

SUPPLEMENTARY INFORMATION

Single Particle Automated Raman Trapping Analysis

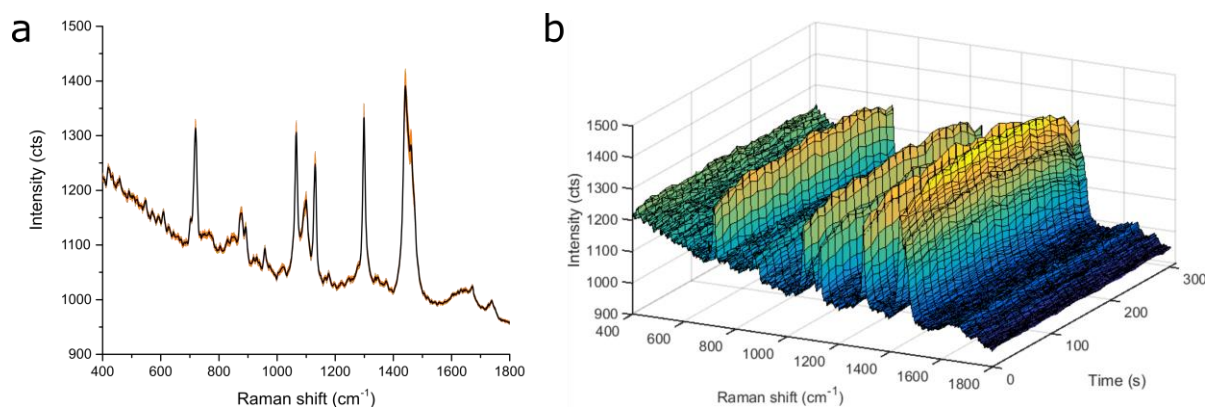
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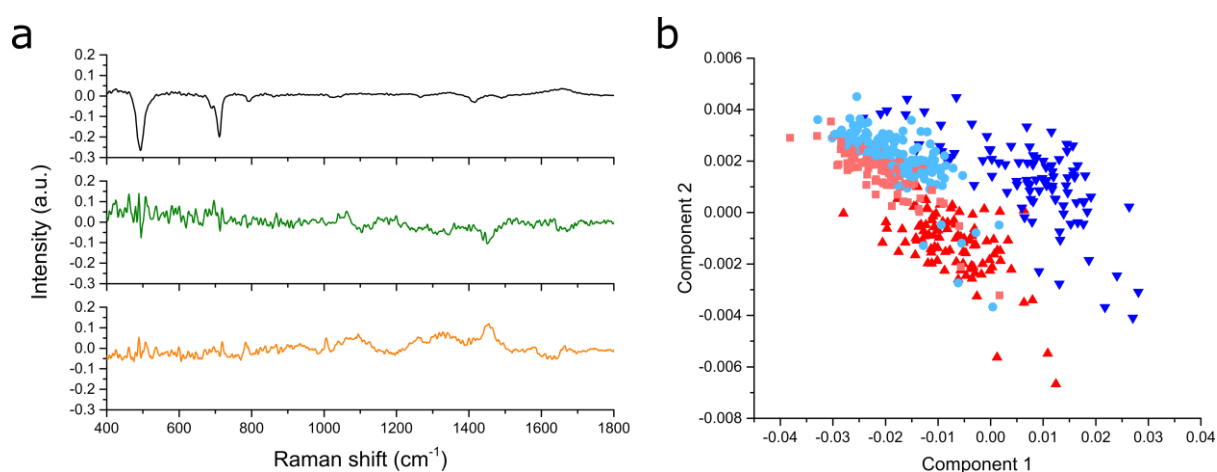
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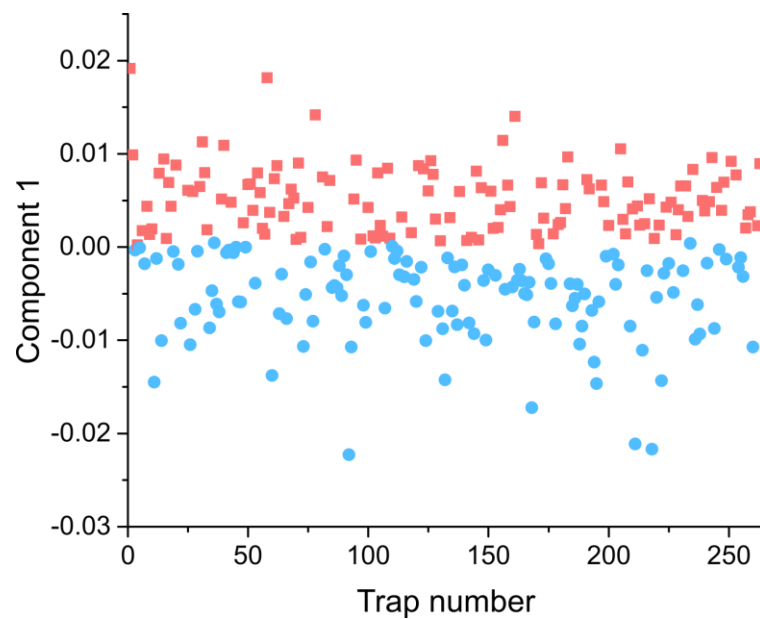
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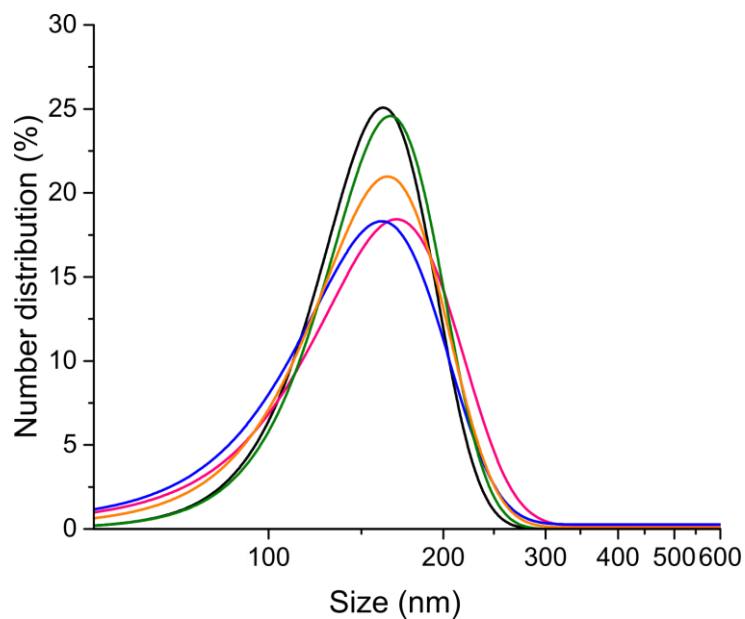
Supplementary Figure 1 | DPPC liposome photostability over time. DPPC liposome held in the optical trap for 5 minutes. (a) Averaged Raman spectra (mean $\pm$ s.d., $n = 26$), (b) surface plot of Raman spectra over time, showing minimal variation over time in the spectrum indicative of photostability.



