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Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our Editorial Policies and the Editorial Policy Checklist.

Please do not complete any field with "not applicable" or n/a. Refer to the help text for what text to use if an item is not relevant to your study.
For final submission: please carefully check your responses for accuracy; you will not be able to make changes later.

Statistics

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

n/a	Confirmed	
☐	☒	The exact sample size (*n*) for each experimental group/condition, given as a discrete number and unit of measurement
☒	☐	A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
☐	☒	The statistical test(s) used AND whether they are one- or two-sided *Only common tests should be described solely by name; describe more complex techniques in the Methods section.*
☐	☒	A description of all covariates tested
☒	☐	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
☐	☒	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
☐	☒	For null hypothesis testing, the test statistic (e.g. *F*, *t*, *r*) with confidence intervals, effect sizes, degrees of freedom and *P* value noted *Give P values as exact values whenever suitable.*
☒	☐	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
☒	☐	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
☐	☒	Estimates of effect sizes (e.g. Cohen's *d*, Pearson's *r*), indicating how they were calculated

Our web collection on statistics for biologists contains articles on many of the points above.

Software and code

Policy information about availability of computer code

Data collection: As indicated in the manuscript text, Images were acquired using Zeiss LSM 880 Airyscan Confocal Microscope with 40X or 63X objective. Images were processed using Zeiss Zen 3.4 (Blue) Imaging software. ImageStudioLite 5.2 (Li-COR Biosciences) and QuantStudio Real-Time PCR software were used for collection of western blot images and qPCR data, respectively.

Data analysis: Statistical analysis was conducted using Graph Pad Prism 9. Western blot quantification was done using ImageStudioLite 5.2 (Li-COR Biosciences). Fluorescence intentisty was quantified using Zeiss Zen 3.4 (Blue) Imaging software. Photoshop was used to overlay images. Pearson's colocalization coefficient were calculated using ImajeJ JACoP plugin. Lipid iterative MS/MS data were annotated with the Agilent Lipid Annotator software. All data files were then analyzed in Skyline-daily (Version 22.2.1.256) to obtain peak areas

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio guidelines for submitting code & software for further information.

Data

Policy information about availability of data

All manuscripts must include a data availability statement. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our policy

Included. Source data files are provided with this paper and all relevant data can be obtained from the corresponding authors Vivek Peche or Nada Abumrad upon reasonable request.

Research involving human participants, their data, or biological material

Policy information about studies with human participants or human data. See also policy information about sex, gender (identity/presentation), and sexual orientation and race, ethnicity and racism.

Reporting on sex and gender	NA
Reporting on race, ethnicity, or other socially relevant groupings	NA
Population characteristics	NA
Recruitment	NA
Ethics oversight	NA

Note that full information on the approval of the study protocol must also be provided in the manuscript.

Field-specific reporting

Please select the one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.

☒ Life sciences ☐ Behavioural & social sciences ☐ Ecological, evolutionary & environmental sciences

For a reference copy of the document with all sections, see nature.com/documents/nr-reporting-summary-flat.pdf

Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

Sample size	For experiments involving quantification n= 3 was chosen as the minimal replicate number. Sample size was determined based on similar studies previously conducted by our group. We confirmed this to be sufficient based on internal controls and low observed variability between samples.
Data exclusions	Data were not excluded from analysis.
Replication	All data presented in this manuscript were quantified from at least 3 biological replicates. Technical replicates were consistent (Coefficient of variability less than 5-10%). All images from at least 3 experiments. Western blots from at least 3 experiments were quantified. All mice experiments, n= at least 3 mice per group/condition/genotype. All attempts at replication were successful.
Randomization	No requirement for randomization. Not relevant to study. Minimized for selection bias.
Blinding	No blinding was performed in the experimental design as conditions were evident.

Behavioural & social sciences study design

All studies must disclose on these points even when the disclosure is negative.

Study description	NA
Research sample	NA
Sampling strategy	NA
Data collection	NA
Timing	NA
Data exclusions	NA
Non-participation	NA
Randomization	NA

Ecological, evolutionary & environmental sciences study design

All studies must disclose on these points even when the disclosure is negative.

