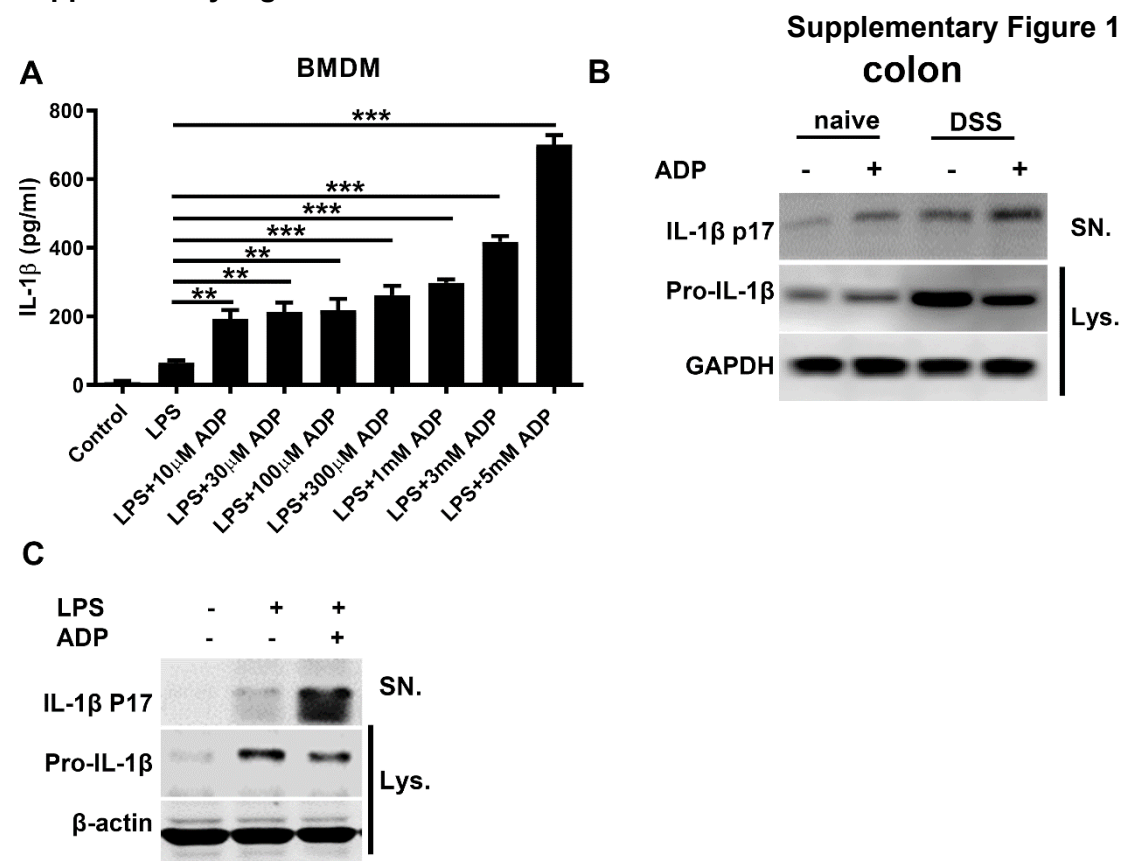


Supplementary Figures:

