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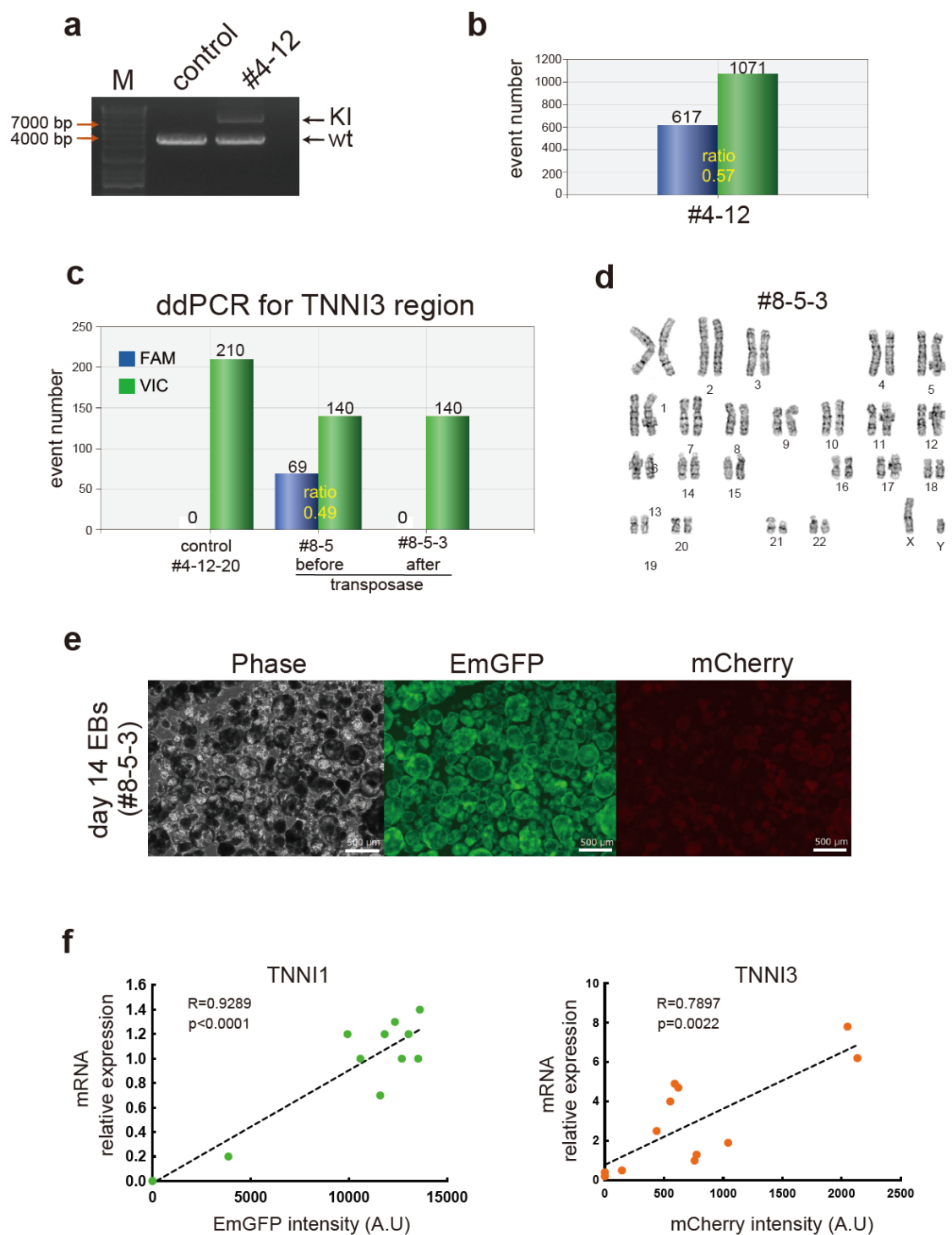
Supplementary Table 1

Set of nucleases and gRNAs.

