

Supplementary Information

Oncogenic zinc finger protein ZNF322A promotes stem cell-like properties in lung cancer through transcriptional suppression of c-Myc expression

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Inventory of supplementary information

Supplementary Data

Supplementary Figure 1 is related to Figure 1.

Supplementary Figure 2 is related to Figure 1.

Supplementary Figure 3 is related to Figure 2.

Supplementary Figure 4 is related to Figure 3.

Supplementary Figure 5 is related to Figure 4.

Supplementary Figure 6 is related to Figure 5.

Supplementary Figure 7 is related to Figure 5.

Supplementary Figure 8 is related to Figures 4 and 5.

Supplementary Figure 9 is related to Discussion.

Supplementary Table 1 is related to Materials and Methods.

Supplementary Table 2 is related to Materials and Methods.

Supplementary Table 3 is related to Materials and Methods.

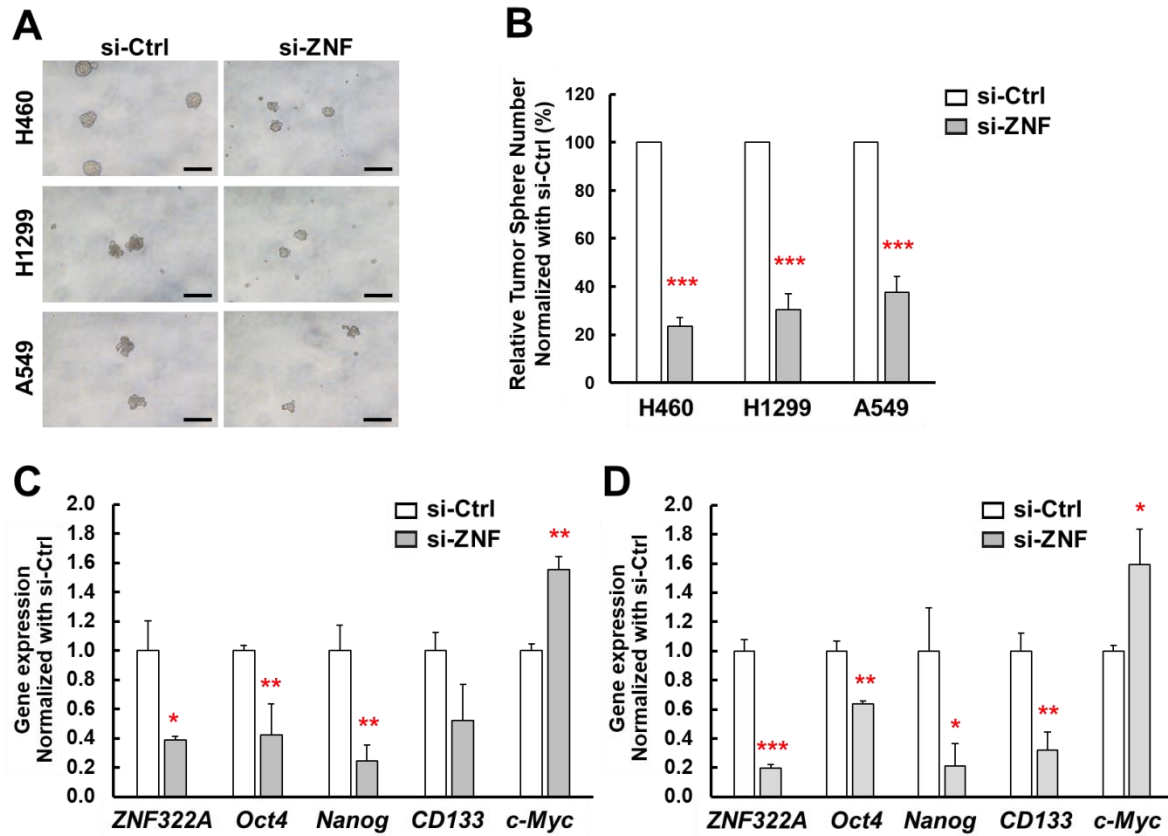
Supplementary Table 4 is related to Figure 1.

Supplementary Table 5 is related to Figure 1.

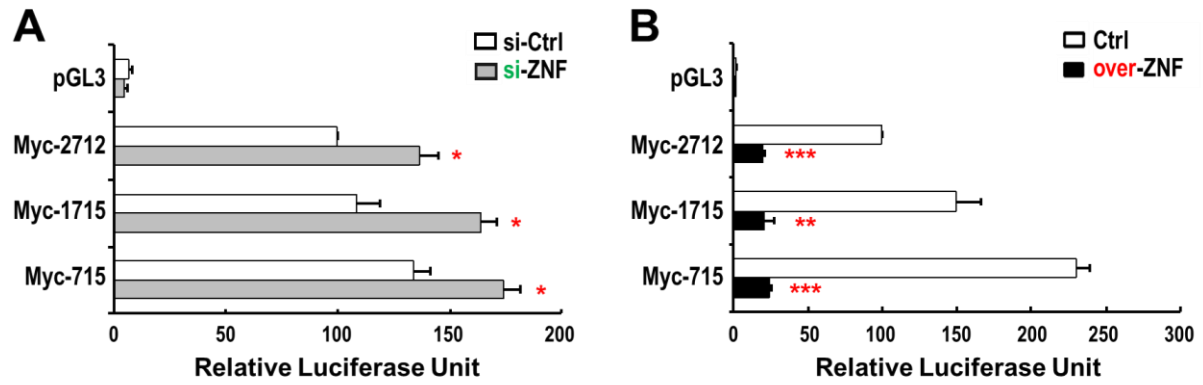
Supplementary Table 6 is related to Figure 1.



