

Supplementary Information

Mutated H3 Histones Drive Human Pre-Leukemic Hematopoietic Stem Cell Expansion

And Promote Leukemic Aggressiveness.

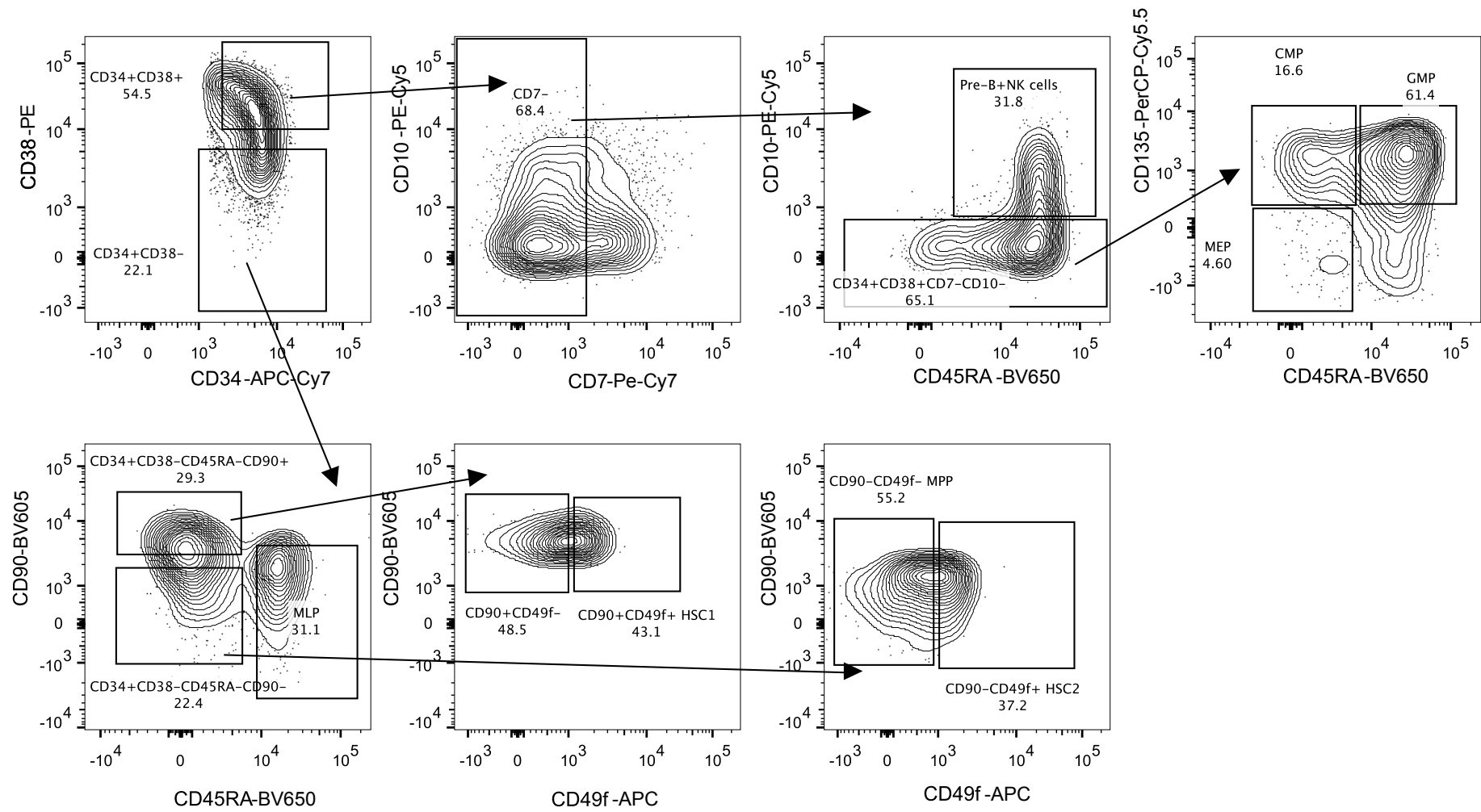
Boileau et al.