Supplementary Table 2

19 Primer sets for ddPCR and PCR.

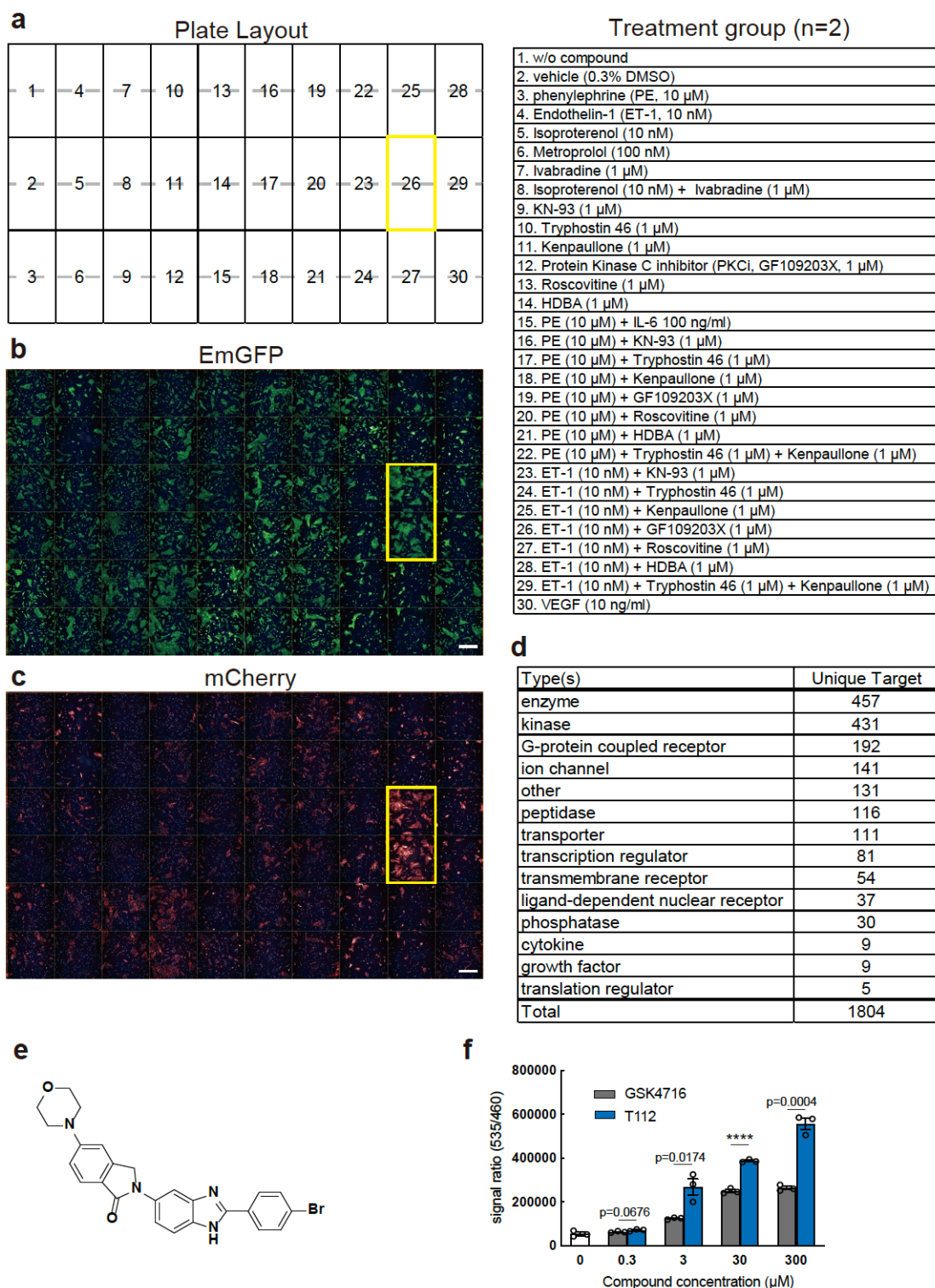
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Supplementary Figure 1 Generation of double reporter hiPSC lines.

a, PCR analysis of the TNNI1 targeted locus indicated in Figure 1A, black arrows. The primer sets used are listed in Supplementary Table 2. **b**, ddPCR of the TNNI1-EmGFP knockin clones before the removal of the transposons. **c**, ddPCR of the TNNI3-mCherry knockin clones before and after treatment with piggyBac