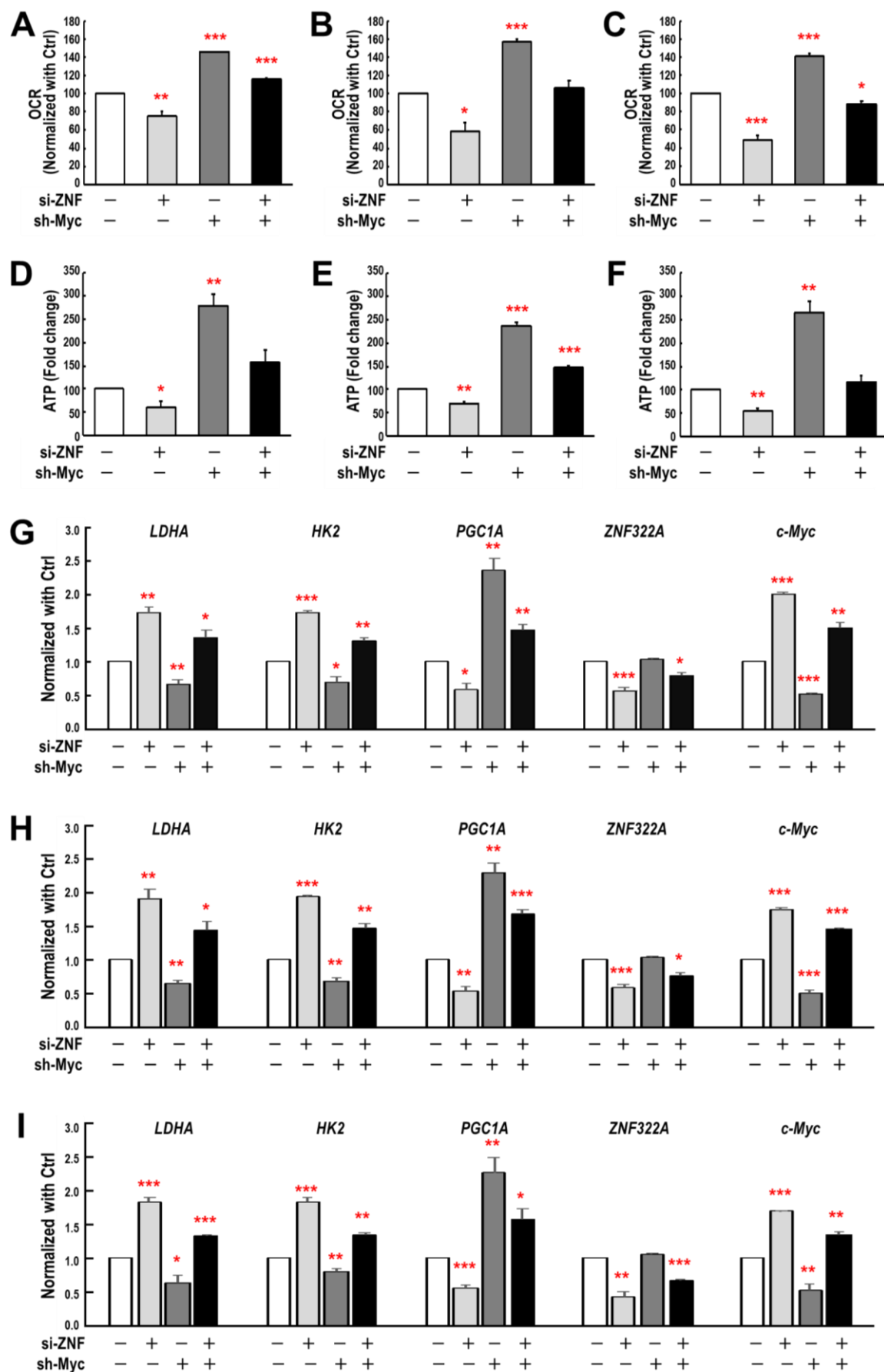
Supplementary Figure 2. Disease biomarkers and gene ontology analyses of ZNF322A-driven transcriptome. **A** Disease biomarkers regulated by ZNF322A. ZNF322A transcriptional target genes, including 128 ZNF322A positively-regulated genes and 118 ZNF322A negatively-regulated genes, were analyzed by MetaCore resource. Top seven disease pathways mapped for genes from ZNF322A positively transcriptional target genes and top five from ZNF322A negatively transcriptional target genes are shown. **B** Gene ontology analysis of ZNF322A target genes by DAVID software.



Supplementary Figure 3. Knockdown of endogenous ZNF322A suppresses self-renewal and stemness-related gene expression in lung cancer cells *in vitro*. **A, B** *In vitro* tumor sphere formation assay of H460, H1299 and A549 lung cancer cells expressing si-control (si-Ctrl) or si-ZNF322A (si-ZNF) oligos were photographed (**A**) and quantified (**B**). Scale bar, 200 nm. **C, D** qRT-PCR analysis of stemness-related genes and ZNF322A expression level in H460 (**C**) and H1299 (**D**) lung cancer cells expressing si-Ctrl or si-ZNF oligos. *GAPDH* was used as internal control. The error bars represent SEM from three independent experiments (* $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$).

