

SUPPLEMENTARY INFORMATION

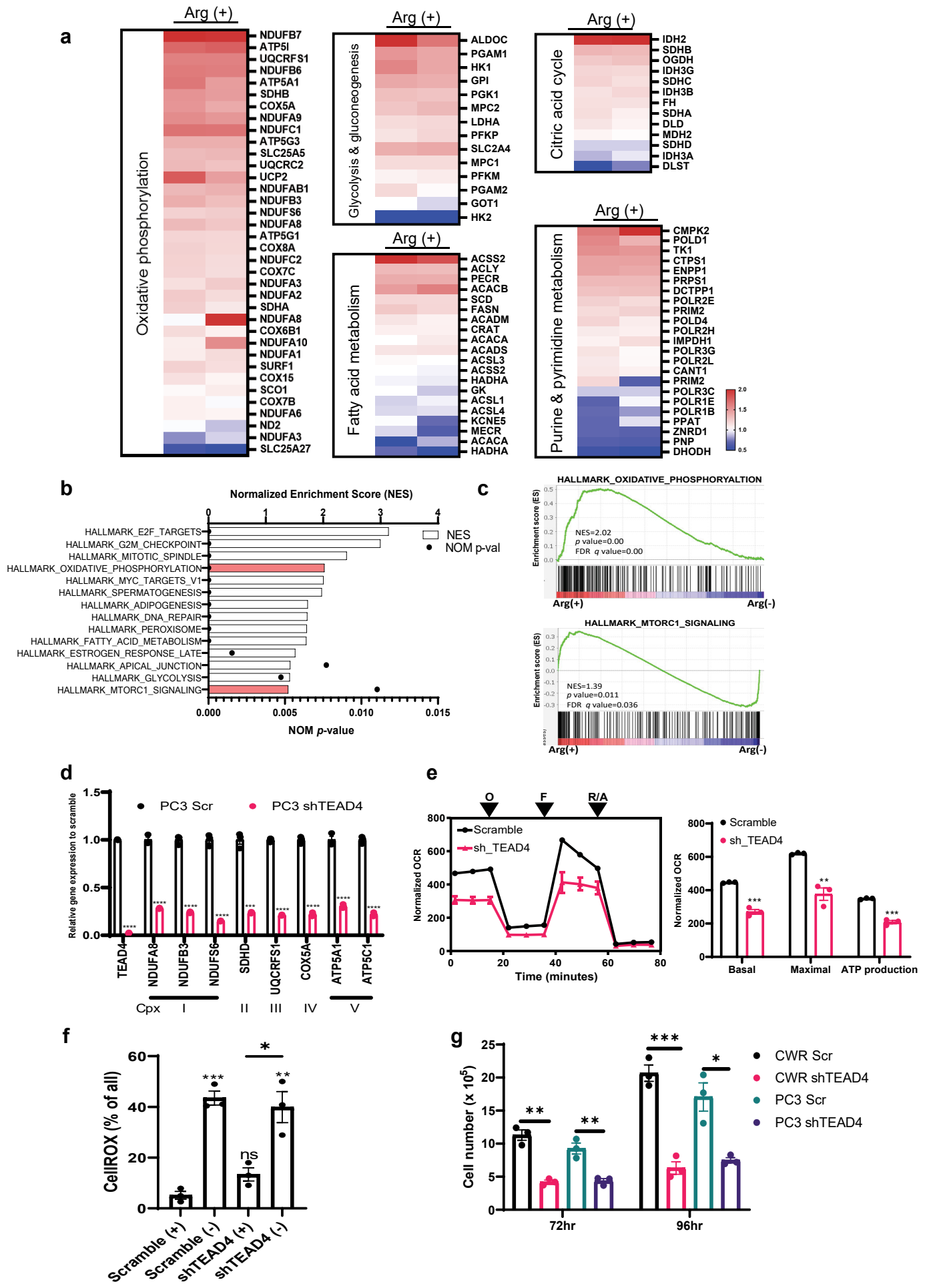
Arginine is an epigenetic regulator targeting TEAD4 to modulate OXPHOS in prostate cancer cells

Chia-Lin Chen, Sheng-Chieh Hsu, Tan-Ya Chung, Cheng-Ying Chu, Hung-Jung Wang, Pei-Wen Hsiao, Shauh-Der Yeh, David K. Ann, Yun Yen, Hsing-Jien Kung

