

Appendix A – Trophic discrimination factors used for mixing model analysis, sensitivity comparisons, and comparison with a range of species-specific trophic discrimination factors found in the literature.

Table 1 – Trophic Discrimination Factors

<u>Model</u>	<u>Category</u>	<u>Source</u>	<u>Carbon ($\delta^{13}\text{C}$)</u>	<u>Nitrogen ($\delta^{15}\text{N}$)</u>	<u>Sulfur ($\delta^{34}\text{S}$)</u>
Invertebrate	Invertebrate (Primary Consumer)	McCutchan et al. 2003	0.4±1.2 ‰	2.3±1.61 ‰	n/a
	Invertebrate (Omnivore)	McCutchan et al. 2003	0.6±1.47 ‰	3.45±1.97 ‰	n/a
	Invertebrate (Predator)	McCutchan et al. 2003	0.8±1.7 ‰	4.6±2.28 ‰	n/a
Fish	Fish (base)	McCutchan et al. 2003	1.1±0.35 ‰	2.8±0.40 ‰	1.9±0.51 ‰
	Fish (base + Inv Omni)	McCutchan et al. 2003	1.7±1.51 ‰	6.25±2.01 ‰	2.65±2.45 ‰
Sensitivity Comparison	Invertebrate (Primary Consumer)	Vander Zanden and Rasmussen 2001	-0.41±1.14 ‰	2.52±2.5 ‰	n/a
	Invertebrate (Predator) [Shrimps]	Vander Zanden and Rasmussen 2001	0.3±1.54 ‰	7.01±2.53 ‰	n/a
	Fish (base + Inv Omni)	Vander Zanden and Rasmussen 2001	0.3±1.54 ‰	7.01±2.53 ‰	n/a
Range of Values Reported From Additional Taxa-Specific Literature	Invertebrate (Primary Consumer)	Sagers & Goggin 2007, de Carvalho et al. 2015, Dionne et al. 2016	-4.9 to 8.5	-0.88 to 7.2	No Data
	Invertebrate (Omnivore)	Mancinelli 2012, Tiselius & Fransson 2016, Remy et al. 2018, Umezawa et al. 2018	0.2 to 2.6	0.6 to 4.48	No Data
	Invertebrate (Predator)	de Carvalho et al. 2015	-1.5 to 1.25	-0.01 to 9.05	No Data
	Fish (base)	Bastos et al. 2017, Blanke et al. 2017	-0.57 to 1.6	1.77 to 4.78	No Data

Table 2 – Sensitivity Results. Average and maximum refer to discrepancy between models based on different trophic discrimination factors.

Subregion	Season	Source	Invertebrates		Fish	
			Average	Maximum	Average	Maximum
ULN	Summer	BuMa_ScAc	0.7%	1.5%	0.2%	1.2%
		CeDe_EgDe	1.4%	2.2%	5.3%	13.1%
		GFA	1.5%	2.6%	2.1%	6.3%
		JuBa_ScCa	1.4%	3.7%	3.0%	19.6%
		MySp	2.5%	3.4%	4.3%	23.1%
		Phyto	2.1%	4.9%	2.5%	7.9%
		Tysp_IrPs	0.5%	0.7%	3.9%	10.5%
UCA	Spring	BD	1.4%	3.8%	1.1%	4.4%
		CeDe_MySp	1.9%	2.6%	19.4%	38.4%
		Phyto	3.8%	7.4%	20.3%	33.8%
		ScCa_L	3.3%	7.3%	2.8%	11.9%
		ScCa_U	2.6%	3.0%	2.4%	16.9%

Bastos, R. F., F. Correa, K. O. Winemiller, and A. M. Garcia. 2017. Are you what you eat? Effects of trophic discrimination factors on estimates of food assimilation and trophic position with a new estimation method. *Ecological Indicators* 75: 234-241. <https://doi.org/10.1016/j.ecolind.2016.12.007>

Blanke, C. M., Y. Chikaraishi, Y. Takizawa, S. A. Steffan, P. S. Dharampal, and M. J. Vander Zanden. 2017. Comparing compound-specific and bulk stable nitrogen isotope trophic discrimination factors across multiple freshwater fish species and diets. *Canadian Journal of Fisheries and Aquatic Sciences* 74(8): 1291-1297. <https://doi.org/10.1139/cjfas-2016-0420>

de Carbalho, A. P. C., B. Gucker, M. Brauns, and I. G. Boechat. 2015. High variability in carbon and nitrogen isotopic discrimination of tropical freshwater invertebrates. *Aquatic Sciences* 77(2) 307-314. <https://doi.org/10.1007/s00027-014-0388-x>

Dionne, K.; F. Dufresne, and C. Nozais. 2016. Variation in $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ trophic enrichment factors among *Hyalella azteca* amphipods from different lakes. *Hydrobiologia* 781(1):217-230. <https://doi.org/10.1007/s10750-016-2846-z>

Mancinelli, G. 2012. On the trophic ecology of Gammaridea (Crustacea: Amphipoda) in coastal waters: A European-scale analysis of stable isotopes data. *Estuarine Coastal and Shelf Science* 114:130-139. <https://doi.org/10.1016/j.ecss.2011.12.003>

McCutchan, J. H., W. M. Lewis, C. Kendall, and C. C. McGrath. 2003. Variation in trophic shift for stable isotope ratios of carbon, nitrogen, and sulfur. *Oikos* 102 (2):378-390. <https://doi.org/10.1034/j.1600-0706.2003.12098.x>.

Remy, F., F. Darchambeau, A. Melchior, and G. Lepoint. 2017. Impact of food type on respiration, fractionation and turnover of carbon and nitrogen stable isotopes in the marine amphipod *Gammarus aequicauda* (Martynov, 1931). *Journal of Experimental Marine Biology and Ecology* 486: 358-367. <https://doi.org/10.1016/j.jembe.2016.10.031>

Sagers, C. L., and F. L. Goggin. 2007. Isotope enrichment in a phloem-feeding insect: influences of nutrient and water availability. *Oecologia* 151(3):464-472. <https://doi.org/10.1007/s00442-006-0603-0>

Tiselius, P. and K. Fransson. 2016. Daily changes in $\delta^{15}\text{N}$ and $\delta^{13}\text{C}$ stable isotopes in copepods: equilibrium dynamics and variations of trophic level in the field. *Journal of Plankton Research* 38(3):751-761. <https://doi.org/10.1093/plankt/fbv048>

Umezawa, Y; Tamaki, A; Suzuki, T; Takeuch, S; Yoshimizu C; and Tayasu, I. 2018. Phytoplankton as a principal diet for callinassid shrimp larvae in coastal waters, estimated from laboratory rearing and stable isotope analysis. *Marine Ecology Progress Series*. 592, pp 141-158

Vander Zanden, M. J., and J. B Rasmussen. 2001. Variation in $\delta^{15}\text{N}$ and $\delta^{13}\text{C}$ trophic fractionation: implications for aquatic food web studies. *Limnology and Oceanography* 46 (8):2061-2066. <https://doi.org/10.4319/lo.2001.46.8.2061>.