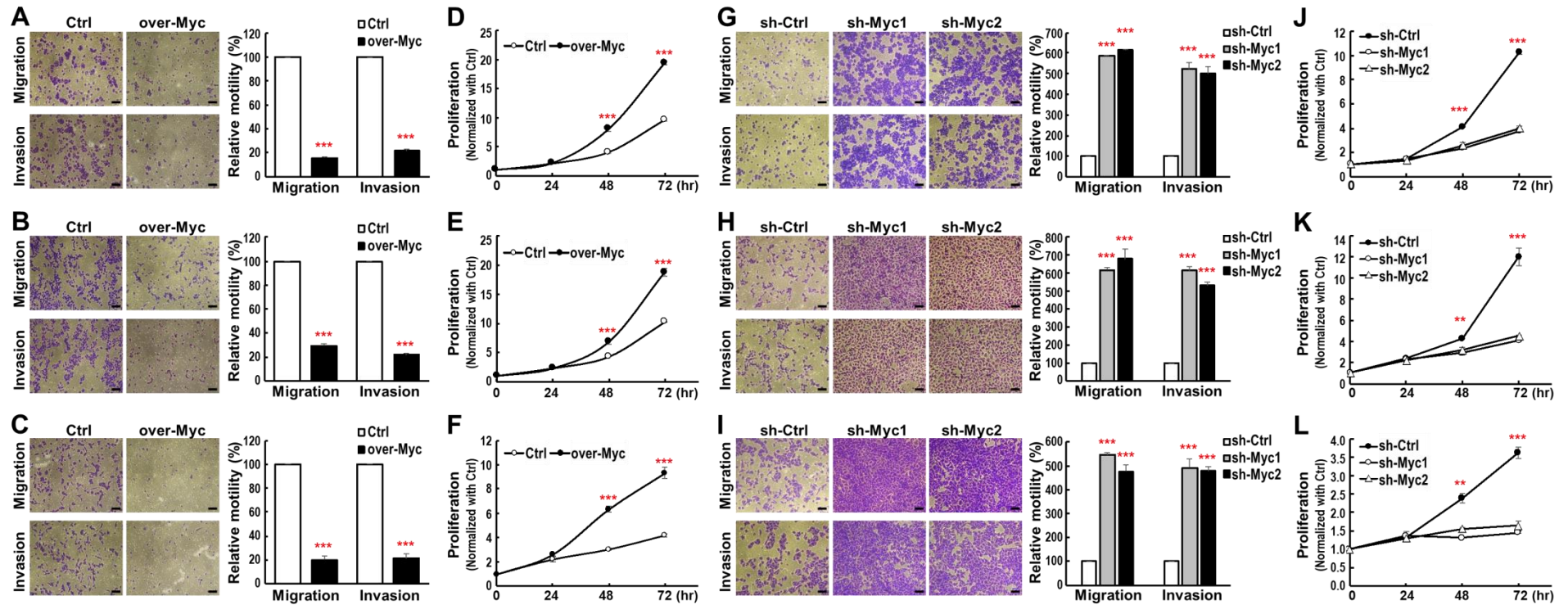


Supplementary Figure 4. ZNF322A negatively regulates *c-Myc* promoter activities. **A, B** Dual luciferase activity assays were performed using firefly luciferase reporter vectors containing -2157/+555, -1160/+555 or -160/+555 fragments of *c-Myc* promoters, which are indicated as Myc-2712, Myc-1715 and Myc-715, respectively. Data represent promoter activity in A549 cells expressing *ZNF322A* expression vector (over-ZNF) (**A**) or si-*ZNF322A* (si-ZNF) oligos (**B**). TSS: Transcription start site. Data are mean $\pm$ SEM (* P < 0.05; ** P < 0.01; *** P < 0.001).

