

SUPPLEMENTARY INFORMATION

Real-time monitoring of PARP1-dependent PARylation by ATR-FTIR-spectroscopy

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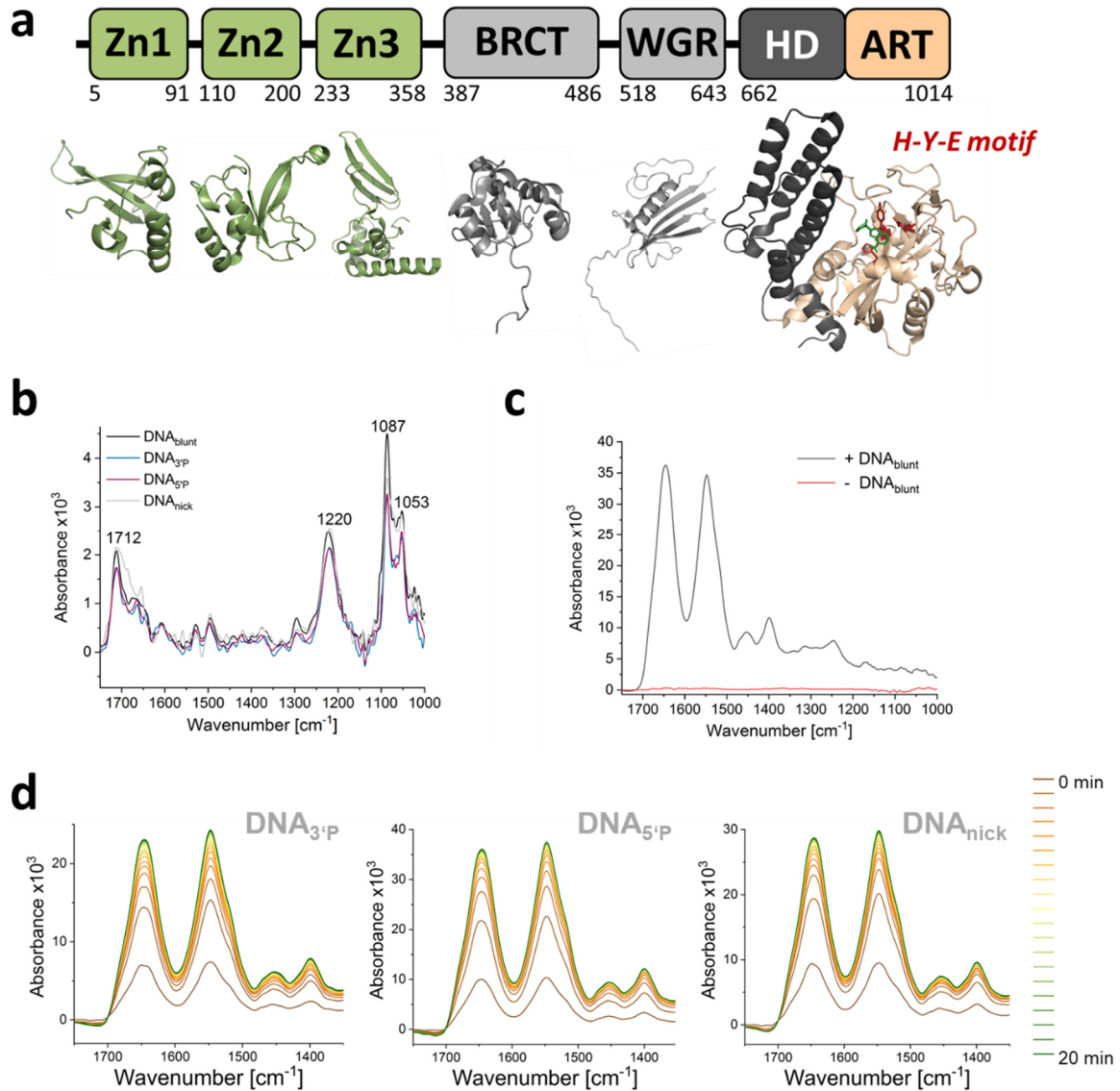
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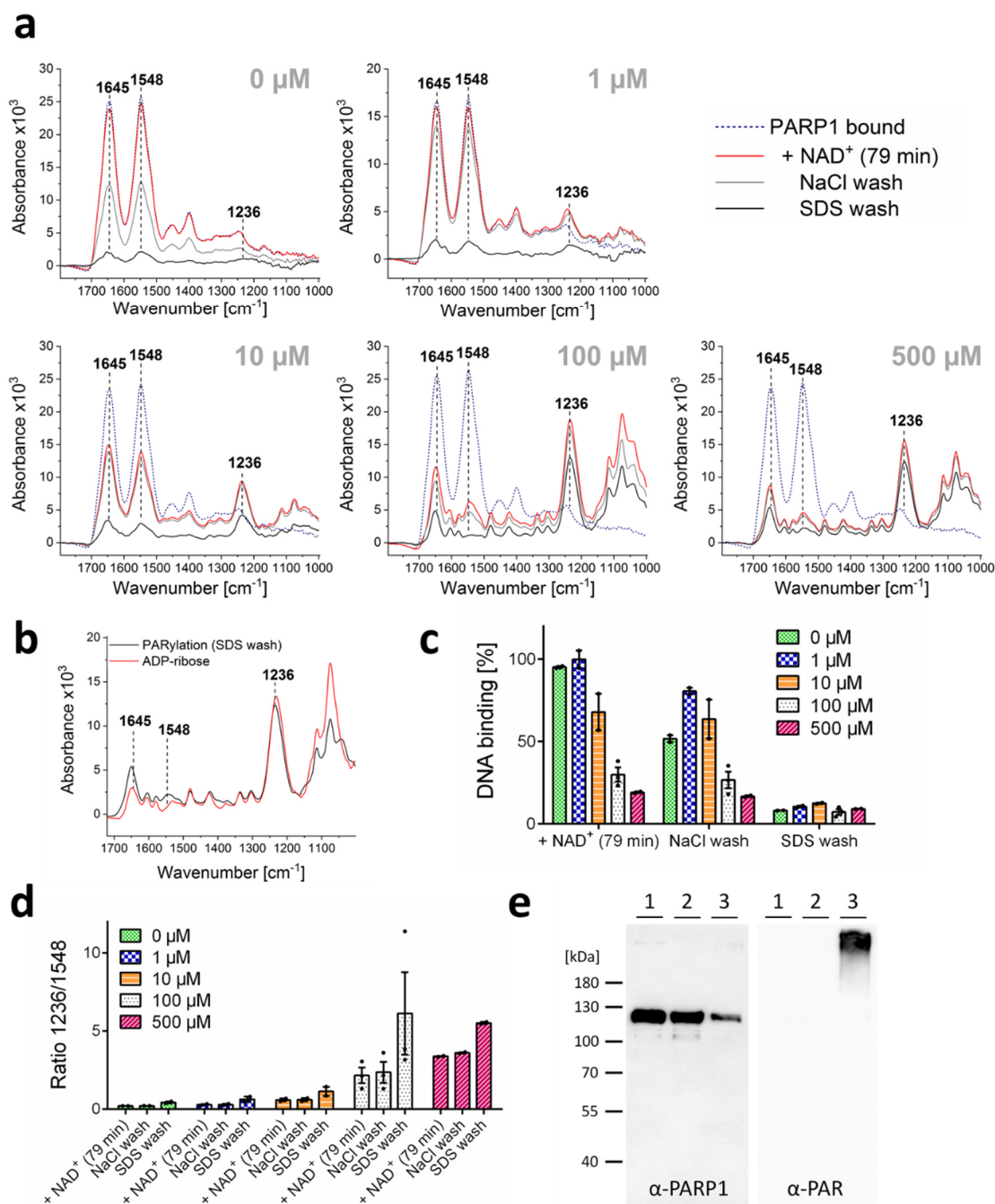
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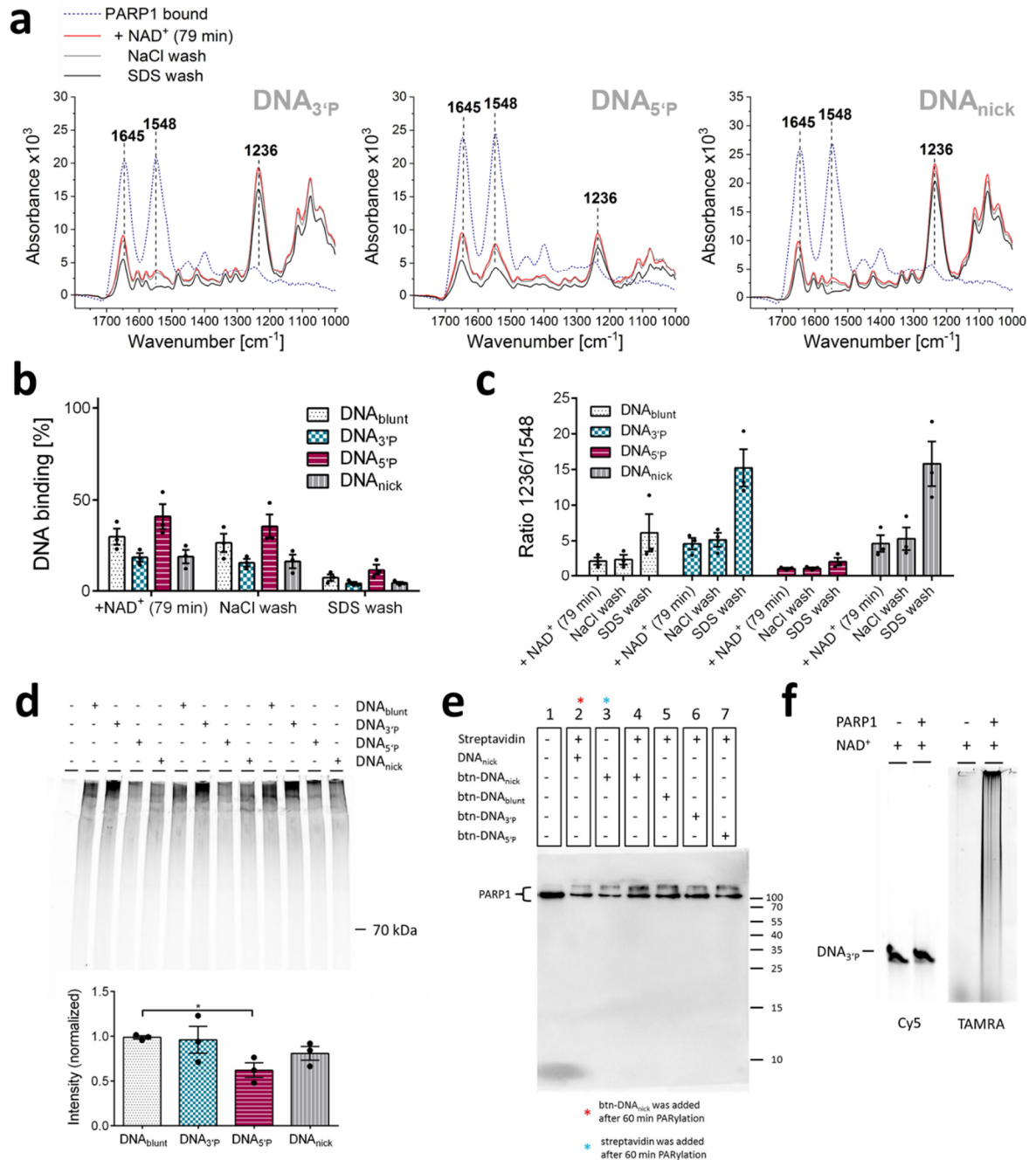