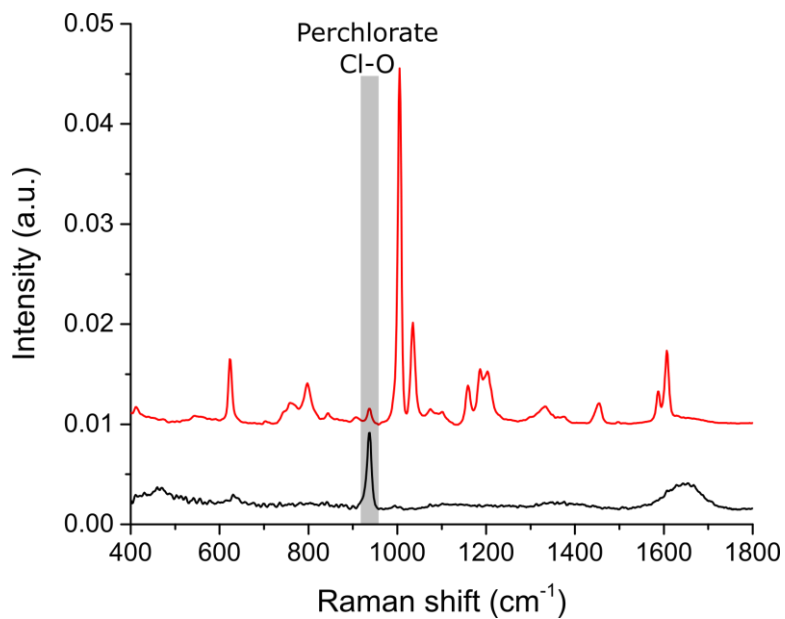
Supplementary Figure 2 | SPARTA polymersome composition analysis, PLSDA classification. 3 component PLSDA classification analysis of a 50-50 v/v % mixture of ABA and ABA-heparin polymersomes ($n = 263$). (a) PLSDA components 1, 2 and 3 (top to bottom), (b) PLSDA scores of component 1 and 2 and classification of ABA (light red) and ABA-heparin (light blue) like polymersomes, modelled using pure ABA (red, $n = 99$) and ABA-heparin (blue, $n = 98$) trapping runs.