Karyotype	AUBMC N (%)	Toronto N (%)
Successful	108 (89)	292 (94)
No data	14 (11)	20 (6)
Normal	34 (31)	148 (51)
t (15;17)	7 (6)	11 (4)
t (8;21)	8 (7)	18 (6)
Inv (16)	7 (6)	15 (5)
t (9;11)	4 (4)	4 (1)
t (9;22)	2 (2)	1 (0)
t (6;9)	4 (4)	2 (1)
Hyper diploid	1 (1)	0 (0)
Complex	20 (19)	31 (11)
Other	40 (37)	62 (21)

Supplementary Table 1 Summary of cytogenetic information of the two cohorts sequenced for H3 histone mutations as determined by karyotypic analysis.

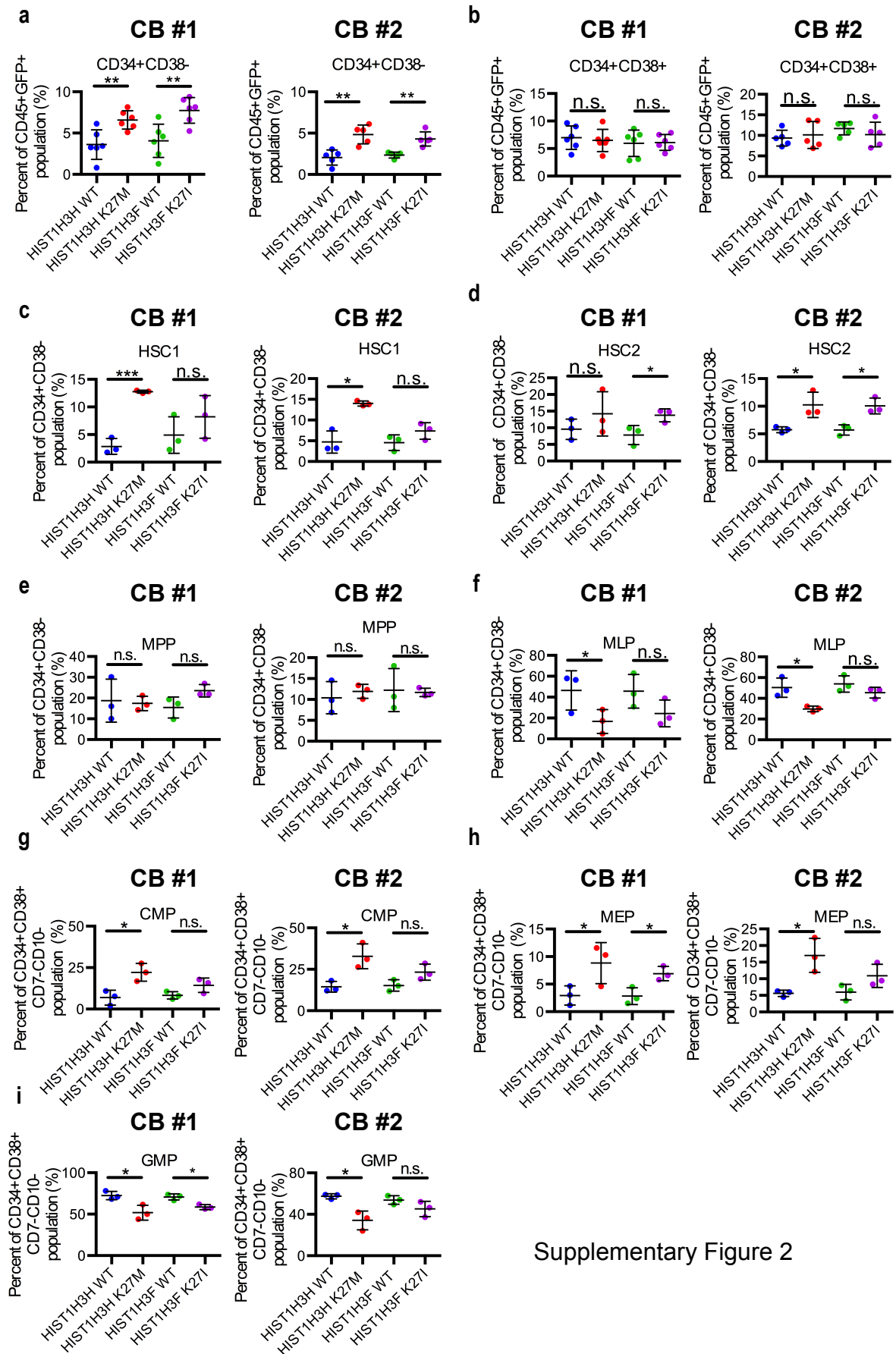
Molecular Abnormalities	AUBMC N (%)	Toronto N (%)
PML-RARA	11 (9)	NA
AML1-ET0	11 (9)	NA
CBF-MYH11	8 (7)	NA
AF9-MLL	4 (3)	NA
BCR-ABL	2 (2)	NA
NPM1c	14 (23)	67 (56)
FLT3-ITD	29 (24)	40 (30)
FLT3-D835	9 (7)	7 (6)
CEBPA	6 (14)	NA
CKIT	1 (2)	NA
IDH1	13 (11)	11 (17)
IDH2	8 (7)	13 (51)

Supplementary Table 2 Summary of abnormalities of the two cohorts sequenced for H3 histone mutations as determined by molecular assays.