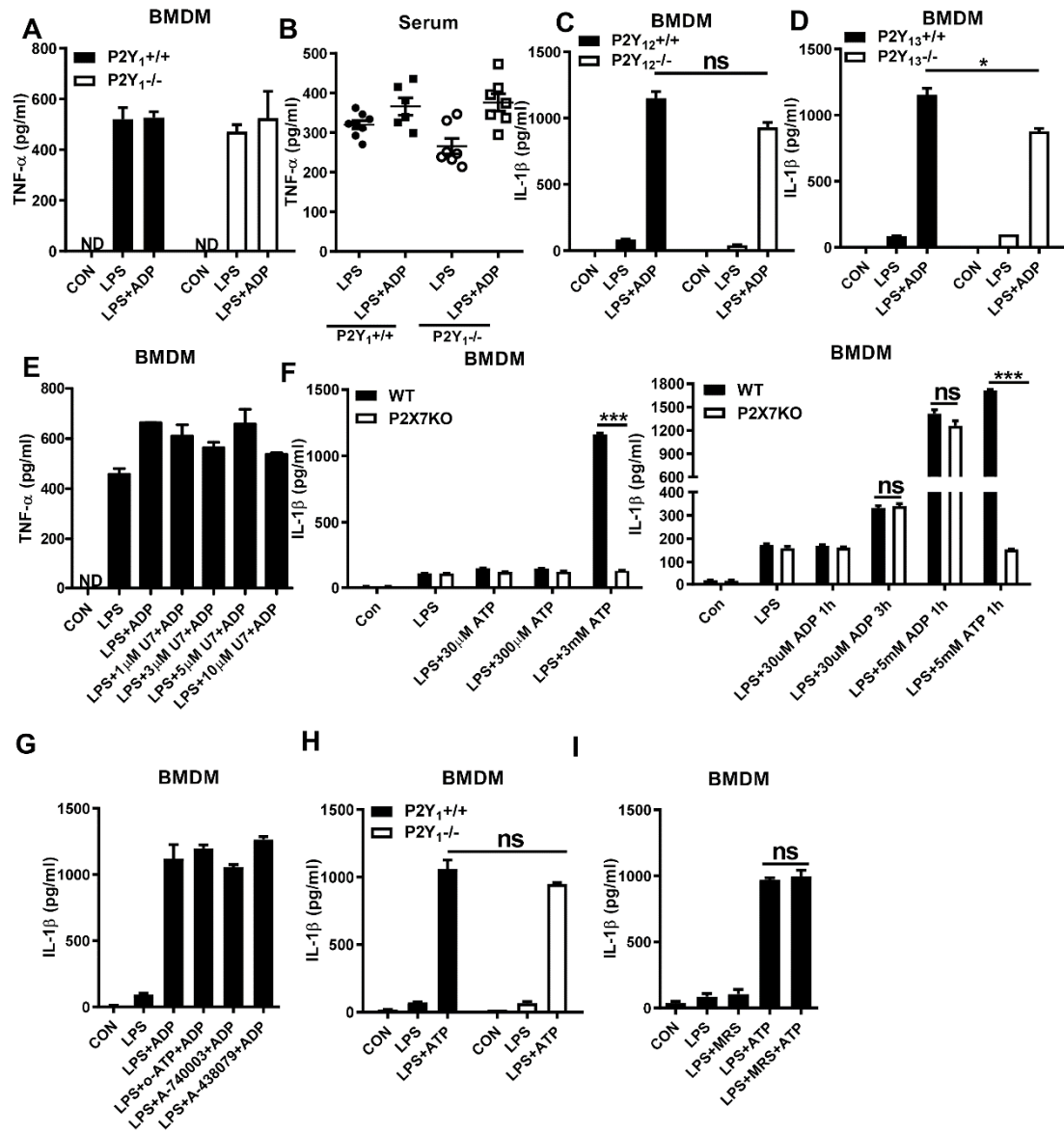


Supplementary Figure 1. ADP promotes the mature of IL-1β but not the expression of pro- IL-1β, related to Figure 2

(A) LPS-primed BMDM cells were stimulated with indicated concentrations of ADP for 3 h, then production of IL-1β was measured by ELISA. (B) WT mice were treated with 2.5% DSS for 7 days and euthanized at day 7, 0.3 g colon sections were cultured in DMEM for 6 h and then stimulated with ADP (5 mM) for 2 h. The expression of pro-IL-1β in tissue lysates and mature IL-1β in culture supernatants (SN) were analyzed by immunoblotting. (C) LPS-primed BMDM cells were stimulated with ADP (5 mM) for 80 min, the expression of pro-IL-1β in cell lysates and mature IL-1β in culture supernatants (SN) were analyzed by immunoblotting. Data are shown as mean ±SEM.