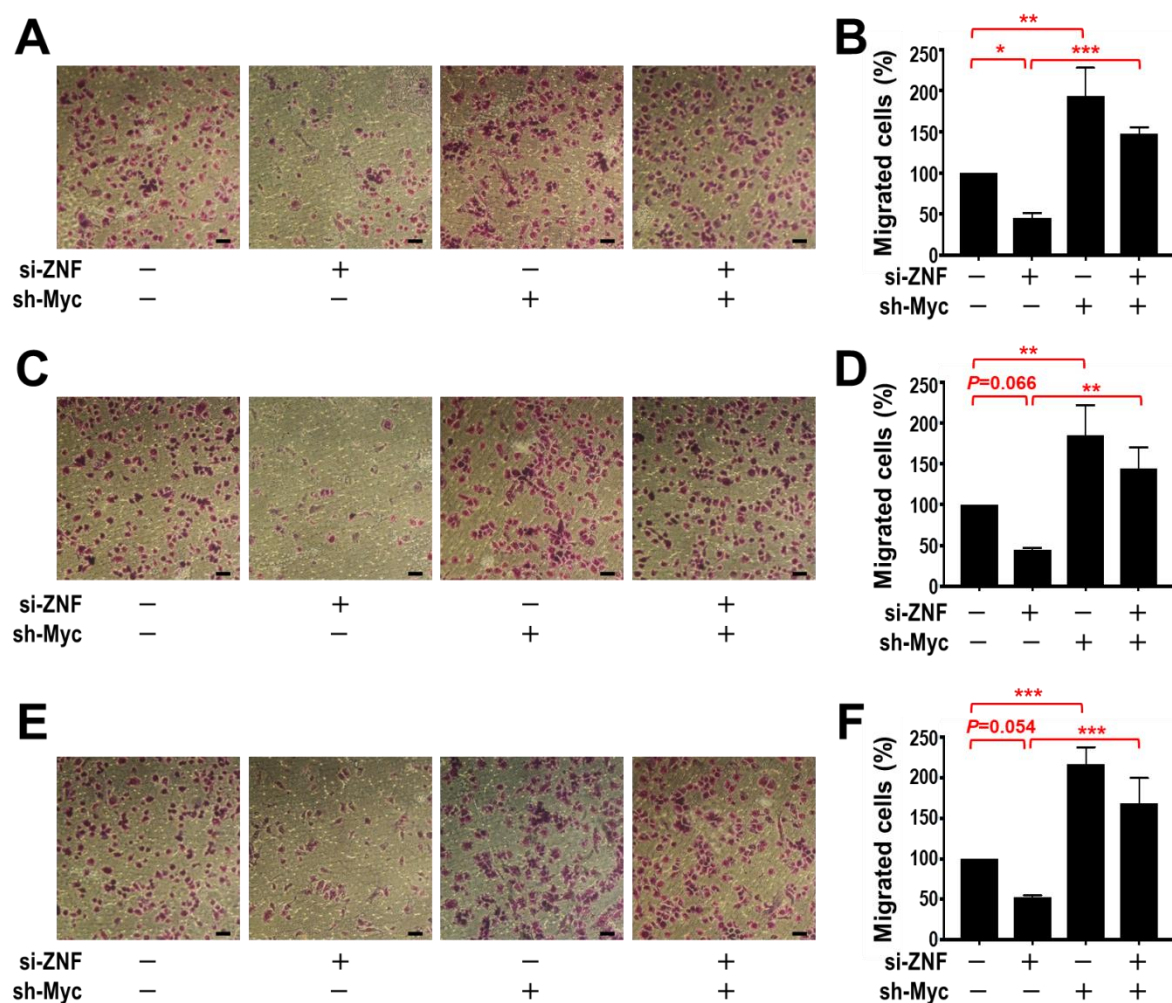


Supplementary Figure 5. Reconstitution experiments by c-Myc knockdown reversed

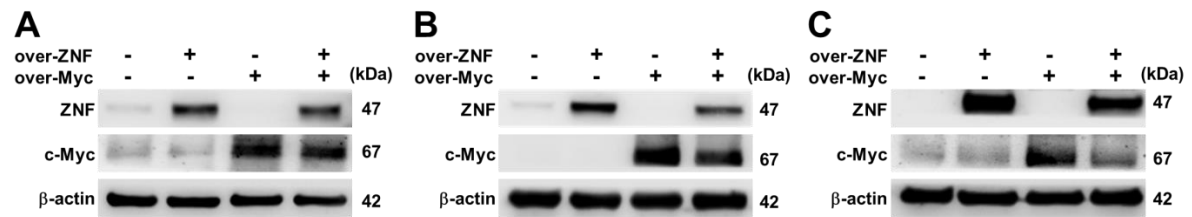
ZNF322A-mediated metabolic reprogramming in lung cancer cells. ZNF322A-mediated c-Myc suppression promotes metabolic reprogramming in lung cancer cells. **A-C** Oxygen consumption rate (OCR) was determined using XF Extracellular Flux Analyzer in single knockdown ZNF322A (si-ZNF), or c-Myc (sh-Myc) or in double knockdown reconstituted cells (si-ZNF/sh-Myc) of H460 (**A**), H1299 (**B**) and A549 (**C**) lung cancer cells. **D-F** ATP content of si-ZNF, sh-Myc, or si-ZNF/sh-Myc H460 (**D**), H1299 (**E**) and A549 (**F**) lung cancer cells was determined. **G-I** qRT-PCR analysis of metabolism-related genes in si-ZNF, sh-Myc, or si-ZNF/sh-Myc of H460 (**G**), H1299 (**H**) and A549 (**I**) lung cancer cells was examined. The error bars represent SEM from three independent experiments (* $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$).



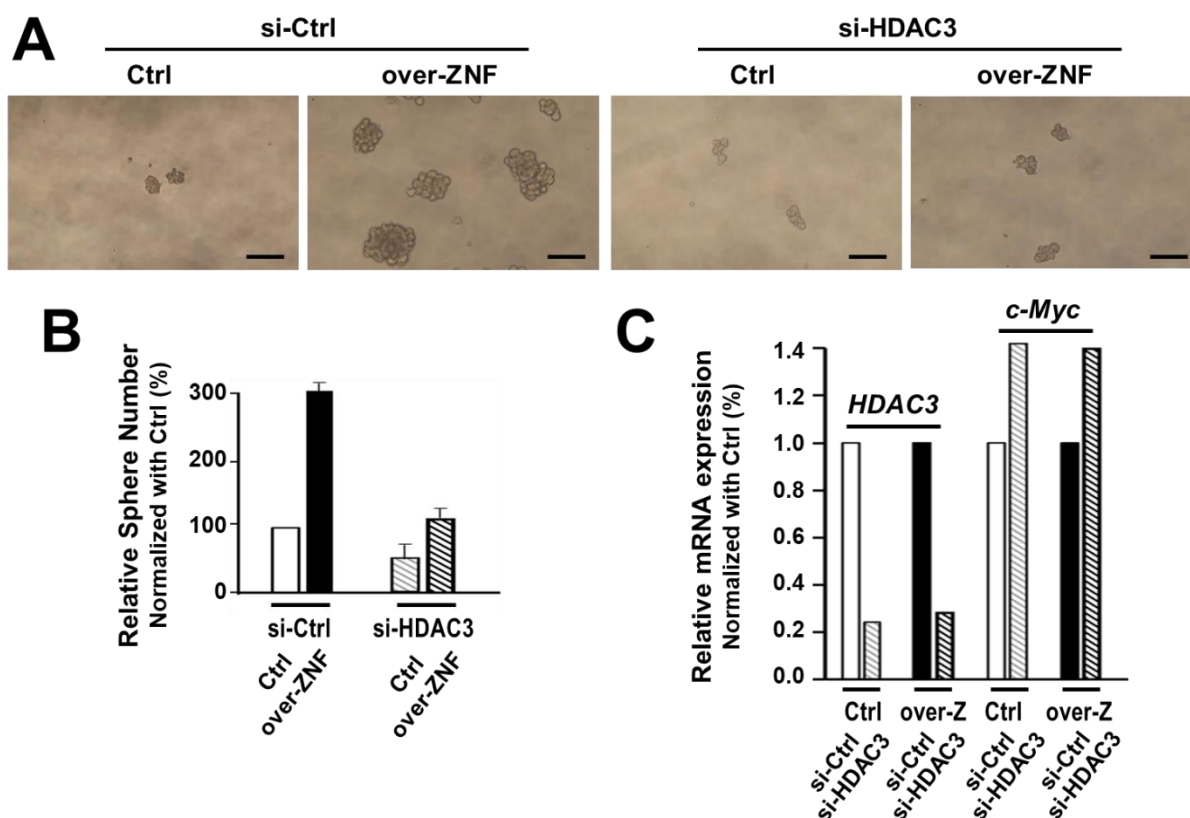
Supplementary Figure 6. c-Myc acts as a metastasis suppressor in lung cancer cells. A-C c-Myc overexpression (over-Myc) suppressed migration and invasion abilities in H460 (A), H1299 (B) and A549 (C) lung cancer cells. Represented figures (*left*) and quantified results (*right*) are shown. Scale bar, 200 nm. D-F c-Myc overexpression (over-Myc) promoted cell proliferation in H460 (D), H1299 (E) and A549 (F) lung cancer cells. G-I c-Myc silenced with two different shRNA clones (sh-Myc1 and sh-Myc2) increased migration and invasion abilities in H460 (G), H1299 (H) and A549 (I) lung cancer cells. Represented figures (*left*) and quantified results (*right*) are shown. Scale bar, 200 nm. J-L c-Myc silenced with two different shRNA clones (sh-Myc1 and sh-Myc2) suppressed cell proliferation in H460 (J), H1299 (K) and A549 (L) lung cancer cells. Data are mean $\pm$ SEM (** $P < 0.01$; *** $P < 0.001$).



