

Supplementary Information

Ultra-photostable Small-molecule Dyes Facilitate Near-infrared Biophotonics

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Supplementary Methods

Materials and reagents

Analytical grade solvents including petroleum ether, dichloromethane, and ethyl acetate were purchased from Titan Scientific. Dulbecco's modified eagle medium (DMEM) cell culture was purchased from Gibco. Fetal bovine serum (FBS) was purchased from ExCell. Trypsin (0.25%) and penicillin-streptomycin solution (100X) were purchased from MesGen Biotechnology. All other chemicals were purchased from major manufacturers in China including TCI, Macklin and J&K. All chemicals were used as received without further purification unless otherwise noted.

Characterization

^1H -NMR and ^{13}C -NMR spectra were obtained on a Bruker AV-400 spectrometer. Chemical shifts are referenced to the residue solvent peaks and given in ppm. MALDI-TOF MS analyses were performed in AB SCIEX 4800 Plus MALDI-TOF/TOFTM mass spectrometer. LC/MS analyses were conducted in LC-20AD linked AB SCIEX Triple TOF 4600. Surface molecular charge, partition coefficient (logD) and hydrophobicity were calculated using MarvinSketch (ChemAxon). Absorbance spectra were acquired with a PerkinElmer Lambda 750S UV-visible-NIR spectrometer. Fluorescence spectra were recorded on Horiba Fluorescence Spectrometer instrument (FluorologTM HORIBA Scientific) with external 808 nm (MDL-H-5W) and 980 nm (MDL-H-5W) semiconductor lasers (Changchun New Industries Optoelectronics Tech. Co., Ltd.) as excitation sources. Quartz cuvettes (1 cm) were used for absorbance and emission measurements. NIR-II cellular intravital imaging was conducted on an epifluorescence microscopy system with 640×512 pixel 2D InGaAs NIRvana camera. In vivo NIR-II fluorescence images were captured by a modified home-built InGaAs NIRvana 640 CCD camera (Princeton Instruments Inc.). Photoacoustic signals and images were captured by VevoLAZR (FujiFilm VisualSonics Inc.) system with Nd: YAG laser with optical parametric oscillator (OPO) (680~970 nm) as excitation source. Femtosecond and nanosecond transient absorption (TA) measurements were carried out on commercial transient absorption spectrometers (Helios Fire Transient absorption spectrometer and Helios-EOS fire, respectively, ultrafast system). X-ray crystallography data sets were collected with Bruker D8 VENTURE X-ray diffractometer at 173(2) K, the structure was solved by direct

methods using SHELXS and refined against F2 on all data by full-matrix least-squares with SHELXL-2014 following established refinement strategies.

Measurement of fluorescence quantum yield (QY)

Quantum yields (Φ_f) were determined in various solvents relative to IR26 ($\Phi_f = 0.05\%$ in DCE),¹ from plots of integrated fluorescence intensity vs. absorbance, according to the following relationship:

$$\Phi_{f.l.s} = \Phi_{f.l.r} \times \frac{n_s^2}{n_r^2} \times \frac{K_s}{K_r} \quad (1)$$

where subscripts r and s denote standard and test sample, respectively, Φ_f is the fluorescence quantum yield, K is the slope of the integrated fluorescence intensity vs. absorbance plot, and n is the refractive index of the solvent. Measurements were performed with the absorbance at 808 nm of all dye solutions <0.1 in order to maximize illumination homogeneity and optical transparency. The 808 nm laser was used as the excitation source and the emission spectrum in the 850-1500 nm region was acquired in fluorescence spectrometer.

Preparation of various bioactive species

GSH: A 20 mM stock solution of GSH was prepared in deionized water.

H₂O₂: A commercial H₂O₂ (30% in water) was diluted before use. The concentration of H₂O₂ stock was determined by measuring the absorbance at 240 nm with molar extinction coefficient of 43.6 M⁻¹ cm⁻¹.

·O₂⁻: A 10 mM stock solution of KO₂ was dissolved in DMSO directly.

·OH: ·OH was prepared by the Fenton reaction between FeCl₂ stock (20 mM) and H₂O₂ stock (20 mM).

ClO⁻: ClO⁻ stock solution (20 mM in deionized water, pH=12) was prepared by dilution of commercial NaClO solution and assayed by measuring the absorbance at 293 nm ($\epsilon_{293 \text{ nm}} = 350 \text{ M}^{-1} \text{ cm}^{-1}$).