** $p < 0.01$; *** $p < 0.001$.

Supplementary Figure 2

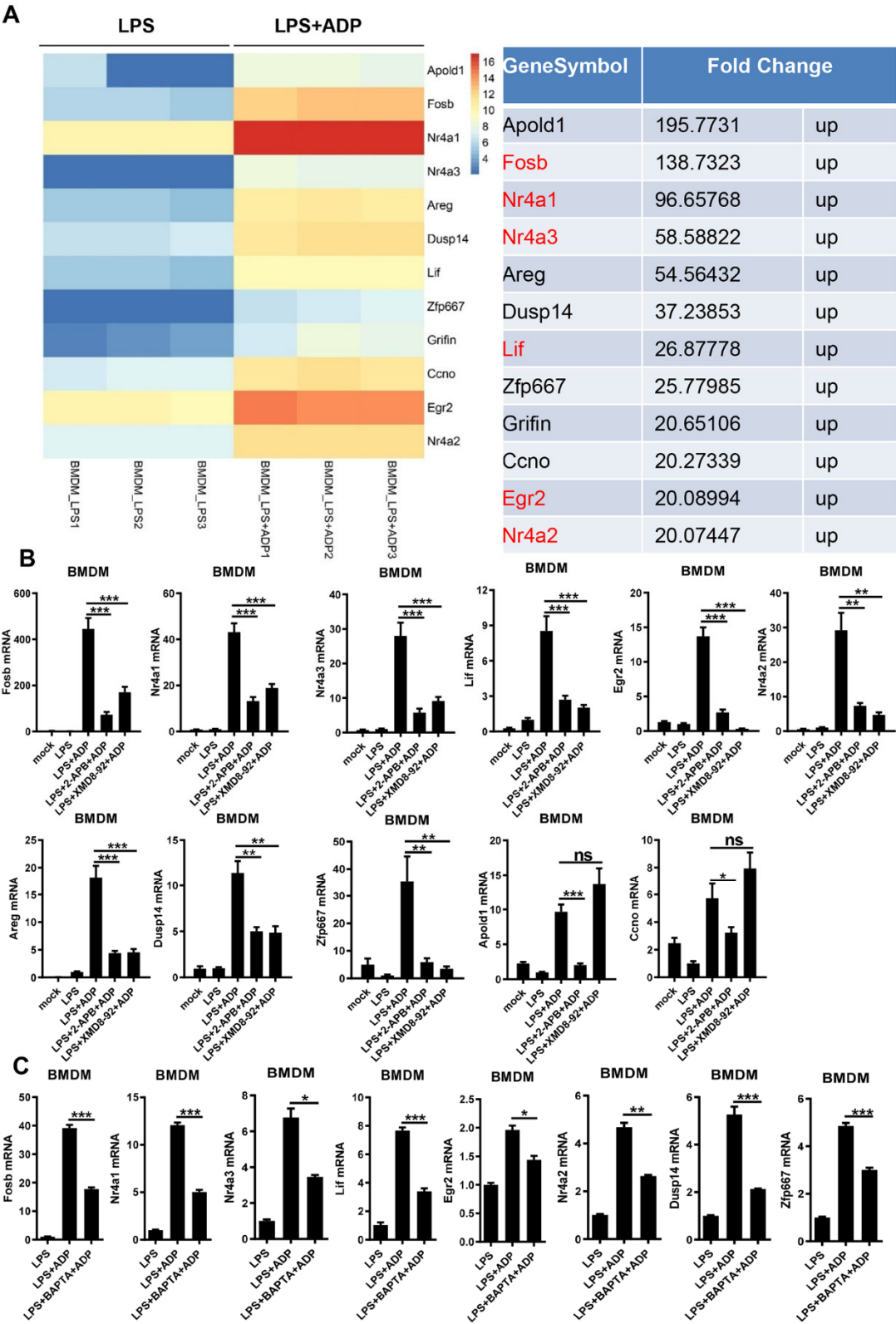


Supplementary Figure 2. P2Y₁₂ and P2Y₁₃ deficiency have little influence on ADP activation of the NLRP3 inflammasome, related to Figure 4