Supplementary Figure 7. Reconstitution experiments by c-Myc knockdown reversed ZNF322A-mediated migration in lung cancer cells. A-C c-Myc knockdown increased cell migration of ZNF322A depleted lung cancer cells (si-ZNF/sh-Myc) in H460 (A), H1299 (B) and A549 (C). Represented figures (*left*) and quantified results (*right*) are shown. Scale bar, 200 nm. Data are mean $\pm$ SEM (* P < 0.05; ** P < 0.01; *** P < 0.001 as determined by One-Way ANOVA).



Supplementary Figure 8. Protein expression level of the manipulated ZNF322A and c-Myc in various lung cancer cells. A-C *ZNF322A* expression vector (over-ZNF) and *c-Myc* expression vector (over-Myc) were transfected into H460 (A), H1299 (B) and A549 (C) cells. Cell lysates were subjected to immunoblotting. β -actin serves as an internal control.



Supplementary Figure 9. Knockdown of HDAC3 reduces sphere formation and leads to re-expression of *c-Myc* mRNA. **A, B** H1299 cells overexpressing ZNF322A (over-ZNF) or empty vector (Ctrl) were transfected with control si-oligo (si-Ctrl) or mixed siRNA oligos against HDAC3 (recognize sequence 5-AGAGGCCUCAACUUCUUGGCAUGG-3, 5-AACUCUGGUCAUCA AUGCCAUCCCG-3, 5-UUAAUUCUCCACAUCGCUUCCUUG-3; # 1299001, Invitrogen, Carlsbad, CA, USA) then subjected for tumor sphere formation assay. Spheres were photographed (**A**) and quantified (**B**). Scale bar, 200 nm. **C** qRT-PCR analyses of *HDAC3* and *c-Myc* mRNA expression levels were measured in H1299 cells expressing ZNF322A (over-Z) and/or si-*HDAC3* (si-HDAC3) oligos.

Supplementary Table S1. Antibodies and their reaction conditions used in the current study

Target	KD	Raised In	Application	Dilution	Source	Catalog No.
ZNF322A	44	Rabbit	Immunoblotting ChIP	1:1000 1:50	LifeSpan	LS-C30280
c-Myc	67	Mouse	Immunoblotting	1:1000	Santa Cruz	sc-42
β -actin	42	Mouse	Immunoblotting	1:5000	Novus Biologicals	NB 600-501
HDAC3	49	Rabbit	ChIP	1:50	Cell signaling	sc-11417
Ac-H3	16	Rabbit	ChIP	1:50	Millipore	06-599
HA-tag (ZNF322A)	— ^a	Rabbit	Immunoblotting ChIP	1:1000 1:50	GeneTex	GTX29110

^a —, Molecular weight is not applicable.

Supplementary Table S2. The primers used in the current study

Gene	Primer	Sequences (5'→3')	Application ^a	PCR size (bp)	Tm (°C)
<i>GAPDH</i> mRNA	Forward	GAG TCA ACG GAT TTG GTC GT	qRT-PCR	238	60
	Reverse	TTG ATT TTG GAG GGA TCT CG			
<i>ZNF322A</i> mRNA	Forward	GTG GTC TGC GTG TGA GAG TGG C	qRT-PCR	226	60
	Reverse	TTC TGA CGC ATG GGG AGG GCT			
<i>ADD1</i> mRNA	Forward	TCC CAG GTT TTG TGT GGT GTA GT	qRT-PCR	150	60
	Reverse	ACG GTC TGT GAG TGT GGT GAA G			
<i>CCND1</i> mRNA	Forward	AAC TAC CTG GAC CGC TTC CT	qRT-PCR	204	60
	Reverse	CCA CTT GAG CTT GTT CAC CA			
<i>p53</i> mRNA	Forward	GTG GAA GGA AAT TTG CGT GT	qRT-PCR	184	60
	Reverse	CCA GTG TGA TGA TGG TGA GG			
<i>DNAJC6</i> mRNA	Forward	CAC CAG AGC AGG TGA AGA AGG T	qRT-PCR	120	60
	Reverse	AGG CAT CAT TGA GCT CCA TGA			
<i>GFRA3</i> mRNA	Forward	TTG CAG CTA AGA TGC GTT TTC A	qRT-PCR	103	60
	Reverse	GGC CTC ACA GCA GGG TTT T			
<i>EDEM1</i> mRNA	Forward	CAA GAA TCC CTT CTA CCT CCA TGT A	qRT-PCR	131	60
	Reverse	GCT CTC CAT CCG GTC TTC TG			
<i>KIF6</i> mRNA	Forward	GCA GAT TCG ATG TCT GTG ATG TG	qRT-PCR	133	60
	Reverse	ATG GAC GAC ACT GGC CTC TT			
<i>IRX2</i> mRNA	Forward	GCC GGC GCC TCA AGA	qRT-PCR	170	60
	Reverse	GTC CAC GTG CAG GCT GAT C			
<i>PCDH10</i> mRNA	Forward	GCC ATC GTC ACC GGT TAC AC	qRT-PCR	182	60
	Reverse	TCT CCA TGA CCA CTG TCC TTA GAG			
<i>SLC44A1</i> mRNA	Forward	GAT AGT CTG CAG CAC AGG TTT AGC T	qRT-PCR	200	60
	Reverse	CAG GGC TCC CAT CAT TGT ATT T			

