

Retrotransposable Element Derepression Distinguishes *DNMT3A*- from *TET2*-Mutant Clonal Hematopoiesis

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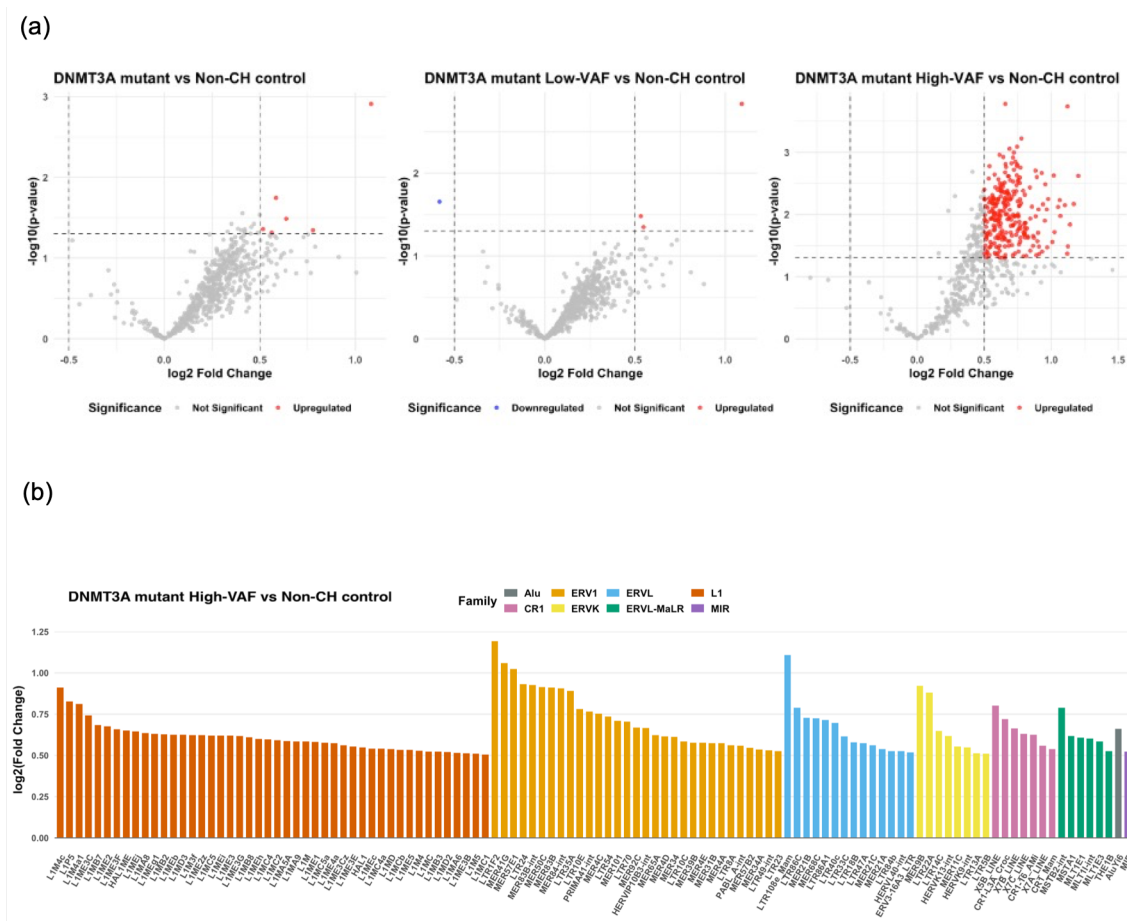
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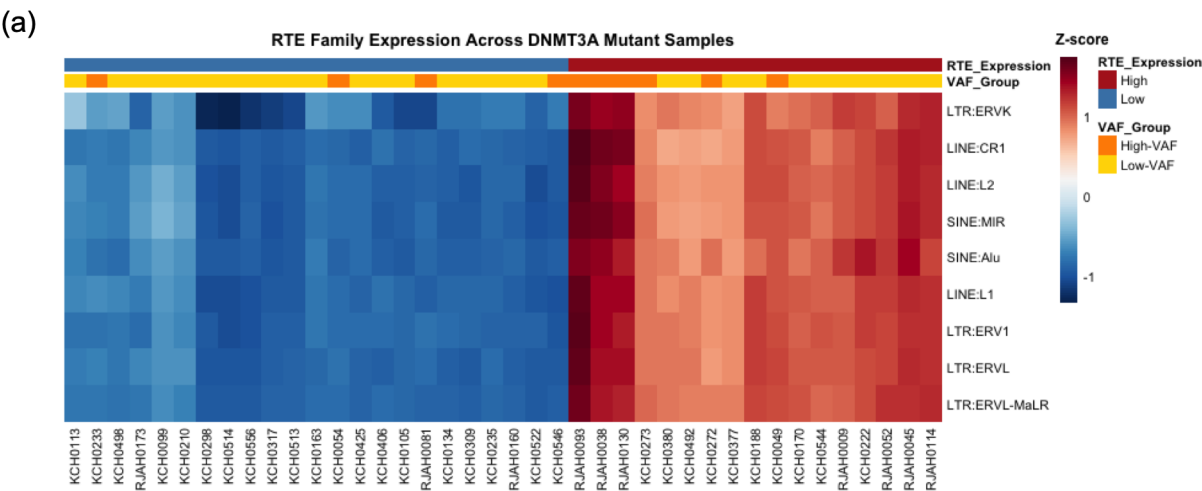