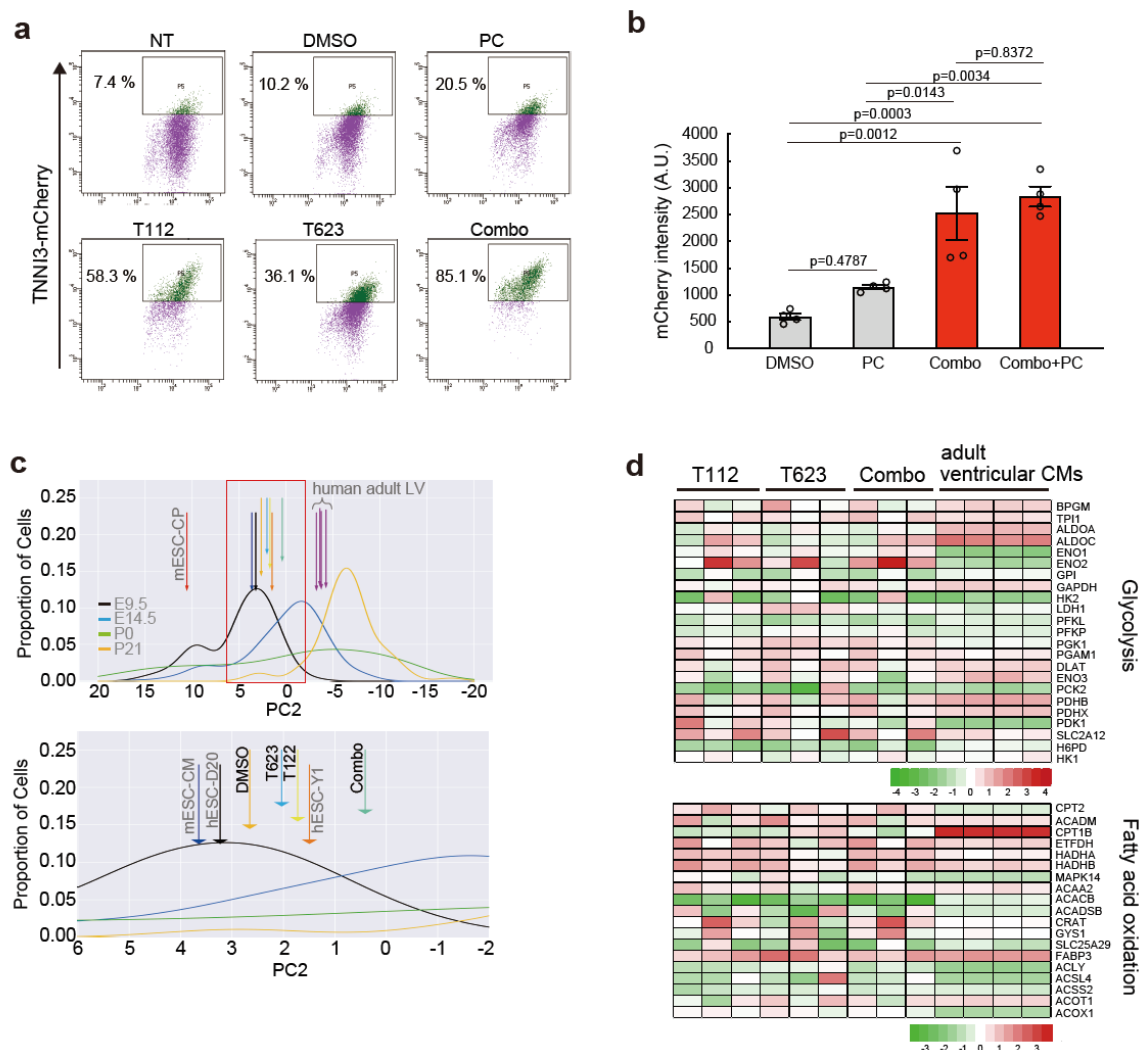
transposase. **d**, Karyotype of TNNI1-EmGFP knockin clone #8-5-3. **e**, Phase-contrast and EmGFP and mCherry fluorescence images of day-14 EBs. Scale bars: 500 μ m. **f**, The relationship between TNNI1 expression and EmGFP intensity (**left**) and between TNNI3 expression and mCherry intensity (**right**). Pearson correlation was used to analyze the correlation between gene expressions and fluorescence intensities, and t-test (two-tailed) was used to test the null hypothesis: $r=0.9289$, $p<0.0001$ (left) and $r=0.7897$, $p=0.0022$ (right). Samples: day (d)0 hiPSCs, d3-no treatment (NT), d6-NT, d8-NT, d10-NT, d10-DMSO, d10-T112, d16-NT, d16-DMSO, d16-T112, d16-T623 and d16-Combo. Each dot indicates the mean of each sample. $n=3$ independent experiments per sample. The expression level of d16-DMSO = 1.0.



