

Table S1. Animal models of mechanoregulation in dentofacial development and remodeling: models, transduction pathways, key evidence, and study limitations

Category	Animal model	Method	Mechanotransduction pathway	Main finding	Limitations
Dental epithelial invagination	Mouse (E11–E13) molar placode/bud stage ^{1,2}	Live time-lapse imaging; suprabasal tissue incision–recoil; Shh inhibition; cell-mechanics analysis	Actomyosin tension–Shh axis; FGF-driven asymmetric division	Suprabasal cells assemble actin bundles to generate tension that bends the epithelium into the mesenchyme; Shh blockade prevents contraction/intercalation, yielding shallow, widened tooth germs; “vertical telescoping” proposed as a new early-invagination mechanism.	No in vivo force quantification; embryo sex mostly unspecified; mechanics inferred indirectly from recoil.
Dental epithelial invagination	Mouse (E11–E15) molar epithelial invagination; ex vivo live imaging ³	4D live imaging; fluorescent cell tracking; Myosin II inhibition; Shh conditional knockout	Myosin II–Shh coordination axis; non-muscle myosin II drives cell adhesion and movement	Myosin II mediates cell adhesion and directed movement and acts with Shh to coordinate epithelial intercalation; Myosin II inhibition or Shh loss disrupts epithelial folding, integrating mechanical and morphogen signaling.	Possible off-target effects of Myosin II inhibitors; embryo sex unspecified; window limited to E11–E15.
Dental epithelial invagination	Mouse (E12–E15) molar bud/cap stage; ex vivo organ culture ⁴	Organ culture; live fluorescence imaging; cell-trajectory tracking; cytoskeletal dynamics	Coordinated cell movement–tissue deformation axis (proliferation/migration/apoptosis coordination)	Epithelial deformation is driven by the coordinated interplay of proliferation, migration and apoptosis rather than a single factor; the coordination of cellular dynamics via actin remodeling plays an important role in tooth epithelial morphogenesis; trajectory analysis reveals collective tissue-scale mechanical behavior.	Ex vivo organ culture, largely descriptive; embryo sex unspecified; no direct force quantification; mechanism mostly correlative.
Dental epithelial invagination	Mouse (E11–E13) dental/salivary ectodermal placode ⁵	In vivo suprabasal cell labeling; laser ablation; quantification of cell intercalation	Suprabasal contraction–cell intercalation–tissue invagination axis	Invagination is driven by active suprabasal contraction; suprabasal intercalation generates an inward mechanical push; the mechanism is conserved between tooth and salivary gland; laser ablation confirms suprabasal prestress.	Ex vivo ectodermal placode model (cross-organ inference, tooth/salivary gland); embryo sex unspecified; mechanics inferred from laser-ablation recoil only.
Mesenchymal condensation & crown morphogenesis	Mouse (E13–E15) molar mesenchyme; in vivo knockout ⁶	In vivo Collagen VI knockout; IHC (Pax9, Msx1); quantification of mesenchymal condensation	LOX–collagen VI–ECM stiffening–RhoA–stem-cell fate axis	In vivo LOX cross-linking of collagen VI stabilizes the condensed mesenchyme; collagen VI loss destabilizes condensation and reduces Pax9/Msx1, showing ECM stiffening governs mesenchymal progenitor fate via RhoA.	Sex unspecified; ECM mechanics inferred from condensation phenotype; no direct force measurement.
Mesenchymal	Rodent (incisor/molar, E13–	Direct tissue-pressure	Proliferation-driven compressive	Proliferation generates an anisotropic stress	Mechanics captured at single time