Supplementary Figure 1-4

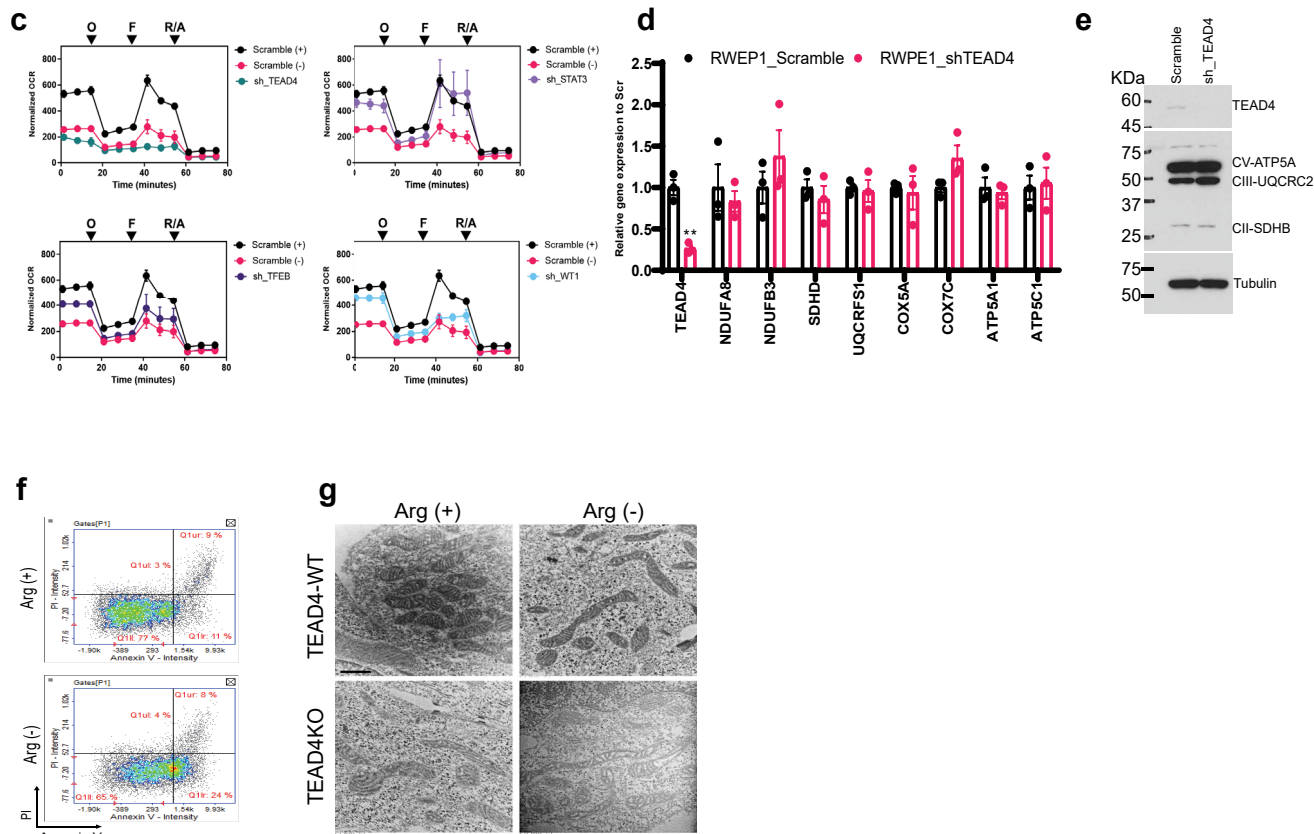
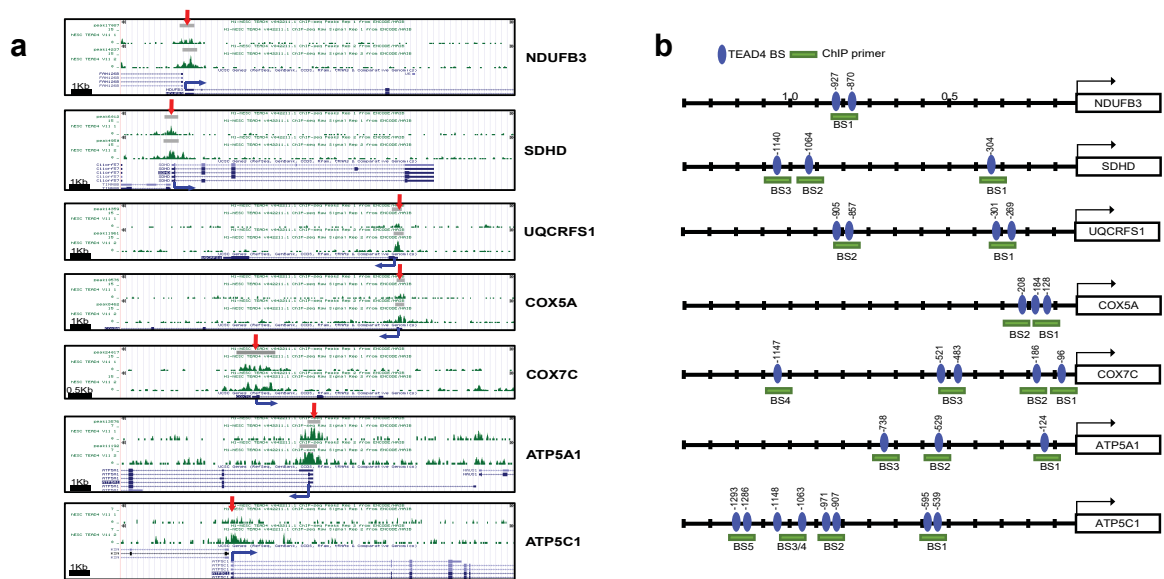
Supplementary Table 1-4

