

Supplementary Information for Vedanayagam et al,
Cancer-associated mutations in DICER1 RNase IIIa and IIIb domains
exert similar effects on miRNA biogenesis

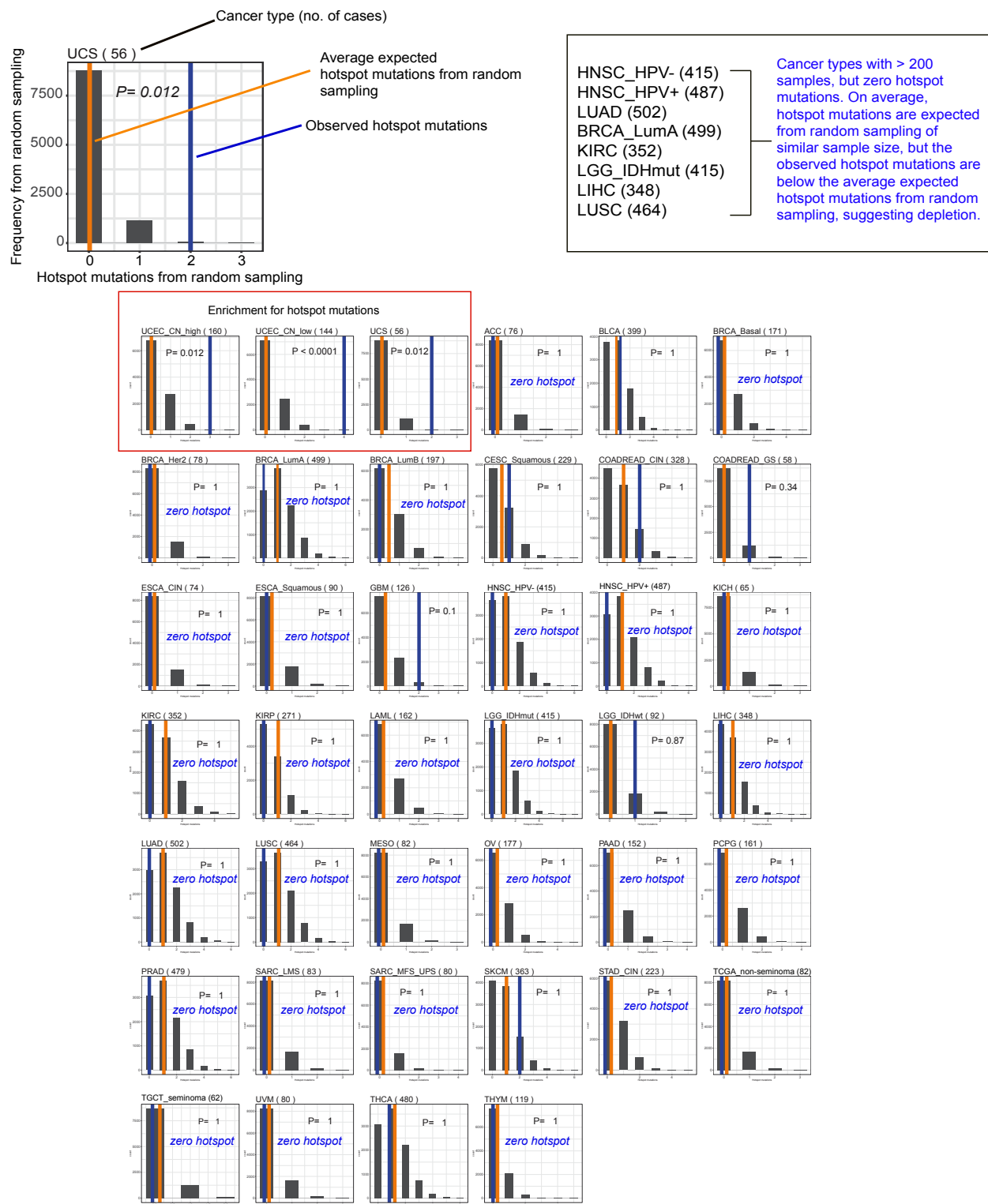
Cancer Hotspots analysis at *DICER1*

Residue		Q-value	Samples
E1813		1.04E-10	11
D1810		8.22E-5	9
D1709		0.0130	10
S1344		0.0281	7