condensation & crown morphogenesis	E15) proliferation-driven compression ⁷	quantification; proliferation inhibition; exogenous pressure rescue; IHC (nuclear YAP, Shh)	stress–Hippo/YAP/TAZ–Shh signaling–center formation	field; the high-pressure center shows reduced nuclear YAP, suppressed proliferation and Shh expression, specifying the enamel knot (EK); inhibition abolishes EK, rescued by exogenous pressure.	points; sex unspecified; exogenous pressure may not match native magnitude.
Mesenchymal condensation & crown morphogenesis	Mouse (E13–E15) molar cap stage; α E-catenin/Yap/Taz cKO ⁸	Conditional knockout (α E-catenin, Yap, Taz); YAP overexpression; histology/IF (Shh, Fgf4)	α E-catenin–cytoskeletal coupling–YAP/TAZ cytoplasmic retention–EK non-proliferative state	α E-catenin links neighboring cytoskeletons to resist proliferation-driven pressure; its loss causes aberrant nuclear YAP and EK defects, rescued by combined Yap/Taz deletion; YAP overexpression mislocates EK and arrests cap stage.	Single cap-stage window; sex unspecified; lacks direct in vivo tension readout.
Mesenchymal condensation & crown morphogenesis	Mouse (PN0–PN14) molar; YAP-overexpression transgenic ⁹	YAP-overexpression transgenic mouse; histology; IF (EK markers Shh); crown morphometry	YAP nuclear activity–EK specification–Hippo dose-effect axis	YAP overexpression causes abnormal crown morphology, altered EK number/position and cusp defects, confirming YAP level is a key dose-dependent determinant of EK specification and crown shaping.	Sex unspecified; endogenous YAP dynamics unresolved.
Mesenchymal condensation & crown morphogenesis	Mouse (E13–PN0) molar; Yap cKO (K14-Cre) ¹⁰	Yap conditional knockout (K14-Cre); histology; EK-marker analysis; cusp morphometry	YAP–primary/secondary EK specification–cusp number axis	Yap loss impairs primary EK specification and produces abnormal secondary EKs and fewer cusps, showing YAP/Hippo controls stage-specific EK appearance and multi-cusp patterning.	Sex unspecified; TAZ compensation not fully excluded.
Mesenchymal condensation & crown morphogenesis	Mouse molar; epithelial YAP1 cKO (Pitx2-Cre) ¹¹	Epithelium-specific YAP1 cKO; RNA-seq; IF (Shh, FGF4)	Epithelial YAP1–Shh/FGF signaling–ameloblast differentiation axis	Epithelial YAP1 promotes ameloblast terminal differentiation via Shh/FGF; YAP1 loss causes enamel defects and reduced ameloblastin, revealing YAP1's role in mechanics–differentiation coupling.	Sex unspecified; single time point for RNA-seq.
Mesenchymal condensation & crown morphogenesis	Mouse $\times$ bank vole molars; ex vivo organ culture + computation ¹²	Ex vivo culture (jaw constraint removed); artificial braces mimicking lateral compression; finite-element modeling	Jaw lateral compression–cusp offset (physical constraint)	Vole molars lose species-specific cusp offset without jaw support; mouse molars gain offset cusps with artificial braces; lateral compression is sufficient to induce offset, defining jaw constraint as essential for tooth morphology.	Ex vivo culture and computational assumptions; donor-embryo sex/age unspecified; species comparison limited to two rodents.
Mesenchymal condensation & crown morphogenesis	Rodent ever-growing incisor; incisor-truncation model ¹³	Unilateral/bilateral incisor clipping; BrdU; growth-rate assay	Occlusal unloading–mesenchymal stem-cell proliferation axis	Bilateral clipping increases mesenchymal stem-cell proliferation and eruption rate, implying occlusal load regulates stem-cell activity; unilateral clipping shows no change, so simple unloading is insufficient;	Contradictory unilateral vs bilateral outcomes; receptor unresolved; sex/age unspecified.