(A) LPS-primed P2Y₁^{+/+} and P2Y₁^{-/-} BMDM cells were treated with ADP (5 mM) for 1 h and production of TNF-α was measured by ELISA. (B) P2Y₁^{+/+} and P2Y₁^{-/-} mice were intraperitoneally treated with 20 mg/kg LPS for 6h and then intraperitoneally challenged with ADP (300mg/kg) for 1 h. The production of TNF-α in serum was measured by ELISA. (C) LPS-primed P2Y₁₂^{+/+} and P2Y₁₂^{-/-} BMDM cells were treated with ADP (5 mM) for 1 h and the production of IL-1β was measured by ELISA. (D) LPS-primed P2Y₁₃^{+/+} and P2Y₁₃^{-/-} BMDM cells were treated with ADP (5 mM) for 1 h and the production of IL-1β was measured by ELISA. (E) LPS-primed BMDM cells were pretreated with indicated concentrations of U73122 for 1 h and then stimulated with ADP (5 mM) for 1 h. The production of TNF-α was measured by ELISA. (F) LPS-primed P2X7^{+/+} and P2X7^{-/-} BMDM cells were stimulated with indicated concentrations of ATP for 1 h or indicated concentrations of ADP for the indicated time; the production of IL-1β was measured by ELISA. (G) LPS-primed BMDM cells were pretreated with P2X7 inhibitors (100 μM oATP, 5 μM A-740003, 5 μM A-438079) for 1 h and then stimulated with ADP (5 mM) for 1 h, the production of IL-1β was measured by ELISA. (H) LPS-primed P2Y₁^{+/+} and P2Y₁^{-/-} BMDM cells were treated with ATP (5 mM) for 1 h and the production of IL-1β was measured by ELISA. (I) LPS-primed BMDM cells were pretreated with MRS2179 (10 μM) or not for 1 h and then stimulated with ATP (5 mM) for 1 h, production of IL-1β was detected by ELISA. Data are shown as mean ±SEM.

p*<0.05; **p*<0.001; ns, not significant; ND, not detected.

