

Supplementary Material

***Thymus* × *citriodorus*: an Emerging Aromatic and Medicinal Hybrid Plant with Relevant Bioactive Potential**

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Table 1 Source, processing features, and chemical characterization of the *Thymus* × *citriodorus* volatile oils (VO) analyzed by GC-MS. The major compound in each VO is represented in **bold**, and underlined if its relative abundance was superior to 50%

Origin	Reported variety	Plant parts	Harvesting stage (period)	Isolation details	Yield	Major components ^b (%)	Ref.
Portugal	<i>T. citriodorus</i>	ND	ND	Steam distillation ^a	ND	Geraniol (27.5) , 1,8-cineole (16.3), thymol (9.2), geranial (4.2), <i>p</i> -cymene (3.7), neral (3.4)	(Oliveira et al. 2022)
Lithuania	<i>T. citriodorus</i>	Leaves and flowers	(June 2020)	Hydrodistillation (Clevenger) of dried plants for 2 h ^a	Variable due to fertilization (% of dry weight): ~ 0.5 – 0.6	Variations dependent on the fertilizers (liquid cattle dung <i>versus</i> hummus): Geraniol (23.78 vs 26.27) ; nerol (14.73 vs 14.20); geranial (11.97 vs 13.04); neral (11.27 vs 10.63); germacrene D-4-ol (6.66 vs 8.74)	(Vaičiulytė et al. 2022)
Lithuania	<i>T. citriodorus</i>	Leaves and flowers	(June 2021)	Hydrodistillation (Clevenger) of dried plants for 2 h ^a	Variable due to fertilization (% of dry weight): ~ 0.2 – 0.35	Variations dependent on the fertilizers (liquid cattle dung <i>versus</i> hummus): Geraniol (23.60 vs 20.55) ; nerol (15.73 vs 12.29); geranial (13.47 vs 12.59); neral (11.27 vs 10.63); germacrene D-4-ol (6.66 vs 8.74)	(Vaičiulytė et al. 2022)
Lithuania	<i>T. citriodorus</i>	Leaf inflorescences	Full flowering (June 2015, 2016 and 2017)	Hydrodistillation (Clevenger) of dried plant for 2 h ^a	0.5-0.7 mL / 100 g	Average (2015-2017): Geraniol (26.53) , nerol (16.55), geranial (13.49), neral (12.90)	(Ložienė et al. 2021)
Greece	<i>T. citriodorus</i>	Aerial parts	Flowering	Hydrodistillation (Clevenger) of dried plant for 3 h. 100 g of plant / 1 L of distilled water	5 mL / 100 g of dry weight	Geraniol (47.08) , geranial (7.94), neral (7.12), geranyl acetate (4.05)	(Ntalli et al. 2020a)

Austria	<i>T. citriodorus</i>	ND	Flowering	ND	1.08% of dry weight	Geraniol (83.34) , neral (6.84), geranial (2.92)	(Malankina et al. 2019)
Russia	<i>T. citriodorus</i> var. <i>variegata</i>	ND	Flowering	ND	ND	Thymol (53.46) , isobornyl propionate (12.92), carvacrol (6.05), α -terpineol (3.44)	(Malankina et al. 2019)
Lithuania	<i>T. citriodorus</i>	Leaves and inflorescences (free from stems)	Flowering (June 2015)	Hydrodistillation according to European Pharmacopeia 6 th Edition with 19 distillation times ^b . 30 g / 0.5 L of water	Variable across distillation times Lowest: 0.28% (m/m) at 5 min; Highest: 0.75% (m/m) at 150 min	Average from different distillation times: Geraniol (36.6) , geranial (17.5), nerol (11.5), neral (10.2)	(Jurevičiūtė et al. 2019)
Republic of Moldova	<i>T. citriodorus</i> “aureus”	Aerial parts	Full flowering (July 2015)	Hydrodistillation (modified Clevenger) of powered plant for 3 h. 100 g / 1 L of double distilled water	1.31 mL of VO / 100 g of dry weight	Geraniol (60.31) , geranial (9.26), neral (7.15), thymol (3.45)	(Aprotosoiaie et al. 2019)
Republic of Moldova	<i>T. citriodorus</i>	Aerial parts	Full flowering (July 2015)	Hydrodistillation (modified Clevenger) of powered plant for 3 h. 100 g / 1 L of double distilled water	1.61 mL of VO / 100 g of dry weight	Lavandulol (54.27) , geranial (12.25), neral (9.00), nerol (3.88)	(Aprotosoiaie et al. 2019)
Turkey	<i>T. citriodorus</i>	Aerial parts	Pre-flowering, flowering and post-flowering (May to June 2012)	Hydrodistillation (Clevenger) of dried plant (20 g) for 3 h	2.0% (m/m) pre-flowering; 1.9% (m/m) flowering; 1.3% (m/m) post-flowering	Variations from pre-flowering to post-flowering stages: Terpinolene (61.20 - 59.0) , α -terpineol (22.84 – 29.56), linalool (3.47 – 1.35), bornyl acetate (1.34 – 1.94), borneol (1.11 – 1.24)	(Toncer et al. 2017)
Turkey	<i>T. citriodorus</i>	ND	(June 2015)	Hydrodistillation (Clevenger) of dried plant (100 g) for 4 h	1.4% (m/m) of dry weight	Geraniol (38.53) , neral (29.20), nerol (5.61), L-borneol (4.94), 3-octanone (2.66)	(Duman and Özcan 2017)