				integrin–YAP may mediate macro-force.	
Mandibular arch & muscle force	Mouse (E10–E12) mandibular arch mesenchyme ¹⁴	Live imaging; vinculin FRET tension sensor; Wnt5a/Piezo1/Yap/Taz KO; oscillatory cortical-force analysis	Wnt5a–YAP/TAZ–Piezo1–Ca ²⁺ transients–actomyosin ordering	Oscillatory cortical forces drive 3D mesenchymal intercalations that shape the mandibular arch; Wnt5a coordinates polarity and actomyosin dynamics; YAP/TAZ and Piezo1 orient intercalations via Ca ²⁺ and actomyosin.	Early embryonic arch only (E10–E12); sex unspecified; force-sensing hard to separate from downstream transcription; no postnatal validation.
Mandibular arch & muscle force	Mouse (E12–E17) embryonic tendon/muscle–bone coordination ¹⁵	Scx-Cre tendon lineage tracing; muscle paralysis; 3D skeletal reconstruction; craniofacial morphometry	Muscle contraction–tendon transmission–skeletal strain–bone morphological adaptation axis	Muscle-derived force transmitted via tendons is required for skeletal morphological adaptation; muscle paralysis yields characteristic craniofacial anomalies; tendon-lineage cells sense mechanical cues to coordinate the muscle–bone interface.	Sex unspecified; downstream force sensor unidentified.
Mandibular arch & muscle force	Mouse muscular dystrophy / tendon-deletion ^{16,17}	Dystrophy mutants; conditional deletion of masseter tenogenic/myogenic components; μ CT; mandibular morphometry	Tendon-transmitted force–bone mechanosensing–mandibular shape axis	Chronically reduced muscle force in dystrophy yields characteristic craniofacial anomalies; loss of masseter tendon/myogenic components causes micrognathia, masseter degeneration and reduced mandibular mechanosensing, proving masseter-tendon force is essential for mandibular shape.	Downstream molecular pathway unspecified; species/sex varied and inconsistently reported.
Tooth root development	Mouse (PN3.5–PN21) molar root; trigeminal ganglion ¹⁸	Sensory-neuron-specific KO; Ca ²⁺ imaging; PDGF-A gain/loss-of-function	Piezo2 (trigeminal sensory neuron)–Ca ²⁺ influx–PDGF-A paracrine axis	Piezo2 ⁺ trigeminal neurons sense root-associated force and secrete PDGF-A in a Ca ²⁺ -dependent manner to coordinate mesenchymal behavior; Piezo2 loss disrupts root morphogenesis, revealing a neuro-mechanical transduction mechanism.	In vivo functional contribution of the human homolog and dose–response limited; sex unspecified.
Tooth root development	Mouse (PN3.5–PN21); Gli1-CreER;Fgfr1(fl/fl) cKO ¹⁹	Tamoxifen-inducible cKO; histology; IF (Piezo2, WNT/ β -catenin); WNT-inhibitor rescue	FGFR1 (Gli1 ⁺ progenitors)–Piezo2– β -catenin/WNT cascade	Fgfr1 loss in Gli1 ⁺ progenitors aberrantly activates Piezo2 and WNT, narrowing the PDL space, ectopic cementum/bone and tooth ankylosis; WNT downregulation rescues ankylosis.	Sex unspecified; outcome assessed at limited time points.
Tooth root development	Rat (PN21) maxillary molar; sustained orthodontic force (7 d) ²⁰	Continuous force delivery (7 d); histology (H&E); Ki67 IHC; HERS morphology	Sustained force–apical proliferation–HERS integrity axis	Prolonged 7-day loading shortens roots, causes apical bending, reduces apical proliferation and disrupts HERS; damage is irreversible after force removal, defining a mechanically sensitive apical window.	Single sex/age cohort; molecular pathway unresolved; clinical extrapolation limited.
Tooth root	Rat (PN21 or 28) maxillary first	Light orthodontic force (3 d);	Light force–odontoblast	Three-day light force deforms dentin,	Short-term, single-dose study; sex

development	molar; light force (3 d) ²¹	μCT; histology; dentin/odontoblast analysis	function–dentin formation axis	disorganizes odontoblasts, reduces apical proliferation and shortens roots, showing the developing root apex is highly sensitive even to short-term light force.	unspecified; no molecular mechanism.
Tooth eruption & replacement	Dog (deciduous/permanent); follicle removal/replacement ²²	Follicle removal; metal/silicone germ replacement; radiology/histology	Bite-force sensing–follicle–polarized osteoclast/osteoblast recruitment–RANKL/OPG remodeling axis	Replacing the germ with a replica still erupts if the follicle is intact; removing the follicle arrests eruption; the root is not required, polarized osteoclast/osteoblast recruitment is the core mechanism.	Modern molecular validation limited; sex/age inconsistently reported across species.
Tooth eruption & Replacement	American alligator (lifelong tooth renewal) ²³	BrdU/EdU multi-mitotic labeling; histology; artificial extraction; IF (β-catenin, sFRP1, Sox2)	Wnt/β-catenin activation–sFRP1 loss–quiescent progenitor activation axis	Quiescent odontogenic progenitors at the distal dental lamina are activated by physiological exfoliation or extraction; cycling correlates with β-catenin activation and sFRP1 loss, revealing a reptilian renewal niche.	Limited relevance to single-generation mammalian dentition; sex/age unspecified; in vivo lineage tracing restricted.
Tooth eruption & Replacement	Miniature pig (E50–PN10) tooth-replacement model ²⁴	Serial H&E 3D reconstruction; μCT; FEA (ANSYS); Flexcell compression; IF (integrin β1, ERK1, RUNX2, Wnt)	Integrin β1–ERK1–RUNX2–mesenchymal-to-epithelial Wnt conversion	Faster deciduous growth than the alveolus accumulates compressive stress that suppresses the SDL via integrin β1-ERK1-RUNX2; eruption releases stress, RUNX2 falls and Wnt shifts to the permanent epithelium, initiating succession.	Large-animal study with limited sample size; single breed; narrow sex/age window; FEA assumptions on tissue properties.
Tooth eruption & Replacement	Miniature pig; Fbln1 KO; SDL quiescence ²⁵	Conditional Fbln1 KO; RUNX2 overexpression; ChIP (RUNX2–Fbln1 enhancer); IF	RUNX2–Fibulin-1 (Fbln1)–SDL niche homeostasis axis (force→transcription→matrix)	RUNX2 binds the Fbln1 enhancer; the biomechanical RUNX2–Fibulin-1 axis maintains SDL niche quiescence; Fbln1 loss causes premature SDL activation, gating replacement timing.	Sex/age unspecified; niche mechanics inferred, not directly measured.
Jaw bone development	Rat ; soft- vs hard-diet ²⁶⁻²⁸	Dietary intervention; μCT; histology; IHC (IGF-1r, PTHrP, Col2a1); mandibular morphometry	PTHrP–Ihh endochondral ossification axis; IGF-1r–chondroblast proliferation	Soft diet reduces masticatory load, lowering condylar bone volume, ramus growth and cartilage thickness with reduced IGF-1r, suppressing endochondral ossification and confirming load is required for growing-jaw development.	Single species/sex (often male) and age cohort; strain not quantified.
Jaw bone development	Rat (adult male); incisal bite-plane TMJ overload ²⁹	Bite-plane loading; μCT; histology (cartilage layers); mandibular length/ramus height	Overload–endochondral ossification suppression–condylar homeostasis axis	Increased TMJ load thins condylar cartilage, suppresses endochondral ossification and reduces total mandibular length and ramus height, inversely mirroring the soft-diet model.	Single species and sex (adult male rat only); one loading magnitude/duration; molecular pathway presumed.
Jaw bone	Mouse (Osterix-Cre;	Osteoblast/chondrocyte	BMP/BMPRI A	Bmpr1a loss shortens the anterior mandible	Single sex/age; interaction effects

