasile C (2014) Pyrolysis Behaviours Of Various Biomasses. *Polym Degrad Stab* 100:1-9 <https://doi.org/10.1016/j.polymdegradstab.2014.01.005>
3. Vercruysse W., Smeets J., Haeldermans T, Joos B, Hardy A, Samyn P, Yperman J, Vanreppelen K., Carleer R, Adriaensens P, Marchal W, Vandamme D (2021) Biochar from raw and spent common ivy: Impact of preprocessing and pyrolysis temperature on biochar properties. *J Anal Appl Pyrolysis* 159:105294. <https://doi.org/10.1016/j.jaap.2021.105294>
4. Poiana M-A, Alexa E, Munteanu M-F, Gligor R, Moigradean D, Mateescu C (2015) Use of ATR-FTIR spectroscopy to detect the changes in extra virgin olive oil by adulteration with soybean oil and high temperature heat treatment. *Open Chem* 13:689–698. <https://doi.org/10.1515/chem-2015-0110>
5. Wang L, Fang Y, Yin J, Li X, Jiang J, Zhang Y, Yang Haiping (2022) Fast pyrolysis of guaiacyl-syringyl (GS) type milled wood lignin: Product characteristics and CH₄ formation mechanism study. *Sci Total Environ* 838:156395. <https://doi.org/10.1016/j.scitotenv.2022.156395>.
6. Kamnev AA, Dyatlova YA, Kenzhegulov OA, Vladimirova AA, Mamchenkova PV, Tugarova AV (2021) Fourier Transform Infrared (FTIR) Spectroscopic Analyses of Microbiological Samples and Biogenic Selenium Nanoparticles of Microbial Origin: Sample Preparation Effects. *Molecules* 26:1146. <https://doi.org/10.3390/molecules26041146>
7. Hustia A, Cantor M, Stefan R, Miclean M, Roman M, Neacsu I, Contiu I, Magyarid K, Baia M (2016) Assessing the Indoor Pollutants Effect on Ornamental Plants Leaves by FT-IR Spectroscopy. *Acta Phys Pol A* 129:142-149.
8. El-Hendawy ANA. (2006) Variation in the FTIR spectra of a biomass under impregnation, carbonization and oxidation conditions. *J. Anal. Appl. Pyrolysis* 75 159–166. <https://doi.org/10.1016/j.jaap.2005.05.004>