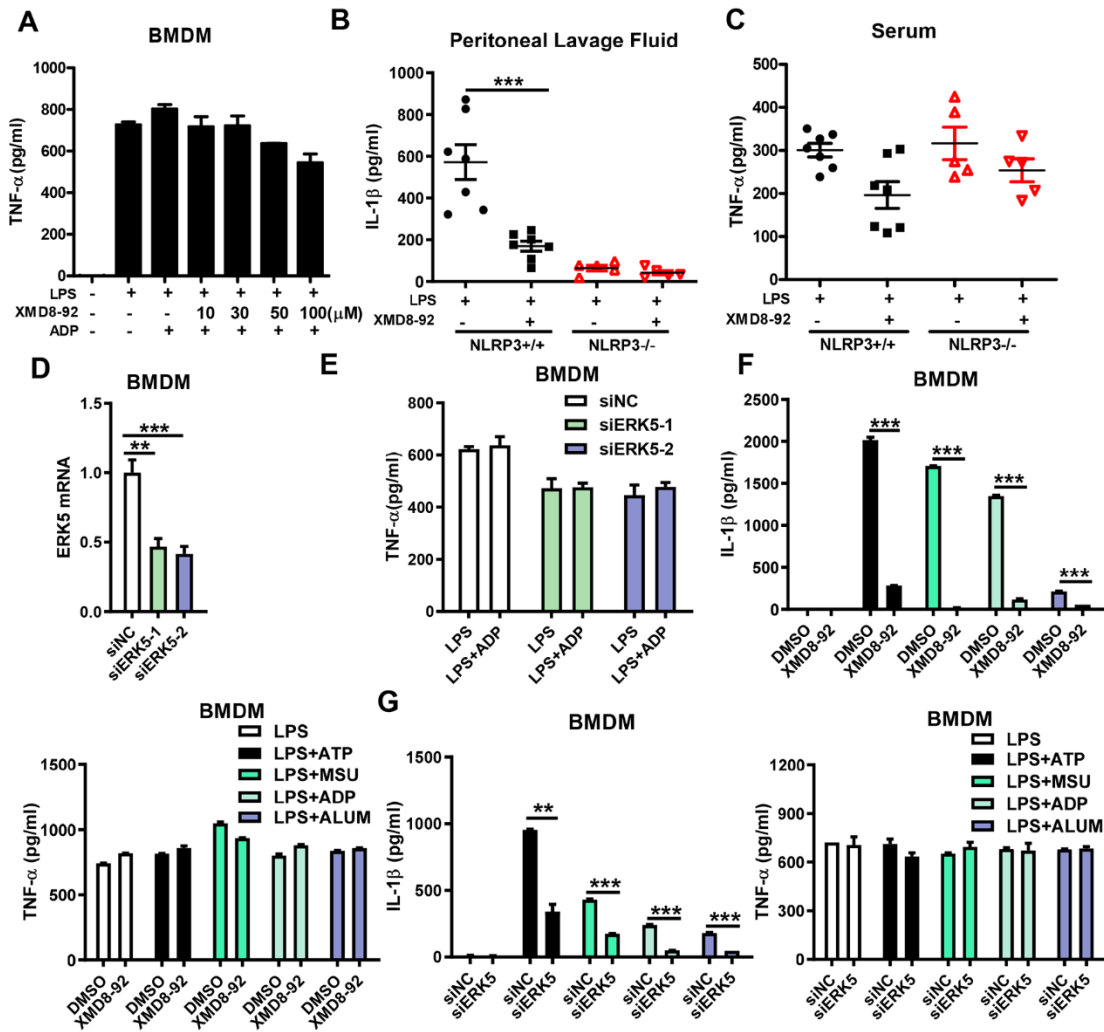
Supplementary Figure 3



Supplementary Figure 3. ERK5 pathway is involved in ADP activation of the NLRP3 inflammasome, related to Figure 6

(A) LPS-primed BMDM cells were stimulated with ADP (5 mM) for 1 h, then mRNA levels were analyzed by microarray. Shown is the heatmap of strongly upregulated genes (Left) and genes upregulated >20-fold change were listed (right). (B) LPS-primed BMDM cells were pretreated with 2-APB (100 μ M) or XMD8-92 (100 μ M) for 1 h and then stimulated with ADP (5 mM) for 1 h. The expression of Fosb, Nr4a1, Nr4a2, Nr4a3, Lif, Egr2, Areg, Dusp14, Zfp667, Apold1 and Ccno mRNA were analyzed by QPCR. (C) LPS-primed BMDM cells were pretreated with 50 μ M BAPTA-AM for 1 h and then stimulated with ADP (5 mM) for 1 h. The expression of Fosb, Nr4a1, Nr4a2, Nr4a3, Lif, Egr2, Dusp14, Zfp667 mRNA were analyzed by QPCR. Data are shown as mean $\pm$ SEM. * p <0.05; ** p <0.01; *** p <0.001; ns, not significant.