development	Bmpr1a(fl/fl)); soft-diet × KO ^{30,31}	Bmpr1a cKO; soft diet; treadmill loading; μ CT; superimposed imaging	mechanosensitivity (posterior vs anterior condyle)	and posteriorly inclines the ramus; BMP governs the posterior condyle while masticatory load governs the anterior part; KO osteoblasts become hypersensitive to load, gaining trabecular bone with exercise.	hard to disentangle; load quantification limited.
Jaw bone development	Zebrafish (craniofacial cartilage); muscle paralysis/Pthlha overexpression ³²	Muscle paralysis (tricaine); Pthlha overexpression; ISH (Col10a1 hypertrophic markers); IHC	Pthlha–muscle contraction–hypertrophic zone–endochondral ossification axis	Pthlha restricts the hypertrophic zone; muscle-derived force is required for normal hypertrophic-marker expression and ossification; paralysis abolishes this, showing muscle input is necessary for craniofacial cartilage ossification.	Zebrafish cartilage biology differs from mammalian bone; paralysis is global; sex/stage inconsistently reported; translation to mammals indirect.
Jaw bone development	Mouse Yap/Taz skeletal-cell cKO ³³	Yap/Taz osteoblast/skeletal cKO; bone histomorphometry; collagen analysis; TRAP	YAP/TAZ–osteoblast activation–matrix collagen organization–osteoclast suppression axis	Combined YAP/TAZ loss yields an osteogenesis-imperfecta-like phenotype with spontaneous fractures; YAP/TAZ synergistically promote postnatal bone development and suppress osteoclasts.	Sex/age cohorts limited; phenotype may mask cell-type-specific roles.
Periodontal ligament & OTM	Mouse (Lepr-Cre lineage); periodontal homeostasis ³⁴	scRNA-seq; Lepr ⁺ PDLSC lineage tracing and ablation; Piezo1 agonist Yoda1 injection; morphometry	Lepr ⁺ perivascular PDLSC–Piezo1 mechanosensing–periodontal homeostasis axis	scRNA-seq identifies Lepr ⁺ perivascular cells as core PDL-homeostasis stem cells; their ablation disrupts homeostasis; Yoda1 markedly promotes periodontal growth, showing Piezo1-mediated occlusal-force sensing.	Sex/age unspecified; translation to human PDL uncertain.
Periodontal ligament & OTM	Mouse (Gli1-CreER lineage); periodontal homeostasis ³⁵	Gli1-CreER lineage tracing; occlusal-force removal (extraction control); in situ Gli1 ⁺ PDLSC marking	Occlusal force–osteocyte sensing–Gli1 ⁺ periodontal stem-cell activation axis	Occlusal force, sensed via osteocytes, regulates Gli1 ⁺ PDLSC proliferation and differentiation; force removal reduces Gli1 ⁺ proliferation; orthodontic force reactivates Gli1 ⁺ cells to drive alveolar remodeling.	Sex/age cohort limited; human PDL translation uncertain.
Periodontal ligament & OTM	Mouse (Gli1-CreER); orthodontic-force-induced alveolar remodeling ³⁶	Gli1-CreER lineage tracing; orthodontic spring; Gli1 ⁺ proliferation/differentiation kinetics; μ CT	Orthodontic force–Gli1 ⁺ progenitor activation–osteoblast recruitment–alveolar remodeling axis	Orthodontic stimulation activates Gli1 ⁺ PDL progenitors and drives osteoblast differentiation; Gli1 ⁺ cells are the main source of tension-side bone formation during OTM.	Sex/age unspecified.
Periodontal ligament & OTM	Rat (orthodontic tooth movement); PDL proteomics ³⁷	Orthodontic appliance loading; PDL tissue proteomics (LC-MS/MS); ECM-protein quantification	Mechanical stress–ECM-remodeling protein expression–PDL structural adaptation axis (specific pathway unresolved)	Orthodontic force markedly alters ECM-remodeling proteins (collagen, fibronectin) in the PDL; proteomics reveals a stress-induced matrix-adaptation program underlying OTM.	Sex/age unspecified; pathway inference correlative.
Periodontal ligament	Rat (orthodontic tooth	In vivo MGF injection/gene	MGF–Fyn-FAK axis–	Occlusal force induces MGF; MGF drives	Sex/age unspecified; regenerative