Supplementary Figure 2 Compound screening information.

a, left: Plate layout of the assay for determining the positive control. **right:** List of factors used in the assay. **b,** EmGFP images of the assay after treatment. Scale bars: 500 μm. **c,** mCherry images of the assay after treatment. Scale

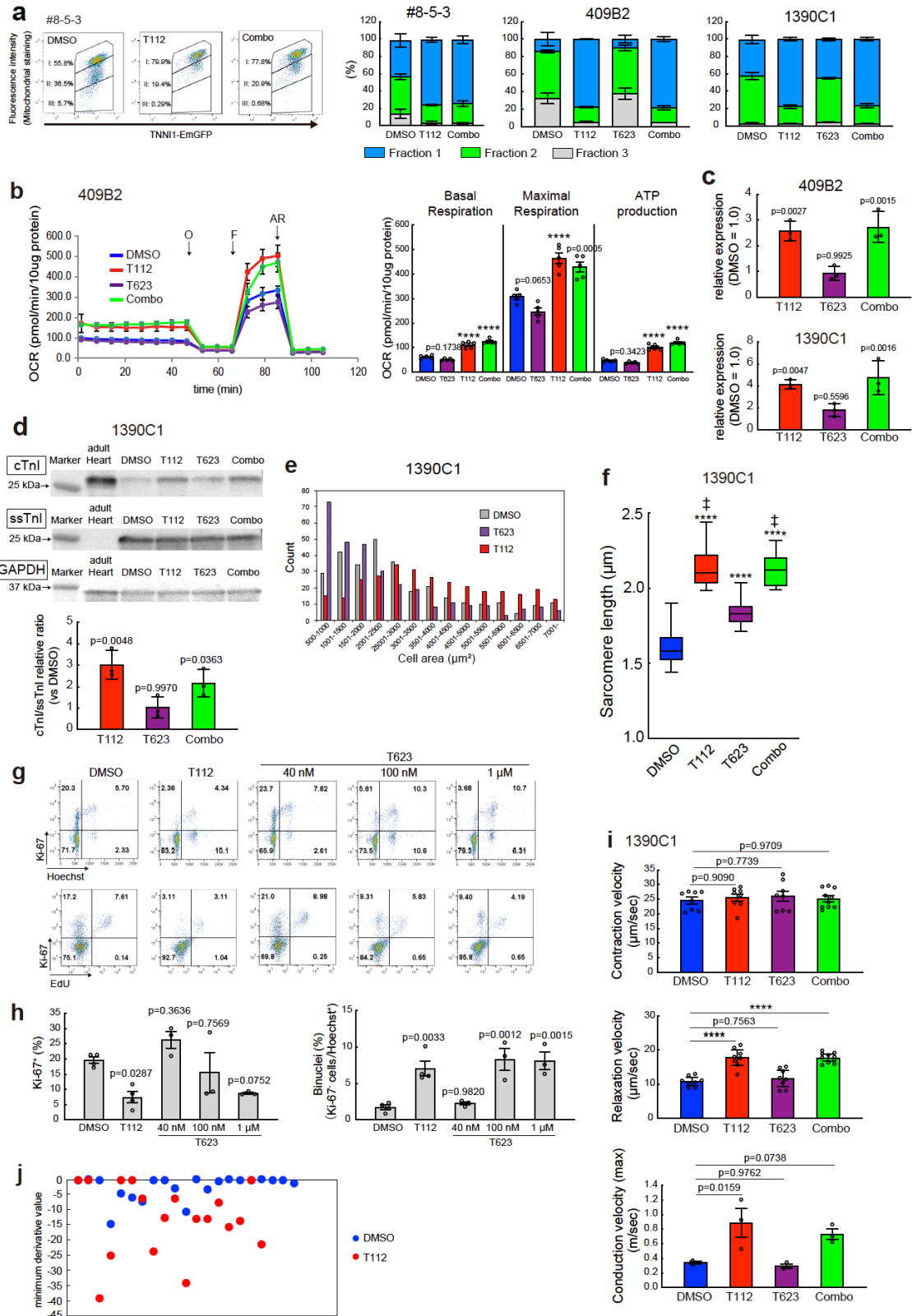
bars: 500 μm . **d**, Our biologically annotated compound library. **e**, The chemical structural formula of T112, 2-[2-(4-bromophenyl)-1H-benzimidazol-5-yl]-5-(morpholin-4-yl)-2,3-dihydro-1H-isoindol-1-one. **f**, The TR-FRET PGC1 α binding assay using T112 and GSK4716. n=3 independent experiments per group. Statistical analysis was done using unpaired two-tailed t-test. ****P < 0.0001.