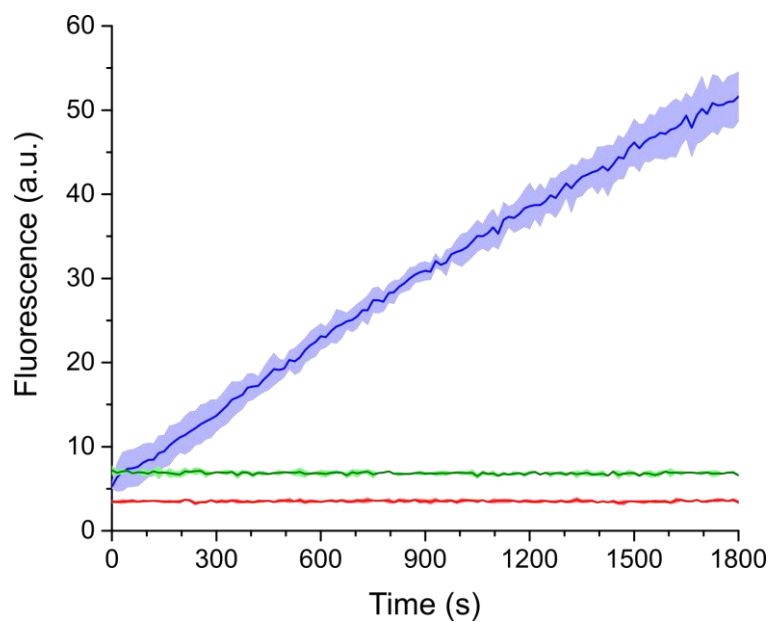
Supplementary Figure 3 | Classification versus trap number. PCA component 1 against the trap number of a 50-50 v/v % mixture of ABA and ABA-heparin polymersomes ($n = 263$), classified by PCA based Ward's clustering into ABA (red) and ABA-heparin (blue) populations. Plot shows a random distribution of trapping of both particles over time.



Supplementary Figure 4 | DLS number distributions of polystyrene particles during serial functionalisation. Gaussian fits of DLS number distributions for PS particles with sulfhydryl functionalisation (mean $\pm$ s.d.) (black, 157.5 ± 34.9 nm), disulphide and nitrobenzoic acid (red, 166.1 ± 46.6 nm) and disulphide and CYY functionalised particles (blue, 156.6 ± 43.5 nm), recovery of sulfhydryl after addition of TCEP to disulphide and nitrobenzoic acid functionalisation (orange, 162.1 ± 36.6 nm), and to CYY functionalisation (green, 160.3 ± 40.9 nm).



Supplementary Figure 5 | Perchlorate Raman spectra. Averaged Raman spectra of PBS (black, $n = 234$) and 200 nm polystyrene beads (red, $n = 300$), each supplemented with 50 mM sodium perchlorate. The perchlorate gives a single, clear solution marker peak at 938 cm^{-1} .



Supplementary Figure 6 | Fluorescence analysis of coumarin dye clicking. Copper catalysed click reaction of 3-Azido-7-hydroxycoumarin (Absorption/Emission = 404/477 nm) to 200 nm polystyrene particles functionalised with propargyl amine resulting in fluorescence when forming the triazole product, ($n = 3$, mean $\pm$ s.d.) clicking conditions (blue), no change in fluorescence in absence of dye (red) or absence of copper sulphate (green).

Supplementary Table 1 | Raman peak vibration assignment.

Figure	Vibration	Raman shift (cm ⁻¹)	Reference
3 a,b,c	<i>C-D</i> stretch	2105	1
3 d	<i>Si-O-Si</i> stretch	490	2
3 d	<i>Si-C</i> sym. stretch	709	2
3 d	<i>C-C</i> stretch glucose/saccharide	930	3
3 d	Glucose/saccharide	1070	3
4 b,c	<i>S-S</i> stretch	452, 512	4, 5
4 b,c	<i>S-H</i> bend	936	5
4 b,c	<i>C=C</i> ring breathing (Tyrosine)	840, 860	3
6 b,c,d	<i>C≡C</i> stretch (Alkyne)	2129	1
6 b,c,d	<i>N≡N</i> stretch (Azide)	2116	1
6 b,c,d	Triazole ring stretch	1331	3, 4
Supplementary Figure 2	<i>Cl-O</i> stretch (Perchlorate)	938	6

Supplementary References

1. Yamakoshi, H. et al. Alkyne-tag Raman imaging for visualization of mobile small molecules in live cells. *J. Am. Chem. Soc.* **134**, 20681-20689 (2012).
2. Cai, D., Neyer, A., Kuckuk, R. & Heise, H.M. Raman, mid-infrared, near-infrared and ultraviolet–visible spectroscopy of PDMS silicone rubber for characterization of polymer optical waveguide materials. *J. Mol. Struct.* **976**, 274-281 (2010).
3. Movasaghi, Z., Rehman, S. & Rehman, I.U. Raman spectroscopy of biological tissues. *Appl. Spectrosc. Rev.* **42**, 493-541 (2007).

4. Yoo, B.K. & Joo, S.-W. In situ Raman monitoring triazole formation from self-assembled monolayers of 1,4-diethynylbenzene on Ag and Au surfaces via “click” cyclization. *J. Colloid Interface Sci.* **311**, 491-496 (2007).
5. Bazylewski, P., Divigalpitiya, R. & Fanchini, G. In situ Raman spectroscopy distinguishes between reversible and irreversible thiol modifications in l-cysteine. *RSC Adv.* **7**, 2964-2970 (2017).
6. Cherney, D.P., Bridges, T.E. & Harris, J.M. Optical trapping of unilamellar phospholipid vesicles: investigation of the effect of optical forces on the lipid membrane shape by confocal-Raman microscopy. *Anal. Chem.* **76**, 4920-4928 (2004).