& OTM	movement); PDL regeneration ³⁸	delivery; FAK/ERK/p38 inhibitors; histology (PDL collagen remodeling)	ERK1/2/p38 MAPK–PDLSC fibroblastic differentiation	PDLSC fibroblastic differentiation and in vivo PDL regeneration via Fyn-FAK-ERK1/2/p38; ERK1/2 is a rapid convergent node across mechanical stimuli.	outcome short-term.
Periodontal ligament & OTM	Mouse (orthodontic tooth movement, in vivo) ³⁹	ATG7 siRNA knockdown; in vivo OTM rate; RANKL/OPG; autophagic-flux assay	ATG7-dependent autophagy–RANKL/OPG–osteoclast 'brake' mechanism	ATG7 knockdown blocks autophagy, raises RANKL/OPG and accelerates OTM in vivo, showing ATG7-dependent autophagy restrains movement rate by limiting osteoclast activity.	Mixed mouse in vivo / human in vitro design; sex/age unspecified; single force magnitude.
Periodontal ligament & OTM	Mouse (orthodontic tooth movement); PDL fibroblasts × sensory neurons, in vivo ⁴⁰	In vivo compressive force; TAC1/SP IF; neuron-activation assay; TRAP	Compression–TAC1/SP secretion–sensory-neuron activation–pro-inflammatory/osteoclast loop (neuro-immune axis)	Compression induces TAC1 expression and SP secretion in PDL fibroblasts, activating sensory neurons that in turn drive inflammation and osteoclastogenesis; this neuro-immune loop underlies orthodontic pain.	Co-culture simplifies in vivo neuro-immune complexity; sex/age unspecified; pain readout indirect.
Periodontal ligament & OTM	Rat (orthodontic tooth movement); CD109/PDL alveolar remodeling ⁴¹	CD109 conditional KO and overexpression; orthodontic spring; TRAP; osteoblast markers; μ CT	Orthodontic force–CD109 (PDLSC surface receptor)–osteogenesis/osteoclastogenesis balance axis	Mechanical force activates CD109 on PDLSCs, balancing osteogenesis and osteoclastogenesis via TGF β signaling; CD109 loss increases resorption and accelerates tooth movement.	Mixed in vivo rat OTM and in vitro PDLSC; CD109 role outside PDL unclear; sex/age unspecified; single force paradigm.
Periodontal ligament & OTM	Mouse; occlusal-force alveolar homeostasis / occlusion-loss control ⁴²	Tooth extraction/occlusal removal; μ CT; endothelial markers (CD31, Endomucin); type-H vessel quantification	Occlusal force–type-H angiogenesis–bone formation coupling axis	Occlusal force maintains bone by promoting type-H vessels (CD31hiEndomucinhi) in alveolar bone; occlusion loss reduces type-H vessels and bone mass; pro-type-H signals partially rescue bone loss.	Vessel–bone causality partly correlative; sex/age unspecified.
Temporomandibular joint (TMJ)	Gli1Cre- mouse, Piezo 1 KO, temporomandibular joint osteoarthritis model ⁴³	Histological and immunohistochemical analyses; assessing the effects of Piezo1 deletion on condylar development and homeostasis	Piezo1 in fibrocartilage stem cells	Piezo1 is a key mechanosensitive ion channel in the cartilage, regulating condylar development and homeostasis. Its deletion in Gli1+ cells disrupts FCSC differentiation and endochondral ossification, leading to developmental deficiencies.	Sex/age effects not fully separated.
Temporomandibular joint (TMJ)	Mouse (TMJ attachment–condyle interface); Gli1+ TMJ progenitors ⁴⁴	Gli1-CreER lineage tracing; multi-timepoint labeling (BrdU/EdU); scRNA-seq	Gli1+ slow-cycling progenitor–fibrocartilage homeostasis–mechanoresponsive potential axis	Gli1+ cells enrich at the TMJ attachment–condyle interface, show slow-cycling clonal expansion and constitute the core fibrocartilage progenitor pool; scRNA-seq reveals multipotency and	Lineage tracing largely descriptive; sex/age unspecified; functional contribution inferred.