Supplementary Figure 3 Expression profiles related to cardiac maturation in hiPSC-CMs.

a, Representative flow cytometry images of the TNNI3-mCherry positive percentage of hiPSC-CMs treated with DMSO, positive control (PC), T112, T623 or both (Combo). **b**, Flow cytometric analysis of hiPSC-CMs treated with DMSO, PC, Combo or Combo+PC. n=4 independent experiments per group. Data are the mean $\pm$ SEM. Statistical analysis was done using one-way ANOVA followed by Tukey's HSD test. **c**, top: Density plot (histogram) of the proportion of murine ventricular CMs at E9.5, E14.5, p0 and p21 based on previously published PCA¹ and transcriptional analyses of the datasets obtained in this study using day-16 hiPSC-CMs treated with DMSO (orange arrow), T623 (aqua arrow), T112 (yellow arrow) or T623 and T112 (Combo, green arrow) and public datasets of mESC-CP (red arrow), mESC-CM (blue arrow), hESC-D20 (black

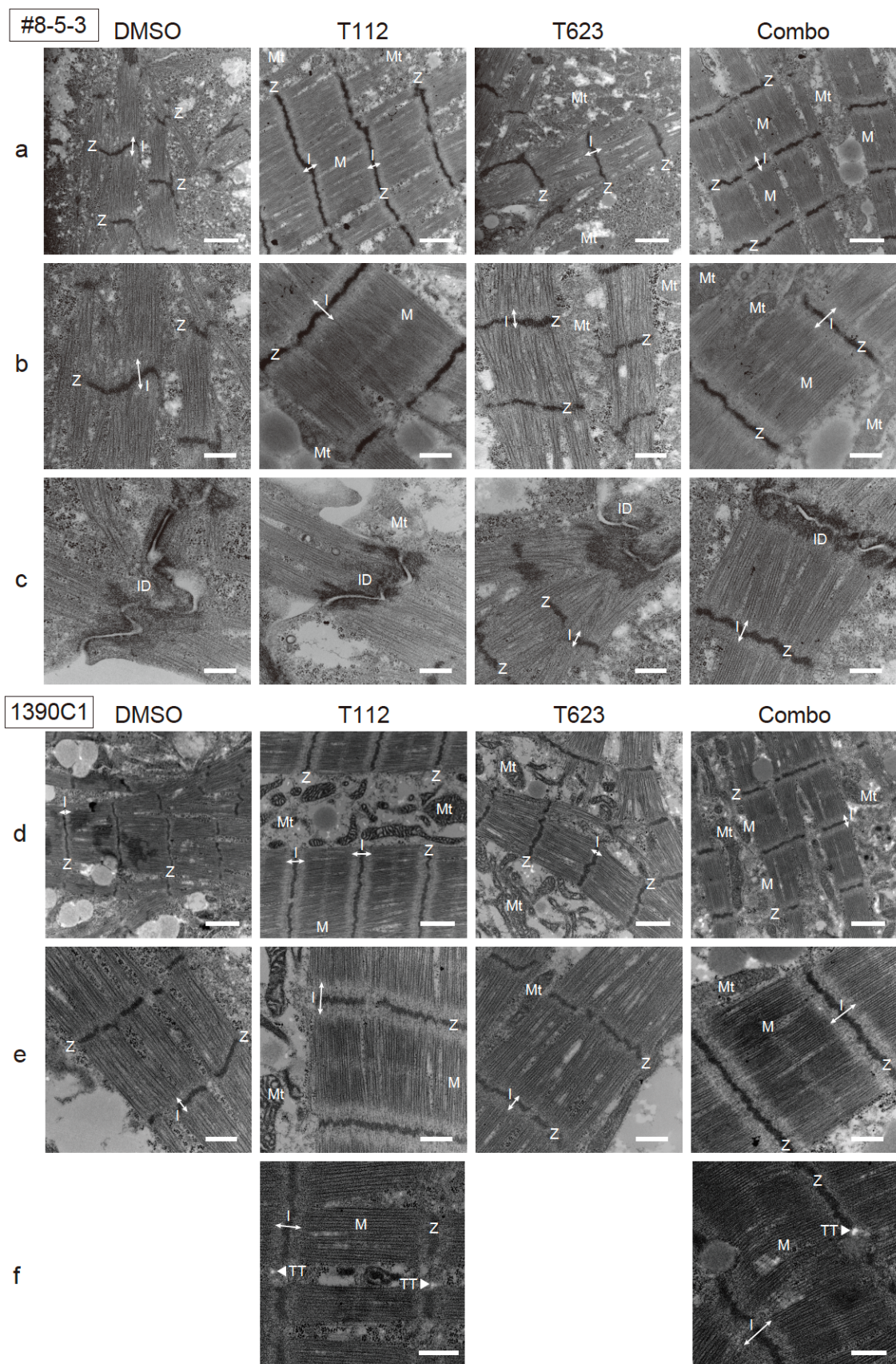
arrow), hESC-Y1 (brown arrow) and adult ventricular CMs (purple arrow)^{2, 3}.
bottom: Magnification of the red box in top. **d**, Heatmap of the expression
changes in glycolysis- and fatty acid oxidation-related genes. Data are
normalized to the DMSO-treated samples.