Gene	Primer	Sequences (5'→3')	Application ^a	PCR size (bp)	T _m (°C)
<i>SEC23A</i> mRNA	Forward	ACA GTG GCG GAA GTC AGG AT	qRT-PCR	181	60
	Reverse	TGA AGG GTT GAC TTT TGA AAG GA			
<i>HBP1</i> mRNA	Forward	AAA TCC TGA CTG TTG GAA GAG GAA	qRT-PCR	161	60
	Reverse	GGT CAC ACG AAT TGA GGA CAA A			
<i>CCR7</i> mRNA	Forward	TCC TTG TCA TTT TCC AGG TAT GC	qRT-PCR	101	60
	Reverse	GCA CAA AGA CTC GAA CAA AGT GTA G			
<i>STXBP1</i> mRNA	Forward	GCA TTA CCA AGG CAC CGT AGA	qRT-PCR	130	60
	Reverse	GAC ATT GGC ATC CAG CAG AA			
<i>ANKDD1A</i> mRNA	Forward	TGG GAA GAG CTT GTC CTT TAA GC	qRT-PCR	177	60
	Reverse	CTG GTG CCT GTC CAC TGT TG			
<i>ARHGAP23</i> mRNA	Forward	CCC CTG TGG GCG ACAA	qRT-PCR	124	60
	Reverse	GAC TTG GCT GAA CTA CAG GTG GTA			
<i>DAGLB</i> mRNA	Forward	TGC AGG AAT ATT CTC AGA GCT TCA	qRT-PCR	175	60
	Reverse	CCA AAC AGT TCG TAC CAC AAA CC			
<i>DLG5</i> mRNA	Forward	ACA TTG CTC CGC ACG CTA TT	qRT-PCR	150	60
	Reverse	GCT CTT TGG AAT GCC TCT GAG T			
<i>MYLK2</i> mRNA	Forward	GAC GGG ATC CTC TTC ATG CA	qRT-PCR	120	60
	Reverse	CCG TGC CAG GCC AAA GT			
<i>PARD6G</i> mRNA	Forward	CCC ACC ATA TCT CCA ACA GTG A	qRT-PCR	150	60
	Reverse	CGG CCT CCT CTC GTT TCT G			
<i>MKNK1</i> mRNA	Forward	CAA CTC CTG TAC CCC CAT AAC C	qRT-PCR	120	60
	Reverse	CAG CGC TTG TCG TAG AAT GTG			
<i>PAK4</i> mRNA	Forward	GCG CCC AGG TGA GCAA	qRT-PCR	151	60
	Reverse	TCT CCG TCC ACC ATC TCA ATC			
<i>E2F2</i> mRNA	Forward	GAA GGC CAA GAA CAA CAT CCA	qRT-PCR	117	60
	Reverse	GCC TGC TCC GTG TTC ATC A			

Gene	Primer	Sequences (5'→3')	Application ^a	PCR size (bp)	T _m (°C)
<i>KPNA6</i> mRNA	Forward	CTG GGC CAT CAC CAA TGC	qRT-PCR	150	60
	Reverse	AGG ATG TTC TCC AGT CCA TTG AG			
<i>NMT1</i> mRNA	Forward	TAT ACG CTG CCC TCC ACC AT	qRT-PCR	170	60
	Reverse	TCC ATG AGA TCC AGT GCA TTG			
<i>ZNF85</i> mRNA	Forward	CTA GGT CTT GTT TTC CCT GCT TTG	qRT-PCR	150	60
	Reverse	CCA CAT CCC TAA ATG TCA ATG GT			
<i>HMGN5</i> mRNA	Forward	GTC GTG CTG CAG TTA GTT CAT TG	qRT-PCR	200	60
	Reverse	ACT GGC ACA AGC ATA GCA GAC A			
<i>SFXN4</i> mRNA	Forward	TTT CCA CAG ATT GGA CAG ATA CAG TAC T	qRT-PCR	187	60
	Reverse	CCA GAA GAC CTG TGG CAA AGT T			
<i>GATA6</i> mRNA	Forward	GAA TTC AAA CCA GGA AAC GAA AAC	qRT-PCR	135	60
	Reverse	GGG AAG TAT TTT TGC TGC AAT CAT			
<i>NEFL</i> mRNA	Forward	CCA AGA CCC TGG AAA TCG AA	qRT-PCR	151	60
	Reverse	TCA CTC TTT GTG GTC CTC AAT TCA			
<i>CCAR1</i> mRNA	Forward	AAG TCA TTG TGG TTA CCT TCT TGA AA	qRT-PCR	200	60
	Reverse	CTA GCA TAT CTT CCT GCA ATG ACT CA			
<i>NR1H4</i> mRNA	Forward	TGC TCT GCT TAC AGC AAT TGT TAT C	qRT-PCR	200	60
	Reverse	CAG CGT GGT GAT GAT TGA ATG			
<i>ELK4</i> mRNA	Forward	AGG CGC ATC GTG TTC GA	qRT-PCR	177	60
	Reverse	CTG CCT GCA AAA GCT TAA ACT G			
<i>ZNF238</i> mRNA	Forward	TGC ATC TGT CTC TCT TAG TCT GCT TT	qRT-PCR	150	60
	Reverse	GTA GCA AAT GTC TAC TAT GGT CTG GAA			
<i>MXII</i> mRNA	Forward	GCC AAA GCA CAC ATC AAG AAA C	qRT-PCR	101	60
	Reverse	TCC AGT CGC CAC TTT AAA AAT CTC			
<i>SGK196</i> mRNA	Forward	GCG AGG AGC TGA GAA CAG AAG	qRT-PCR	200	60
	Reverse	ATA GCC AAG CAG CGT GAC AA			