				mechanoresponsive transcription.	
Temporomandibular joint (TMJ)	Mouse (TMJ osteoarthritis); Gli1 ⁺ /YAP-Hhip axis ⁴⁵	Surgically induced TMJ OA; Gli1-CreER lineage tracing; YAP cKO; Hhip functional analysis; subchondral µCT	Gli1 ⁺ osteochondral progenitor–YAP–Hhip–aberrant subchondral remodeling axis	In TMJ OA, Gli1 ⁺ osteochondral progenitors show elevated YAP activity, driving Hhip expression and aberrant subchondral remodeling; YAP loss or Hhip inhibition attenuates the OA phenotype.	Sex/age unspecified; cell-type specificity within Gli1 ⁺ pool unclear.
Temporomandibular joint (TMJ)	Porcine TMJ disc; in vitro mechanical loading ^{46,47}	In vitro disc loading; conductivity measurement; glucose diffusion coefficient; cell viability	Mechanical strain–reduced water/porosity–nutrient diffusion limit–cell metabolic stress axis	Strain lowers disc water content and porosity, reducing glucose diffusion; abnormal loading causes metabolic dysfunction and stem-pool exhaustion, driving TMJ OA, implicating disc biomechanics in degeneration.	Sex/age unspecified; does not capture chronic in vivo degeneration.

E, embryonic day; *PN*, postnatal day; *PDL/PDLSC*, periodontal ligament (stem cells); *EK*, enamel knot; *SDL*, successional dental lamina; *HERS*, Hertwig's epithelial root sheath; *OTM*, orthodontic tooth movement; *TMJ*, temporomandibular joint; *cKO*, conditional knockout; *KO*, knockout; *FEA*, finite-element analysis; *HH*, Hamburger–Hamilton stage.

References

- Li, J., Economou, A. D., Vacca, B. & Green, J. B. A. Epithelial invagination by a vertical telescoping cell movement in mammalian salivary glands and teeth. *Nature Communications* **11**, doi:10.1038/s41467-020-16247-z (2020).
- Calamari, Z. T., Hu, J. K. H. & Klein, O. D. Tissue Mechanical Forces and Evolutionary Developmental Changes Act Through Space and Time to Shape Tooth Morphology and Function. *BioEssays* **40**, doi:10.1002/bies.201800140 (2018).
- Du, W. *et al.* Myosin II mediates Shh signals to shape dental epithelia via control of cell adhesion and movement. *PLoS Genet* **20**, e1011326, doi:10.1371/journal.pgen.1011326 (2024).
- Schubert, M. *et al.* Coordination of Cellular Dynamics Contributes to Tooth Epithelium Deformations. *Plos One* **11**, doi:10.1371/journal.pone.0161336 (2016).
- Nusse, R., Panousopoulou, E. & Green, J. B. A. Invagination of Ectodermal Placodes Is Driven by Cell Intercalation-Mediated Contraction of the Suprabasal Tissue Canopy. *PLOS Biology* **14**, doi:10.1371/journal.pbio.1002405 (2016).
- Mammoto, T. *et al.* Mesenchymal condensation-dependent accumulation of collagen VI stabilizes organ-specific cell fates during embryonic tooth formation. *Dev Dyn* **244**, 713–723, doi:10.1002/dvdy.24264 (2015).
- Shroff, N. P. *et al.* Proliferation-driven mechanical compression induces signalling centre formation during mammalian organ development. *Nature Cell Biology* **26**, 519–529, doi:10.1038/s41556-024-01380-4 (2024).
- Li, C.-Y. *et al.* αE-catenin inhibits YAP/TAZ activity to regulate signalling centre formation during tooth development. *Nature Communications* **7**, doi:10.1038/ncomms12133 (2016).
- Liu, M., Zhao, S. & Wang, X. P. YAP Overexpression Affects Tooth Morphogenesis and Enamel Knot Patterning. *Journal of Dental Research* **93**, 469–474, doi:10.1177/0022034514525784 (2014).
- Kwon, H.-J. E., Li, L. & Jung, H.-S. Hippo pathway/Yap regulates primary enamel knot and dental cusp patterning in tooth morphogenesis. *Cell and Tissue Research* **362**, 447–451, doi:10.1007/s00441-015-2267-8 (2015).
- Zheng, Y. *et al.* Epithelial YAP1 Regulates Ameloblast Differentiation through SHH/FGF Signaling. *Journal of Dental Research*, doi:10.1177/00220345251336164 (2025).
- Renvoisé, E. *et al.* Mechanical constraint from growing jaw facilitates mammalian dental diversity. *Proceedings of the National Academy of Sciences* **114**, 9403–9408, doi:10.1073/pnas.1707410114 (2017).
- An, Z. *et al.* A quiescent cell population replenishes mesenchymal stem cells to drive accelerated growth in mouse incisors. *Nature Communications* **9**, doi:10.1038/s41467-017-02785-6 (2018).
- Tao, H. *et al.* Oscillatory cortical forces promote three dimensional cell intercalations that shape the murine mandibular arch. *Nature Communications* **10**, doi:10.1038/s41467-019-09540-z (2019).
- Sharir, A., Stern, T., Rot, C., Shahar, R. & Zelzer, E. Muscle force regulates bone shaping for optimal load-bearing capacity during embryogenesis. *Development* **138**, 3247–3259, doi:10.1242/dev.063768 (2011).
- Jones, D. C., Zelditch, M. L., Peake, P. L. & German, R. Z. The effects of muscular dystrophy on the craniofacial shape of *Mus musculus*. *Journal of Anatomy* **210**, 723–730, doi:10.1111/j.1469-7580.2007.00730.x (2007).
- Liu, C. *et al.* Disrupted tenogenesis in masseter as a potential cause of micrognathia. *Int J Oral Sci* **14**, 50, doi:10.1038/s41368-022-00196-y (2022).