Italy	<i>T. citriodorus</i>	Above ground tissues	(May 2014)	Steam distillation of fresh plants (≈ 1 kg) ^a	0.16 mL / 100 g	Citronellol (40.1) , geranyl acetate (27.5), geranial (5.7), neral (4.4), nerol (3.9)	(Checcucci et al. 2017)
Italy	<i>T. citriodorus</i>	ND	ND	Hydrodistillation (Clevenger) of dried plant for 2 h	Variable across drought stress conditions: 0.6 mL – 1 mL/ 100 g of dry weight	Variations across different levels of PEG-6000 induced drought stress ^c : Geraniol (4.2 – 21.9) , pseudophytol (12.1 – 5.0), carvacrol (0 – 31.7), thymol (0 – 4.4)	(Tátrai et al. 2016)
Portugal	<i>T. citriodorus</i>	Leaves and small branches of aerial parts	Autumn	Hydrodistillation (Clevenger) of the dried plant (30 g) for 3 h, according to the European Pharmacopeia 6 th edition	1.26% (m/m)	Geraniol (76.1) , geranial (5.28), neral (3.65), caryophyllene (1.57), nerol (1.44)	(Teixeira and Rodrigues 2014)
ND	<i>T. citriodorus</i>	Leaves	ND	Steam distillation ^a	ND	Borneol (28.82) , thymol (14.43), linalool (8.26), α -terpineol (8.23), camphor (5.1)	(Wu et al. 2013)
Italy (accessions with different origins)	<i>T. citriodorus</i>	ND	ND	Hydrodistillation for 4 h ^a	ND	Variation depending of the accessions' origin: Geraniol (62.04 - 27.88) , citral ^d (13.75 – 1.18), <i>trans</i> -caryophyllene (5.80 – 2.42), citronellol (9.57 – 0.82), linalool (2.00 – 0.11), myrcene (12.00 – 0.13)	(Lisi et al. 2011)
Italy	<i>T. citriodorus</i> “anderson’s gold”	Aerial parts	Full blooming	Hydrodistillation (Clevenger) of air-dried plant for 2 h according to Italian Pharmacopoeia XI	ND	Geraniol (61.4) , geranial (14.3), neral (9.1)	(Bertoli et al. 2010)

				edition. 10.0 g - 12.5 g / 2 L distilled water			
Italy	<i>T. citriodorus</i> “silver queen”	Aerial parts	Full blooming	Hydrodistillation (Clevenger) of air-dried plant for 2 h according to Italian Pharmacopoeia XI edition. 10.0 g - 12.5 g / 2 L distilled water	ND	Geraniol (67.1) , geranial (9.7), neral (4.8), 3-octanol (3.4)	(Bertoli et al. 2010)
Iran	<i>T. citriodorus</i>	Aerial parts	Pre-flowering, beginning of flowering, full flowering and fruit set (June to July)	Hydrodistillation (Clevenger) of dried plants (90 g) for 4 h according to the European Pharmacopoeia 2 nd edition	Before flowering: 1.8% (m/m); Beginning of flowering: 2.21% (m/m); Full flowering: 1.45% (m/m); Fruit set: 1.66% (m/m)	Variations from pre-flowering to fruit stages: geraniol (72.5 - 54.2) , geranial (3.2 – 11.9), neral (2.2 – 7.1), 3-octanol (2.7 – 2.6), nerol (1.3 – 2.4)	(Omidbaigi et al. 2009)
Hungary	<i>T. citriodorus</i>	ND	(July 2002)	Steam distillation of air- dried material for 3 h, according to European Pharmacopoeia 4 th edition	0.36% of dry weight	Geraniol (39.2) , carvacrol (15.4), geranial (9.2), neral (7.1), <i>p</i> -cymene (4.5)	(Horváth et al. 2006)
Hungary	<i>T. citriodorus</i> ‘archer’s gold’	ND	(July 2002)	Steam distillation of air- dried material for 3 h, according to European Pharmacopoeia 4 th edition	0.29% of the dry weight	Carvacrol (43.5) , <i>p</i> -cymene (21.1), 1,8- cineole (7.5), β -caryophyllene (3.5)	(Horváth et al. 2006)

