

Supplementary Information

Mass spectrometry-based ligand binding assays on adenosine A₁ and A_{2A} receptors

Purinergic Signalling

A. Massink, M. Holzheimer, A. Hölscher, J. Louvel, D. Guo, G. Spijksma, T. Hankemeier, A. P. IJzerman

Division of Medicinal Chemistry, LACDR, Leiden University, Leiden, The Netherlands (A.M., M.H., A.H., J.L., D.G., A.P.IJ.); Division of Analytical Biosciences, LACDR, Leiden University, Leiden, The Netherlands (G. S., T.H.)

Corresponding author: Prof. Dr. Adriaan P. IJzerman, ijzerman@lacdr.leidenuniv.nl

Synthesis of deuterated internal standards [²H]DPCPX and [²H]ZM-241,385

Results

*Synthesis of [²H₄]DPCPX **2***

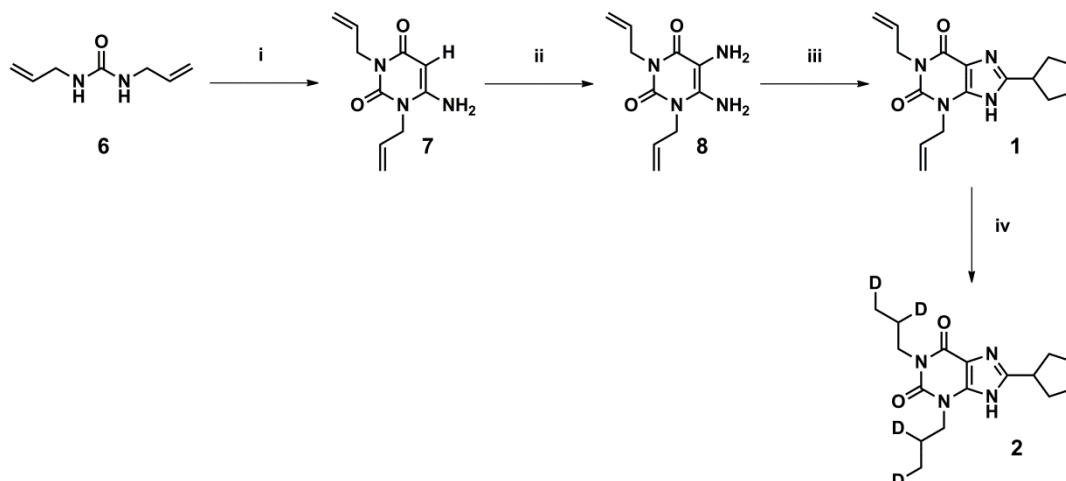
[²H₄]DPCPX **2** was prepared according to the synthetic route shown in Scheme 3 and was adopted from previously described syntheses of non-deuterated DPCPX. Starting from the commercially available diallyl urea **6**, 1,3-diallyl-6-aminouracil **7** was synthesized by condensation with cyanoacetic acid and subsequent base promoted ring closure [1]. **7** was then nitrosylated on the 5-position and subsequently reduced to the diamine **8** [2]. Cyclization of **8** with cyclopentanecarbonyl chloride gave the [²H₄]DPCPX precursor **1** [3]. Finally, the allylic double bonds of compound **1** were reductively deuterated in the presence of Wilkinson's catalyst with NaBD₄ as a deuterium source generating deuterium gas *in situ* upon addition of D₂O [7]. The mass spectrum showed a mass range for the (M+H⁺) species from 305.20 ([²H₀] isotopologue) to 313.27 ([²H₈] isotopologue) in a Gaussian distribution with the desired [²H₄]DPCPX **2** as most abundant isotopologue generating the main mass peak at 309.33.

*Synthesis of [²H₄]ZM-241,385 **5***

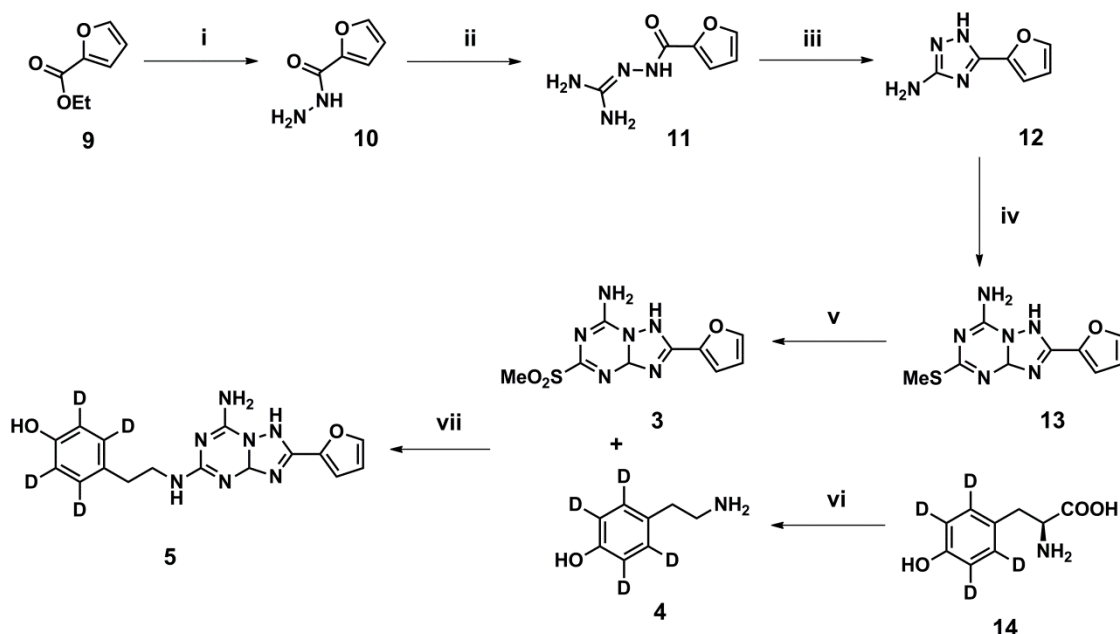
[²H₄]ZM-241,385 **5** was prepared according to the synthetic route shown in Scheme 4 and was adopted from previously described syntheses of non-deuterated ZM-241,385. Commercially available ethyl furan-2-carboxylate **9** was reacted with hydrazine hydrate to give furan-2-carbohydrazide **10**. Through a substitution reaction of **10** with methyl isothioureia hemisulfate diamide **11** was obtained, which underwent a cyclization in water to give the 1,2,4-triazole **12**. In a subsequent cyclization of **12** with dimethyl cyanocarbonimidothioate the triazolotriazine **13** was obtained [4] and the corresponding methylsulfone **3** was prepared by oxidation of the methylthioether with *m*CPBA [5]. For the synthesis of the deuterated building block, [²H₄]tyrosine **14** was enzymatically decarboxylated using tyrosine decarboxylase and pyridoxal-5-phosphate as cofactor to give [²H₄]tyramine **4** [6]. Finally, reaction of the deuterated tyramine **4** with methylsulfone compound **3** yielded the final product [²H₄]ZM-241,385 **5** [4]. MS analysis showed a mass of 342.7 (M+H⁺) and confirmed the incorporation of 4 deuterium atoms in the final product.