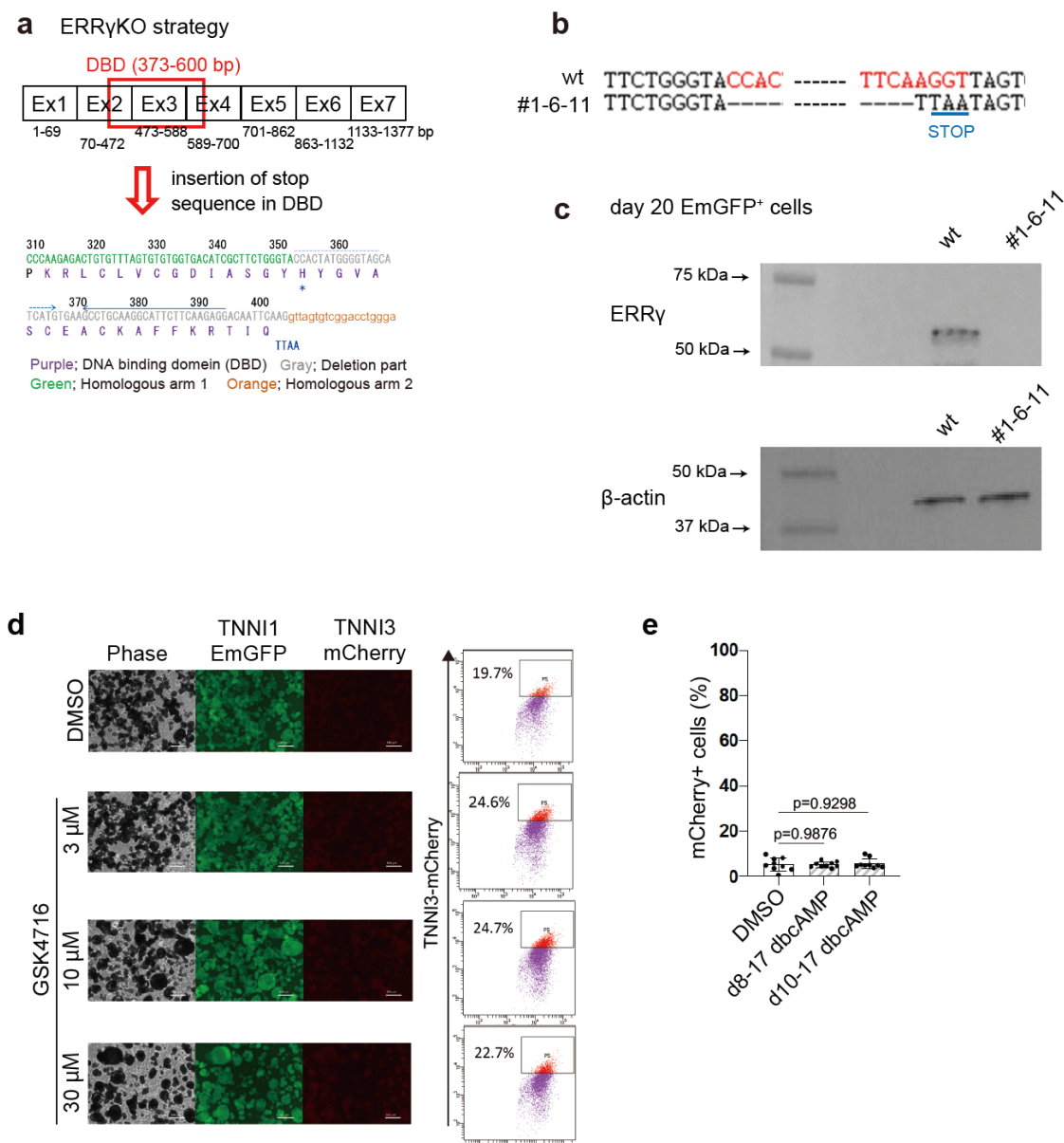
Supplementary Figure 4 T112 accelerates metabolic and sarcomere maturation and contractile properties.