Supplementary reference

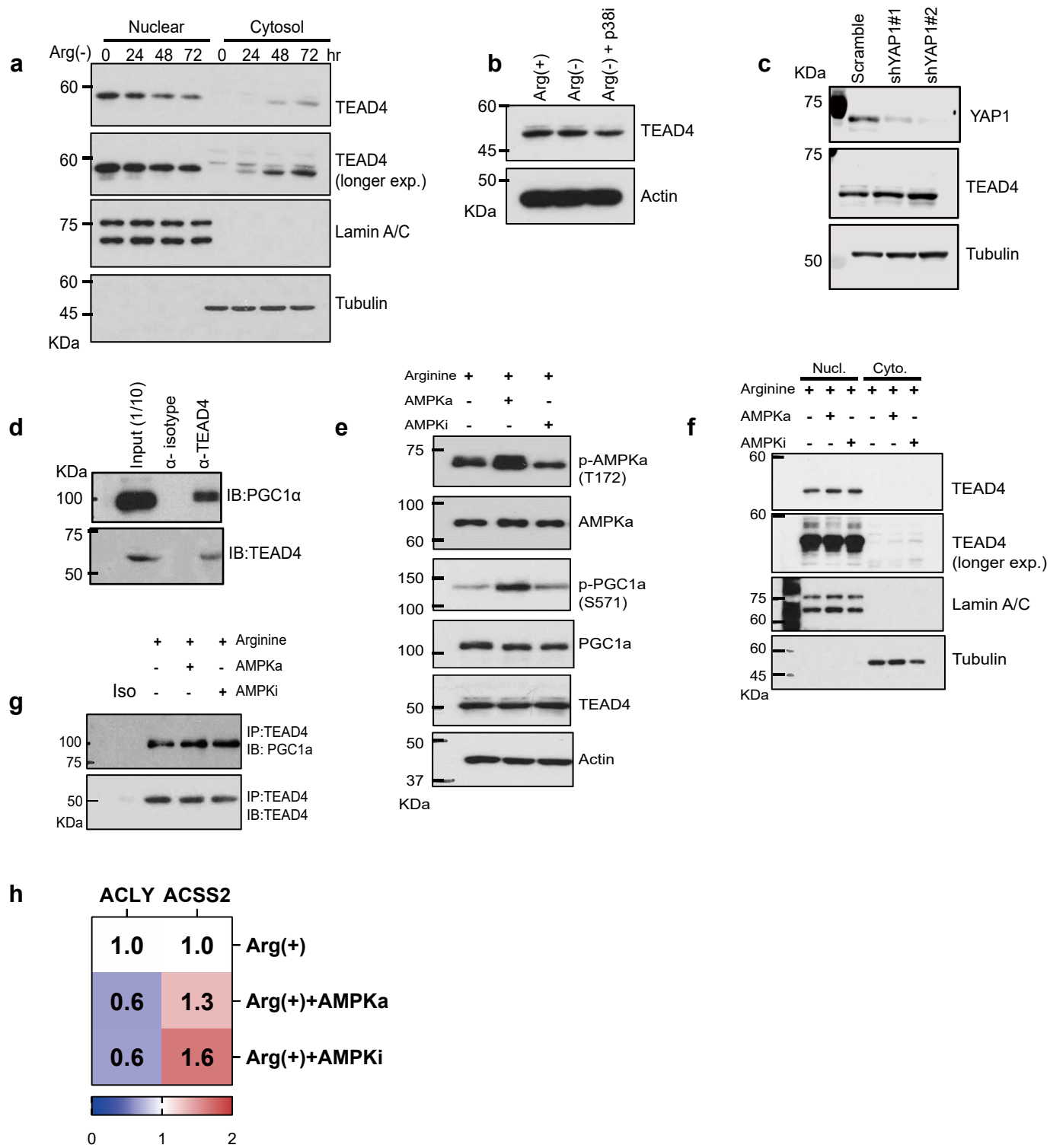


Supplementary Figure 1 Arginine epigenetically modulates mitochondrial OXPHOS genes

