

Supplementary material for

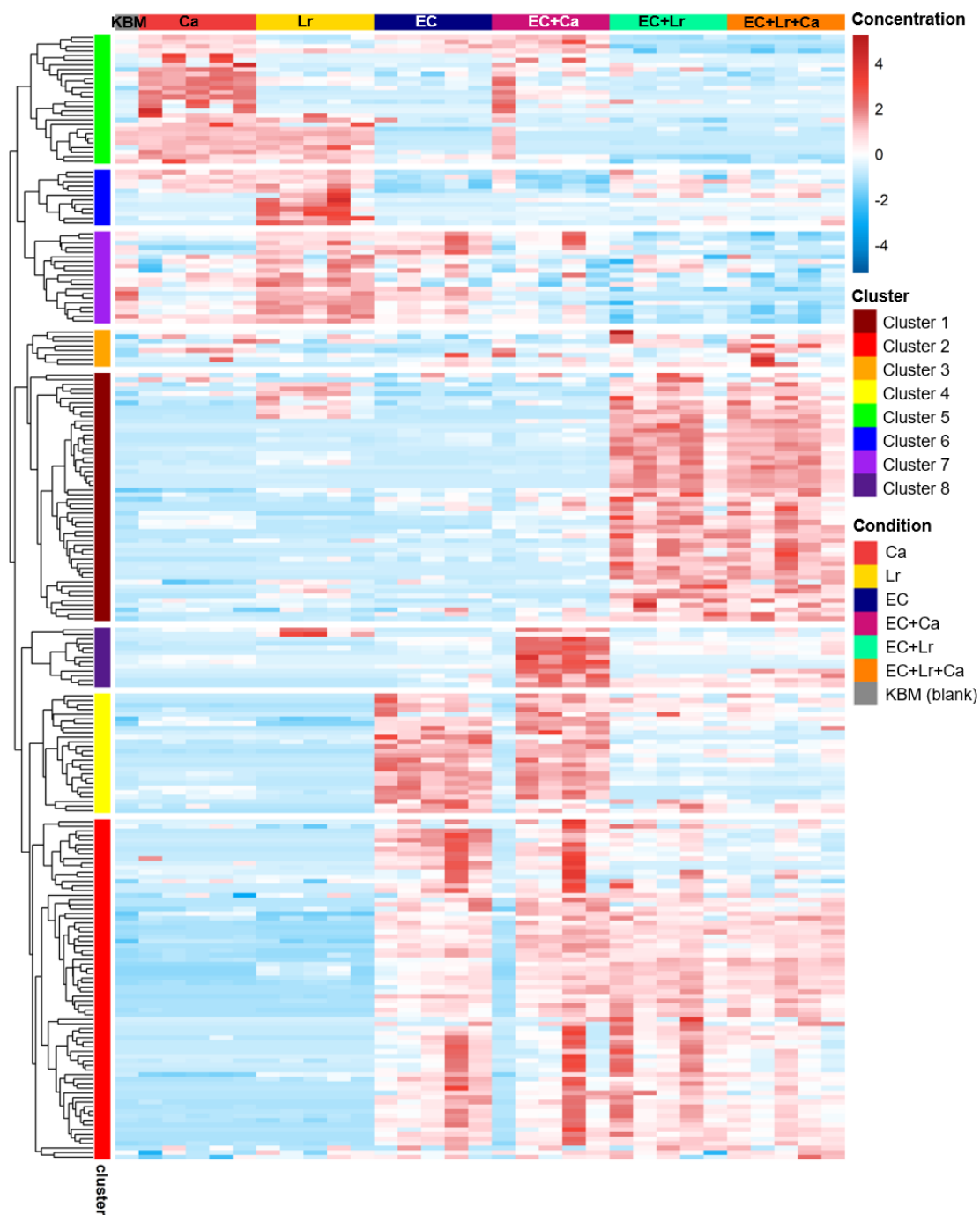
***Lactobacillus rhamnosus* colonisation antagonizes *Candida albicans* by forcing metabolic adaptations that compromise pathogenicity**

Raquel Alonso-Roman¹, Antonia Last¹, Mohammad H. Mirhakkak², Jakob L. Sprague¹, Lars Möller¹, Peter Großmann², Katja Graf^{1,3}, Rena Gratz¹, Selene Mogavero¹, Slavena Vylkova⁴, Gianni Panagiotou^{2,5}, Sascha Schäuble², Bernhard Hube^{1,6}, Mark S. Gresnigt⁷