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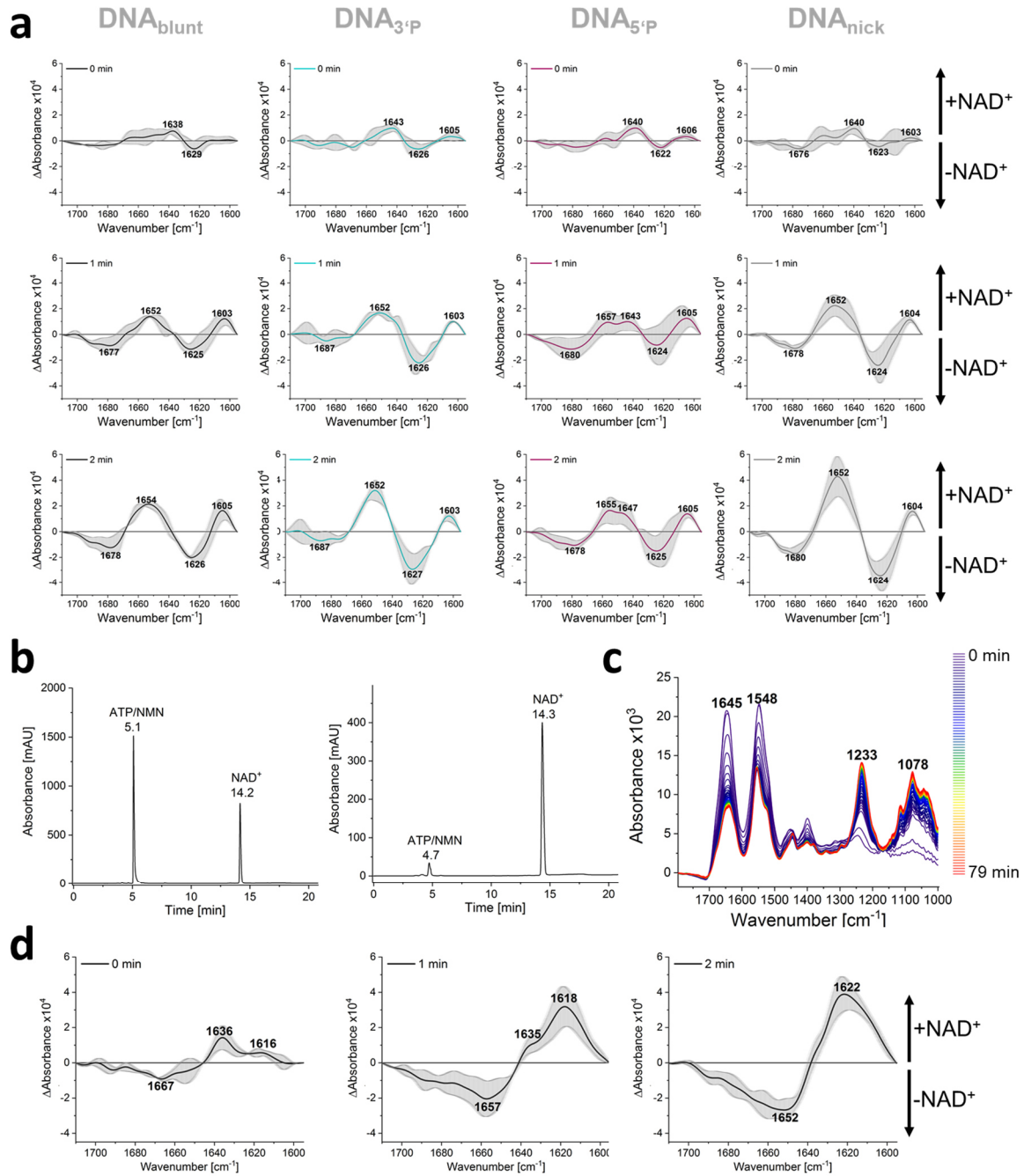
Supplementary Fig. 1. **a** Domain organization of PARP1. PARP1 contains three zinc-finger domains Zn1 (pdb: 3oda), Zn2 (pdb: 3ode) and Zn3 (pdb: 2jvn), a BRCT domain (pdb: 2le0), a WGR domain (pdb: 2cr9) and a catalytic domain harbouring an autoinhibitory HD domain and the catalytically active ART domain (pdb: 3gn7). The catalytic domain is in complex with veliparib. The catalytic conserved H-Y-E motif is highlighted. **b** Representative spectra of immobilized DNA_{blunt}, DNA_{3'P}, DNA_{5'P} and DNA_{nick}. Spectra of streptavidin were subtracted, respectively. **c** Comparison of PARP1 binding to DNA_{blunt}, which was immobilized via streptavidin (+DNA_{blunt}), or streptavidin alone, i.e., without DNA_{blunt} (-DNA_{blunt}). **d** Representative time-dependent spectra of PARP1 binding to immobilized DNA_{3'P}, DNA_{5'P} and DNA_{nick}. Source data are provided as a Source Data file.