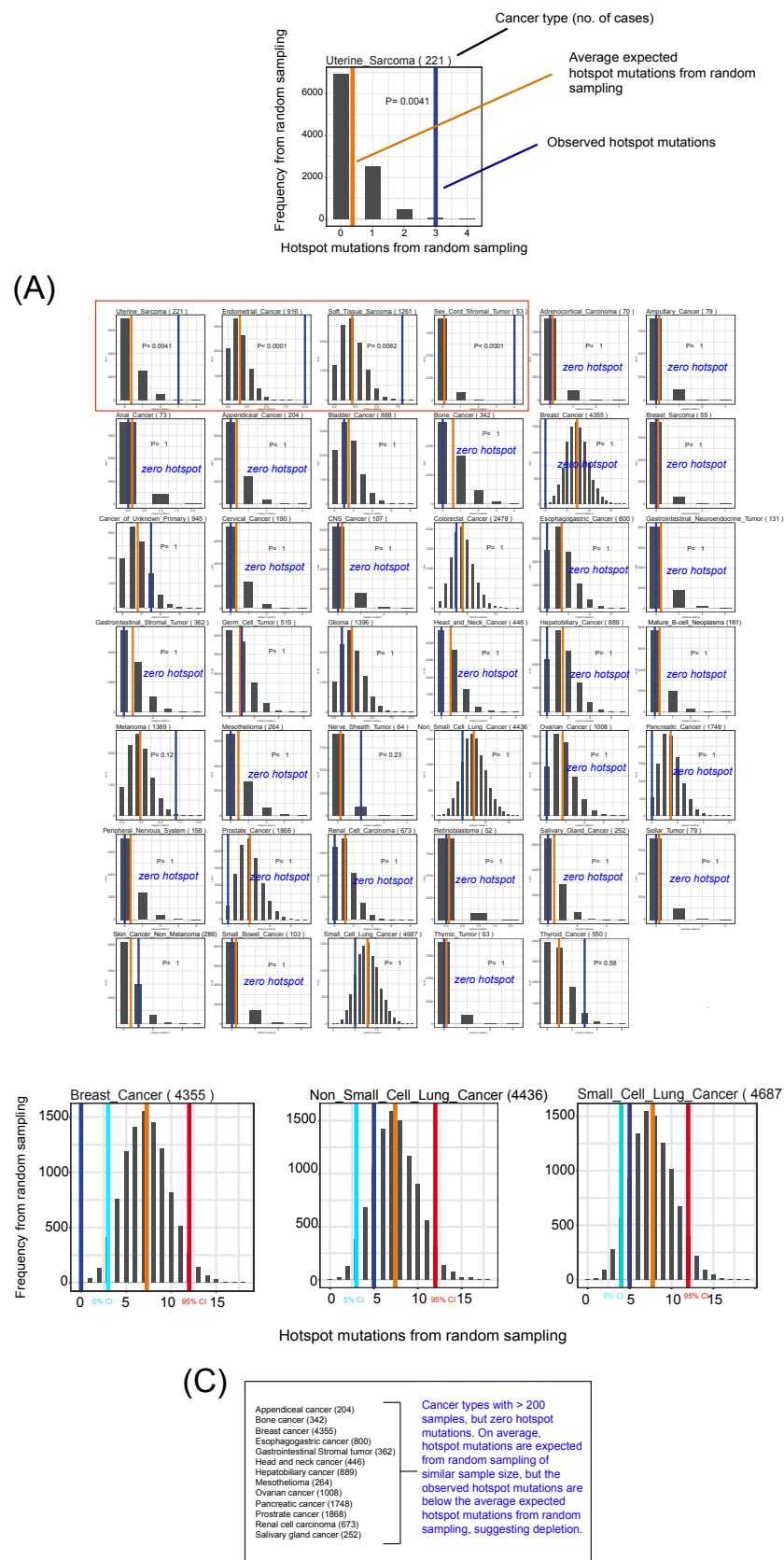
(filtered from 1,165 total mutations from 24,592 tumor samples)

Supplementary Figure 1. Both RNase IIIa and RNase IIIb domains of DICER1 harbor statistically significant hotspot mutations in cancer. Allele distributions and Q-values of three RNase IIIb hotspot residues (E1813, E1810 and D1709) and an uncharacterized RNase IIIa hotspot residue (S1344), from <https://www.cancerhotspots.org>.