Schemes

Scheme 3 Synthesis of [$^2\text{H}_4$]DPCPX (**2**). Reagents and conditions: *i* 1. Cyanoacetic acid, Ac_2O , MW, 80 °C, 4 h, 2. EtOH, NaOH 2M; *ii* 1. NaNO_2 , 42% AcOH, rt, 1 h, 2. $\text{Na}_2\text{S}_2\text{O}_4$, EtOAc/ H_2O = 2/1, rt, 0.5 h, *iii* 1. Cyclopentanecarbonyl chloride, pyridine, dry CH_2Cl_2 , N_2 -atm, 5 °C to rt, 2 h, 2. NaOH 3M, reflux, 1 h; *iv* $\text{Rh}(\text{PPh}_3)_3\text{Cl}$, NaBD_4 , D_2O , dry THF, 60 °C, 3.5 h



Scheme 4 Synthesis of [$^2\text{H}_4$]ZM-241,385 (**5**). Reagents and conditions: *i* Hydrazine hydrate, EtOH, reflux, 18 h; *ii* Methyl isothioureia hemisulfate, 1% NaOH, rt, 23 h; *iii* H_2O , MW, 160 °C, 1.5 h; *iv* Dimethyl cyanocarbonimidodithioate, 180 °C, 16 h; *v* mCPBA, CH_2Cl_2 , -5 °C to rt, 18 h; *vi* Tyr-decarboxylase, pyridoxal-5-phosphate, acetate buffer 0.1 M (pH 5.5), H_2O , 37 °C, 17 h; *vii* Et_3N , MeCN, MW, 70 °C, 3 h



References

1. Devi I, Bhuyan PJ (2005) An expedient method for the synthesis of 6-substituted uracils under microwave irradiation in a solvent-free medium. *Tetrahedron Lett* 46 (34):5727-5729. doi:doi:10.1016/j.tetlet.2005.06.075
2. Soriano A, Ventura R, Molero A, Hoen R, Casadó V, Cortés A, Fanelli F, Albericio F, Lluís C, Franco R, Royo M (2009) Adenosine A_{2A} receptor-antagonist/dopamine D₂ receptor-agonist bivalent ligands as pharmacological tools to detect A_{2A}-D₂ receptor heteromers. *J Med Chem* 52 (18):5590-5602. doi:10.1021/jm900298c
3. Erickson RH, Hiner RN, Feeney SW, Blake PR, Rzeszutarski WJ, Hicks RP, Costello DG, Abreu ME (1991) 1,3,8-trisubstituted xanthines. Effects of substitution pattern upon adenosine receptor A₁/A₂ affinity. *J Med Chem* 34 (4):1431-1435
4. Jörg M, Agostino M, Yuriev E, Mak F, Miller N, White J, Scammells P, Capuano B (2013) Synthesis, molecular structure, NMR spectroscopic and computational analysis of a selective adenosine A_{2A} antagonist, ZM 241385. *Struct Chem* 24 (4):1241-1251. doi:10.1007/s11224-012-0151-7
5. Jörg M, Shonberg J, Mak FS, Miller ND, Yuriev E, Scammells PJ, Capuano B (2013) Novel adenosine A_{2A} receptor ligands: a synthetic, functional and computational investigation of selected literature adenosine A_{2A} receptor antagonists for extending into extracellular space. *Bioorg Med Chem Lett* 23 (11):3427-3433. doi:10.1016/j.bmcl.2013.03.070
6. Ntai I, Phelan VV, Bachmann BO (2006) Phosphonopeptide K-26 biosynthetic intermediates in *Astrosporangium hypotensionis*. *Chem Commun (Camb)* (43):4518-4520
7. Adair GRA, Kapoor KK, Scolan ALB, Williams JMJ (2006) Ruthenium catalysed reduction of alkenes using sodium borohydride. *Tetrahedron Lett* 47 (50):8943-8944. doi:10.1016/j.tetlet.2006.10.026