Supplementary Figure 1. Upregulated RTE subfamilies in DNMT3A-mutant clonal haematopoiesis.

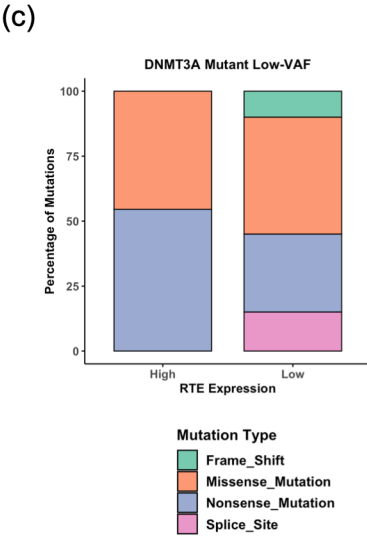
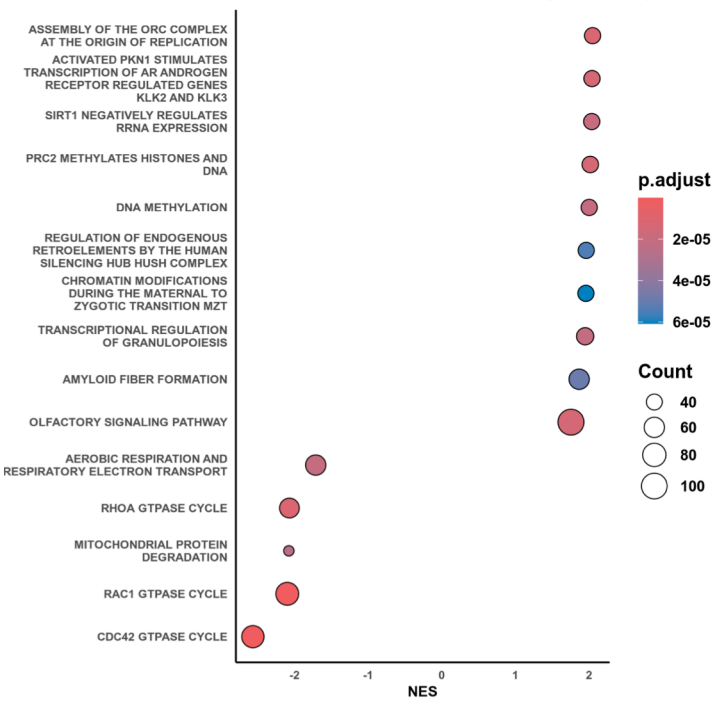
(a) Sensitivity analysis using genomic DNA-derived VAF to classify DNMT3A-mutant cases into high- and low-VAF groups. The overall pattern of RTE activation remained consistent with the primary analysis and identified an even greater number of