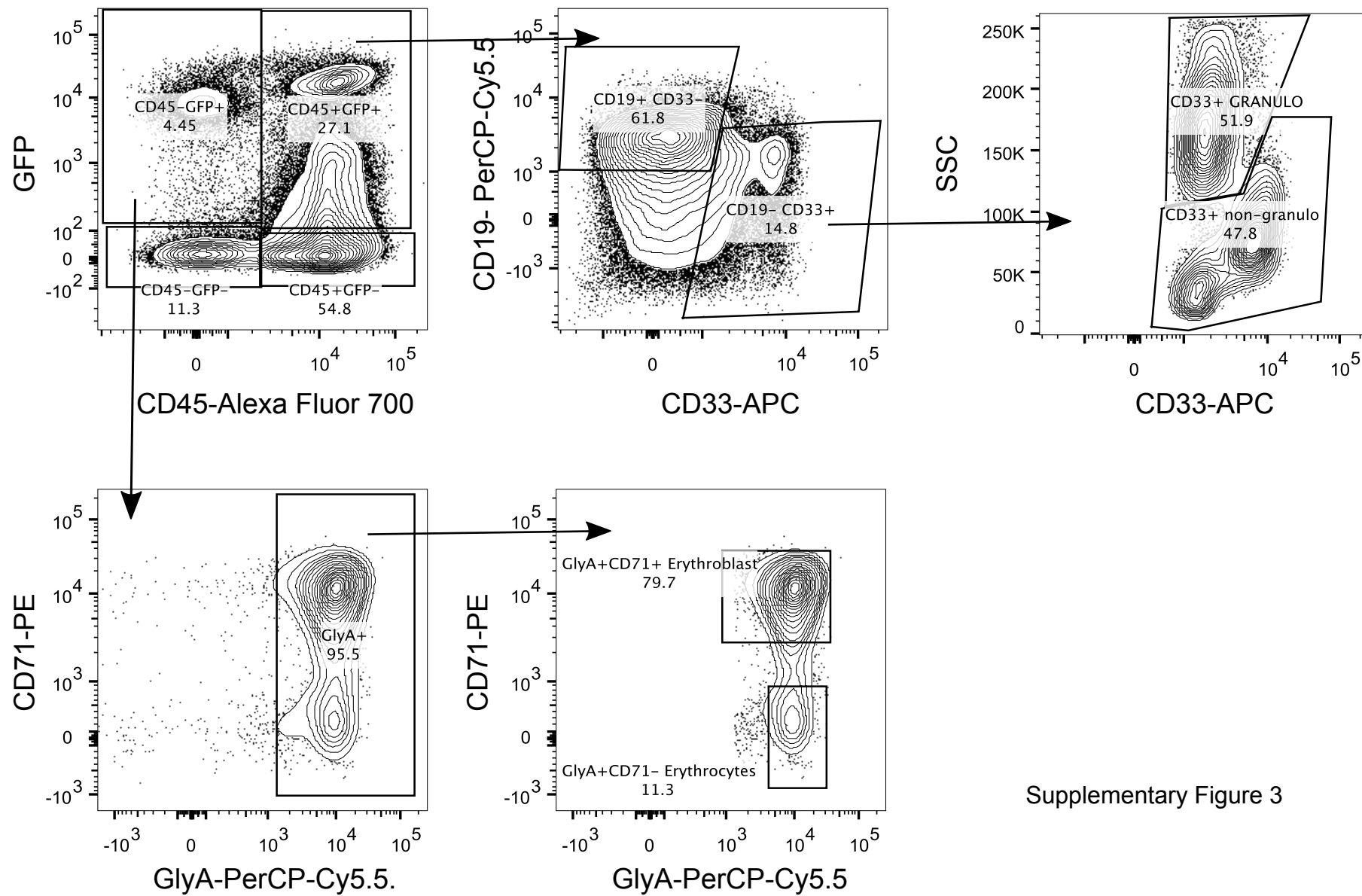
Supplementary Figure 1

Supplementary Figure 1 The gating scheme for defining HSC1(CD34+CD38-CD45RA-CD90+CD49f+), HSC2 (CD34+CD38-CD45RA-CD90-CD49f+), multipotent progenitor (MPP-CD34+CD38-CD90-CD49f-), multilymphoid progenitor (MLP-CD34+CD34-CD90-CD45RA+), common myeloid progenitor (CMP-CD34+CD38+CD7-CD10-CD135+CD45RA-), megakaryocyte-erythroid progenitor (MEP-CD34+CD38+CD7-CD10-CD135-CD45RA-) and granulocyte-macrophage progenitor (GMP-CD34+CD38+CD7-CD10-CD135+CD45RA+) from a representative lineage and mouse depleted sample of the bone marrow of a mouse xenotransplanted with cord blood cells overexpressing HIST1H3H K27M.



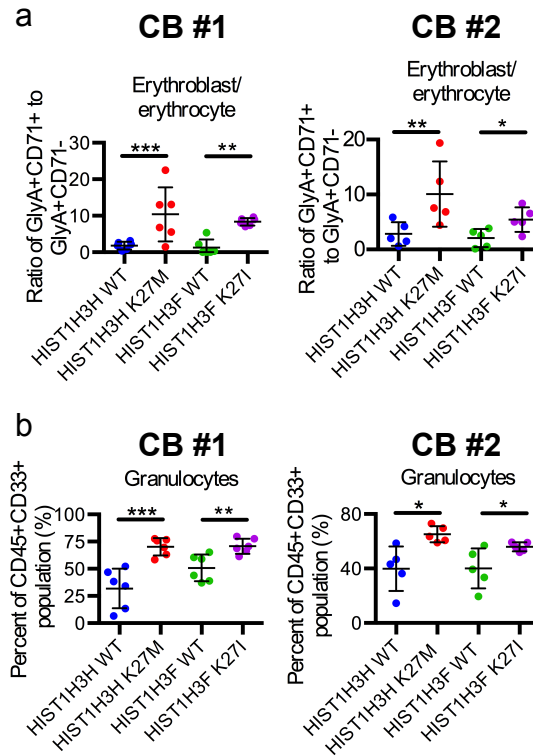
Supplementary Figure 2

Supplementary Figure 2 H3.1 K27 mutations alter HSC frequency and hematopoietic differentiation *in vivo*. Flow cytometry analysis of **(a)** CD34+CD38⁻ cells, **(b)** CD34+CD38⁺ cells, **(c)** HSC1, **(d)** HSC2, **(e)** MPP, **(f)** MLP, **(g)** CMP, **(h)** MEP and **(i)** GMP populations from the bone marrow of the injected femur of mice injected with CD34+CD38⁻ human cord blood transduced with HIST1H3H WT/K27M or HIST1H3F WT/K27I after 12-14 weeks. Both biologically replicated experiments are shown (CB#1 and CB#2). Data represents the mean and the error bars are the standard deviation.; n=6 (CB#1), n=5 (CB#2). Statistical analysis was performed by two-way Student's t-tests. *p≤0.05, **p≤0.01, ***p≤0.005, ****p≤0.0001