Italy	<i>T. citriodorus</i>	ND	ND	Steam distillation ^a	ND	Geraniol (36.4) , geranyl acetate (22.4), nerol (5.31), geranial (4.99), neral (4.89)	(Sacchetti et al. 2005)
Iran	<i>T. citriodorus</i>	Leaves	Full flowering (June)	Hydrodistillation (Clevenger) of dried plants (90 g) for 4 h according to the European Pharmacopoeia 2 nd edition	1.4% (m/m) of dry weight	Geraniol (54.4) , geranial (13.9), nerol (5.2), 3-octanone (3.3), borneol (3.2)	(Omidbaigi et al. 2005)
Hungary	<i>T. citriodorus</i>	Aerial parts	ND	Steam distillation of the air dried material, according to European Pharmacopoeia 4 th edition	0.36% of dry weight	Geraniol (43) , carvacrol (6.1), thymol (0.8)	(Horváth et al. 2004)
Hungary	<i>T. citriodorus</i> “archer’s gold”	Aerial parts	ND	Steam distillation of the air dried material, according to European Pharmacopoeia 4 th edition	0.29% of dry weight	Carvacrol (55.1) , thymol (0.7)	(Horváth et al. 2004)
Germany	<i>T. citriodorus</i>	Aerial parts	(July to October 1992)	Hydrodistillation ('Karlsruher apparatus') of the fresh plant for 4h using n-hexane (2 mL) as solvent ^a	ND	Geraniol (61.3) , geranial (8.2), neral (5.5), β -caryophyllene (3.7), nerol (2.8)	(Stahl-Biskup and Holthuijzen 1995)

^a Details of the distillation method were not reported; ^b 5, 10, 15, 20, 25, 30, 35, 40, 45, 50, 55, 60, 70, 80, 90, 100, 110, 120 and 150 min; ^c variations due to drought stress induced with PEG-6000 from 0% (control) to 4%; ^d Isomeric distribution is not described; ND – Information not disclosed in the original paper

Table 2 Source, processing and extraction features and chemical characterization of *Thymus × citriodorus* extracts

Source	Preparation	Plant parts	Harvesting stage (period)	Extraction method and details	Yield	Analyzed by	Major components	Ref.
Greece	Water extract	ND	ND	<i>Aqueous extraction under sonication</i> 5 g of plant powder / 25 mL of water and sonication for 15 min. The obtained extract was filtered through cotton	ND	GC-MS ^a	Traces of thymol	(Ntalli et al. 2020b)
Portugal	Aqueous extract	Upper part of stems and their leaves	Post-blooming, end fructification (October 2014)	<i>Aqueous decoction</i> 0.5 g of lyophilized plant / 150 mL of distilled water and boiled with agitation for 20 min. The extract was filtered twice and the volume was reduced by concentration in a rotary evaporator	~9% (m/m)	HPLC-DAD/ESI-MS	In mg/g of extract Rosmarinic acid (51.80 ± 1.86), luteolin-(?)-O-hexuronide (13.22 ± 0.36), salvianolic acid A (11.91 ± 0.96), chrysoeriol-(?)-O-hexoside (7.06 ± 0.65), salvianolic acid K (5.21 ± 1.5)	(Taghouti et al. 2020)
Portugal	Hydroethanolic extract	Upper part of stems and their leaves	Post-blooming, end fructification (October 2014)	<i>Hydroethanolic extraction</i> 0.5 g of lyophilized plant material / 50 mL of 80 % ethanol solution. The mixture was agitated for 1 h and centrifuged for 5 min, followed by a second extract of the pellet. The extracts were filtered and then concentrated in a rotary evaporator	~14% (m/m)	HPLC-DAD/ESI-MS	In mg/g of extract Rosmarinic acid (86.23 ± 13.88), salvianolic acid I (25.87 ± 3.90), luteolin-(?)-O-hexuronide (17.25 ± 3.13) chrysoeriol-(?)-O-hexoside (11.96 ± 1.83), salvianolic acid A (8.32 ± 0.62)	(Taghouti et al. 2020)
Greece	Hydrosol	Aerial parts	Flowering	<i>Condensation water from hydrodistillation</i> Hydrodistillation (Clevenger) of dried plant for 3 h. 100 g of plant / 1 L of distilled water. The obtained hydrosol was filtered	ND	GC-MS ^b	In percentage (%) ^c Geraniol (44.06), carvacrol (4.39), thymol (2.22)	(Ntalli et al. 2020a)