Study description	NA
Research sample	NA
Sampling strategy	NA
Data collection	NA
Timing and spatial scale	NA
Data exclusions	NA
Reproducibility	NA
Randomization	NA
Blinding	NA

Did the study involve field work? ☐ Yes ☒ No

Field work, collection and transport

Field conditions	NA
Location	NA
Access & import/export	NA
Disturbance	NA

Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems

n/a	Involved in the study
☐	☒ Antibodies
☐	☒ Eukaryotic cell lines
☒	☐ Palaeontology and archaeology
☐	☒ Animals and other organisms
☒	☐ Clinical data
☒	☐ Dual use research of concern
☒	☐ Plants

Methods

n/a	Involved in the study
☒	☐ ChIP-seq
☒	☐ Flow cytometry
☒	☐ MRI-based neuroimaging

Antibodies

Antibodies used	See below
Validation	All antibodies used for western blots and immunostaining have been validated by previous studies conducted by the authors who routinely use them and references are included. In addition, antibodies were validated in this study in knockout animal models for specificity. All the antibodies are validated (see validation statements, citations on the manufacturer's website)

Antibodies used Anti-Mouse CD36: Research and Diagnostic Systems (R&D) #AF2519 (WB 1:100, Immunofluorescence, IF 1:50), Anti-Human CD36: R&D #AF1955 (1:100, 1:50), Caveolin 1 Rabbit monoclonal: Cell Signaling Technologies (CST)#3238 (1:1000, 1:100), pCaveolin1Y14: mouse monoclonal (mAb): BD Biosciences#611338 (1:1000), Phospho-Akt (Ser473) (D9E) XP Rabbit mAb: CST#4060 (WB 1:1000), Akt (pan) (C67E7) Rabbit mAb: CST#4691 (WB 1:1000), Anti-β-Actin (C4): mouse mAB, SantaCruz (SC) # sc-47778 (1:1000), Phospho-Ezrin (Thr567)/Radixin (Thr564)/Moesin (Thr558): Rabbit polyclonal: CST#3141 (1:1000), Ceramide: mouse mAB Simga#C8104 (IF 1:100), Src Family (36D10) : Rabbit mAb, CST#2109 (1:1000), Phospho-Cofilin (Ser3) (77G2): Rabbit mAb CST#3313 (1:1000), Cofilin mouse mAb Sigma#SAB2702206 (1:1000), Rab7 (D95F2) XP®: Rabbit mAb CST#9367 (1:1000), CD9 (E8L5J): Rabbit mAb CST#98327 (WB 1:1000), Anti-CD81 (B-11): mouse mAb sc-166029 (1:1000), Calnexin: Rabbit polyclonal Enzo Life Sci#ADI-SPA-860 (1:1000), Secondary Abs for IF: Donkey anti-Goat, -Rabbit, -Mouse Alexa Fluor 488, 647 and 594 (Invitrogen, 1:500).

Eukaryotic cell lines

Policy information about cell lines and Sex and Gender in Research

Cell line source(s)	Primary human dermal microvascular cells (hMEC) and primary mouse lung microvascular (mMEC) cells were used.
Authentication	hMEC were purchased from Lonza and mouse primary cells were isolated from lungs using magnetic beads. Both cells types were confirmed with VE-cadherin (EC marker).
Mycoplasma contamination	All cells were tested and were mycoplasma negative.
Commonly misidentified lines (See ICLAC register)	NA

Palaeontology and Archaeology

Specimen provenance	NA
Specimen deposition	NA
Dating methods	NA

☐ Tick this box to confirm that the raw and calibrated dates are available in the paper or in Supplementary Information.

Ethics oversight	NA

Note that full information on the approval of the study protocol must also be provided in the manuscript.