(a) Heat map of Affymetrix microarray analysis shows arginine (Arg+) globally induced metabolic pathways, including oxidative phosphorylation pathway, glucose, fatty acid and DNA metabolic pathways in PC3 prostate cancer cells. The color key indicates the fold change to control. (b) Gene set enrichment assay (GSEA) of PC3 microarray data was performed using hallmark gene set collection. Normalized enrichment scores (NES) for pathways indicates the significantly difference in arginine stimulation group. (c) GSEA shows the genes associated with oxidative phosphorylation (upper panel) and mTORC1 pathway (lower panel) are enriched in arginine stimulation Arg (+) group. (d) Real-time PCR analysis of OXPHOS genes expression after silencing of TEAD4 *via* shRNA in PC3 cells. (e) Seahorse assay for mitochondrial respiration activities after silencing TEAD4 *via* shRNA in PC3 cells. O: oligomycin, F: FCCP, R/A: rotenone /antimycin. (f) Advance image cytometer analysis shows silencing of TEAD4 increased cellular ROS production in PC3 cells. (g) Cell number after silencing of TEAD4 by shRNA in CWR22Rv1 and PC3 cells at different time points in the presence of arginine. Data are presented as mean values + SEM of independent experiments (n=3 in d-g). * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, **** $p < 0.0001$, ns: not significant, using unpaired two-tailed Student's t test. Source data are provided as a Source Data file.



Supplementary Figure 2 Arginine targets TEAD4 to modulate the mitochondrial functions (a) TEAD4 ChIP-seq distribution at representative OXPHOS genes loci from ENCODE database (ENCFF000OWA and ENCFF000OWD), visualized by UCSC Genome Browser (<https://genome.ucsc.edu>). (b) Predication of TEAD4 binding sites (BS in blue) on OXPHOS promoter regions by FIMO program. The green box indicates the target sites of ChIP-qPCR. (c) Individual shRNA plot for seahorse assay in Figure 3c. (d) TEAD4 was knock-down in non-arginine auxotrophic epithelial prostate cells, RWPE1, *via* lentivirus shRNA infection for 24 hours, following antibiotics selection for one week. OXPHOS genes expression was determined by quantitative real-time PCR and the protein level was evaluated by immunoblot (e). (f) Apoptosis analysis by double staining of annexin V and propidium iodide (PI) staining after arginine deprivation for 48-hours. (g) Ultrastructure of mitochondrial morphology after 48-hours arginine deprivation in wild type (WT) or TEAD4 knock-out (TEAD4KO) cells (scale bar=500nm). Data are presented as mean values + SEM of independent experiments (n=3 in d). ** $p < 0.01$, using unpaired two-tailed Student's t test. Source data are provided as a Source Data file.