Portugal	Supercritical fluid extract	Leaves and small branches	Autumn	<i>Supercritical CO₂ fluid extraction</i> Compressed CO ₂ (40 °C) was added to 30.0 g ± 0.1 g of plant material in an in-house built equipment for 3 h.	0.05% (m/m)	GC-MS	In percentage (%) Geraniol (66.47 ± 0.03), geranial (8.63 ± 0.06), thymol (7.63 ± 0.09)	(Teixeira and Rodrigues 2014)
Poland	Methanol extract	Fresh plant leaves	ND	<i>Methanolic extraction</i> 0.2 g of fresh plant / 30% methanol (with 1 % acetic acid and 1% ascorbic acid) for 20 min under sonication with occasional shaking. After centrifugation the supernatant was filtered	ND	UPLC-MS	In mg/100 g of fresh wight Gallic acid (2754.3 ± 2.4), (+) – catechin (1414.1 ± 1.4), kaempferol 3-O-rutinoside (718.8 ± 3.9)	(Kolniak-Ostek and Oszmiański 2015; Paśławska et al. 2020)
Poland	Methanol extract	Dried plant Leaves	ND	<i>Methanolic extraction</i> 0.2 g of powered dry (by CD, FB, MFB or MV) plant / 30% methanol (with 1 % acetic acid and 1% ascorbic acid) for 20 min under sonication with occasional shaking. After centrifugation the supernatant was filtered	ND	UPLC-MS	Variable due to different drying techniques (mg/100 g of dry matter) (+) - Catechin (290.0 ± 0.6 – 1592.3 ± 2.1), <i>cis</i> -5-caffeolquinic acid (174.7 ± 0.4 – 3603.3 ± 6.2), B-type procyanidin dimer (85.7 ± 0.7 – 1131.1 ± 0.3), procyanidin C1 (30.3 ± 0.2 – 403.1 ± 4.2)	(Kolniak-Ostek and Oszmiański 2015; Paśławska et al. 2020)
Portugal	Aqueous infusions	Apical leaves	Summer months	<i>Aqueous infusion</i> 0.3 g of dry plant material / 100 mL of heated distilled water (85 °C). The infusion was left to stand for 7 min until filtration	ND	HPLC-DAD-ESI/MS	Variable accordingly to lots in µg/mL (standard lot <i>versus</i> reserve lot) Rosmarinic acid (78 ± 2 vs 66 ± 2), luteolin 7-O-glucuronide (39 ± 1 vs 41.2 ± 0.9), lithospermic acid A (19.4 ± 0.6 vs 18.8 ± 0.7)	(Rita et al. 2018)
Lithuania	Methanol extract	Above-ground parts	From floral budding to	<i>Methanolic extraction</i> 0.5 g sieved plant / 10 mL of 100% methanol in an ultrasonic bath for 30	ND	HPLC–UV	Variation from floral bud to end vegetation (µg/g, dry weight):	(Raudone et al. 2017)

			the end of vegetation (May and September 2016)	min. The extracted samples were centrifuged and filtered			Oleonolic acid ($1820 \pm 29 - 1506 \pm 24$), ursolic acid ($4869 \pm 86 - 3965 \pm 70$)	
Lithuania	Ethanol extract	Above- ground parts	From floral budding to the end of vegetation (May and September 2016)	<i>Ethanolic extraction</i> 0.1 g of sieved plant material / 10 mL of 70 % ethanol in an ultrasonic bath for 30 min. The extracted samples were centrifuged and filtered	ND	HPLC–UV	Variation through different phenological phases ($\mu\text{g/g}$, dry weight): Rosmarinic acid ($27.273 \pm 170 - 9008 \pm 18$), caffeic acid ($146 \pm 1 - 138 \pm 1$), luteolin-7-rutinoside ($581 \pm 7 - 72 \pm 1$), luteolin-7-glucoside ($148 \pm 2 - 5 \pm 0.1$)	(Raudone et al. 2017)
Portugal	Ethanol extract	Leaves and stems	ND	<i>Ethanolic extraction</i> 5 g of aerial parts (previously defatted 3 times with 150 mL n- hexane for 30 min at room temperature) / 150 mL of ethanol solution (80% (v/v)) at room temperature, for 30 min. The resulting mixture was filtered and the residue was re-extracted for four times.	ND	HPLC/DAD - ESI/MS	In mg/g extract Luteolin-7- <i>O</i> - α -glucuronide (12 ± 2), rosmarinic acid (10.4 ± 0.6), apigenin-7- β - <i>O</i> -glucuronide (9 ± 2)	(Pereira et al. 2013b)(Per eira et al. 2013a)
Portugal	Hydrolate	ND	ND	<i>Condensation of water from steam distillation^e</i>	ND	GC-MS ^d	Relative amount in volatile part (%) 1,8-Cineole (26.3), linalool (24.3), geraniol (13.9), α -terpineol (11.2) Relative amount in hydrolate (%): 1,8-Cineole (0.0145), linalool (0.0134), geraniol (0.0077), α - terpineol (0.0062)	(Oliveira et al. 2022)