significantly upregulated RTE subfamilies in the high-VAF group, demonstrating that the findings are robust to the method used for estimating clonal burden.

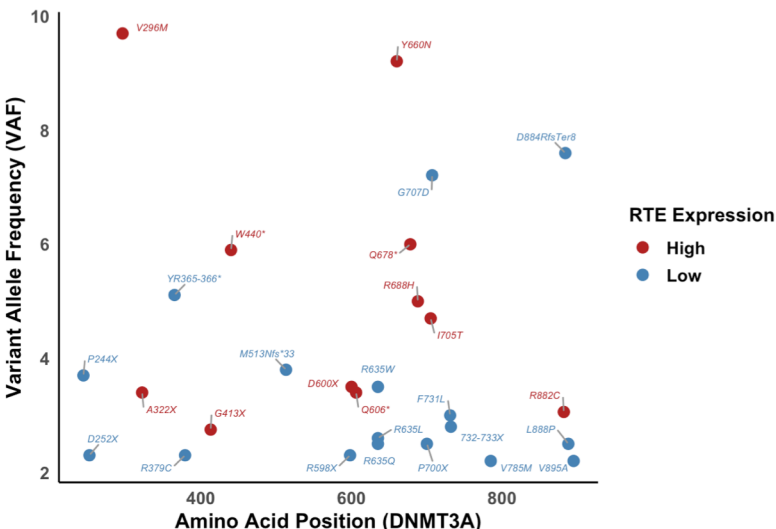
(b) Lists of RTE subfamilies upregulated in DNMT3A-mutant high-VAF vs. non-CH controls. The DNMT3A-mutant high-VAF group showed the largest number of upregulated subfamilies, particularly within the LINE:L1, LTR:ERV1, and LTR:ERVL classes ($\log_2$ fold change > 0.5, $p < 0.05$).