Corresponding author: bernhard.hube@hki-jena.de

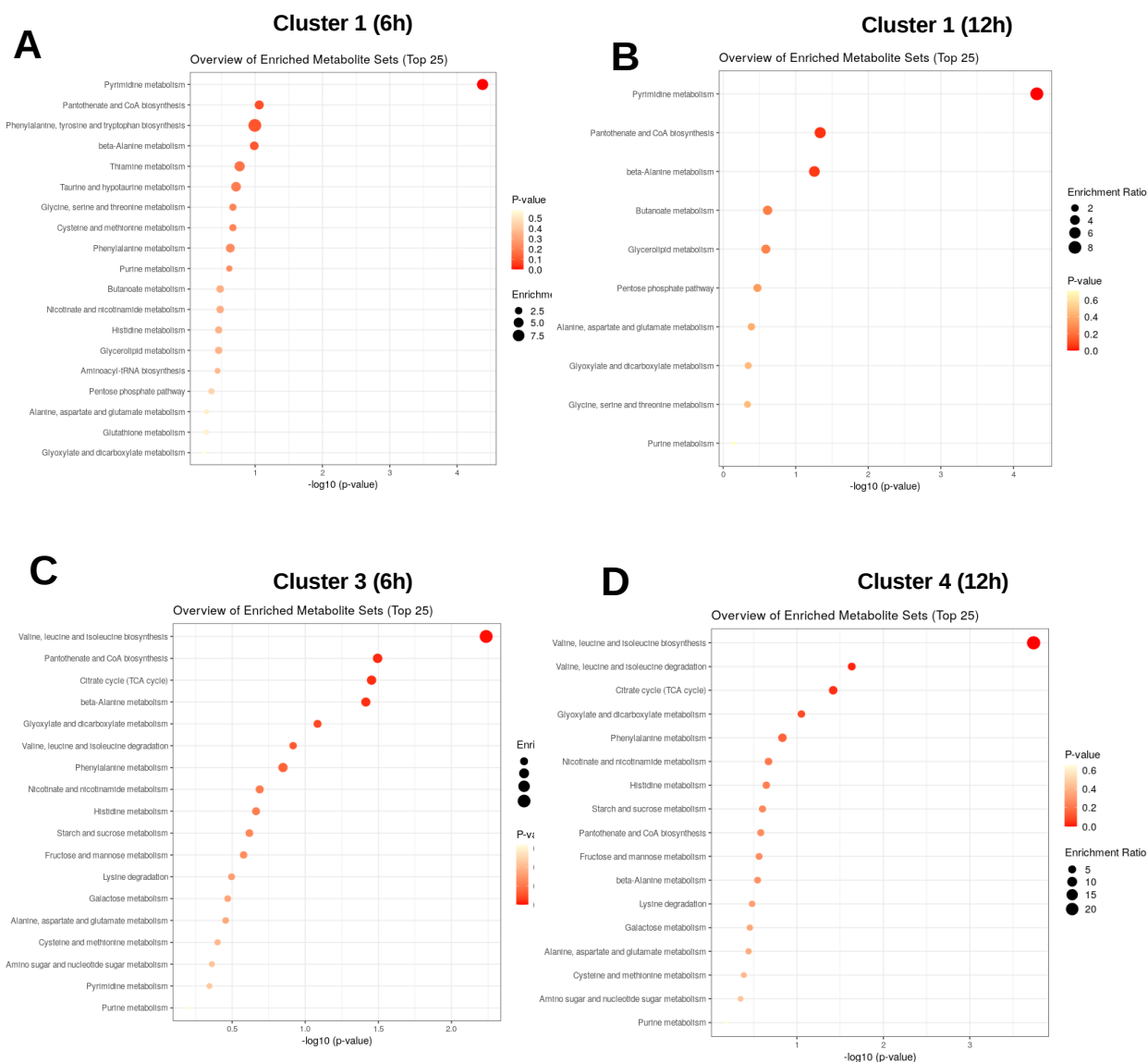
Affiliations:

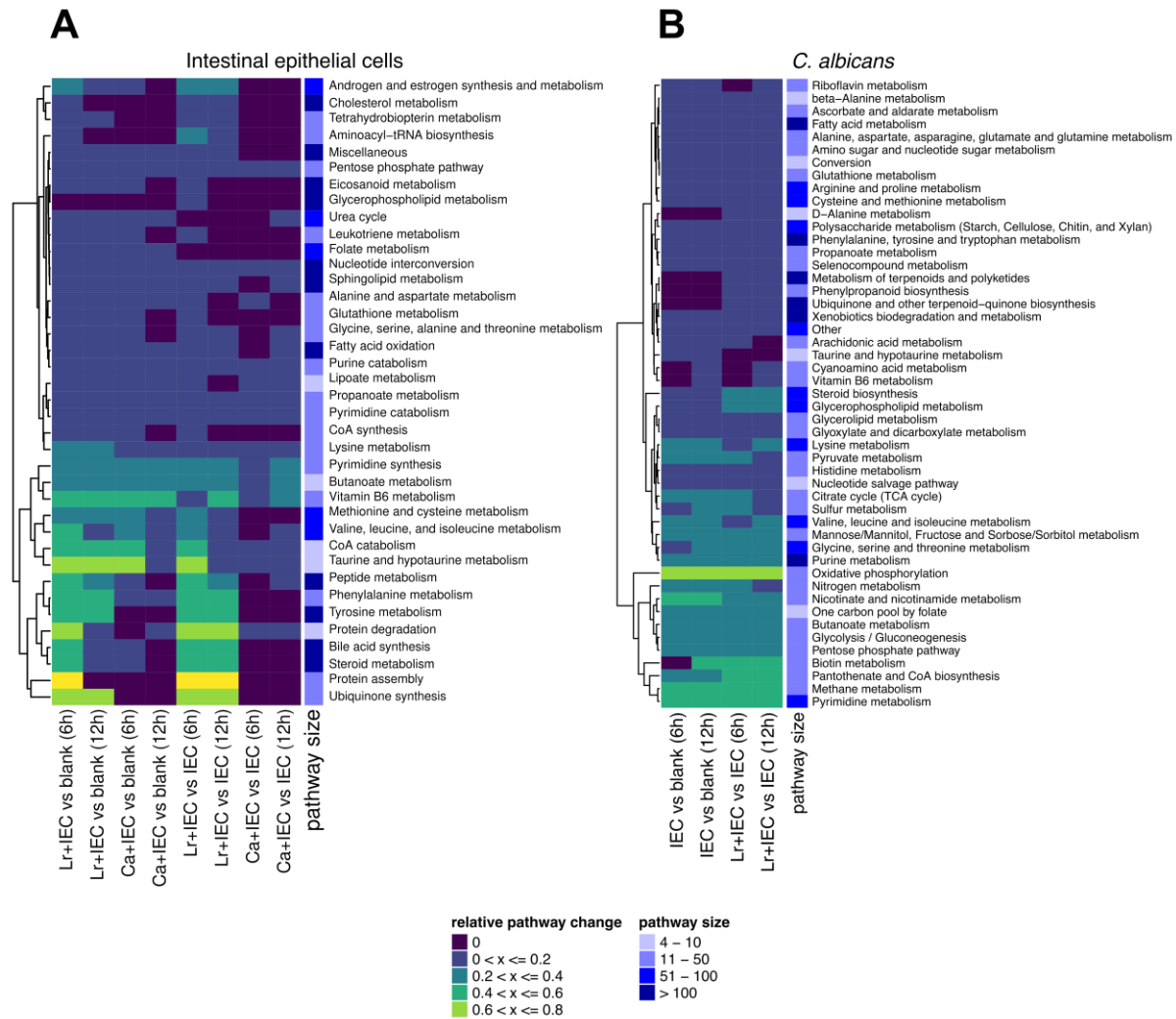
- 1: Department of Microbial Pathogenicity Mechanisms, Leibniz Institute for Natural Product Research and Infection Biology - Hans-Knoell-Institute, Jena, Germany
- 2: Systems Biology and Bioinformatics Unit, Leibniz Institute for Natural Product Research and Infection Biology - Hans-Knoell-Institute, Jena, Germany
- 3: Dynamic42 GmbH, Jena, Germany
- 4: Septomics Research Centre, Leibniz Institute for Natural Product Research and Infection Biology – Hans-Knoell-Institute, Jena, Germany
- 5: Department of Medicine and State Key Laboratory of Pharmaceutical Biotechnology, University of Hong Kong, China
- 6: Institute of Microbiology, Friedrich Schiller University, Jena, Germany
- 7: Junior Research Group Adaptive Pathogenicity Strategies, Leibniz Institute for Natural Product Research and Infection Biology - Hans-Knoell-Institute, Jena, Germany