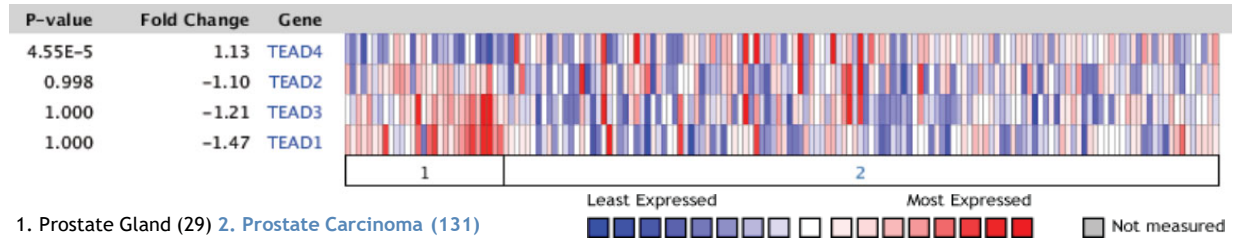


Supplementary Figure 3 Arginine mediated nuclear retention of TEAD4 (a) Immunoblotting shows TEAD4 location after arginine deprivation at different time pointed (related to figure 4e). (b) Immunoblotting of total TEAD4 expression after treatment (related to figure 4f). (c) YAP1 was knocked down by two individual YAP1 shRNA clones via lentivirus transduction for 24 hours, following antibiotics selection for one week. Total TEAD4 protein expression was evaluated by immunoblotting. (d) The interaction of TEAD4 with PGC1a was evaluated by immunoprecipitation. Total lysate was first incubated with isotype or TEAD4 antibody overnight and then pull-down by magnetic beads. Interaction was evaluated by PGC1a antibody. 10% of total protein as internal control. (e) In the presence of arginine, cells were treated with either AMPK activator, Metformin (10mM), or AMPK inhibitor, Dorsomorphin (10 μ M), for 24hours. The downstream target, phosphor-PGC1a (S571) was confirmed by immunoblot. However, the total level of TEAD4 was not affected. (f) The immunoblot of cell fractionation shows AMPK did not affection TEAD4 translocation in the presence of arginine. (g) The immunoprecipitation shows AMPK did not affect the interaction of TEAD4 with PGC1a in the presence of arginine. (h) The quantitative real-time PCR shows AMPK did not correlate with the expression of ACLY or ACSS2 in the presence of arginine. The color key indicates the relative gene expression to control. Source data are provided as a Source Data file.

a

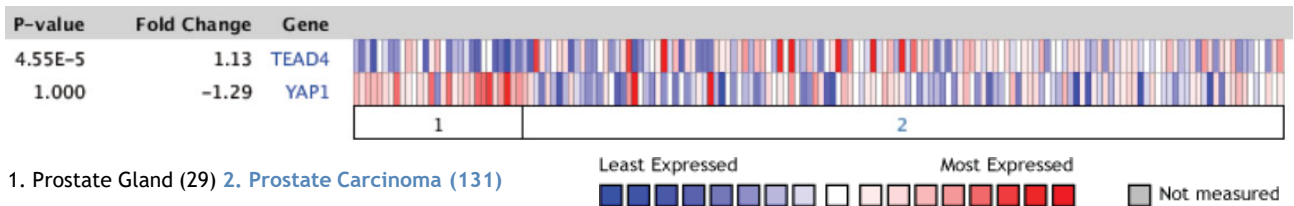
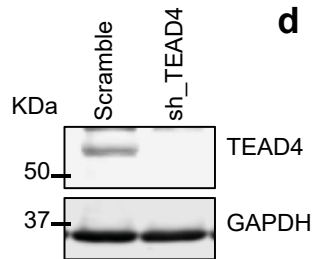
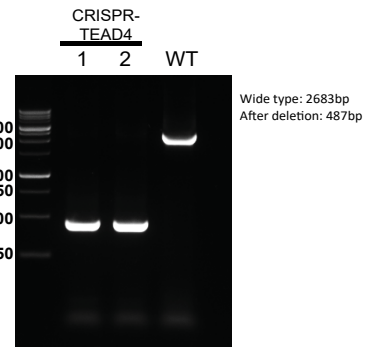
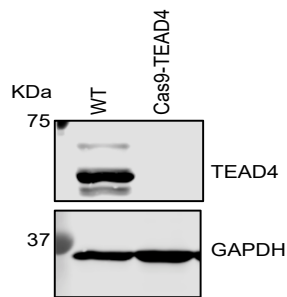
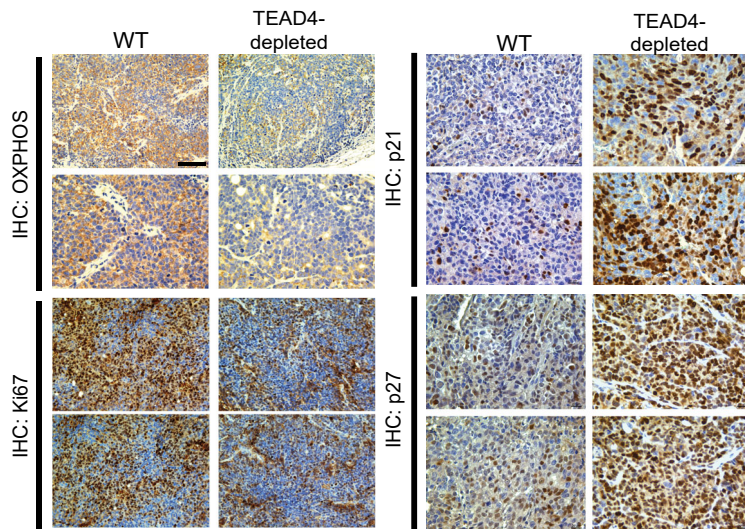
Comparison of TEAD1, TEAD2, TEAD3 and TEAD4 in Taylor Prostate 3