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Supplementary Figure 1

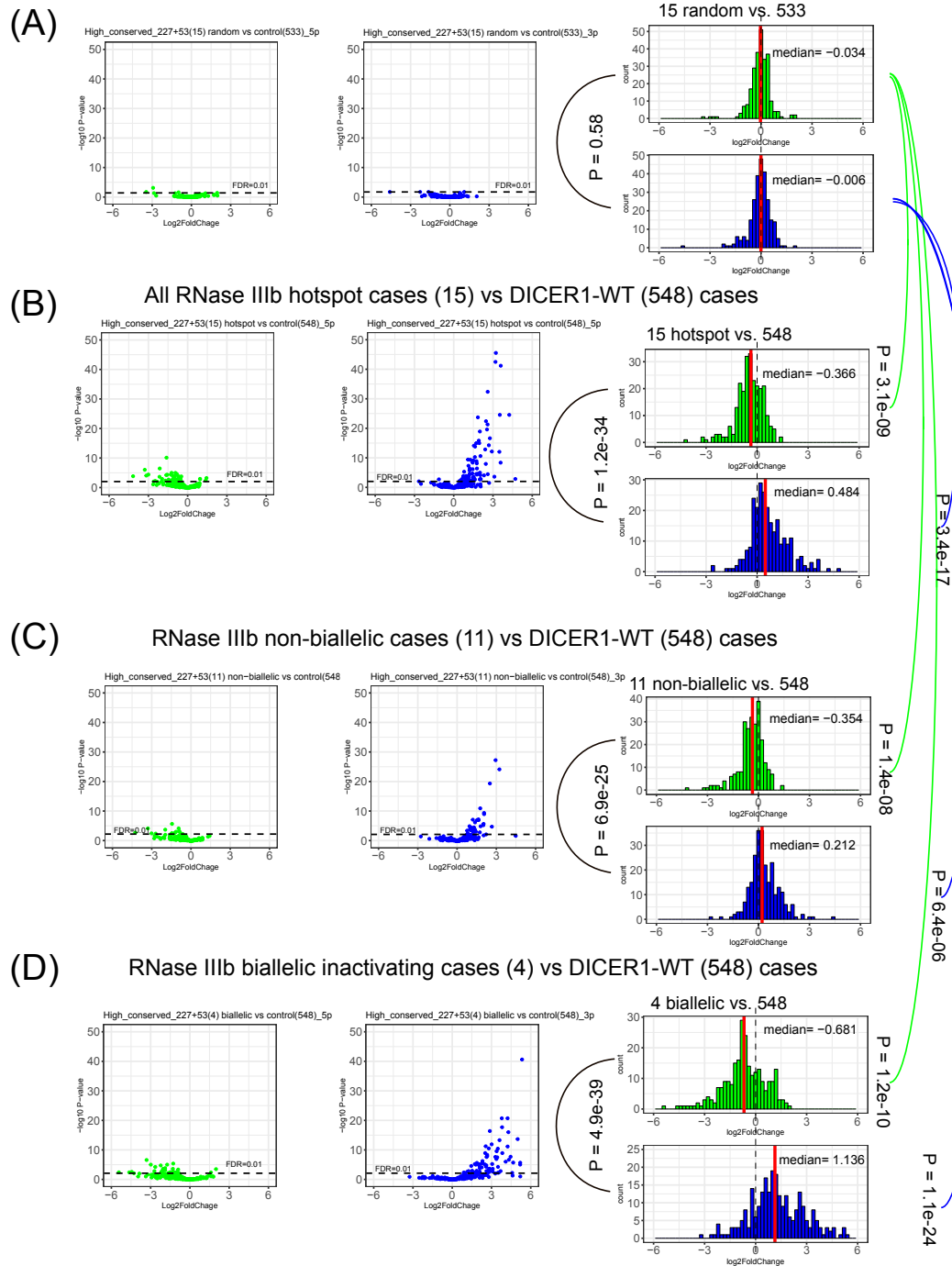


Supplementary Figure 2: Bootstrap resampling analysis for enrichment of hotspot mutations in TCGA PanCan data. Three Uterine cancer types, Uterine Corpus Endometrial Carcinoma (CN high and CN low), and uterine carcinosarcoma are enriched for hotspot mutations (red box). For every cancer type, frequency of hotspot mutations observed from random sampling is shown as a histogram (grey bars). The average expected hotspot mutations from 10000 bootstrap replicates is shown (orange line), along with the actual observed hotspot mutations for every TCGA cancer type (blue line). TCGA cancer types with zero hotspot mutations are highlighted and these make up 70% of TCGA cancer types with > 50 samples. Cancer types with sample size > 200 shown in the box on the right corner illustrates dearth of hotspot mutations in cancers that have similar or higher samples compared to hotspot enriched uterine cancers, indicating depletion of hotspot mutations for most TCGA cancer types. Note that for all cases with zero hotspot mutations, we were not able to estimate statistical significance for depletion due to highly left skewed distribution from resampling, as a result of overall low abundance of hotspot mutations in TCGA PanCan data (31 out of 9919 samples). However, for all these cases, the observed hotspot mutations (blue line) are well below the average hotspot mutations expected from 10000 bootstrap replicates.