Supplementary Fig. 1: Untargeted metabolomics of *in vitro* intestinal epithelial *L. rhamnosus* colonization and *C. albicans* infection

Hierarchical clustering of the presence and absence of metabolites in the *in vitro* model at 12 hours post infection (hpi).

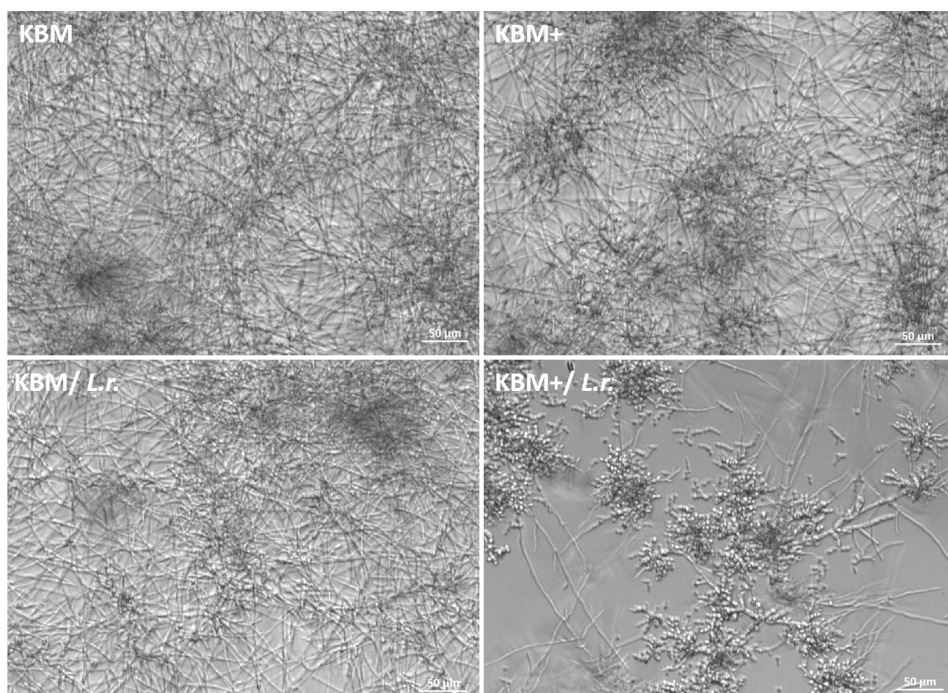




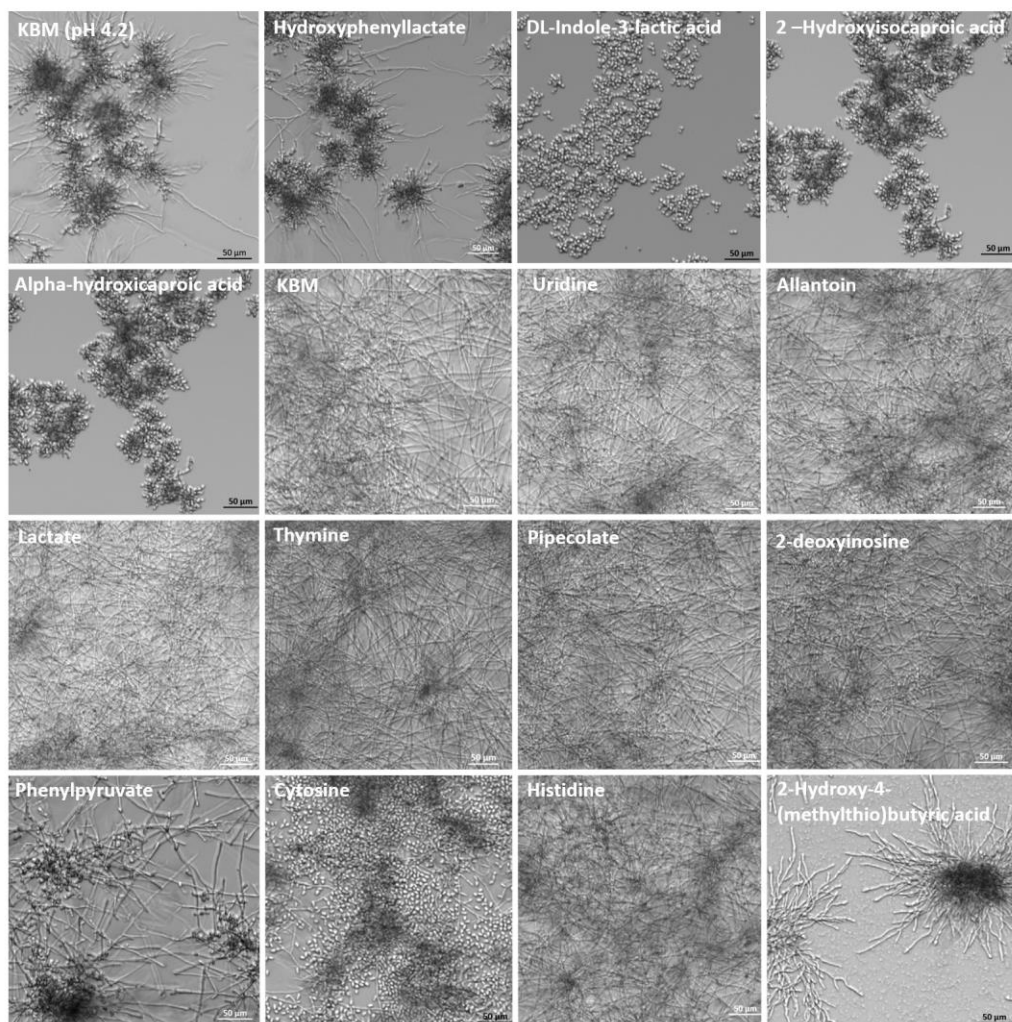
Supplementary Fig. 3: Pathway flux activity of IECs and *C. albicans*

Simulation of pathway activity levels in IECs (A) and in *C. albicans* (B) compared between different setups as indicated. Relative pathway change was determined by identifying the number of pathway-specific reactions for which feasible flux ranges differ according to flux variability analysis.