Over-expression in Prostate Carcinoma vs. Normal
log2 median-centered intensity

**b**

Comparison of TEAD4 and YAP1 in Taylor Prostate 3

Over-expression in Prostate Carcinoma vs. Normal
log2 median-centered intensity

**c****d****e**

Supplementary Figure 4 TEAD4 could be a potential target for cancer therapy (a) Oncomine database (www.oncomine.org) shows TEADs expression in Taylor Prostate cancer library. (b) Oncomine database shows TEAD4 and YAP1 expression in Taylor prostate cancer library. (c) Western blotting shows the knock-down efficiency of shRNA_TEAD4 CWR22Rv1 cells before subcutaneous injection. (d) Knock-out efficiency of CRISPR-Cas9 TEAD4 (left panel) CWR22Rv1 cells before subcutaneous injection as well as PCR confirmation (right panel). (e) IHC staining of pan-OXPHOS protein, CDK inhibitors, p21 and p27, and proliferation marker, Ki67 in mouse tumor samples (scale bar=50 μ m). Source data are provided as a Source Data file.

Supplementary Table 1 Target OXPHOS genes in TEAD4 ChIP-seq datasets from the ENCODE dataset

Complex	I		II	III	IV	V
Targets	NDUFA1	NDUFB1	SDHA	UQCR10	COX10	ATP5B
	NDUFA2	NDUFB2	SDHAF1	UQCR11	COX11	ATP5C1
	NDUFA3	NDUFB3	SDHAF2	UQCRB	COX14	ATP5D
	NDUFA4	NDUFB5	SDHAF3	UQCRC1	COX15	ATP5E
	NDUFA4L2	NDUFB6	SDHAF4	UQCRC2	COX16	ATP5O
	NDUFA5	NDUFB7	SDHAP1	UQCRFS1	COX17	ATP5F1
	NDUFA6	NDUFB9	SDHAP3	UQCRH	COX18	ATP5G1
	NDUFA7	NDUFB10	SDHB	UQCRHL	COX19	ATP5G2
	NDUFA8	NDUFB11	SDHC	UQCRQ	COX20	ATP5G3
	NDUFA9	NDUFC1	SDHD	CYC1	COX4I1	ATP5H
	NDUFA10	NDUFC2			COX5A	ATP5I
	NDUFA11	NDUFC2-			COX5B	ATP5J
	NDUFA12	KCTD14			COX6A1	ATP5J2
	NDUFA13	NDUFS1			COX6B1	ATP5J2-
	NDUFAB1	NDUFS3			COX6C	PTCD1
	NDUFAF1	NDUFS4			COX7A2	ATP5L
	NDUFAF2	NDUFS5			COX7A2L	ATP5L2
	NDUFAF3	NDUFS6			COX7B	ATP5S
	NDUFAF4	NDUFS7			COX7C	ATP5SL
	NDUFAF5	NDUFS8			COX8A	
	NDUFAF6	NDUFV1				
	NDUFAF7	NDUFV2				
		NDUFV3				

Supplementary Table 2 Primer list For ChIP-histone H3 acetylation on OXPPOS promoter region

Promoter region	Forward primer sequence (5' to 3')	Reverse primer sequence (5' to 3')
NDUFB3-pro	GAT GCT ACC TGG TTT ACG AGA G	GGA GGC AAA GTG CTA GAT GAA
NDUFS6-pro	GAA CCA CTG GGT ACT CCT AAT C	TGC AGC CTC ATG CAG TT
SDHD-pro	CTA ACT AGT CTC CCG TAC CCT T	GCT CAG GTG CCA GGT ATA TAA G
UQCRCF1-pro	CAA GAT GTG AAG CAG GGT AAT C	GAG CTC TGT GCA TCC TCT AA
COX5A-pro	TGT GGC TCA TGC CTG TAAT C	ACC ACG CCT GGC TAA TTT
COX7C-pro	CCC TTC TAG TGC TCC TTG ATA C	TTC CCA ACA TGG CTG AGT T
ATP5A1-pro	ACC ACA ACT CCC AGA AAT CC	CCG CCA CTT TAC TAG GAA CTC
ATP5C1-pro	GGT ACA GTG TAG GTG CTC AAT AA	AGA AAC AGG GTG GTT AGG ATT C