18 Meng, L. *et al.* Piezo2⁺ mechanosensory neurons orchestrate postnatal development through mechano-chemo-transduction of PDGFA signaling. *Proc Natl Acad Sci U S A* **122**, e2504103122, doi:10.1073/pnas.2504103122 (2025).

19 Pei, F. *et al.* FGF signaling modulates mechanotransduction/WNT signaling in progenitors during tooth root development. *Bone Research* **12**, doi:10.1038/s41413-024-00345-5 (2024).

20 Zhao, L., Matsumoto, Y., Ono, T. & Iseki, S. Effects of mechanical force application on the developing root apex in rat maxillary molars. *Archives of Oral Biology* **101**, 64-76, doi:10.1016/j.archoralbio.2019.03.010 (2019).

21 Zhao, L., Matsumoto, Y., Iseki, S. & Ono, T. Effects of short-term orthodontic force application on the root at different developmental stages in rat maxillary molars. *Am J Orthod Dentofacial Orthop* **163**, 531-539.e532, doi:10.1016/j.ajodo.2022.04.016 (2023).

22 Marks, S. C., Jr. & Cahill, D. R. Experimental study in the dog of the non-active role of the tooth in the eruptive process. *Arch Oral Biol* **29**, 311-322, doi:10.1016/0003-9969(84)90105-5 (1984).

23 Wu, P. *et al.* Specialized stem cell niche enables repetitive renewal of alligator teeth. *Proc Natl Acad Sci U S A* **110**, E2009-2018, doi:10.1073/pnas.1213202110 (2013).

24 Wu, X. *et al.* Biomechanical stress regulates mammalian tooth replacement via the integrin β 1-RUNX2-Wnt pathway. *The EMBO Journal* **39**, doi:10.15252/embj.2019102374 (2019).

25 Li, G. *et al.* Fibulin-1 Regulates Initiation of Successional Dental Lamina. *Journal of Dental Research* **102**, 1220-1230, doi:10.1177/00220345231182052 (2023).

26 Mavropoulos, A., Kiliaridis, S., Bresin, A. & Ammann, P. Effect of different masticatory functional and mechanical demands on the structural adaptation of the mandibular alveolar bone in young growing rats. *Bone* **35**, 191-197, doi:10.1016/j.bone.2004.03.020 (2004).

27 Hichijo, N. *et al.* Effects of the masticatory demand on the rat mandibular development. *Journal of Oral Rehabilitation* **41**, 581-587, doi:10.1111/joor.12171 (2014).

28 Elsalanty, M. *et al.* Effects of Decreased Occlusal Loading during Growth on the Mandibular Bone Characteristics. *Plos One* **10**, doi:10.1371/journal.pone.0129290 (2015).

29 Honda, T. *et al.* Effects of mechanical load on mandibular condylar cartilage and subchondral bone of male rats. *Archives of Oral Biology* **177**, doi:10.1016/j.archoralbio.2025.106341 (2025).