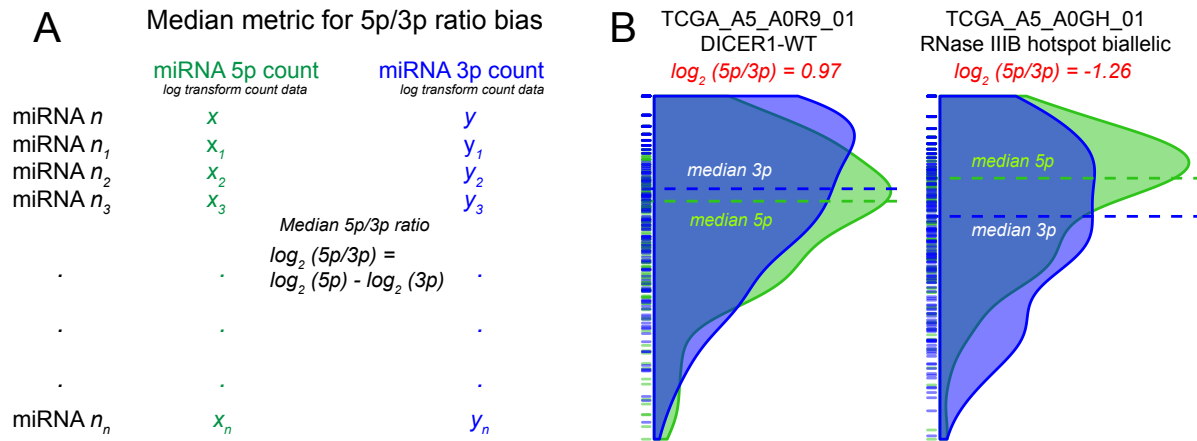


miRNA-5p/3p expression in Uterine Corpus Endometrial Cancer (UCEC)

Control comparison: Random DICER1-wt cases (15) vs DICER1-WT (533) cases



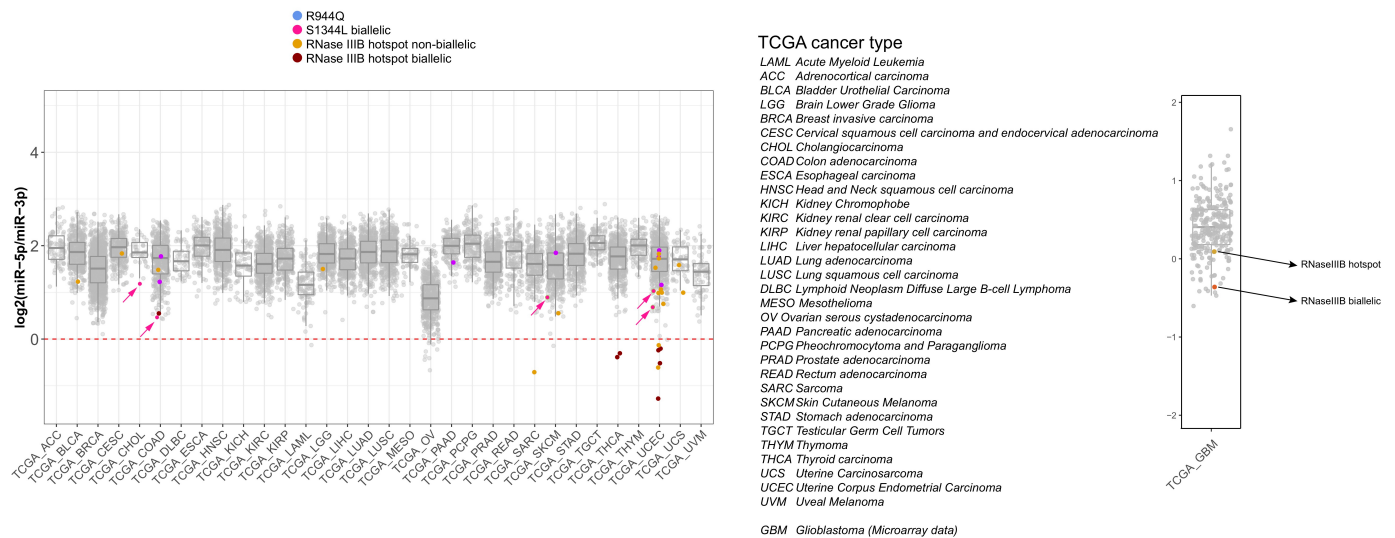
Supplementary Figure S4: miRNA-5p/3p expression in Uterine Corpus Endometrial Cancer (UCEC), showing miRNA processing asymmetry alterations across small RNA datasets. For all comparisons, a volcano plot, and a barplot are shown for miRNA-5p (green), and -3p (blue), respectively. (A) miRNA-5p/3p expression in randomly selected non-hotspot endometrial cases compared to remainder of DICER1-wt, in addition to other endometrial WT cases (hereafter, together referred as WT). In control comparisons some miRNAs exhibited apparent up/downregulation, but almost none were significant. On the other hand, when miRNA-5p/3p expression was compared between WT cases and either all RNaseIIIb mutations (15 cases) (B), or just the non-allelic RNase IIIb hotspot mutations (11 cases) (C), or only the biallelic hotspot mutations (4 cases) (D), we find significantly elevated miRNA-3p reads, and substantial reduction of miRNA-5p reads in all three comparisons. Of note, only biallelic hotspot mutations represented most drastic bias in miRNA-5p/3p expression. False discovery rate for significantly different expression was determined using the Benjamini-Hochberg method at a FDR of 0.01. Other P-values for all comparisons are from a Wilcoxon Rank-Sum test.