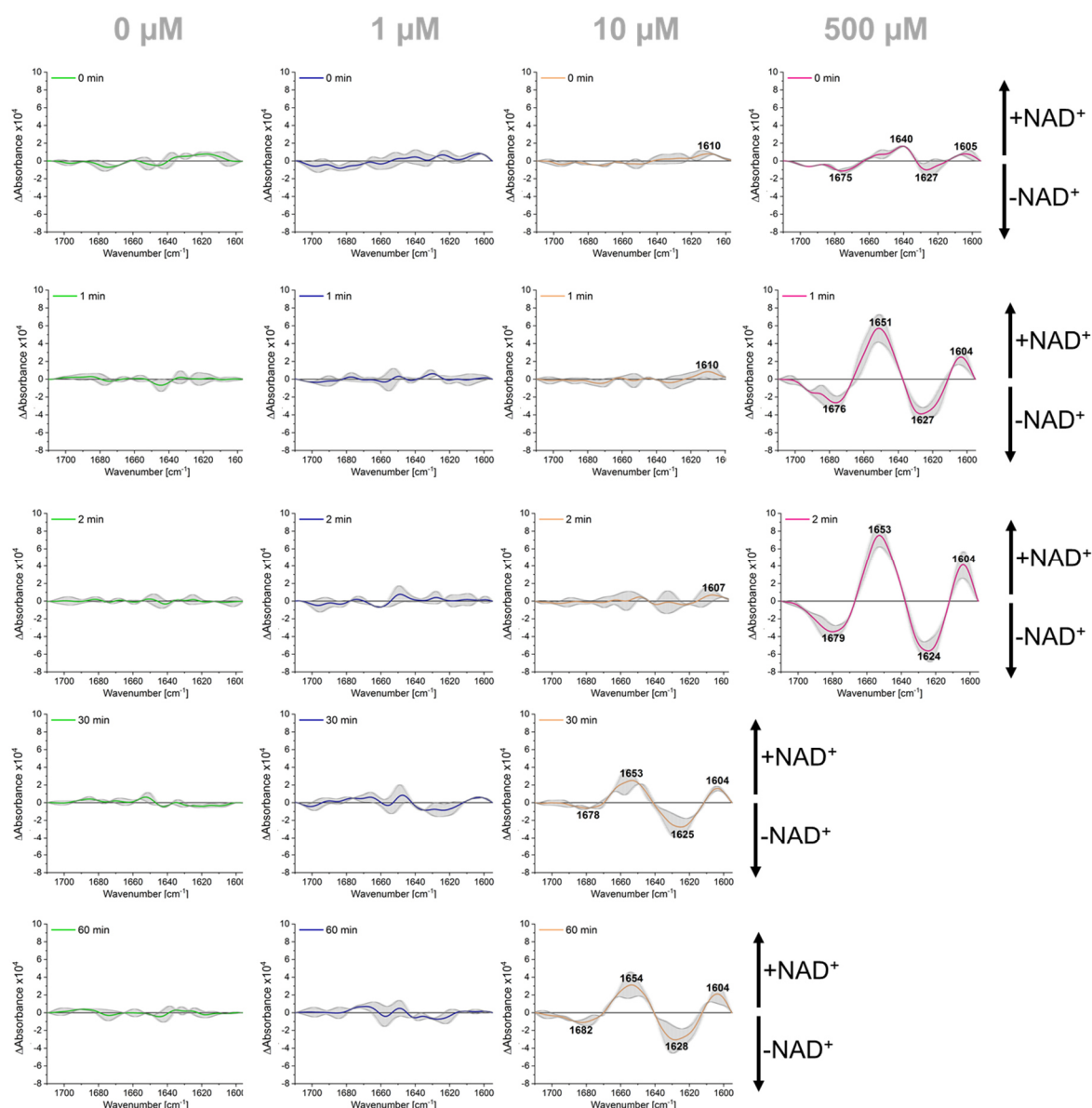
Supplementary Fig. 2. a Representative spectra of PARP1 bound to DNA_{blunt} before ('PARP1 bound') and after the addition of various concentrations (0-500 μM) of NAD⁺ ('79 min') and subsequent washing with 1 M NaCl ('NaCl wash') and 1% SDS ('SDS wash'). **b-d** Evaluation of **a**. **b** Comparison of the spectrum of **a**, bottom right panel (addition of 500 μM NAD⁺, 'SDS wash'), with the spectrum of ADP-ribose (Fig. 3a). **c** Quantification of PARP1 binding to DNA_{blunt}. Intensities of amide II bands (1548 cm^{-1}) were analysed. 'PARP1 bound' was set to 100%. **d** Analysis of 'trans-PARylation'. Ratio of anti-symmetric phosphate vibration of PAR (1236 cm^{-1}) and amide II band of PARP1 (1548 cm^{-1}) was calculated. **c, d** Means $\pm$ SEM of $n=3$ (100 μM) and $n=2$ (0, 1, 10 and 500 μM) independent experiments respectively. **e** Analysis of the supernatant from an ATR-FTIR spectroscopic experiment by immunoblotting. PARP1 was detected via CII10 antibody and PAR was detected via 10H antibody. Lanes 1+2: Analysis of the efficiency of PARP1 binding to immobilized DNA_{blunt} [before (1) and after (2) addition of PARP1 to immobilized DNA_{blunt}]. Lane 3: Analysis of the extent of auto-modification of PARP1 after NAD⁺ addition (79 min). Source data are provided as a Source Data file.