Ukraine	Alcoholic extract	ND (var. “silver queen”)	(June to October 2017–2021)	Alcoholic extraction (1:10) ^e	ND	GC-MS	In percentage (%) Oleic acid (10.35 ± 1.42), caryophyllene oxide (9.62 ± 0.97), matricarin (7.59 ± 0.86), geranial (6.35 ± 0.75), β -bisabolene (6.15 ± 0.75), thymol (5.46 ± 0.70)	(Steshenko and Mazulin 2022)
Japan	Aromatic extract	ND	ND	Solvent extraction Overnight extraction with diethyl ether ^e		GC-MS	In percentage (%) Geraniol (50), geranial (5.6), caryophyllene (4)	(Hirai et al. 2022)
USA	Methanolic extract	Edible parts of fresh plant	ND	Methanolic extraction 10 g / 0.2 L of methanol at room temperature (twice). The extracts were centrifuged and the supernatants were filtered		HPLC	Variation among plants purchased with 2-3 months interval (mg / g, wet weight): Rosmarinic acid ($5.60 - 9.70$) ^f	

ND – Information not disclosed in the original paper; ^a To proceed to GC-MS chemical analysis, water extracts and hydrosols were subjected to an extraction with two portions of petroleum ether (25 mL each); ^b submitted to chemical analysis after 1/1 (v/v) fractionation in petroleum ether; ^c Percentage in relation to the volatile fraction isolated from the hydrolate; ^d Prior GC-MS analysis, the sample (100 mL) was added of *n*-dodecane as internal standard and submitted to liquid/liquid partition with *n*-pentane/diethyl oxide [93/7 (v/v)]. The organic fraction was concentrated at low pressure and room temperature and further adjusted to 10 mL with *n*-pentane; ^e Further details were not disclosed; ^f Only rosmarinic acid was quantified. MFB - microwave- assisted fluidized bed; MV - microwave-vacuum; CD - convective drying; FB - fluidized bed

Table S3 Reported relevant activities (in bold) of *Thymus × citriodorus* volatile oils (VO) and extracts and the major compounds described for each preparation

Preparation	Source	Main compounds	Reported activity	Ref.
VO	Greece	Geraniol, geranial, neral	Nematicidal activity (<i>M. incognita</i> and <i>M. javanita</i>)	(Ntalli et al. 2020a)
Water extracted (filtered)	Greece	ND	Nematicidal activity (<i>M. incognita</i> and <i>M. javanita</i>)	(Ntalli et al. 2020a)
Hydrolate	Greece	Geraniol, carvacrol, thymol	Nematicidal activity (<i>M. incognita</i> and <i>M. javanita</i>)	(Ntalli et al. 2020a)
Water extract (from powder plant)	Greece	Traces of thymol	Nematicidal activity (<i>M. incognita</i> and <i>M. javanita</i>)	(Ntalli et al. 2020b)
Plant powder	Greece	ND	Bio-fertilizing properties	(Ntalli et al. 2020a)
VO from <i>T. citriodorus</i> “aureus” cultivar	Republic of Moldova	Geraniol, geranial, neral	Antifungal activity (<i>Aspergillus flavus</i>); Antioxidant capacity (RP, DPPH, ABTS ⁺)	(Aprotosoiaie et al. 2019)
VO	Republic of Moldova	Lavandulol, geranial, neral	Antifungal activity (<i>Aspergillus flavus</i>); Antioxidant capacity (DPPH, ABTS ⁺)	(Aprotosoiaie et al. 2019)
VO	Italy	Geraniol, geranyl acetate, nerol	Antimicrobial activity (<i>C. albicans</i> , <i>R. glutinis</i> , <i>S. cerevisiae</i> , <i>S. pombe</i> , <i>Y. lipolytica</i>); Antioxidant capacity (DPPH, β -carotene)	(Sacchetti et al. 2005)
Ethanol extracts of frozen and dried plant	Slovenia	ND	Antimicrobial activity (<i>S. aureus</i> , <i>L. monocytogenes</i> , <i>C. jejuni</i> , Verotoxin <i>E. coli</i>)	(Abramovič et al. 2018)
Methanol–water extracts of leaves and rhizomes	Poland	ND	Antibacterial activity (<i>Bacillus subtilis</i>)	(Orłowska et al. 2015)
VO	Portugal	Geraniol, 1,8-cineole, thymol	Antibacterial activity (<i>S. aureus</i> , <i>S. epidermidis</i> , <i>C. acnes</i>); Anti-biofilm activity (<i>C. acnes</i>); Anti-inflammatory activity on macrophages cell line (Raw 264.7); Antioxidant capacity (DPPH); Environmental toxicity on freshwater crustacean <i>Daphnia magna</i>	(Oliveira et al. 2022)
Hydrolate	Portugal		Antibacterial activity (<i>S. aureus</i> , <i>S. epidermidis</i> , <i>C. acnes</i>); Anti-biofilm activity (<i>C. acnes</i>); Anti-inflammatory activity on macrophages cell line (Raw 264.7); Antioxidant capacity (DPPH)	(Oliveira et al. 2022)