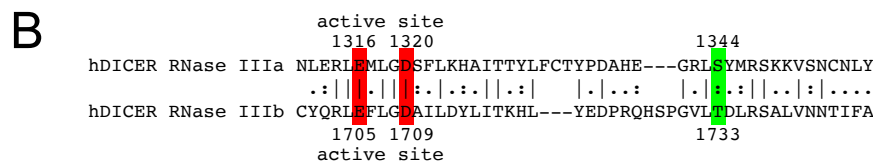
Supplementary Figure 5. Schematic and utilization of a scoring metric to evaluate relative production of miRNA-5p and -3p strands across a library.

(A) We focused on a set of 280 well-conserved and/or broadly-expressed miRNAs (see Supplementary Table 5). We summarize the $\log_2$ expression counts of all 5p and 3p strands separately. Because many miRNAs exhibit highly asymmetric maturation of 5p and 3p strands, we found it advantageous to calculate the median expression of 5p and 3p strands in a library, as opposed to the mean expression, which was more biased by highly expressed loci. The median 5p/3p ratio m_{53}^i is given as $\log_2(5p/3p)$.

(B) The shift in 5p:3p arm distribution between an arbitrarily selected DICER-wt sample and an RNase IIIb hotspot with a biallelic inactivating mutation is illustrated, with the resulting effect on the m_{53}^i metric.



Supplementary Figure 6. Distribution of mi53 scores across TCGA samples segregated by cancer types. Bear in mind that the mi53 metric is influenced by miRNA expression. Thus, the overall averages for some cancers is lower than others; e.g., acute myeloid leukemia (LAML) and Ovarian serous cystadenocarcinoma (OV), Uveal Melanoma (UVM). Nevertheless, for the majority of cancers, the mi53 metric is relatively stable, allowing outliers to be defined. Note that many RNase IIb hotspot mutants, well as the RNase IIIa hotspot (S1344L) mutants, are distinctly amongst the cases with the lowest mi53 scores in each cancer cohort. On the other hand, the seemingly recurrent, but functionally inert R944Q cases did not exhibit any strong bias. Moreover, the RNase IIb hotspot mutants with biallelic inactivating mutations comprise the lower scoring cases amongst the entire ~10,000 TCGA cases analyzed. Glioblastoma (GBM) was the only legacy TCGA cohort only with microarray miRNA data available; thus there was limited power to analyze matching 5p/3p miRNA sets. However, an RNase IIb case with biallelic inactivating mutation was amongst the lowest mi53 scoring cases in this cohort.



3D ribbon diagram of the protein structure showing the binding site of the inhibitor. The protein is represented by a grey ribbon. The inhibitor is shown as a stick model with orange and green spheres. Key residues are highlighted: E1705, D1709, D1810, E1813 (orange), T1733 (blue), S1344 (magenta), and Mg²⁺ (green). A vertical double-headed arrow indicates the distance between T1733 and S1344, labeled "Homologous residues".

(C) Modeling based on a DICER1 RNase III homodimeric structure (PDB:2eb1) shows that cancer-prone S1344 in the RNase IIIa domain is homologous to T1733 in RNase IIIb domain, which in turn lies in close proximity to RNase IIIb metal ion-binding residues that form part of the active site (and are also cancer hotspot mutations). This provides evidence that S1344, although it resides within RNase IIIa domain, is important for proper function of the RNase IIIb domain.