Gene	Primer	Sequences (5'→3')	Application ^a	PCR size (bp)	T _m (°C)
<i>HNRNPF</i> mRNA	Forward	TCC AGA AGT GTC TCC CAC TGA A	qRT-PCR	107	60
	Reverse	CCC AGC ATC ATG GAC ACT TG			
<i>GNB3</i> mRNA	Forward	GGG ACC TGC CGT CAG ACT T	qRT-PCR	175	60
	Reverse	TGA TGC CGC AGA TGA TGC T			
<i>CTNNA3</i> mRNA	Forward	GCA AAG GAC CAC TAA AGC ATA CAA	qRT-PCR	152	60
	Reverse	CTG TTC CAG GTA GGC CAA CAA			
<i>RIT1</i> mRNA	Forward	CAT GAT CCC ACC ATT GAA GAT G	qRT-PCR	175	60
	Reverse	AAC TTC GAC GAT CCG TGA TAG AG			
<i>PFN2</i> mRNA	Forward	AGG AGG CCG CCA TTG TC	qRT-PCR	120	60
	Reverse	CTT CCC GGT CTT TTC CTA CAA TC			
<i>CSNK2A2</i> mRNA	Forward	AGG CCA TGG AGC ACC CAT A	qRT-PCR	120	60
	Reverse	ACC CGT CGC TTT CCA GTC TT			
<i>CAPN2</i> mRNA	Forward	TTG ATG ATG GAT TCA GGA GAC TGT	qRT-PCR	175	60
	Reverse	CCG TCC GAA TCT AGC ATG TCA			
<i>Oct4</i> mRNA	Forward	CGA AAG AGA AAG CGA ACC AG	qRT-PCR	157	60
	Reverse	GCC GGT TAC AGA ACC ACA CT			
<i>Nanog</i> mRNA	Forward	CTG TGA TTT GTG GGC CTG AA	qRT-PCR	190	60
	Reverse	TCT TCC TTT TTT GCG ACA CTC TT			
<i>Sox2</i> mRNA	Forward	ACA ACT CGG AGA TCA GCA	qRT-PCR	183	60
	Reverse	GCA GCG TGT ACT TAT CCT TC			
<i>CD133</i> mRNA	Forward	ACA GTT TGC CCC CAG GAA AT	qRT-PCR	150	60
	Reverse	ATC CAT TCC CTG TGC GTT GA			
<i>ABCB1</i> mRNA	Forward	GGT GCT GCT TTC CTG CTG AT	qRT-PCR	140	60
	Reverse	CCA ACA CTA AAA GCC CCA ATT AA			
<i>ABCG2</i> mRNA	Forward	CCA TTG CAT CTT GGC TGT CA	qRT-PCR	180	60
	Reverse	CGA TGC CCT GCT TTA CCA AA			

Gene	Primer	Sequences (5'→3')	Application ^a	PCR size (bp)	T _m (°C)
<i>c-Myc</i> mRNA	Forward	CTC TCC GTC CTC GGA TTC TCT	qRT-PCR	157	60
	Reverse	TTC CAC AGA AAC AAC ATC GAT TTC			
<i>HDAC3</i> mRNA	Forward	AGC ACC CGC ATC GAG AAT C	qRT-PCR	141	60
	Reverse	GGA CAC TAG GTG CAT GGT TCA G			
<i>GLUT1</i> mRNA	Forward	CGG GTT GTG CCA TAC TCA TG	qRT-PCR	203	60
	Reverse	CCA GTT GGA GAA GCC TGC AA			
<i>LDHA</i> mRNA	Forward	GAG AGT GCT TAT GAG GTG ATC AAA CT	qRT-PCR	181	60
	Reverse	CCA AAA TGC AAG GAA CAC TAA GG			
<i>ENO1</i> mRNA	Forward	TGA ACG AGA AGT CCT GCAACT G	qRT-PCR	132	60
	Reverse	TCT CCC CCG AAC GAT GAG			
<i>HK2</i> mRNA	Forward	TCC GTAACA TTC TCA TCG ATT TCA	qRT-PCR	178	60
	Reverse	CAG GTG CTC TCA AGC CCT AAG T			
<i>PFKM</i> mRNA	Forward	ACC AGA CAG ATT TTG AGC ATC GA	qRT-PCR	189	60
	Reverse	TAA TCT ATT CCC CTC ACT CCA GAG A			
<i>PGC1A</i> mRNA	Forward	TTG AAG AGC GCC GTG TGA T	qRT-PCR	160	60
	Reverse	CAG GTA TAA CGG TAG GTA ATG AAA CCA			
<i>ADD1</i>	Forward	GCT GCA GTG AGC CAT GGT T	ChIP-qPCR	180	60
	Reverse	TAT ATA CCC CAA AGA AAT GAA AAC AGC TA			
<i>CCND1</i>	Forward	GCC CAT TCT GCC GGC TTG GA	ChIP-qPCR	132	60
	Reverse	GGG GTG AGG TGG AGG TGG CT			
<i>p53</i>	Forward	GCA CCA GGT CGG CGA GAA TCC	ChIP-qPCR	131	60
	Reverse	TGC GAG GCT CCT GGC ACA AA			
<i>IRX2</i>	Forward	GCT TCT GGC AAA TTA AGA TCT CTC A	ChIP-qPCR	198	60
	Reverse	TCT GGT GGT GTG CAA CTG TAG TC			
<i>PCDH10</i>	Forward	GGA GGT TGC AGT AAG CCA AGA	ChIP-qPCR	186	60
	Reverse	GTG TTT TTG AAT TTT CCT TGA GAT GTC			