A

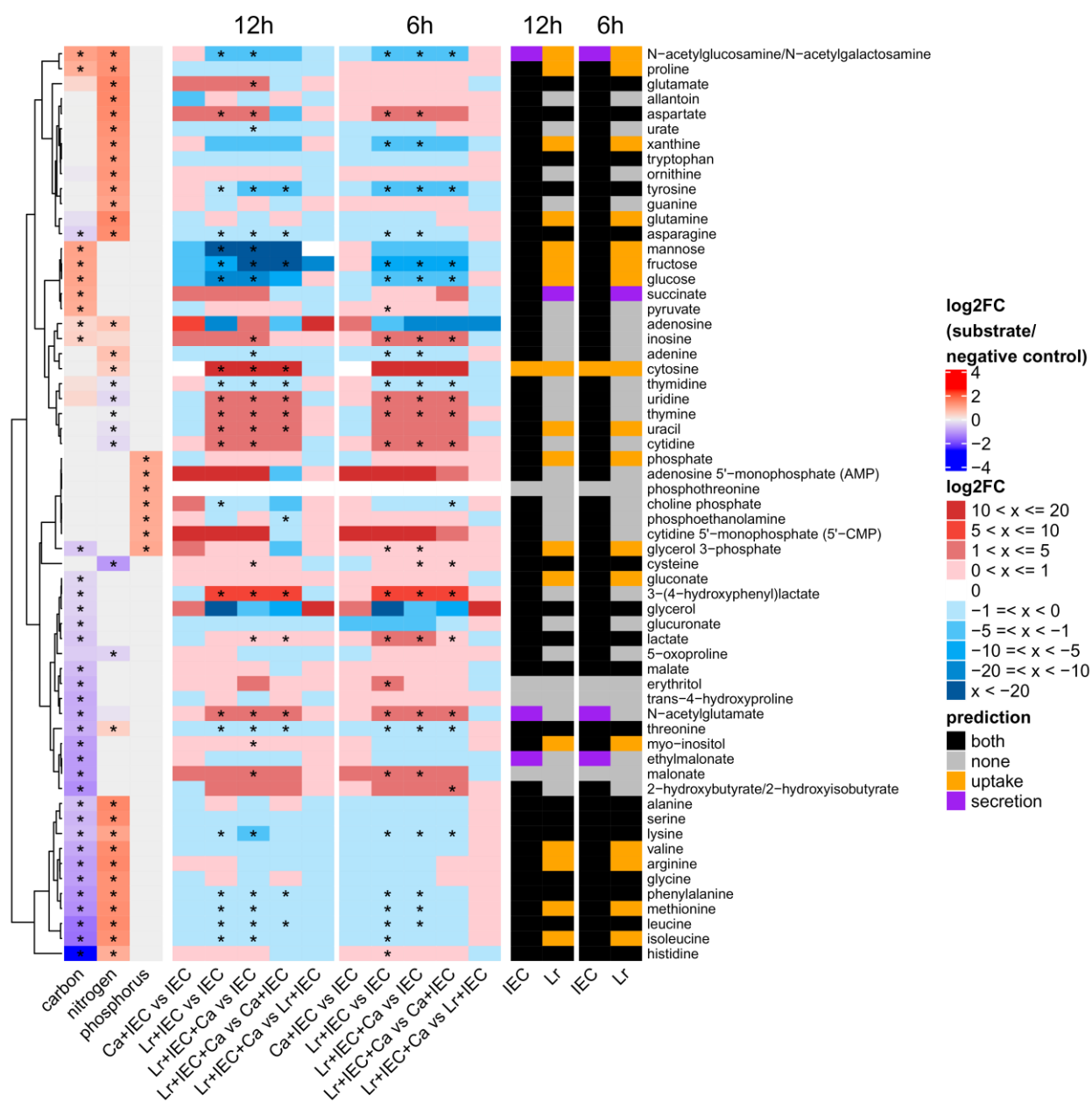


B



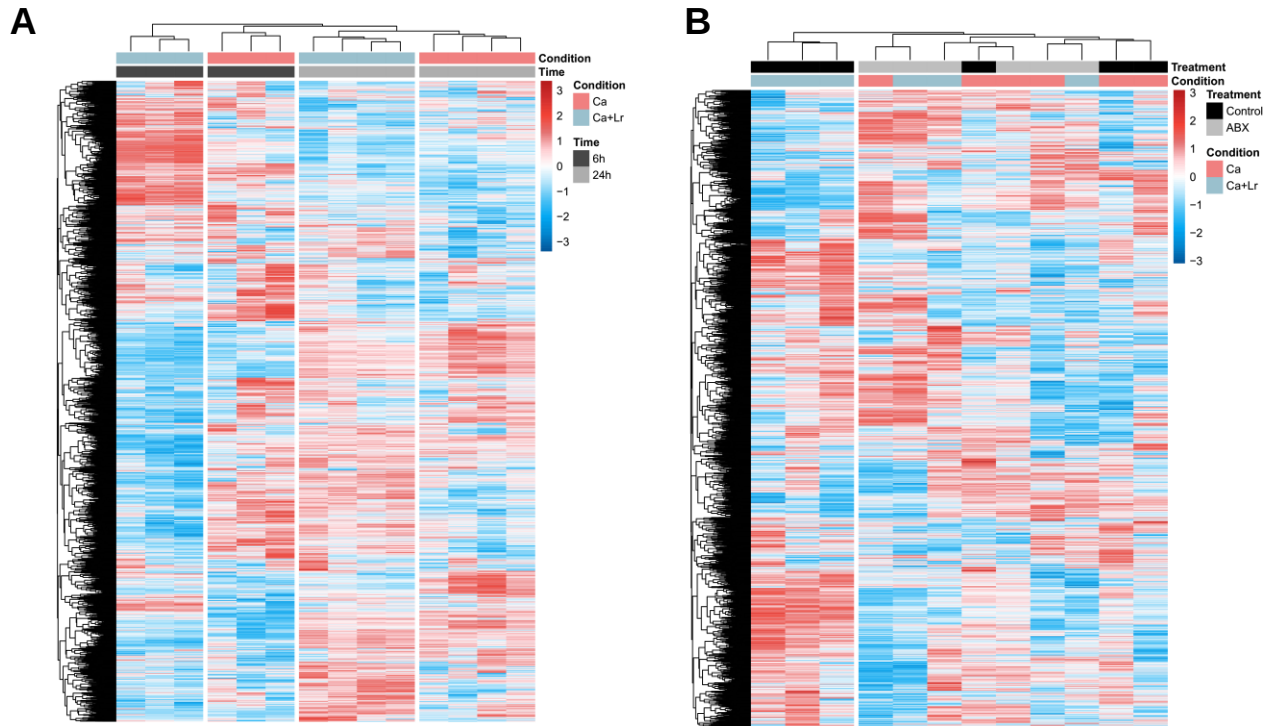
Supplementary Fig. 4: Impact of *L. rhamnosus*-colonization derived metabolites on *C. albicans*

(A) Representative images of *C. albicans* morphology changes grown in *L. rhamnosus*-conditioned or unconditioned media, after 20 h at 37°C with 5% CO₂ (n=3 biological replicates). (B) Representative images of *C. albicans* morphology grown in presence of all the metabolites tested, at neutral or acidic pH, after 20 h at 37°C with 5% CO₂ (n=2 biological replicates).



Supplementary Fig. 5: *L. rhamnosus* colonization creates an unfavourable metabolic environment for *C. albicans*

Phenotypic microarray growth experiments for wild-type *C. albicans* on a carbon, nitrogen or phosphorous source (left). Metabolome data measured at 6 and 12 h as well as metabolic modelling predictions (right) are indicated for the whole panel of metabolites. For metabolic modelling, media was adapted from metabolome data derived from IECs spent media or blank. Uptake or secretion was determined by identifying feasible flux ranges for metabolite-specific exchange reactions alongside optimization for biomass. ANOVA (two-sided) was performed for phenotypic microarrays, Wilcoxon test (two-sided) for metabolomics, with FDR correction (* = $p_{adj} \leq 0.05$).



Supplementary Fig. 6: Changes in *C. albicans* gene expression upon infection of *L. rhamnosus* colonized IECs

(A) Hierarchical clustering based on Euclidean distance of *C. albicans* gene expression at 6 and 24 h during *in vitro* infection of IECs in the presence and absence of *L. rhamnosus* colonization. Data summarized from n=3 and n=4 independent experiments at 6 and 24 hpi respectively. (B) Hierarchical clustering based on Euclidean distance of *C. albicans* gene expression during *in vitro* infection of IECs in the presence and absence of *L. rhamnosus* colonization and in the presence and absence of antibiotics. Data summarized from n=3 at 24 hpi.