VO from <i>T. citriodorus</i> “silver queen” cultivar	Ukrain	ND	Antimicrobial activity (<i>S. aureus</i> , <i>E. coli</i> , <i>P. aeruginosa</i> , <i>C. albicans</i>)	(Steshenko et al. 2021)
Hydroethanolic extract	Portugal	Rosmarinic acid, salvianolic acid I, luteolin-(?)-O-hexuronide	Anti-proliferative activity on human colon adenocarcinoma cell line (Caco-2); Antioxidant capacity (ABTS ⁺ , •OH, NO•)	(Taghouti et al. 2020)
Aqueous decoction	Portugal	Rosmarinic acid, luteolin-(?)-O-hexuronide, salvianolic acid A	Anti-proliferative activity on human colon adenocarcinoma cell line (Caco-2); Antioxidant capacity (ABTS ⁺ , •OH, NO•)	(Taghouti et al. 2020)
Aqueous infusions	Portugal	Rosmarinic acid, luteolin 7-O-glucuronide, lithospermic acid A	Antioxidant capacity (RP, DPPH, β-carotene)	(Rita et al. 2018)
Ethanolic extract	Portugal	Luteolin-7-O-α-glucuronide, rosmarinic acid, apigenin-7-β-O-glucuronide	Cytoprotective activity on human hepatoblastoma cell line (HepG2); Antioxidant capacity (RP, DPPH, ROS)	(Pereira et al. 2013a)
VO	ND	Borneol, thymol, linalool, α-terpineol	Antiproliferative activity on human hepatoblastoma cell line (HepG2)	(Wu et al. 2013)
Extract ^a	ND	ND	Anti-aging properties by stimulation of Emilin-1 (normal human skin fibroblasts cell line)	(Fitoussi et al. 2011)
Methanolic extract	Palestina	ND	Pesticidal (antifungal) properties (<i>Ustilago tritici</i> and <i>Ustilago nuda</i>)	(Karsou and Samara 2021)
Ethanolic extract	Turkey	ND	Antibacterial activity (<i>S. aureus</i> , <i>E. faecalis</i> , <i>P. aeruginosa</i> , <i>E. coli</i>)	(Kerem et al. 2023)
Hexane extract	Turkey	ND	Antibacterial activity (<i>S. aureus</i>)	(Kerem et al. 2023)
Chloroformic extract	Turkey	ND	Antibacterial activity (<i>S. aureus</i> , <i>E. faecalis</i> , <i>P. aeruginosa</i> , <i>E. coli</i>)	(Kerem et al. 2023)

^aDetails on the type of extract were not available; ND – information not disclosed in the original work; RP – reducing Power; DPPH – 2,2-Diphenyl-1-picrylhydrazyl radical-scavenging activity; β-carotene – β-carotene bleaching inhibition; ABTS^{•+} – ABTS radical cation scavenging; •OH – hydroxyl radicals scavenging assay; NO• – nitric oxide radical scavenging activity; ROS – Reactive oxygen species

Table S4 Reported antimicrobial activity of *Thymus* × *citriodorus* volatile oils (VO) and extracts

Preparation	Source	Antimicrobial activity	Ref.
VO from <i>T. citriodorus</i> “aureus”	Republic of Moldova	<i>Aspergillus flavus</i> MUCL 19006 MIC: 0.5 µL/mL; MLC: 1 µL/mL	(Aprotosoia et al. 2019)
VO	Republic of Moldova	<i>Aspergillus flavus</i> MUCL 19006 MIC: 1 µL/mL; MLC: 1 µL/mL	(Aprotosoia et al. 2019)
VO	Italy	<i>Candida albicans</i> ATCC 48274 - MIC ^a : 0.06 mg/mL <i>Rhodotorula glutinis</i> ATCC 16740 - MIC ^a : 0.09 mg/mL <i>Saccharomyces cerevisiae</i> ATCC 60232 - MIC ^a : 0.06 mg/mL <i>Schizosaccharomyces pombe</i> ATCC 2365 - MIC ^a : 0.06 mg/mL <i>Yarrowia lipolytica</i> ATCC 16617 - MIC ^a : 0.03 mg/mL	(Sacchetti et al. 2005)
Ethanol extracts of frozen and dried plant	Slovenia	MIC (mg CAE / mL growth media) <i>Staphylococcus aureus</i> Frozen: 621 ± 217; Dried: 490 ± 172 <i>Listeria monocytogenes</i> Frozen: 1064 ± 0; Dried: 1190 ± 413 <i>Campylobacter jejuni</i> Frozen: 288 ± 252; Dried: 306 ± 286 <i>Verotoxin Escherichia coli</i> Frozen: 3900 ± 868; Dried: 6158 ± 1371	(Abramovič et al. 2018)
Methanol–water extracts of leaves and rhizomes	Poland	Inhibition zone (cm ²) ^b <i>Bacillus subtilis</i> ATCC 6633 Leaves: 1.44 (±0.07); Rhizomes: 0.78 (±0.06)	(Orłowska et al. 2015)
VO	Portugal	<i>Staphylococcus aureus</i> ATCC 6538 MIC: 0.125% (v/v); MLC: 0.250% (v/v) <i>Staphylococcus epidermidis</i> DSM 28764 MIC: 0.06% (v/v); MLC: 0.250% (v/v)	(Oliveira et al. 2022)