(b) GSEA Reactome: DNMT3A Mutant High-VAF - High vs Low RTEs



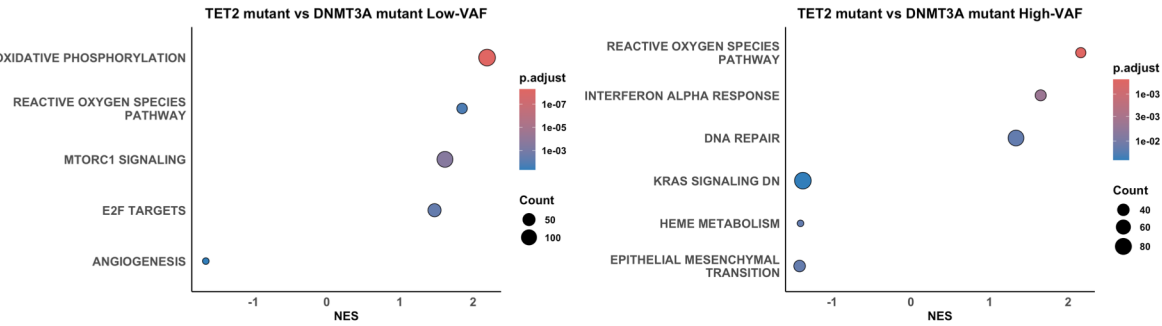
(d) DNMT3A Mutant Low-VAF



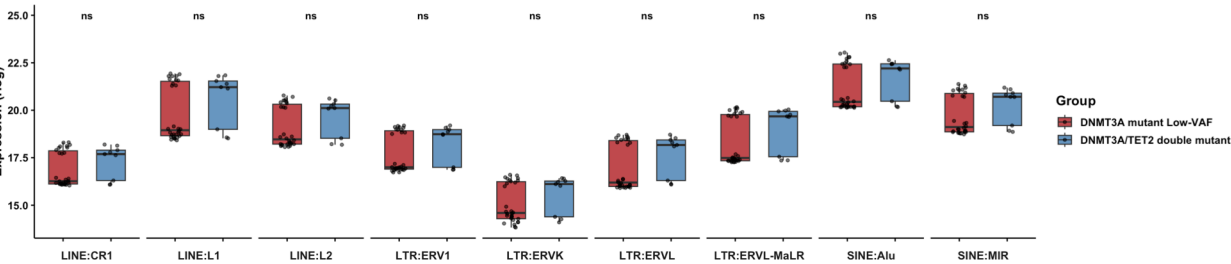
Supplementary Figure 2. Stratification of *DNMT3A*-mutant cases and additional analyses of low-VAF and high-VAF subgroups.

- (a) *DNMT3A*-mutant cases were stratified into high vs. low RTE expression groups based on RTE family expression.
- (b) Gene set enrichment analysis (GSEA) of high VAF *DNMT3A* clones comparing high vs. low RTE expression showing enrichment of pathways involved in HUSH & PRC2-mediated silencing and SIRT1-mediated rRNA regulation.
- (c) Enrichment analysis of mutation types in *DNMT3A*-mutant low-VAF samples comparing high vs. low RTE expression, showing an overrepresentation of nonsense mutations.
- (d) Scatterplot of amino acid position vs. variant allele frequency (VAF) in *DNMT3A*-mutant low-VAF samples, colour-coded by RTE expression group.

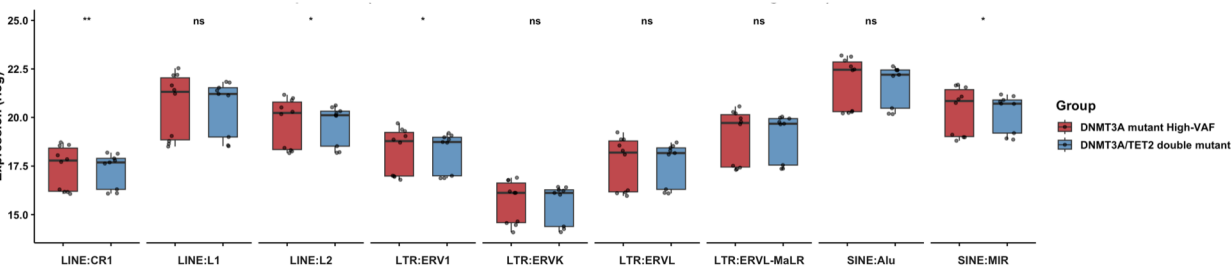
(a)



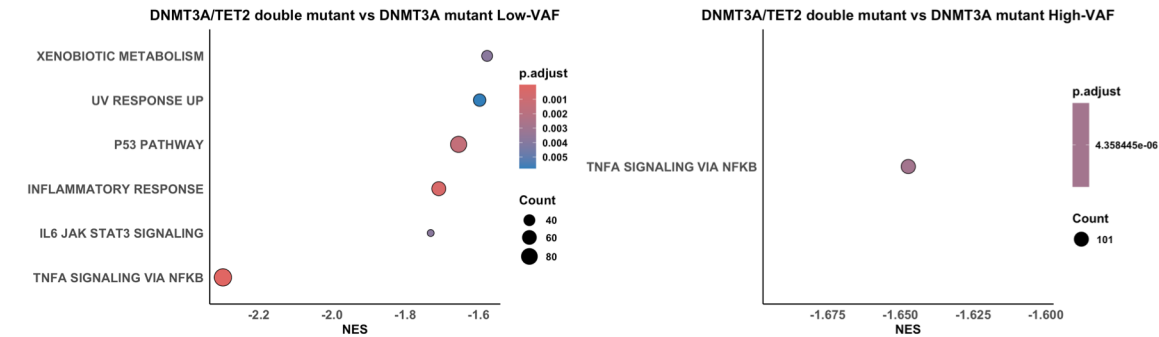
(b)