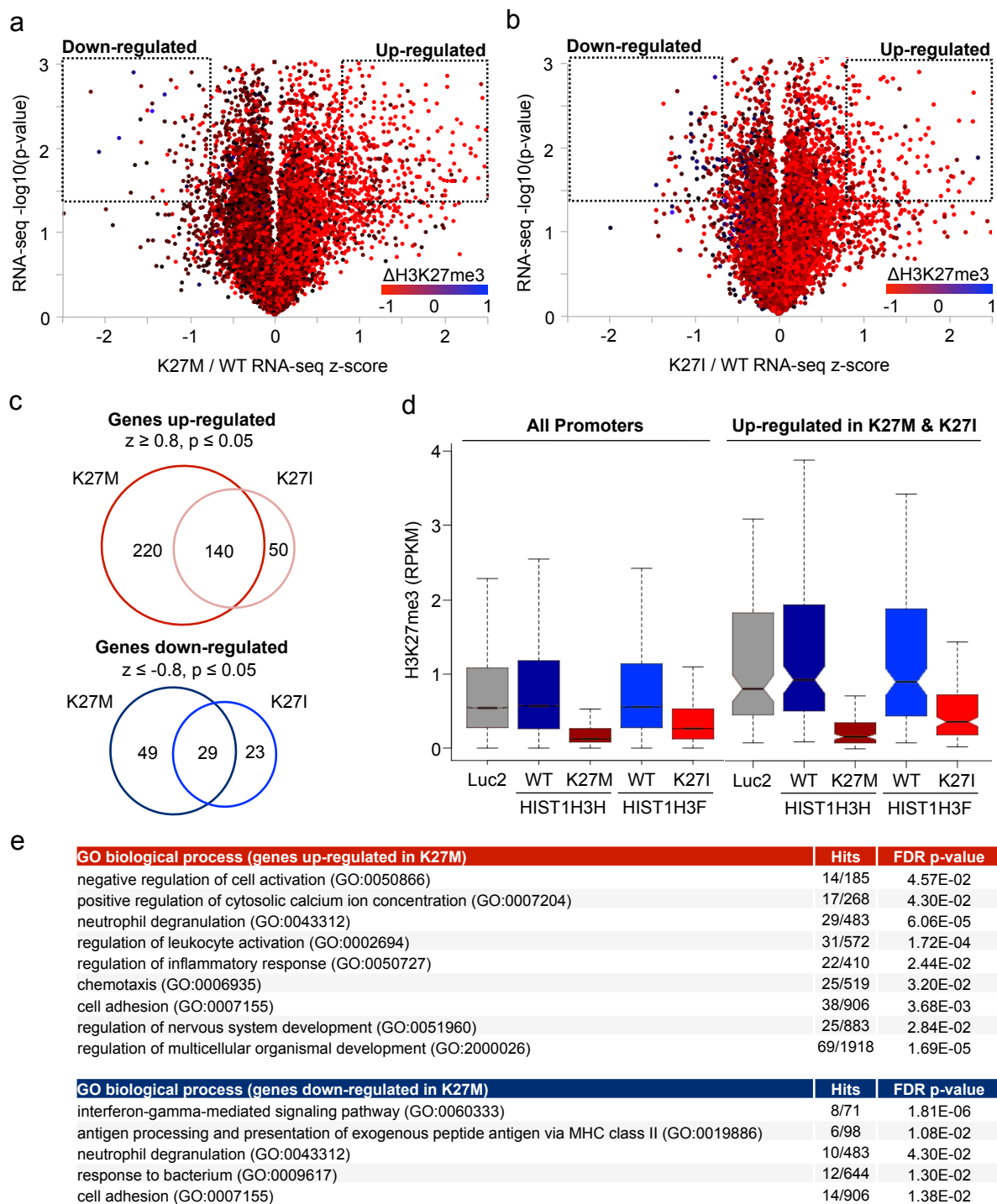


Supplementary Figure 3

Supplementary Figure 3 The gating scheme for defining granulocytes (CD45+CD33+SSChigh), erythroblasts (CD45-GlyA+CD71+) and erythrocytes (CD45-GlyA+CD71-) from a representative sample of the bone marrow of a mouse xenotransplanted with CD34+CD38- cord blood cells overexpressing HIST1H3H K27M.