		<i>Cutibacterium acnes</i> DSM 1897 MIC: 0.06% (v/v); MLC: 0.125% (v/v)	
Hydrolate	Portugal	<i>Staphylococcus aureus</i> ATCC 6538 MIC: > 50% (v/v); MLC: > 50% (v/v) <i>Staphylococcus epidermidis</i> DSM 28764 MIC: > 50% (v/v); MLC: > 50% (v/v) <i>Cutibacterium acnes</i> DSM 1897 MIC: 50% (v/v); MLC: > 50% (v/v)	(Oliveira et al. 2022)
VO from <i>T. citriodorus</i> ‘silver queen’	Ukraine	Growth inhibition zones (mm) <i>Staphylococcus aureus</i> ATCC 25923 – 14.60 ± 1.5 <i>Escherichia coli</i> ATCC 259220 – 19.60 ± 1.8 <i>Pseudomonas aeruginosa</i> ATCC 90270 – 0 (no activity) <i>Candida albicans</i> ATCC 885/6530 – 29.30 ± 2.8	(Steshenko et al. 2021)
Ethanolic extract	Turkey	Growth inhibition zones (mm) depend on applied volume (15 µL, 20 µL or 25 µL, respectively) <i>Enterococcus faecalis</i> ATCC 29212 – 18; 22; 14.5 <i>Staphylococcus aureus</i> ATCC 29213 – 45; 43; 32.5 <i>Pseudomonas aeruginosa</i> ATCC 2753 – 9; 11.5; 9 <i>Escherichia coli</i> ATCC 25922 – 3.5; 4; 9	(Kerem et al. 2023)
Hexane extract	Turkey	Growth inhibition zones (mm) depend on applied volume (15 µL, 20 µL or 25 µL, respectively) <i>Staphylococcus aureus</i> ATCC 29213 – 5.5; 17; 8.5	(Kerem et al. 2023)
Chloroformic extract	Turkey	Growth inhibition zones (mm) depend on applied volume (15 µL, 20 µL or 25 µL, respectively) <i>Enterococcus faecalis</i> ATCC 29212 – 17; 19; 20.5 <i>Staphylococcus aureus</i> ATCC 29213 – 30; 50; 16.5 <i>Pseudomonas aeruginosa</i> ATCC 2753 – 0; 4; 0 <i>Escherichia coli</i> ATCC 25922 – 0; 0; 3.5	(Kerem et al. 2023)

MIC – Minimum inhibitory concentration; MLC – Minimum lethal concentration; ^a MIC was considered as the lowest concentration of each VO showing a clear zone of inhibition by standard discs diffusion technique; ^b by Dot-blot Test

Table S5 Bioactive properties of *Thymus × citriodorus* volatile oils (VO) and extracts regarding their antiproliferative and anti-inflammatory activities

Preparation	Source	Anti-proliferative / cytotoxic activity (IC ₅₀)	Anti-inflammatory activity (EC ₅₀)	Ref.
Hydroethanolic extract	Portugal	Caco-2 cells 24h: 128.2 µg/mL ± 5.75 µg/mL 48h: 114.6 µg/mL ± 4.38 µg/mL HepG2 cells 24h: > 500 µg/mL 48h: > 500 µg/mL		(Taghouti et al. 2020)
Aqueous decoction	Portugal	Caco-2 cells 24h: 223.70 µg/mL ± 8.38 µg/mL 48h: 159.40 µg/mL ± 16.85 µg/mL HepG2 cells 24h: > 500 µg/mL 48h: > 500 µg/mL		(Taghouti et al. 2020)
VO	Portugal	Raw 264.7 cells 24h: 0.01% (v/v)	NO production (Raw 264.7 cells) EC ₅₀ : 0.0019% (v/v)	(Oliveira et al. 2022)
Hydrolate	Portugal	Raw 264.7 cells 24h: 16.90% (v/v)	NO production (Raw 264.7 cells) EC ₅₀ : 2.76% (v/v)	(Oliveira et al. 2022)