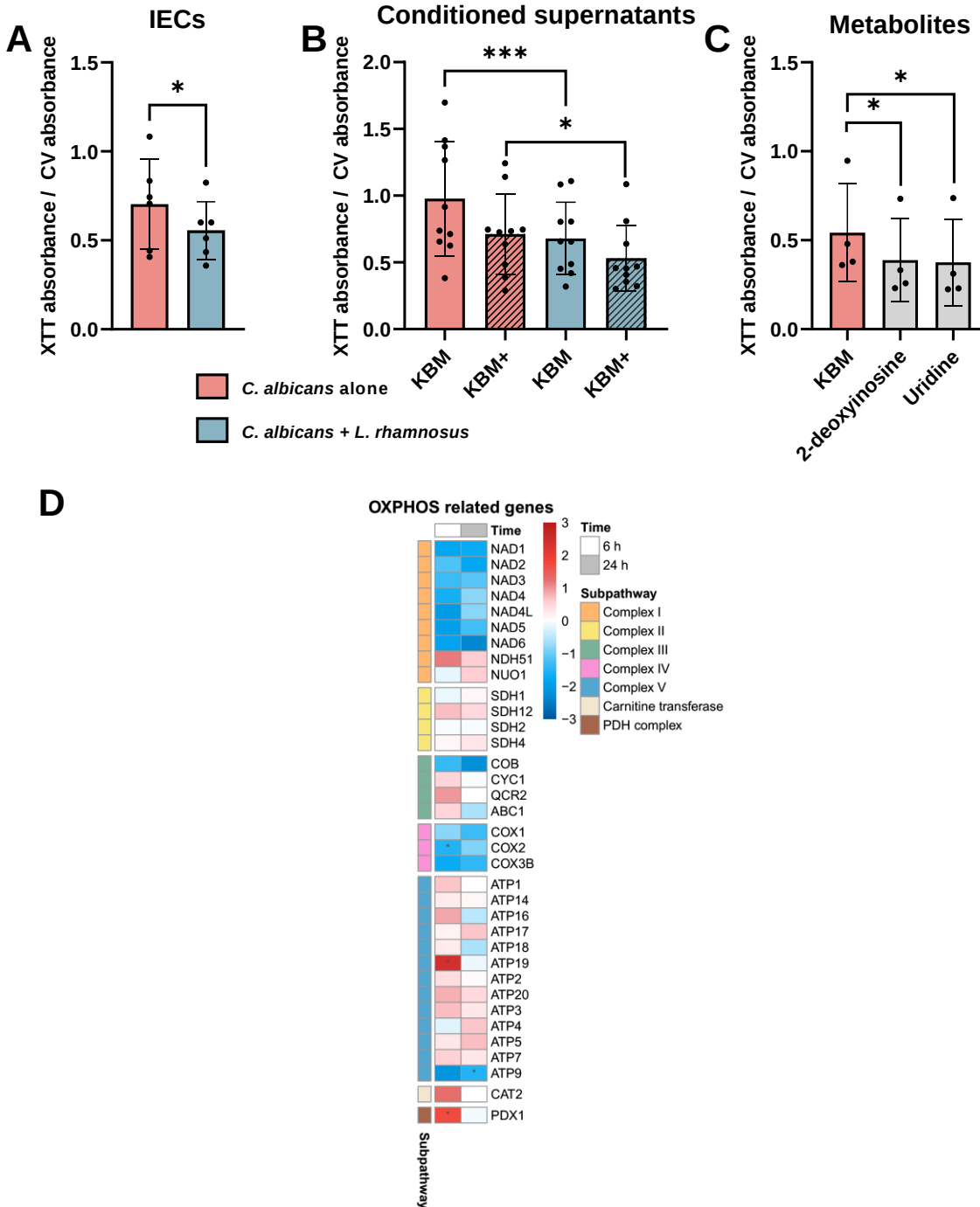
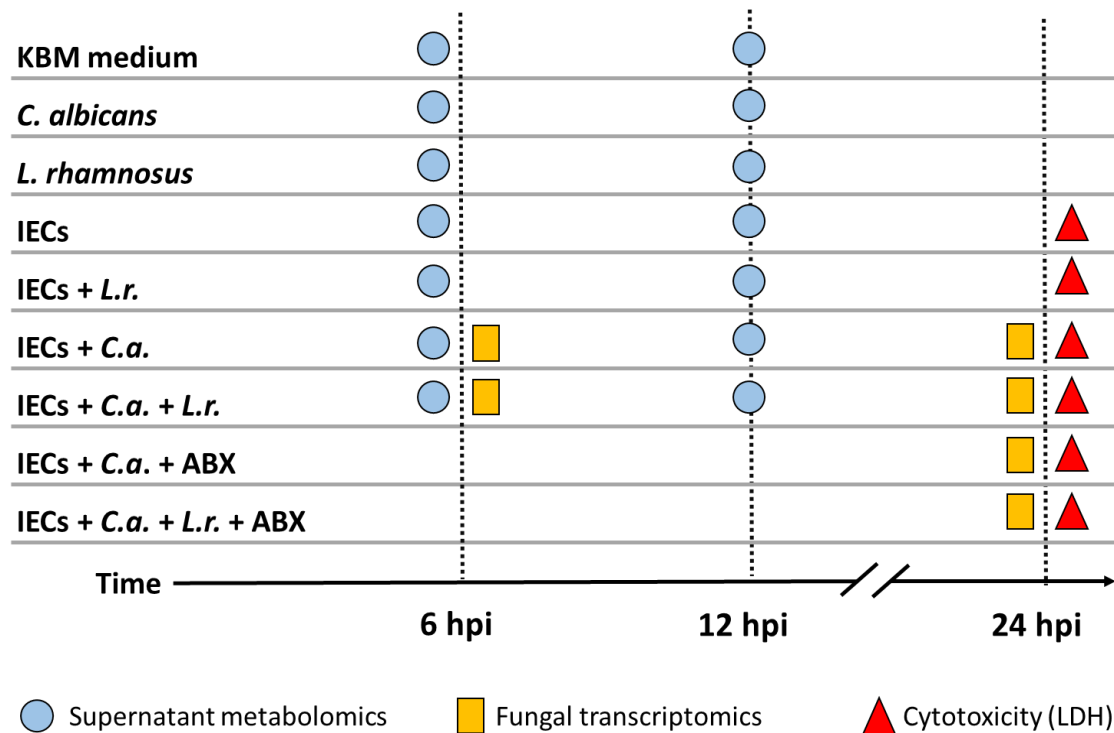


Fig. S7 *L. rhamnosus* modulates activity of *C. albicans* mitochondrial dehydrogenases

(A) Mitochondrial activity, represented as conversion of XTT by mitochondrial dehydrogenases (XTT absorbance) normalized with biomass (crystal violet (CV) absorbance), of *C. albicans* incubated in supernatants of *L. rhamnosus*-colonized or uncolonized IECs (1h, 37°C, 5% CO₂) (n=6, * = *p* 0.0350), or (B) in *L. rhamnosus*-conditioned or unconditioned KBM or KBM+ medium independently of host cells (1h, 37°C, 5% CO₂) (n=10, *** = *p* 0.0007 for KBM and * = *p* 0.0170 for KBM+), or (C) in presence or absence of 50 mM uridine (* = *p* 0.0470) or 2-deoxyinosine (* = *p* 0.0278) (24h, 37°C, 5% CO₂) (n=4). Bars represent the mean and error bars show standard

deviation (SD), dots represent the mean of technical replicates of the individual experiments, data were compared for significance using a paired t-test (two-tailed), * = $p \leq 0.05$, ** = $p \leq 0.01$, *** = $p \leq 0.001$. Source data are provided as a Source Data file. (D) Heatmap showing the transcriptional regulation of *C. albicans* OXPHOS genes as a result of *L. rhamnosus* colonization prior to infection at 6 and 24 hpi. Legend colour represents the Log₂ fold change of the regulation in presence vs. absence of *L. rhamnosus*. The asterisks (*) represent significance, based on the criteria of a Log₂ fold change of > 1 or < -1 and a moderated t-test Bonferroni corrected p -value of < 0.05. Data summarized from n=3 and n=4 independent experiments at 6 and 24 hpi respectively.



Supplementary Fig. 8: Study design

Overview of the samples, taken at different time points for transcriptional, metabolic, and cytotoxicity analysis in the *in vitro* model, in the presence of intestinal epithelial cells (IECs), *L. rhamnosus* (*L.r.*), *C. albicans* (*C.a.*) and antibiotics (ABX).

Supplementary Table 1: Metabolites of the clusters 1-8 shown in Fig. 2B (6 hpi).

[illegible]

Supplementary Table 2: Metabolites of the clusters 1-8 shown in Fig. S1 (12 hpi).