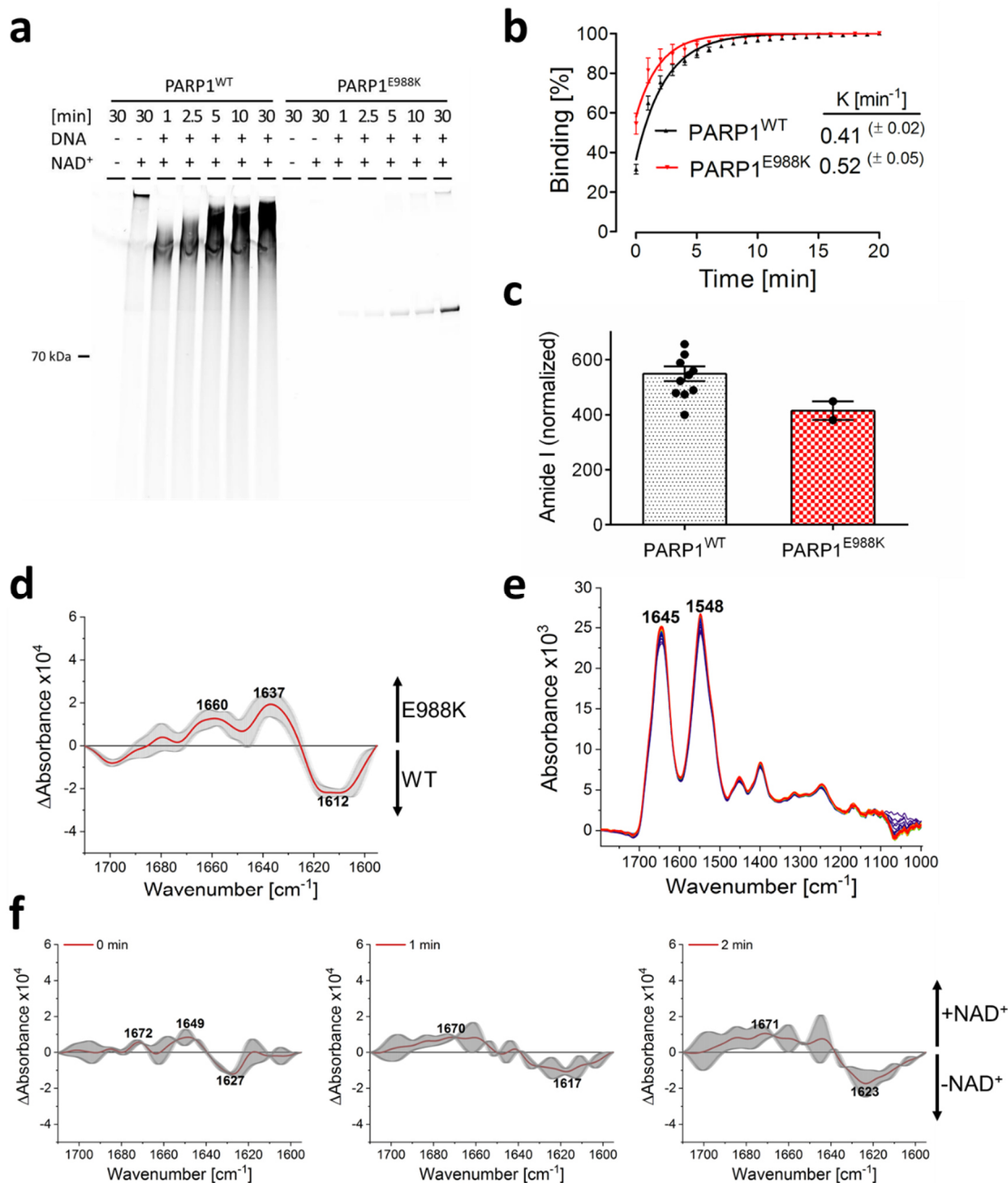
Supplementary Fig. 3. a Representative spectra of PARP1 bound to DNA_{3'P}, DNA_{5'P} or DNA_{nick} before ('PARP1 bound') and after the addition of 100 μ M of NAD⁺ ('79 min') and subsequent washing with 1 M NaCl ('NaCl wash') and 1% SDS ('SDS wash'). **b,c** Evaluation of **a** and Supplementary Fig. 2a (100 μ M NAD⁺). **b** Quantification of PARP1 binding to DNA_{blunt}, DNA_{3'P}, DNA_{5'P} or DNA_{nick}. Intensities of amide II bands (1548 cm⁻¹) were analysed. 'PARP1 bound' was set to 100%. **c** Analysis of 'trans-PARylation'. Ratio of anti-symmetric phosphate vibration of PAR (1236 cm⁻¹) and amide II band of PARP1 (1548 cm⁻¹) was calculated. **b,c** Means $\pm$ SEM of n=3 independent experiments. **d** Analysis of auto-modification of PARP1 via a gel-based assay using fluorescently labelled NAD⁺. PARylation was started by the addition of 100 μ M TAMRA-labelled NAD⁺. Upper panel: Representative gel with three technical replicates. Lower panel: Densitometric quantification of signal intensities. Means $\pm$ SEM of n=3 independent experiments. '*' indicates p<0.05, statistical analysis was performed using an unpaired two-sided t-test. **e** Immunodetection of PARP1 corresponding to Fig. 4d. **f** Analysis of covalent PARylation of DNA_{3'P} via a gel-based assay detecting Cy5-labelled DNA_{3'P} ('Cy5') or TAMRA-labelled NAD⁺ ('TAMRA'). Source data are provided as a Source Data file.