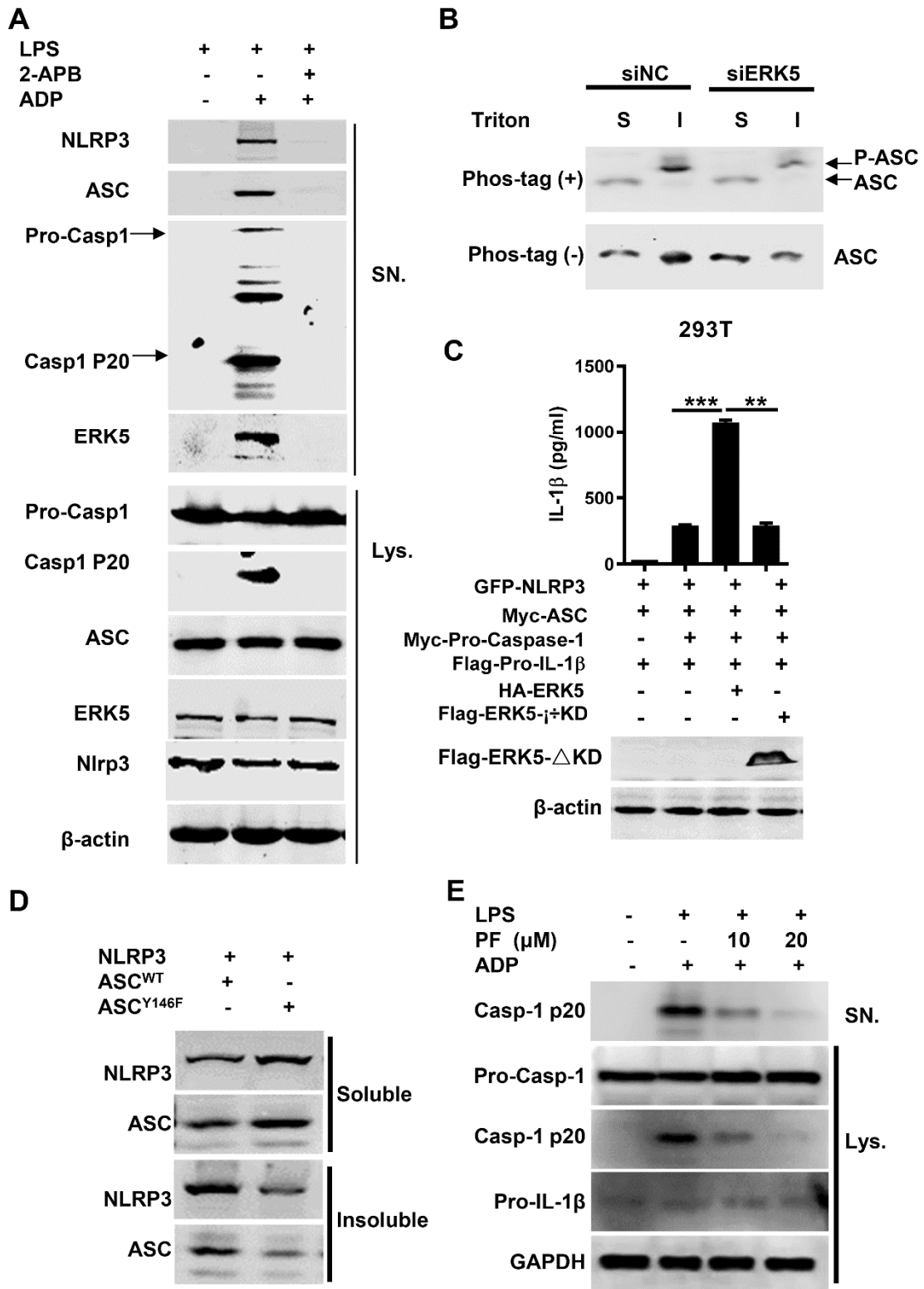
Supplementary Figure 4



Supplementary Figure 4. Blocking ERK5 attenuated NLRP3 inflammasome activity, related to Figure 6

(A) LPS-primed BMDM cells were pretreated with indicated concentrations of XMD8-92 for 1 h and then stimulated with ADP (5 mM) for 1 h. The production of TNF- α was detected by ELISA. (B-C) NLRP3^{+/+} and NLRP3^{-/-} mice were intraperitoneally treated with XMD8-92 (50 mg/kg) and LPS (20 mg/kg) for 6h, and levels of IL-1 β in peritoneal lavage fluid and TNF- α in serum were measured by ELISA. (D) BMDM cells were transfected with negative control siRNA or ERK5-specific siRNA for 30 h, and expression of ERK5 mRNA was analyzed by QPCR. (E) BMDM cells were transfected with negative control siRNA or ERK5-specific siRNA for 36 h and then primed with LPS (500 ng/ml) for 6h. The cells were then treated with 5 mM ADP for 1 h, and production of TNF- α was detected by ELISA. (F) LPS-primed BMDM cells were pretreated with XMD8-92 (100 μ M) for 1 h and then stimulated with indicated NLRP3 inflammasome inducers. The production of IL-1 β and TNF- α was measured by ELISA. (G) BMDM cells were transfected with negative control siRNA or ERK5-specific siRNA for 36 h and then primed with LPS (500 ng/ml) for 6h. The cells then stimulated with indicated NLRP3 inflammasome inducers, and production of IL-1 β and TNF- α was measured by ELISA. Data are shown as mean $\pm$ SEM. ** p <0.01; *** p <0.001.