30 Uptegrove, A. *et al.* Influence of bone morphogenetic protein (BMP) signaling and masticatory load on morphological alterations of the mouse mandible during postnatal development. *Archives of Oral Biology* **169**, doi:10.1016/j.archoralbio.2024.106096 (2025).

31 Roeder, R. K. *et al.* Mechanical Loading Synergistically Increases Trabecular Bone Volume and Improves Mechanical Properties in the Mouse when BMP Signaling Is Specifically Ablated in Osteoblasts. *Plos One* **10**, doi:10.1371/journal.pone.0141345 (2015).

32 Hoyle, D. J., Dranow, D. B. & Schilling, T. F. Pthlh and mechanical force control early patterning of growth zones in the zebrafish craniofacial skeleton. *Development* **149**, doi:10.1242/dev.199826 (2022).

33 Kegelman, C. D. *et al.* Skeletal cell YAP and TAZ combinatorially promote bone development. *The FASEB Journal* **32**, 2706-2721, doi:10.1096/fj.201700872R (2018).

34 Zhang, D. *et al.* Lepr-Expressing PDLSCs Contribute to Periodontal Homeostasis and Respond to Mechanical Force by Piezo1. *Adv Sci (Weinh)* **10**, e2303291, doi:10.1002/advs.202303291 (2023).

35 Men, Y. *et al.* Gli1⁺ Periodontium Stem Cells Are Regulated by Osteocytes and Occlusal Force. *Dev Cell* **54**, 639-654 e636, doi:10.1016/j.devcel.2020.06.006 (2020).

36 Liu, A. Q. *et al.* Mechanosensing by Gli1(+) cells contributes to the orthodontic force-induced bone remodelling. *Cell Prolif* **53**, e12810, doi:10.1111/cpr.12810 (2020).

37 Thant, L. *et al.* Extracellular Matrix-Oriented Proteomic Analysis of Periodontal Ligament Under Mechanical Stress. *Frontiers in Physiology* **13**, doi:10.3389/fphys.2022.899699 (2022).

38 Zhao, Y. *et al.* Mechanochemical coupling of MGF mediates periodontal regeneration. *Bioengineering & Translational Medicine* **9**, doi:10.1002/btm2.10603 (2023).

39 Yang, Z. *et al.* Inhibition of ATG7 promotes orthodontic tooth movement by regulating the RANKL/OPG ratio under compression force. *Open Medicine* **20**, doi:10.1515/med-2025-1252 (2025).

40 Symmank, J., Löffler, L., Schulze-Späte, U. & Jacobs, C. Bidirectional substance P signaling between periodontal ligament fibroblasts and sensory neurons under mechanical stress. *Frontiers in Molecular Neuroscience* **18**, doi:10.3389/fnmol.2025.1583908 (2025).

41 Li, Y. *et al.* Mechanical force-activated CD109 on periodontal ligament stem cells governs osteogenesis and osteoclast to promote alveolar bone remodeling. *Stem Cells Transl Med* **13**, 812-825, doi:10.1093/stcltm/szae035 (2024).

42 Chen, Y. *et al.* Occlusal Force Maintains Alveolar Bone Homeostasis via Type H Angiogenesis. *J Dent Res* **102**, 1356-1365, doi:10.1177/00220345231191745 (2023).

43 Wang, X., Wang, J., Zhang, Y., He, Y. & Chen, S. Piezo1 regulates fibrocartilage stem cell in cartilage growth and osteoarthritis. *Osteoarthritis and Cartilage* **33**, 980-991, doi:10.1016/j.joca.2025.04.013 (2025).

44 Correia Cavalcante, R., Zhang, H., Ma, P. X. & Mishina, Y. Gli1⁺ Cells Exhibit Clonogenicity and Slow-Cycling Features at the Temporomandibular Joint (TMJ) Enthesis–Condyle Interface. *International Journal of Molecular Sciences* **27**, 3324 (2026).

45 Xiao, Y. *et al.* Targeting YAP-Hhip-mediated osteogenesis in Gli1⁺ osteogenic progenitors suppresses aberrant subchondral bone remodeling in osteoarthritis. *Osteoarthritis and Cartilage* **33**, 1082-1094, doi:<https://doi.org/10.1016/j.joca.2025.05.007> (2025).

46 Kuo, J., Wright, G. J., Bach, D. E., Slate, E. H. & Yao, H. Effect of mechanical loading on electrical conductivity in porcine TMJ discs. *J Dent Res* **90**, 1216-1220, doi:10.1177/0022034511415275 (2011).

47 Cisewski, S. E. *et al.* The effects of oxygen level and glucose concentration on the metabolism of porcine TMJ disc cells. *Osteoarthritis Cartilage* **23**, 1790-1796, doi:10.1016/j.joca.2015.05.021 (2015).