Animals and other research organisms

Policy information about studies involving animals; ARRIVE guidelines recommended for reporting animal research, and Sex and Gender in Research

Laboratory animals	All studies used cohorts of adult C57Bl6 mice models, 12-20 week, mice were littermates matched for age and sex.
Wild animals	The study did not involve wild animals.
Reporting on sex	The study involved both male and female C57Bl6 animals .
Field-collected samples	The study did not involve samples collected from the field.
Ethics oversight	The mouse work was performed under the study protocol approved by the Washington University Institutional Animal Care and Use Committee.

Note that full information on the approval of the study protocol must also be provided in the manuscript.

Clinical data

Policy information about clinical studies

All manuscripts should comply with the ICMJE guidelines for publication of clinical research and a completed CONSORT checklist must be included with all submissions.

Clinical trial registration	NA
Study protocol	NA
Data collection	NA
Outcomes	NA

Dual use research of concern

Policy information about dual use research of concern

Hazards

Could the accidental, deliberate or reckless misuse of agents or technologies generated in the work, or the application of information presented in the manuscript, pose a threat to:

No	Yes	
[X]	[ ]	Public health
[X]	[ ]	National security
[X]	[ ]	Crops and/or livestock
[X]	[ ]	Ecosystems
[X]	[ ]	Any other significant area

Experiments of concern

Does the work involve any of these experiments of concern:

No	Yes	
[X]	[ ]	Demonstrate how to render a vaccine ineffective
[X]	[ ]	Confer resistance to therapeutically useful antibiotics or antiviral agents
[X]	[ ]	Enhance the virulence of a pathogen or render a nonpathogen virulent
[X]	[ ]	Increase transmissibility of a pathogen
[X]	[ ]	Alter the host range of a pathogen
[X]	[ ]	Enable evasion of diagnostic/detection modalities
[X]	[ ]	Enable the weaponization of a biological agent or toxin
[X]	[ ]	Any other potentially harmful combination of experiments and agents

Plants

Seed stocks	NA
Novel plant genotypes	NA
Authentication	NA

ChIP-seq

Data deposition

[ ] Confirm that both raw and final processed data have been deposited in a public database such as GEO.

[ ] Confirm that you have deposited or provided access to graph files (e.g. BED files) for the called peaks.

Data access links *May remain private before publication.*	NA
Files in database submission	NA
Genome browser session (e.g. UCSC)	NA

Methodology

Replicates	NA
Sequencing depth	NA
Antibodies	NA
Peak calling parameters	NA
Data quality	NA
Software	NA

Flow Cytometry

Plots

Confirm that:

☐ The axis labels state the marker and fluorochrome used (e.g. CD4-FITC).

☐ The axis scales are clearly visible. Include numbers along axes only for bottom left plot of group (a 'group' is an analysis of identical markers).

☐ All plots are contour plots with outliers or pseudocolor plots.

☐ A numerical value for number of cells or percentage (with statistics) is provided.

Methodology

Sample preparation	NA
Instrument	NA
Software	NA
Cell population abundance	NA
Gating strategy	NA

☐ Tick this box to confirm that a figure exemplifying the gating strategy is provided in the Supplementary Information.

Magnetic resonance imaging

Experimental design

Design type	NA
Design specifications	NA
Behavioral performance measures	NA

Imaging type(s)	NA
Field strength	NA
Sequence & imaging parameters	NA
Area of acquisition	NA

Diffusion MRI ☐ Used ☐ Not used

Preprocessing

Preprocessing software	NA
Normalization	NA
Normalization template	NA
Noise and artifact removal	NA
Volume censoring	NA

Statistical modeling & inference

Model type and settings	NA
Effect(s) tested	NA

Specify type of analysis: ☐ Whole brain ☐ ROI-based ☐ Both

Statistic type for inference

NA

(See Eklund et al. 2016)

Correction

NA

Models & analysis

n/a	Involved in the study
[x]	[ ] Functional and/or effective connectivity
[x]	[ ] Graph analysis
[x]	[ ] Multivariate modeling or predictive analysis

Functional and/or effective connectivity

NA

Graph analysis

NA

Multivariate modeling and predictive analysis

NA