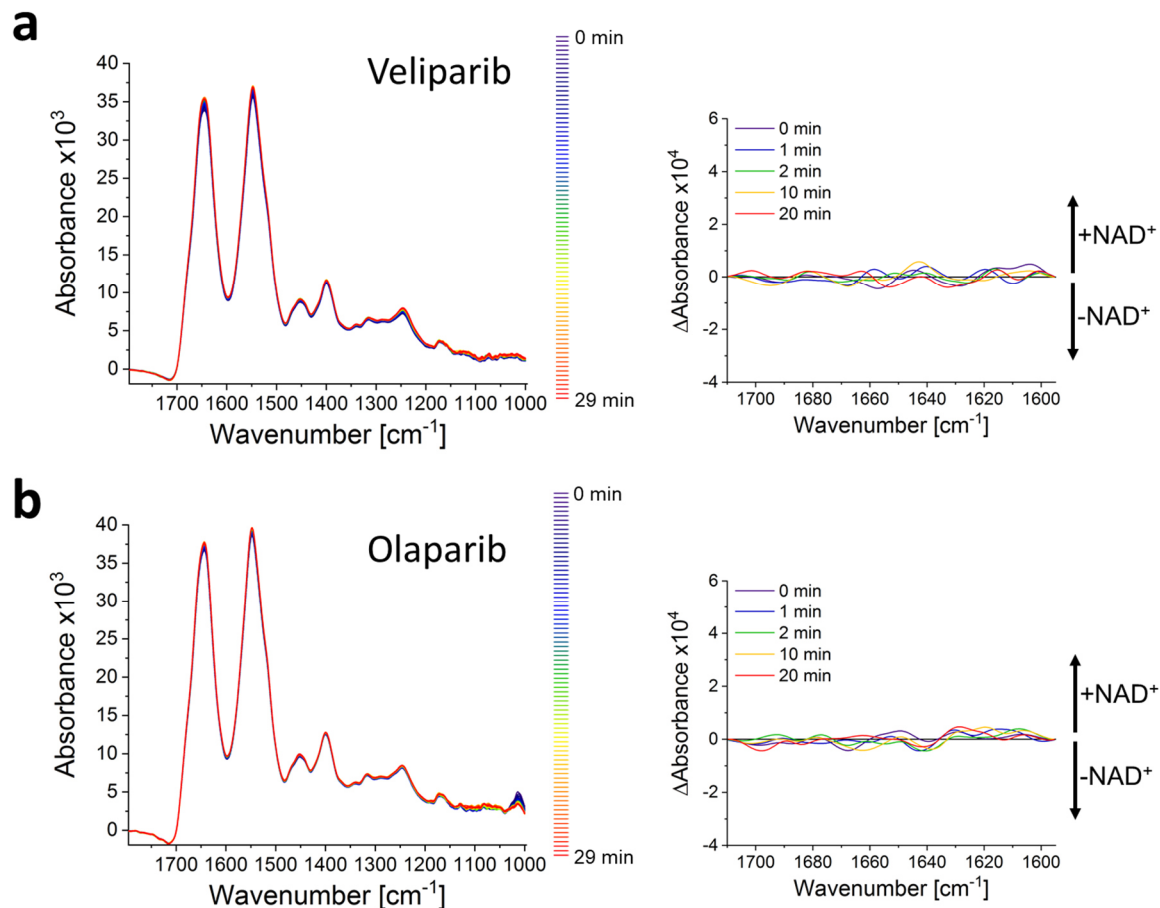
Supplementary Fig. 4. a Individual average curves including SDs (grey), which correspond to data shown in Fig. 5a. **b** Analysis of enzymatic synthesis of $^{13}\text{C}, ^{15}\text{N}$ -NAD $^+$ via HPLC. $^{13}\text{C}, ^{15}\text{N}$ -ATP and NMN were incubated with NMNAT1 in the absence (*left panel*) or in the presence (*right panel*) of PPase. **c** Representative time-dependent spectra following the addition of 100 μM $^{13}\text{C}, ^{15}\text{N}$ -NAD $^+$ to PARP1 bound to immobilized $\text{DNA}_{\text{blunt}}$. Amide I (1645 cm^{-1}) and amide II (1548 cm^{-1}) bands of PARP1 and anti-symmetric (1233 cm^{-1}) and symmetric (1078 cm^{-1}) phosphate vibrations of generated isotopically labelled PAR are indicated. **d** Average curves and SDs (grey), which correspond to data shown in Fig. 5d. Source data are provided as a Source Data file.



Supplementary Fig. 5. Secondary structure analysis of PARP1 upon addition of various NAD^+ concentrations. Difference spectra of amide I bands of PARP1 (1710-1595 cm^{-1}) before and after the addition of NAD^+ (0, 1, 2, 30 and 60 min) were calculated. Average curves and SDs (grey) of $n=2$ independent experiments are plotted, respectively. Source data are provided as a Source Data file.



Supplementary Fig. 6. Analysis of PARP1 variant E988K. **a** Comparison of auto-modification of PARP1^{WT} and PARP1^{E988K} via a gel-based assay using fluorescently labelled NAD⁺. PARylation was started by the addition of 100 μ M TAMRA-labelled NAD⁺. **b** Evaluation of time-dependent binding of PARP1^{WT} and PARP1^{E988K} to DNA_{blunt}. '0 min' refers to start of measurements. Signal intensities of amide I bands (1645 cm^{-1}) at 20 min were set to 100%. Binding kinetics were calculated via a mono-exponential fit