Table S6 Antioxidant activity of *Thymus × citriodorus* volatile oils (VO) and extracts evaluated by different methods including: reducing Power (RP), DPPH radical-scavenging activity (DPPH), β -carotene bleaching inhibition (β -carotene), lipid peroxidation inhibition (TBARS), ABTS radical cation scavenging (ABTS•+), hydroxyl radicals scavenging assay (•OH) and nitric oxide radical (NO•) scavenging activity

Preparation	Source	RP	DPPH	β -carotene	TBARS	ABTS•+	•OH	NO•	Ref.
Aqueous extract	Portugal					1.21 $\pm$ 0.07 mmol Trolox eq. / g extract Or 0.11 $\pm$ 0.03 07 mmol Trolox eq. / g dry plant	(% inhibition) +EDTA: 37.97 $\pm$ 1.12% -EDTA: 30.59 $\pm$ 2.08%	(% inhibition) 41.15 $\pm$ 3.64%	(Taghouti et al. 2020)
Hydroethanolic extract	Portugal					1.52 $\pm$ 0.21 mmol Trolox eq. / g extract Or 0.22 $\pm$ 0.05 mmol Trolox eq. / g dry plant	ND	ND	(Taghouti et al. 2020)
Aqueous infusions	Portugal	EC ₅₀ (mg/mL) Standard: 0.228 $\pm$ 0.003; Reserve: 0.393 $\pm$ 0.002	EC ₅₀ (mg/mL) Standard: 0.34 $\pm$ 0.01; Reserve: 0.32 $\pm$ 0.02	EC ₅₀ (mg/mL) Standard: 1.01 $\pm$ 0.06; Reserve: 0.73 $\pm$ 0.03	EC ₅₀ (mg/mL) Standard: 0.118 $\pm$ 0.001; Reserve: 0.117 $\pm$ 0.001				(Rita et al. 2018)
VO	Republic of Moldova		EC ₅₀ (mg/mL) 34.281 $\pm$ 0.653			EC ₅₀ (mg/mL) 0.522 $\pm$ 0.011			(Aprotosoai e et al. 2019)
VO from <i>T. citriodorus</i> “aureus”	Republic of Moldova	EC ₅₀ (mg/mL) 3.057 $\pm$ 0.151	EC ₅₀ (mg/mL) 1.565 $\pm$ 0.045			EC ₅₀ (mg/mL) 0.014 $\pm$ 0.001			(Aprotosoai e et al. 2019)
Ethanolic extract	Portugal	EC ₅₀ (mg/mL) 0.0882 $\pm$ 0.0008	EC ₅₀ (mg/mL) 0.0117 $\pm$ 0.0015						(Pereira et al. 2013)

VO	Italy	25 % compared with 1 mM trolox ($28.2 \pm 0.20\%$)	50 % compared with BHA ($86.74 \pm 0.61\%$)		(Sacchetti et al. 2005)
VO	Portugal	EC ₅₀ (% v/v) 0.071 ± 0.002		ND	(Oliveira et al. 2022)
Hydrolate	Portugal	EC ₅₀ (% v/v) 20.08 ± 1.25		ND	(Oliveira et al. 2022)

ND – Not determined due to low activity

Table S7 Nematicidal potential of *Thymus × citriodorus* volatile oils (VO) and extracts

Preparation type	Source	Nematicidal activity		Ref.
		<i>Meloidogyne incognita</i> EC ₅₀	<i>Meloidogyne javanica</i> EC ₅₀	
VO	Greece	Day 1: 0.091 ± 0.087% (v/v) Day 2: 0.084 ± 0.079% (v/v)	Day 1: 0.093 ± 0.091% (v/v) Day 2: 0.092 ± 0.090% (v/v)	(Ntalli et al. 2020a)
Water extracted (filtered)	Greece	Day 1: 2.860 ± 2.470% (v/v) Day 2: 8.419 ± 7.137% (v/v)	Day 1: 3.892 ± 3.104% (v/v) Day 2: 6.197 ± 4.401% (v/v)	(Ntalli et al. 2020a)
Hydrolate	Greece	Day 1: 38.950 ± 26.704% (v/v) Day 2: 7.464 ± 5.968% (v/v)	Day 1: > 50% (v/v) Day 2: 4.353 ± 3.703% (v/v)	(Ntalli et al. 2020a)
Water extract (from powder plant)	Greece	Day 1: 35.974 ± 1.066 µL/mL Day 2: ND Day 3: ND	Day 1: 38.057 ± 3.736 µL/mL Day 2: 27.914 ± 2.557 µL/mL Day 3: ND	(Ntalli et al. 2020b)

ND – EC₅₀ values were outside the tested concentration range