(c)



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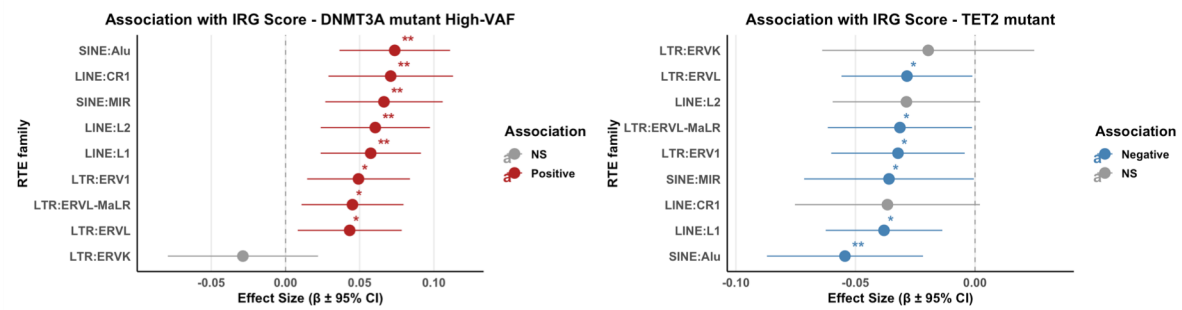
Supplementary Figure 4. Pathway and RTE expression analyses in *TET2* and *DNMT3A/TET2* double mutants.

(a) Gene set enrichment analysis (GSEA) of Hallmark gene sets comparing (1) *TET2-mutants* vs. non-CH controls, (2) *TET2-mutants* vs. *DNMT3A*-mutant low-VAF, and (3) *TET2-mutants* vs. *DNMT3A*-mutant high-VAF. Oxidative phosphorylation was enriched in comparisons 1 and 2, whereas reactive oxygen species (ROS) and interferon- α response pathways were enriched in comparison 3.

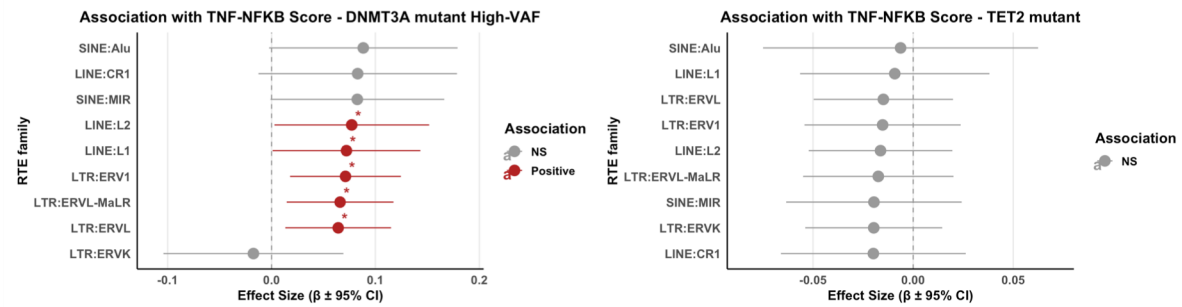
(b–c) Boxplots of RTE family expression comparing (a) *DNMT3A*-mutant low-VAF vs. *DNMT3A/TET2* double mutants, and (b) *DNMT3A*-mutant high-VAF vs. *DNMT3A/TET2* double mutants. CR1, L2, ERV1, and MIR families were significantly downregulated in panel b.