Supplementary Figure 5



Supplementary Figure 5. ERK5 is released from cells along with the NLRP3 inflammasome, related to Figure 7

(A) LPS-primed BMDM cells were pretreated with 2-APB (100 μ M) for 1 h and then stimulated with ADP (5 mM) for 1 h. NLRP3, ASC, Pro-caspase-1 and ERK5 levels in cell culture supernatants (SN) or in cell lysates were analyzed by immunoblotting. (B) PEMs were transfected with negative control siRNA or ERK5-specific siRNA for 36 h and then primed with LPS (500 ng/ml) for 6h. The cells were then treated with 5 mM ADP for 1 h, and phosphorylated and total ASC in the Triton X-soluble and Triton X-insoluble fractions was analyzed by SDS-PAGE with (+) or without (–) Phos-tag. (C) The NLRP3 inflammasome was reconstituted in ERK5^{-/-} 293T cells by transfection of HA-ERK5 or Flag-ERK5- Δ KD, GFP-NLRP3, Myc-ASC, Myc-pro-caspase-1 and Flag-Pro-IL-1 β for 30 h. The production of IL-1 β was detected by ELISA; expression of Flag-ERK5- Δ KD was assessed by immunoblotting. (D) The NLRP3 inflammasome was reconstituted in 293T cells by transfection of GFP-NLRP3, Myc-ASC^{WT} and Myc-ASC^{Y146F} for 30 h, and ASC in Triton X-soluble and Triton X-insoluble fractions was analyzed by immunoblotting. (E) LPS-primed BMDM cells were pretreated with 10 or 20 μ M PF-431396 for 1 h, then stimulated with ADP (5 mM) for 80 min. Cleaved caspase-1 (p20) in culture supernatants (SN) or in cell lysates and pro-caspase-1, pro-IL-1 β in cell lysates were assessed by immunoblotting. Data are shown as mean $\pm$ SEM.

** p <0.01; *** p <0.001.

Supplementary table 1. Primers used for Q-PCR

Gene name	Primers (5'-3')
β -actin	Forward: GTACGCCAACACAGTGCTG
	Reverse: CGTCATACTCCTGCTTGCTG
Fosb	Forward: CTCCAGCTGTTGACCCTTAT
	Reverse: GGGTAAGTGTCTCTTCTCGG
Nr4a1	Forward: CCGGTGACGTGCAACAATTT
	Reverse: CGGGTTTAGATCGGTATGCCA
Nr4a3	Forward: TATGGCTCGGAATACACCACA
	Reverse: GCCCTCCATGAAGGTACTGAA
Lif	Forward: AACCAGATCAAGAATCAACTGGC
	Reverse: TGTTAGGCGCACATAGCTTTT
Egr2	Forward: GCAAATGATGACCGCCAAGG
	Reverse: GTTGATCATGCCATCTCCCG
Nr4a2	Forward: TCGGTTTACTACAAGCCCTCT
	Reverse: GGGGCGACTGCTTAAAGGA
Areg	Forward: GGGGACTACGACTACTCAGAG
	Reverse: TCTTGGGCTTAATCACCTGTTC
Dusp14	Forward: TTGCTCAGATCACCTCCTCTC
	Reverse: AGTACAGTCTAATGGGGGCAT
Zfp667	Forward: TCCAAGGCACCAAGTGACTTTT
	Reverse: GGCAAGACCAACTGAGACTAGA
Apold1	Forward: GAAAGCCACCCGAAGAGTCA
	Reverse: AGGTGGCTAAGTCACACGAC
Ccno	Forward: CTACGACTTCCGAAAGGCG
	Reverse: TCCAGAGTATTCACGGTCAGAC
ERK5	Forward: CCGGTGAGGCGGAA
	Reverse: CGTCAAAGGTCACGTCGAA