Cluster 1 “Secreted by <i>L. rhamnosus</i> colonized IECs”	Cluster 2 “Secreted by IECs” continued	Cluster 4 “Secreted by IECs and consumed by <i>L. rhamnosus</i> ”	cluster 7 “Consumed by <i>L. rhamnosus</i> colonized IECs”
(N(1) + N(8))-acetylspermidine	cholesterol	1-ribosyl-imidazoleacetate	adenine
1-carboxyethyltyrosine	choline phosphate	3-methyl-2-oxobutyrate	arginine
2'-deoxyinosine	creatine	3-methyl-2-oxovalerate	asparagine
2-hydroxy-3-methylvalerate	creatinine	4-methyl-2-oxopentanoate	biotin
2-hydroxy-4-(methylthio)butanoic acid	cystathionine	5-methylthioadenosine (MTA)	cystine
2-hydroxybutyrate/2- hydroxyisobutyrate	dimethylarginine (SDMA + ADMA)	acetylcarnitine (C2)	fructosyllsine
2-hydroxyglutarate	erythritol	aconitate [cis or trans]	glucose
2R,3R-dihydroxybutyrate	erythronate	carnitine	glutamine
3-(4-hydroxyphenyl)lactate	ethylmalonate	carnosine	HEPES
3-formylindole	fumarate	citrate	isoleucine
3-hydroxyoctanoate	gamma-glutamylthreonine	deoxycarnitine	leucine
3-indoleglyoxylic acid	glucuronate	fructose	lysine
3-ureidopropionate	glycerol 3-phosphate	gamma-glutamylalanine	mannitol/sorbitol
5-methyluridine (ribothymidine)	glycerophosphoethanolamine	gamma-glutamylglutamine	mannose
alpha-hydroxyisocaproate	Glycerophosphoglycerol	gamma-glutamylhistidine	methionine
alpha-hydroxyisovalerate	Glycerophosphoinositol	gamma-glutamylisoleucine	phenylalanine
cytidine	glycerophosphorylcholine (GPC)	gamma-glutamylleucine	serine
cytosine	Glycerophosphoserine	gamma-glutamylmethionine	Threonine
dihydroorotate	Gulonate	gamma-glutamylserine	tryptophan
glycerate	Homoarginine	gamma-glutamylvaline	valine
indolelactate	Hypotaurine	hippurate	
mevalonate	Hypoxanthine	isocitrate	
mevalonolactone	Lactate	N-acetylglucosamine/N- acetylgalactosamine	Cluster 8 “Secreted by <i>C. albicans</i> infected IECs”
N-acetylglutamate	malate	N-acetylmethionine	cysteine-glutathione disulfide
N-acetylthreonine	mannitol/sorbitol	nicotinamide	indoleacetate
N-acetyltryptophan	methionine sulfone	S-1-pyrroline-5-carboxylate	isovalerate (i5:0)
N-acetyltyrosine	N1-methyladenosine	xanthine	nicotinamide ribonucleotide (NMN)
N-carbamoylaspartate	N1-methylinosine		
nicotinate ribonucleoside	N6,N6,N6-trimethyllysine	Cluster 5 “Secreted by <i>C. albicans</i> ”	
phenyllactate (PLA)	N-acetylalanine	2,3-dihydroxyisovalerate	
pipecolate	N-acetyl-isoputresanine	2'-deoxyadenosine	
sedoheptulose	N-acetylleucine	3-sulfo-L-alanine	
thymine	N-acetylneuraminate	4-methylthio-2-oxobutanoate	
uracil	nicotinamide riboside	adenosine	
uridine	N-monomethylarginine	alpha-lipoate	
	ornithine	arabitol/xylitol	
	Palmitoyl sphingomyelin (d18:1/16:0)	aspartate	
Cluster 2 “Secreted by IECs”	pantothenate	benzoate	
1-carboxyethylisoleucine	p-cresol sulfate	cysteine s-sulfate	
1-carboxyethylleucine	phenol sulfate	glycerol	
1-carboxyethylphenylalanine	pseudouridine	imidazole lactate	
1-carboxyethylvaline	pyridoxal	kynurenate	
1-methylnicotinamide	pyridoxamine	N-acetylleucine	
1-palmitoyl-2-oleoyl-GPC (16:0/18:1)	pyridoxate	N-acetylputrescine	
1-stearoyl-2-oleoyl-GPC (18:0/18:1)	ribonate	phenylalanylhydroxyproline	
2'-deoxycytidine	Sulfate	ribitol	
2'-deoxyuridine	taurine		
2'-O-methylcytidine	trans-4-hydroxyproline		
3-hydroxy-3-methylglutarate	Urate		
3-hydroxybutyrate (BHBA)		Cluster 6 “Secreted by <i>L. rhamnosus</i> alone”	
3-hydroxyisobutyrate	Cluster 3 “Inconclusive”	adenosine	
4-imidazoleacetate	1-palmitoyl-2-linoleoyl-GPC (16:0/18:2)	argininate	
7-methylguanine	3-hydroxybutyrylglycine	diacetylspermidine	
Alanine	benzoate	gluconate	
Allantoin	cholesterol	glutamine conjugate of C6H10O2 (2)	
alpha-ketoglutarate	imidazole lactate	Inosine	
arabonate/xylonate	methysuccinate	malonate	
argininosuccinate	N-acetylaspartate (NAA)	N-acetylisoleucine	
Benzoate	N-acetylproline	N-acetylphenylalanine	
beta-alanine		N-acetylserine	
betaine		phosphothreonine	