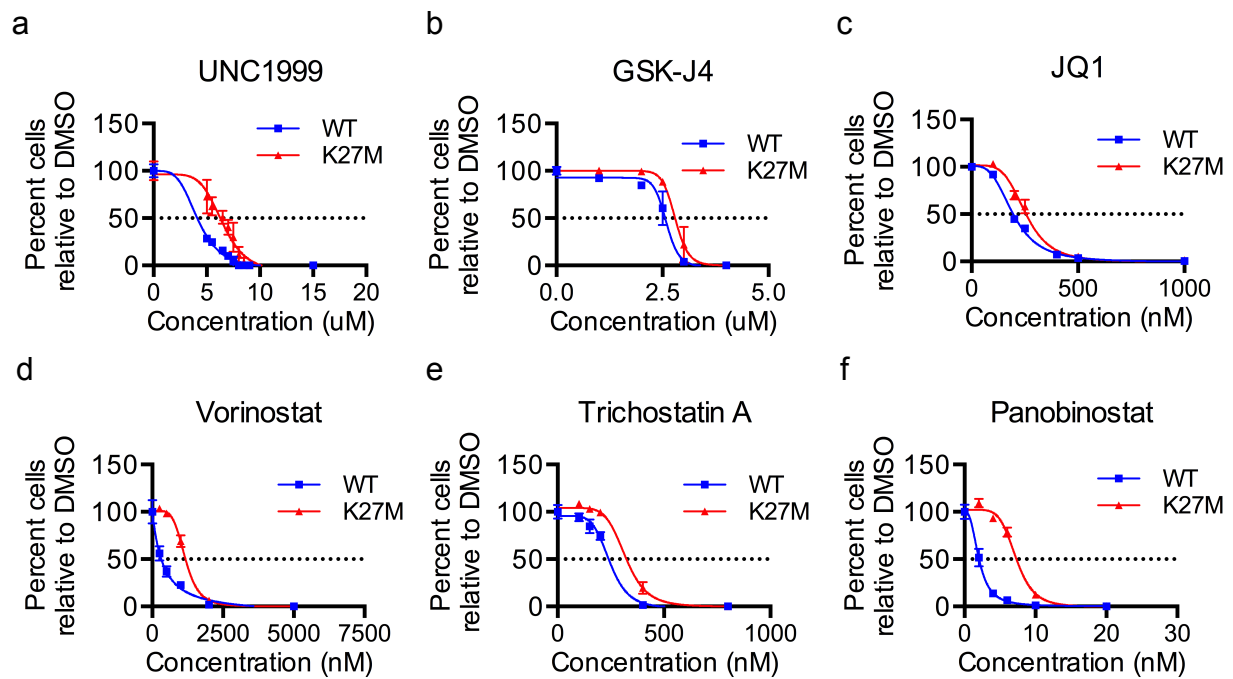


Supplementary Figure 4 H3.1 K27M/I mutations alter erythroid and granulocyte differentiation of human cord blood. Flow cytometry analysis of **(a)** erythroid and **(b)** granulocyte populations from the bone marrow of the injected femur of mice xenotransplanted with CD34+CD38- human cord blood cells transduced with HIST1H3H WT/K27M or HIST1H3F WT/K27I after 12-14 weeks. Both biologically replicated experiments are shown (CB#1 and CB#2). Data represents the mean and error bars are the standard deviations; n=6 (CB#1), n=5 (CB #2). Statistical analysis was performed by two-way Student's t-tests. *p<0.05, **p<0.01, ***p<0.001, ****p<0.0001.



Supplementary Figure 6

Supplementary Figure 6. Expanded analysis of gene expression changes using a z-score cut-off of ± 0.8 . **(a)** Volcano plot depicting genes showing differential expression in K27M relative to HIST1H3H WT and **(b)** K27I relative to HIST1H3F WT. H3K27me3 change of K27M/WT and K27I/WT is overlaid as a heatmap, with red and blue representing loss and gain, respectively. Dashed gates indicate genes called as being significantly down- or up-regulated, using threshold of $|z\text{-score}| \geq 0.8$, $p\text{-value} \leq 0.05$. **(c)** Venn diagram showing overlap of significantly up- and down-regulated genes observed in K27M and K27I cells. **(d)** Box-whisker plot showing change of promoter-specific H3K27me3 in the TEX cells, comparing all annotated genes. The lower and upper whisker represents the minimum and maximum, respectively, after removing outliers where the upper whisker = $\min(\max(x), Q3 + 1.5 * IQR)$ and lower whisker = $\max(\min(x), Q1 - 1.5 * IQR)$, where $IQR = Q3 - Q1$. The two 'hinges' are versions of the first and third quartile. The notches extend to $\pm 1.58 IQR/\sqrt{n}$ representing a confidence interval. Center line indicates median. Note that the up-regulated genes show significantly higher level of H3K27me3 at the promoter in the WT compared to the average promoter. **(e)** Gene ontology enrichment analysis of significantly up- and down-regulated-genes in K27M / WT. Hits represent the total number of genes identified over the total number of genes annotated for the specific GO term. FDR: Fisher's exact test, corrected for multiple testing.



Supplementary Figure 7 TEX cells overexpressing H3.1 K27M are resistant to compounds that modulate histone methylation and acetylation. **(a-f)** TEX cells overexpressing HIST1H3H WT or K27M were treated with **(a)** UNC1999, **(b)** GSK-J4, **(c)** JQ1, **(d)** vorinostat, **(e)** trichostatin A, and **(f)** panobinostat for 5 days at the indicated concentrations. Viability was assessed by flow cytometry. Data represents the mean and error bars are standard deviation; n=3 and is representative of at least 2 independent experiments.