(d) GSEA of Hallmark gene sets comparing (1) *DNMT3A/TET2* double mutants vs. *DNMT3A*-mutant low-VAF, and (2) *DNMT3A/TET2* double mutants vs. *DNMT3A*-mutant high-VAF. Pathway analysis revealed attenuation of TNF–NF κ B and interferon programs compared with *DNMT3A-mutant* cases.

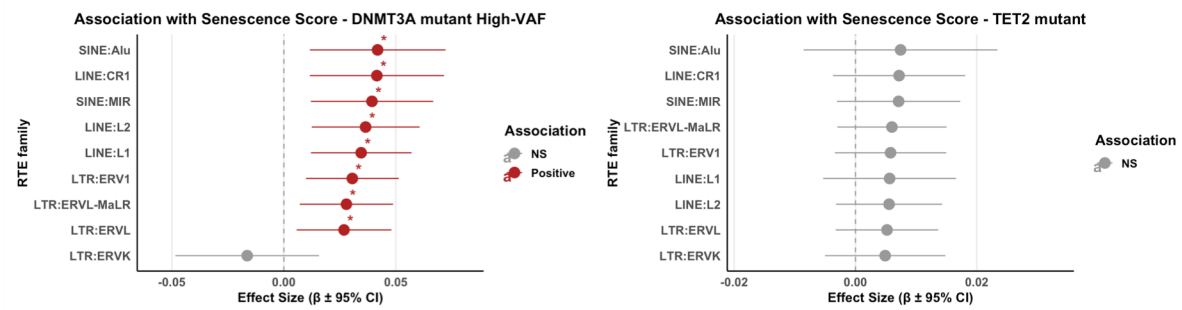
(a)



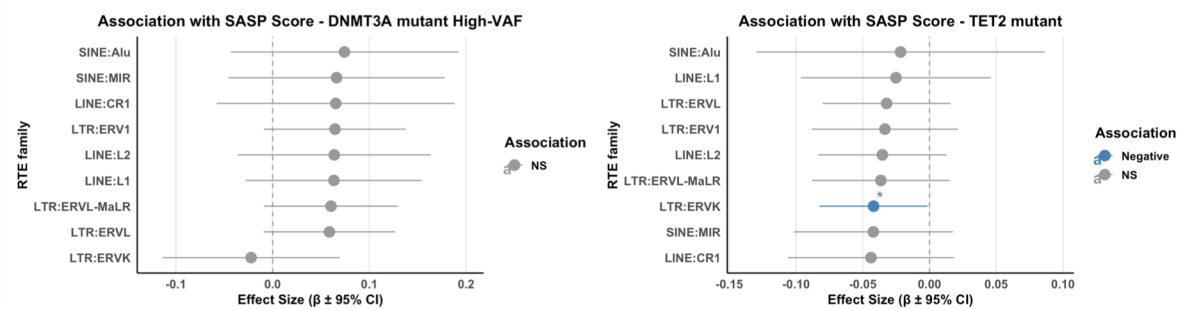
(b)



(c)



(d)



Supplementary Figure 5. Validation of associations between RTE family expression and gene signatures in an independent cohort.

Validation cohort ($n = 13$) from the study by Smith *et al.*, including 7 *DNMT3A*-mutant high-VAF and 6 *TET2*-mutant cases, supporting the findings shown in Figure 4. Forest plots show associations between RTE families and gene signature scores for *DNMT3A*-mutant high-VAF (left) and *TET2*-mutant (right) cases across four panels: (a) interferon regulatory genes (IRG), (b) TNF–NF κ B, (c) senescence, and (d) senescence-associated secretory phenotype (SASP). Positive associations were observed for IRG, TNF–NF κ B, and senescence signatures in *DNMT3A*-mutant high-VAF cases, while SASP showed only a trend.