Gene	Primer	Sequences (5'→3')	Application ^a	PCR size (bp)	T _m (°C)
<i>SLC44A1</i>	Forward	TGA GAT GGA TTT TCA CTC GTG TTG	ChIP-qPCR	188	60
	Reverse	AGC ATG GAG AAA CCC TGT CTC TA			
<i>SEC23A</i>	Forward	GTG GAG ATC CAG TCT TTG TGT GAA	ChIP-qPCR	201	60
	Reverse	CAT GGG AGA TCA CAA ATA CAA GAA TAG			
<i>ANKDD1A</i>	Forward	AGT TCC AGG AGG ACG CTA AGC	ChIP-qPCR	200	60
	Reverse	TGG GCA AAA TGG TGA GAC CTA			
<i>DAGLB</i>	Forward	ATG TCC ACG TCA AGG GTT TTT T	ChIP-qPCR	195	60
	Reverse	CAC CCC TGG CTC ATT TTT GT			
<i>DLG5</i>	Forward	TCA TCC TGG AGG TGA GTT CTG A	ChIP-qPCR	106	60
	Reverse	GCT GTA CTC CTC CGT CTT TGG T			
<i>HMG5</i>	Forward	CCC GCC TCC GCC TTT	ChIP-qPCR	127	60
	Reverse	GGT GCTA TTC GAG ATT CCC TAT TAA A			
<i>SFXN4</i>	Forward	CCT GGG AAG GTA GGT CAA AGC	ChIP-qPCR	206	60
	Reverse	CTT CCT GGC CTC ACG CAA T			
<i>GATA6</i>	Forward	CTT TTT TTG TTG AGA GGG AGT CTT G	ChIP-qPCR	185	60
	Reverse	ACA CGG TGA AAC TCG GTC TGT			
<i>CCAR1</i>	Forward	TGT GCT ATG ATT GCA CCT GTG A	ChIP-qPCR	177	60
	Reverse	GAC TGA CAG ACA TTC TCT GCT TGA C			
<i>DIO2</i>	Forward	GAC AAG GCA GGT GGA TCA TTG	ChIP-qPCR	196	60
	Reverse	GGC GCC ATC TCG CCT TA			
<i>ELK4</i>	Forward	GGA TCA TAG GAG TGA GCC AAT GT	ChIP-qPCR	127	60
	Reverse	TGG GAC GCT CGC TTG AG			
<i>SGK196</i>	Forward	GGA GTG CAA AGA AAA GAA GAA AAT AGA	ChIP-qPCR	125	60
	Reverse	ACA CAC AGA GAC TCG GGA GAA AC			
<i>HNRNPF</i>	Forward	CGA CGG AAA TGA GAC CTT GTC	ChIP-qPCR	125	60
	Reverse	AAC TCC CCA CAA GGA AGT CTT G			

Gene	Primer	Sequences (5'→3')	Application ^a	PCR size (bp)	Tm (°C)
<i>MYC</i>	Forward	ACC CTT GCC GCA TCC A	ChIP-qPCR	186	60
	Reverse	CGT CTA AGC AGC TGC AAG GA			
<i>ACTB</i>	Forward	CAC CAC CAT GTA CCC TGG CAT	ChIP-qPCR	170	60
	Reverse	CAG TGA GGA CCC TGG ATG TGA C			
<i>Myc-2712</i> promoter	Forward	GCG CAG ATC TAC TTT TTC CTC CAG TAA CTC CTC TT	Construction	-- ^b	65
	Reverse	GCG CAA GCT TCT GGT TTT CCA CTA CCC G			
<i>Myc-1715</i> promoter	Forward	GCG CAG ATC TGA GTT AAC GGT TTT TTT CAC AAG GG	Construction	-- ^b	65
	Reverse	GCG CAA GCT TCT GGT TTT CCA CTA CCC G			
<i>Myc-715</i> promoter	Forward	GCG CAG ATC TCG GCT GAG TCT CCT CCC C	Construction	-- ^b	70
	Reverse	GCG CAA GCT TCT GGT TTT CCA CTA CCC G			
<i>Myc-1-mut</i>	Forward	GAA AAA AGA TCC TAG AGC GCT AAT CTC CGC	Site-direct mutagenesis	-- ^b	71
	Reverse	GCG GAG ATT AGC GCT CTA GGA TCT TTT TTC			
<i>Myc-2-mut</i>	Forward	CTC TCT CGC TAA TAG AAG CCC ACC GGC CCT	Site-direct mutagenesis	-- ^b	71
	Reverse	AGG GCC GGT GGG CTT CTA TTA GCG AGA GAG			
<i>Myc+1-mut</i>	Forward	CCA GGA CCC GCT TAG AGG AAA GGC TCT CCT	Site-direct mutagenesis	-- ^b	71
	Reverse	AGG AGA GCC TTT CCT CTA AGC GGG TCC TGG			
<i>Myc+2-mut</i>	Forward	TCT CTG AAA GGC TAG AAT TGC AGC TGC TTA	Site-direct mutagenesis	-- ^b	65
	Reverse	TAA GCA GCT GCA ATT CTA GCC TTT CAG AGA			

^a qRT-PCR, quantitative reverse-transcriptase polymer chain reaction; ChIP-qPCR, quantitative chromatin-immunoprecipitation coupled with polymer chain reaction.

^b --, Not applicable.

Supplementary Table S3. The plasmids and their characteristics used in the current study.

Plasmid	Target	Insert (bp)	Function	Source
pCMV-HA	None	— ^a	Vector control	Homemade
HA-ZNF322A	Wild type ZNF322A	1083	Overexpression	Homemade
pGL3-vector	None	— ^a	Vector control	Promega
pGL3-Myc-2712	<i>c-Myc</i> promoter from -2712 ~ -1	2712	Promoter activity assay	Homemade
pGL3-Myc-1715	<i>c-Myc</i> promoter from -1715 ~ -1	1715	Promoter activity assay	Homemade
pGL3-Myc-715	<i>c-Myc</i> promoter from -715 ~ -1	715	Promoter activity assay	Homemade
Myc-1-mut	Myc-715 promoter with ZNF322A binding element 2 mutation	715	Promoter activity assay	Homemade
Myc-2-mut	Myc-715 promoter with ZNF322A binding element 2 mutation	715	Promoter activity assay	Homemade
Myc+1-mut	Myc-715 promoter with ZNF322A binding element 2 mutation	715	Promoter activity assay	Homemade
Myc+2-mut	Myc-715 promoter with ZNF322A binding element 2 mutation	715	Promoter activity assay	Homemade

^a —, Molecular weight is not applicable.

Supplementary Table S4. List for ZNF322A direct targeting genes is included in the excel file. The accession number for ChIP-seq dataset is GSE94656.

Supplementary Table S5. Gene list for 128 ZNF322A positively-regulated downstream targets is included in the excel file. The accession number for RNA-seq dataset is GSE94537.

Supplementary Table S6. Gene list for 118 ZNF322A negatively-regulated downstream targets is included in the excel file. The accession number for RNA-seq dataset is GSE94537.