Supplementary Table 3: Abbreviations used in Fig.6

Abbreviation	Full name
Reactions	
ACO	Aconitate Hydratase
AKGDH	alpha-Ketoglutarate Dehydrogenase
ALPL	2-Acetolactate Pyruvate-Lyase
CAR2	Ornithine Aminotransferase
ccoN	Cytochrome c Oxidase
CS	Citrate Synthase
DLD	D-lactate Dehydrogenase
ENO	Enolase
FBA	Fructose-Bisphosphate Aldolase
fbcH	Ubiquinol-Cytochrome c Reductase
FBP	Fructose-1,6-Bisphosphatase
FUM	Fumarate Hydratase
G6PD	Glucose-6-phosphate 1-Dehydrogenase
GAPDH	Glyceraldehyde 3-Phosphate Dehydrogenase
GPI	Glucose-6-Phosphate Isomerase
HK	Hexokinase
HPPR	Hydroxyphenylpyruvate Reductase
IDH	Isocitrate Dehydrogenase
ISL	Isocitrate Lyase
LDH	L-Lactate Dehydrogenase
MDH	Malate Dehydrogenase
MS	Malate Synthase
nuoA	NADH-Quinone Oxidoreductase
PCK	Phosphoenolpyruvate Carboxykinase
PDH	Pyruvate Dehydrogenase
PFK	6-Phosphofructokinase
PGAM	Phosphoglycerate Mutase
PGD	6-Phosphogluconate Dehydrogenase
PGK	Phosphoglycerate Kinase
PGL	6-Phosphogluconolactonase
PGM	Phosphoglucomutase
PK	Pyruvate Kinase
PMA	Plasma Membrane ATPase
PRO1	gamma-glutamyl kinase
PRO2	gamma-glutamyl phosphate reductase
RPE	Ribulose-Phosphate 3-Epimerase
RPI	Ribose 5-Phosphate Isomerase
SDH	Succinate Dehydrogenase
SUCD	Succinyl-CoA Synthetase
TAL	Transaldolase
TK	Transketolase
TPI	Triosephosphate Isomerase
Metabolites	
2PG	2-Phospho-D-glycerate
34HPLAC*	3,4-Dihydroxyphenylpyruvate (mapped to Phenylpyruvate measurements)
34HPPYR*	3,4-Dihydroxyphenyllactate (mapped to Phenyllactate measurements)
3PG	3-Phospho-D-glycerate
ACAC	Acetoacetate

ACCOA	Acetyl-CoA
ADP	Adenosine 5'-diphosphate
ALA	Alanine
ALAC	2-Acetolactate
AMP	Adenosine 5'-monophosphate
ATP	Adenosine 5'-triphosphate
BPG	D-Glycerate 1,3-diphosphate
DHAP	Dihydroxyacetone phosphate
F6P	beta-D-Fructose 6-phosphate
FDP	beta-D-Fructose 1,6-bisphosphate
Ficyt c	Ferricytochrome c
Focyt c	Ferrocycytochrome c
G1P	alpha-D-Glucose 1-phosphate
G3P	D-Glycerate 3-phosphate
G6P	alpha-D-Glucose 6-phosphate
GLC	Glucose
GLU	Glutamate
H2O	Water
H+	Hydrogen
ILE	Isoleucine
LAC	Lactate
LEU	Leucin
LYS	Lysine
MET	Methionine
NADH	Nicotinamide adenine dinucleotide (reduced)
NAD+	Nicotinamide adenine dinucleotide
O2	Oxygen
ORN	Ornithine
PEP	Phosphoenolpyruvate
PHE	Phenylalanine
PLAC	Phenyllactate
PPYR	Phenylpyruvate
PRCOA	Propanoyl-CoA
PRPP	5-Phosphoribosyl diphosphate
PUT*	Putrescine (mapped to N-acetylputrescine)
PYR	Pyruvate
St	Sulfate
SUCC	Succinate
THR	Threonine
TRP	Tryptophan
TYR	Tyrosine
VAL	Valine

Supplementary Table 4: Overview of screened deletion mutants for damage potential

Damage phenotype of deletion mutants from the Noble collection (Noble *et al.* 2010 *Nat Genetics*) corresponding to differentially regulated genes during infection of *L. rhamnosus* colonized IECs. Mutant phenotype is classified as hypovirulent or hypervirulent when the damage caused to IECs was significantly lower or higher than the wild type, respectively, and non-significant (ns) when it was not.

KO gene	Mutant phenotype	P value	KO gene	Mutant phenotype	P value
ace2	Hypovirulent	0,004	orf19.5565	ns	0,9998
ahr1	Hypovirulent	<0,0001	orf19.6874	ns	0,9807
aox1	ns	0,9984	orf19.7328	Hypervirulent	0,0838
aro80	ns	0,3589	orf19.7370	ns	0,9008
asm3	ns	0,9993	orf19.7516	ns	0,9996
cfl2	ns	0,9993	orf19.7554	ns	0,9832
cht2	ns	0,9913	pdk2	Hypervirulent	0,054
cip1	ns	0,9818	pex4	ns	0,9989
cyb5	ns	0,9364	pga13	ns	0,999
fcr1	ns	0,5483	pga45	ns	0,9912
gal4	ns	0,9987	pga7	ns	0,9842
gis2	ns	0,9917	pra1	ns	0,8833
het1	ns	0,4653	prk1	ns	0,9997
hgt6	ns	0,9985	prn2	ns	0,9999
hyr1	Hypervirulent	<0,0001	prn4	ns	0,904
kre5	Hypovirulent	0,0003	ptp3	Hypovirulent	<0,0001
mal2	ns	0,9991	pwp1	ns	0,3467
mec3	ns	0,9996	rbe1	Hypervirulent	0,0242
mid1	ns	>0,9999	rbt4	ns	0,9817
nik1	ns	0,9983	rgs2	Hypervirulent	0,0511
opt4	ns	0,9997	rim13	ns	0,9985
opt7	Hypervirulent	0,0572	sef1	ns	0,9608
orf19.1314	ns	0,8752	snq2	ns	0,9981
orf19.173	ns	0,999	sod6	ns	0,9999
orf19.215	ns	0,9995	swi4	ns	0,9661
orf19.3156	ns	0,9995	tye7	ns	0,9997
orf19.3395	ns	>0,9999	uga32	ns	0,9991
orf19.3720	ns	0,9815	vid27	ns	0,9994
orf19.3982	ns	>0,9999	wor2	ns	0,9991
orf19.4014	ns	0,9982	ycp4	ns	0,6831
orf19.4292	Hypovirulent	<0,0001	yhb1	ns	0,923
orf19.4445	ns	0,9993	ypt7	Hypervirulent	0,0766
orf19.4459	Hypervirulent	0,0009	ywp1	ns	0,9996
orf19.449	ns	0,9993	zcf27	Hypervirulent	<0,0001
orf19.4843	ns	0,624			