function. PARP1^{E988K}: Means $\pm$ SEM of n=2 independent experiments. Data of PARP1^{WT} was taken from Fig. 2. **c** Comparison of the amount of PARP1^{WT} and PARP1^{E988K} bound to DNA_{blunt} after 20 min of incubation (data from same experiments as in b). Amide I bands (1645 cm^{-1}) were normalized to the amount of immobilized DNA (1220 cm^{-1}). Data of PARP1^{WT} was taken from Fig. 2. **d** Secondary structure analysis. Difference spectra of amide I bands of PARP1^{WT} and PARP1^{E988K} bound to DNA_{blunt} were calculated. Average curve and SD (grey) of 6 difference spectra are plotted (PARP1^{WT}: n=3 independent experiments; PARP1^{E988K}: n=2 independent experiments). **e** Representative time-resolved spectra subsequent to the addition of 100 μ M NAD⁺ to PARP1^{E988K} bound to immobilized DNA_{blunt}. Amide I (1645 cm^{-1}) and amide II (1548 cm^{-1}) bands are indicated. **f** Secondary structure analysis of PARP1^{E988K} upon addition of 100 μ M NAD⁺. Difference spectra of amide I bands (1710-1595 cm^{-1}) before and after the addition of NAD⁺ (0, 1 and 2 min) were calculated. Average curves and SDs (grey) of n=2 independent experiments. Source data are provided as a Source Data file.



Supplementary Fig. 7. a Addition of 100 μM NAD⁺ to PARP1 bound to immobilized DNA_{blunt} after preincubation with veliparib **a** or olaparib **b**. *Left panels:* Time-dependent IR spectra. *Right panels:* Secondary structure analysis of PARP1. Difference spectra of amide I bands (1710-1595 cm^{-1}) before and after the addition of NAD⁺ (0, 1, 2, 10 and 20 min) were calculated and do not indicate significant structural changes. These control experiments show that structural changes of PARP1 are specifically observed after binding of NAD⁺ to the catalytic centre. Source data are provided as a Source Data file.