Supplementary Table 3 Primer list for ChIP-TEAD4 on OXPPOS promoter region

TEAD4 binding site	Forward primer sequence (5' to 3')	Reverse primer sequence (5' to 3')
NDUFB3_TEAD4_BS1	GGA GGA AGG GAT CTA CCA ACT A	GGG TCA AGG CTG ACA ACA T
SDHD_TEAD4_BS1	GGA CAG GAG AGC CAT AAC TTT G	ACC TTC CTT GGT GGT CCT
SDHD_TEAD4_BS2	TGA ACC AAC TAG GGA ACC AAT TA	ATG GAA GAA TTA CCT TGT CTA GGG
SDHD_TEAD4_BS3	AGA TCC TCG TCC CTA CAG ATT	TTG GTT CCC TAG TTG GTT CAC
UQCRCF1_TEAD4_BS1	GCGAT TACTA TGTGC TGGAC T	GGCCG ACTTT ACGAT CCTTT A
UQCRCF1_TEAD4_BS2	ACT CTA GAC TCA GTT CTC TGC T	TTG GCC TCA GTT TGA TCC TAA T
COX5A_TEAD4_BS1	ACA AGA TCA CGG TCG GTT G	CCT TGT CTC TGG TCA GGT G
COX5A_TEAD4_BS2	GCT GGG ATT ACA GGC ATG A	CCC AGG AGG GCA CTT AAT C
COX7C_TEAD4_BS1	CGA AAC TAC ATT TCC CAC AAT CC	CTT GAG GAA ACG CGA CAA AG
COX7C_TEAD4_BS2	AGG GAT GGG TTT ATT GTC CTT T	CCC GAA GGA TTG TGG GAA AT
COX7C_TEAD4_BS3	TTC CCT GCC CTT TGT TCT T	GCA CAA AGG ACT TCA GAG AGA
COX7C_TEAD4_BS4	GGC ATG GTT GTT CTC TTC CA	ATT GCC TCA CGT GTC TCA TTT A
ATP5A1_TEAD4_BS1	CCT GGG AAC TGC TGC TTA G	TGG TCC TGG CAT TCG TTA TAC
ATP5A1_TEAD4_BS2	AGC TGG GAA TCT TTA ACC TGA G	TCA GAC CCT CTG AGT CGA AA
ATP5A1_TEAD4_BS3	ATG ACT CTT CTA GGC CTC CTT	ACC TAG GTG ATG GGA TGA TAG T
ATP5C1_TEAD4_BS1	CCG GGC GTC TAG TAT TGC	CAG CCT GTG TTC TGC GA
ATP5C1_TEAD4_BS2	AGA CTC CTT GAG CCA CTC TAT	CTT ACC AAC CGT GTG GTC TT
ATP5C1_TEAD4_BS3	GCA TTC AAT CAA CCA CCT CTT T	TAG AGT GGC TCA AGG AGT CT
ATP5C1_TEAD4_BS4	CTT CCT TCA CTT GCA GAA ACA C	GTG CAG GCT AGT GAT GTG AT
ATP5C1_TEAD4_BS5	CAC CCA GTC TAC TGC ACT TTA T	TGC AAG TGA AGG AAG AGG TAT T

Supplementary Table 4 Primer list for qPCR assay

Genes	Forward primer sequence (5' to 3')	Reverse primer sequence (5' to 3')
NDUFA8	CCT TTA CCG GAG AAT CCC TAT C	CCA CGG ACC CAT CTT TAC TT
NDUFAB1	TAT AGC GAC ATG CCT CCT TTG	GAA AGC TTC TCT GGG TCA ATC T
NDUFB3	GGA CAT GAG CAT GGA CAT CAT A	GCC AGC TTC TTC TGG ATA GTT
NDUFS6	CAC ACT GGC CAG GTT TAT GA	GGG CTG CTC TGC TAT CAA AT
SDHD	CGA GAG GGT TGT CAG TGT TT	CCA GTG ACC ATG AAG AGT GAG
UQCRCF1	CCG ATA TTC CAG AAG GCA AGA A	TGG GTC CCT CAA CTG TGA TA
COX5A	GCT CGC TGG GTA ACA TAC TT	GGG CTC TGG AAC CAT ATC ATA G
COX7C	GTC CGT AGG AGC CAC TAT GA	GTG TAG CAA ATG CAG ATC CAA AG
ATP5A1	GAG GAC TGT GTG GTG CTA TTC	CTT TCC CAG CTG CTG TTA GT
ATP5C1	GAG GAC TGT GTG GTG CTA TTC	CTT TCC CAG CTG CTG TTA GT
ACSS1	GGT ACT GGG AGA CAG TAG AGA G	CCC AGG CAT CAC CGT ATT T
ACSS2	GCT GCA GTC TTC TCA TCA CTA C	GAA ACC CTT CTC CTG ACA CTT C
ACLY	AAC ATC CTG GCC TCC AAA C	CGT GGT CCG AGA GAT GAT ATT G
HAT1	TAT TGC TGG TAG CCT GTC AAC	CCT CAA CAT CAT CTG CCT CTA C
KAT2A	GGG TTT CTC CAA GGA CAT CAA	GAT TCA GCT CAC ACT CCA TCA G
KAT2B	GCC GTG TTA TTG GTG GTA TCT	TAG CCC TTG ACT TGC TCA TTT
KAT3A	GGA CCC AGA GTC ATT ACC TTT C	GAG AGG TCC ATG GGA TTC TTT AC
KAT3B	GCA GCA ACA GGT GCT TAG TA	CAG GGA TGG GTT GTG GAT TAG
KAT5	TCC TCC AGG CAA TGA TAT TTA C	AGA CAC AGG TTC TGG GAA TAA C
KAT6A	ATG GAT GTG CCT TCC GTA TC	TCG TAA CTG CTT GGG TTC TC
KAT6B	AAA GGG ACA CCC GAG TTA TG	CTA GAG GAG GAG GAG TGA GAA A
KAT7	CTG ACC CAA GGA GTT CTG TTA TG	AGC ACT AGT ACC ACG CTT AGA
KAT8A	CGA TCA CCA AGG TGA AGT ATG T	CTT GGG CTG TTT CCC ATA GT
KAT9	CCA AAC GTG GGA CTA GAA AGA G	CGA ATC ACC AGG GTA GGA TAG A
TEAD1	CTG AGT CGC AGT TAC CAC CA	AGC CTG GAG CCT TTT CAA G
TEAD2	ACA TGA TGA ACA GCG TCC TG	CAG CAG TTC CTG GGT GTC TC
TEAD3	CAT CGA GCA GAG CTT CCA G	CGT GCA ATC AAC TCA TTT CG
TEAD4	GCC TTC CAC AGT AGC ATG G	AAA GCT CCT TGC CAA AAC C
YAP1	GCA AAT TCT CCA AAA TGT CAG G	CGG GAG AAG ACA CTG GAT TT