a, left: Representative flow cytometry images of hiPSC (#8-5-3)-CMs treated with DMSO, T112 or Combo stained with mitochondrial dyes. **right:** Percentage of each fraction in each group of hiPSC-CMs in multiple hiPSC lines (#8-5-3, 409B2 and 1390C1). n=3 independent experiments per group. Data are the mean $\pm$ SEM. **b, left:** Mitochondrial respiration rates of hiPSC (409B2)-CMs treated with DMSO, T112, T623 or Combo. n=5 biologically independent samples per group. Data are the mean $\pm$ SEM. **right:** Basal and maximal OCR and ATP production. n=5 biologically independent samples per group. Statistical analysis was compared to DMSO using one-way ANOVA followed by Dunnett's test. ****P < 0.0001. **c,** TNNI3 expression level in hiPSC-CMs treated with DMSO, T112, T623 or Combo. **top:** 409B2, **bottom:** 1390C1. n=3 independent experiments per group. Data are the mean $\pm$ SEM. Statistical analysis was compared to DMSO using one-way ANOVA followed by Dunnett's test. **d, top:** Representative western blots of cTnI, ssTnI and GAPDH proteins in hiPSC (1390C1)-CMs treated with DMSO, T112, T623 or Combo. **bottom:** cTnI/ssTnI relative ratio. n=3 independent experiments per group. Data are the mean $\pm$ SEM. Statistical analysis was compared to DMSO using one-way ANOVA followed by Dunnett's test. **e,** Cell area of hiPSC (1390C1)-CMs treated with DMSO, T112 or T623 measured by high content imaging. For each group, n=300 cells over 3 independent experiments. **f,** Sarcomere length in hiPSC (1390C1)-CMs treated with DMSO, T112, T623 or Combo. Boxes represent 25th–75th percentiles; whiskers represent the minimum and maximum ranges; horizontal lines indicate the median values. n=15, one measurement per TEM image from 15 TEM images over 2 consecutive experiments; ****P < 0.0001 compared to DMSO, $\pm$ P < 0.0001 compared to T623 using one-way ANOVA followed by Tukey's HSD test. **g,** Representative flow cytometry images of day-16 cells stained with Ki-67, EdU and Hoechst. **h,** Percentage of Ki-67⁺ (**left**) and Ki-67⁻/Hoechst⁺ cells (**right**) in hiPSC (#8-5-3)-CMs treated with DMSO (n=4), T112 (n=4) or T623 (n=3). n=independent experiments per group. Data are the mean $\pm$ SEM. Statistical analysis was compared to DMSO using one-way ANOVA followed by Dunnett's test. **i,** Contractile properties of hiPSC (1390C1)-CMs treated with DMSO (n=8), T112 (n=8), T623 (n=8) or Combo (n=10). **top:** Contraction velocity. **middle:** Relaxation velocity. n=biologically independent samples examined over 3 independent experiments. **bottom:** Maximum conduction velocity. n=3 independent experiments. Data are the mean $\pm$ SEM. Statistical analysis was compared to DMSO using one-way ANOVA followed by