Supplementary references for Bioinformatic packages

ChIP-seq analysis¹⁻⁸ and Motif analysis⁹⁻¹¹

1. Bolger, A.M., Lohse, M. & Usadel, B. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30, 2114-2120 (2014).
2. Langmead, B., Trapnell, C., Pop, M. & Salzberg, S.L. Ultrafast and memory-efficient alignment of short DNA sequences to the human genome. *Genome Biol* 10, R25 (2009).
3. Langmead, B. & Salzberg, S.L. Fast gapped-read alignment with Bowtie 2. *Nat Methods* 9, 357-359 (2012).
4. Zhang, Y. et al. Model-based analysis of ChIP-Seq (MACS). *Genome Biol* 9, R137 (2008).
5. Feng, J., Liu, T., Qin, B., Zhang, Y. & Liu, X.S. Identifying ChIP-seq enrichment using MACS. *Nat Protoc* 7, 1728-1740 (2012).
6. Ross-Innes, C.S. et al. Differential oestrogen receptor binding is associated with clinical outcome in breast cancer. *Nature* 481, 389-393 (2012).
7. Yu, G., Wang, L.G. & He, Q.Y. ChIPseeker: an R/Bioconductor package for ChIP peak annotation, comparison and visualization. *Bioinformatics* 31, 2382-2383 (2015).
8. Ramirez, F. et al. deepTools2: a next generation web server for deep-sequencing data analysis. *Nucleic Acids Res* 44, W160-165 (2016).
9. Bailey, T.L. DREME: motif discovery in transcription factor ChIP-seq data. *Bioinformatics* 27, 1653-1659 (2011).
10. Grant, C.E., Bailey, T.L. & Noble, W.S. FIMO: scanning for occurrences of a given motif. *Bioinformatics* 27, 1017-1018 (2011).
11. Machanick, P. & Bailey, T.L. MEME-ChIP: motif analysis of large DNA datasets. *Bioinformatics* 27, 1696-1697 (2011).

Ingenuity of pathway analysis (IPA)¹², Oncomine¹³, ENCODE^{14, 15} and GSEA¹⁶

12. Kramer, A., Green, J., Pollard, J., Jr. & Tugendreich, S. Causal analysis approaches in Ingenuity Pathway Analysis. *Bioinformatics* 30, 523-530 (2014).
13. Rhodes, D.R. et al. ONCOMINE: a cancer microarray database and integrated data-mining platform. *Neoplasia* 6, 1-6 (2004).
14. Consortium, E.P. An integrated encyclopedia of DNA elements in the human genome. *Nature* 489, 57-74 (2012).
15. Davis, C.A. et al. The Encyclopedia of DNA elements (ENCODE): data portal update. *Nucleic Acids Res* 46, D794-D801 (2018).

16. Subramanian, A. et al. Gene set enrichment analysis: a knowledge-based approach for interpreting genome-wide expression profiles. *Proc Natl Acad Sci U S A* 102, 15545-15550 (2005).