108 Dunnett's test. **** $P < 0.001$. j, The minimum derivative value of AP traces in
109 hiPSC (#8-5-3)-CM treated with DMSO (blue, n=21) or T112 (red, n=18) over 3
110 independent experiments.
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112



Supplementary Figure 5 Ultrastructural properties of hiPSC-CMs treated with DMSO, T112, T623 or Combo. **a-c**, #8-5-3. **d-f**, 1390C1. **a** and **d**, Typical low magnification TEM images. Scale bars: 1000 nm. **b**, **c**, **e** and **f**, Typical high magnification TEM images. Scale bars: 500 nm. I: I-band; ID: intercalated disc; M: M-line; Mt: Mitochondria; TT: T-tubule; Z: Z line.



Supplementary Figure 6 Targeted homologous recombination strategy using the CRISPR/Cas9 system and analysis of the cardiac maturation status.

a, The ERRyKO strategy. **b**, Sequence analysis of the targeted region. **c**, Western blotting of ERRy and β-actin in day-20 CMs derived from WT or KO (#1-6-11) clones. **d**, **left**: Fluorescence images of day-16 EBs treated with GSK4716. Scale bars: 500 μm. **right**: Representative flow cytometry images of hiPSC-CMs treated with DMSO or GSK4716. TNNI3-mCherry positive percentages are shown. **e**, Flow cytometric analysis of ERRy KO hiPSC-CMs treated with 0.5 mM dbcAMP from day 8 to 17 or from day 10 to 17. n=9 biologically independent samples examined over 3 independent experiments.

132 Data are the mean $\pm$ SEM. Statistical analysis was done using one-way ANOVA
133 followed by Dunnett's test.
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Supplementary Table 1. Set of nucleases and gRNAs.

	Nuclease	gRNA	
		name	sequence
TNNI1-EmGFP	Cas9 (D10A)	AS194	CTCTAGGTACCTCTATTGTGAGG
		S254	TGGGGTCCATCAGAGTCTAGAGG
TNNI3-mCherry	Cas9 VQR variant	S4	CAAGAAAAAGTTTGAGAGCTGA
ERR γ KO	Cas9 (D10A)	AS345	CCACTATGGGGTAGCATCATG
		S393	GCCTGCAAGGCATTCTTCAAGAGG

138 **Supplementary Table 2 Primer sets for ddPCR and PCR.**

ddPCR TNNI1	TNNI1out_F1021	GGGCGCTTCTCACCTACAGAT
	TNNIout_P1043 (VIC)	AAGGACCTGAAGCTGAA
	TNN1out_R1083	AACTTCCCACGGAGGTCCA
ddPCR TNNI3	TNNI3_HA2out_Fwd	AGCTAGTCAGCATCTGGCAATG
	TNNI3_HA2out_probe (VIC)	ATGGCTGCAATGGTT
	TNNI3_HA2out_Rev	CCGGCCTCAAGATAAGCAATAT
ddPCR donor vector	Donor_upTaqF345-363	GGCACTTGGCGCTACACAA
	Donor_TaqP368-382 (FAM)	CCTCTGGCCTCGCAC
	Donor_upTaqR404-385	CCTACCGGTGGATGTGGAAT
PCR TNNI1	TNNI1outF1	GGCTTCAAGGGTGGACAATTTAAGGTG
	TNNI1outR4	TGCAGCGCATGAATCCCTGACTTAC
PCR TNNI3	TNNI3 dPCR Fwd	GATGCCTGAAACCATGGATTG
	TNNI3 HA2 out R	CCGGCCTCAAGATAAGC
PCR ERRyKO	ESRRGoutF3	GAAACAAAGTGATTGAGGATATATTCCC
	HA2-nest-Rv	CCTCATCTATAAATTGAAGGCATT

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References

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2. Wamstad, J.A. *et al.* Dynamic and coordinated epigenetic regulation of developmental transitions in the cardiac lineage. *Cell* **151**, 206-220 (2012).
3. Kuppusamy, K.T. *et al.* Let-7 family of microRNA is required for maturation and adult-like metabolism in stem cell-derived cardiomyocytes. *Proc Natl Acad Sci U S A* **112**, E2785-2794 (2015).