ONOO⁻: Peroxynitrite stock (10 mM in 0.01 M NaOH) was obtained following the literature procedure² and the concentration was determined by measuring the absorbance at 302 nm ($\epsilon_{302 \text{ nm}} = 1670 \text{ M}^{-1} \text{ cm}^{-1}$).

Cell viability

Mouse breast cancer cells (4T1, catalog no.: TCM32) were provided by Cell Bank, Chinese Academy of Science. All cells were cultured in Dulbecco's Modified Eagle medium (DMEM) supplemented with 10% FBS and 1% Penicillin-Streptomycin at 37 °C in a humidified atmosphere of 5% CO₂. 4T1 cells were cultured in a 96-well plate (8×10³ cells/well) after 24 h incubation, the medium was replaced with 100 μL of fresh DMEM containing AF2 and AF3 with concentrations of 0, 0.2, 0.5, 1.0, 2.0, 5.0, 10 and 20 μM, respectively. Cells were incubated further for 4 h. To detect the cytotoxicity, 100 μL Cell Counting Kit-8 (CCK-8) solution (CCK-8: DMEM= 1: 9) was added to each well of the microliter plate and the plate was incubated in the CO₂ incubator for additional 4 h. Enzyme dehydrogenase in living cells was oxidized by this kit to orange carapace. The quality was assessed calorimetrically by using a multi-reader (TECAN, Infinite M200, Germany). The measurements were based on the absorbance values at 450 nm. Following formula was used to calculate the viability of cell growth:

$$V = \frac{Abs_t - Abs_b}{Abs_c - Abs_b} \times 100\% \quad (2)$$

Where *V* is cell viability, subscripts t, b and c represent treatment group, blank group and control group, respectively.

Synthetic procedures

1) General procedure for preparation of compound 1 via C-N cross-coupling

The following procedure for **1c** is representative. A vial was charged with 3,6-dibromo-9-fluorenone (338 mg, 1 mmol), Pd₂(dba)₃ (22.8 mg, 25 μmol, 0.025 eq), and tBuONa (576 mg, 6 mmol, 6 eq). The vial was sealed and vacuated/backfilled with nitrogen (3×). Anhydrous toluene (10 mL) was added, followed by addition of PtBu₃ (15 mg, 75 μmol, 10% wt in toluene, 0.075eq) and dimethylamine (2 mL, 4 mmol, 2 M in THF, 4 eq). The reaction was then stirred and refluxed at 100 °C for 4 h. It was subsequently cooled to room temperature, diluted with DCM, deposited onto Celite, and concentrated to dryness. The crude mixture was dispersed with ethyl acetate (3 mL), filtrated to afford the title compound as an orange solid (228 mg, 86%).

3,6-bis(*tert*-butyl carbamate)-9H-fluoren-9-one (1a): 3,6-dibromo-9-fluorenone (338 mg, 1 mmol), Pd₂(dba)₃ (22.8 mg, 25 μmol, 0.025 eq), tBuONa (576 mg, 6 mmol, 6 eq), PtBu₃ (15 mg, 75 μmol, 10% wt in

toluene, 0.075eq) and tert-butyl carbamate (468 mg, 4 mmol, 4 eq) were used, purified with silica gel chromatography (PE/EA, 3/1, v/v), affording **1a** as a yellow solid (295 mg, 72%). ¹H NMR (400 MHz, CDCl₃) δ 7.72 (d, *J* = 1.5 Hz, 2H), 7.53 (d, *J* = 8.1 Hz, 2H), 7.11 (dd, *J* = 8.1, 1.9 Hz, 2H), 6.82 (s, 2H), 1.54 (s, 18H); ¹³C NMR (101 MHz, CDCl₃) δ 152.24 (C), 145.62 (C), 144.42 (C), 129.77 (C), 125.23 (C), 117.75 (C), 110.31 (C), 28.42 (CH₃).

3,6-bis(tert-butyl methylcarbamate)-9H-fluoren-9-one (1b): 3,6-dibromo-9-fluorenone (338 mg, 1 mmol), Pd₂(dba)₃ (22.8 mg, 25 μmol, 0.025 eq), tBuONa (576 mg, 6 mmol, 6 eq), PtBu₃ (15 mg, 75 μmol, 10% wt in toluene, 0.075eq) and tert-butyl methylcarbamate (524 mg, 4 mmol, 4 eq) were used, purified with silica gel chromatography (PE/EA, 3/1, v/v), affording **1b** as a yellow solid (276 mg, 63%). ¹H NMR (400 MHz, CDCl₃) δ 7.60 (d, *J* = 8.0 Hz, 2H), 7.47 (d, *J* = 1.7 Hz, 2H), 7.15 (dd, *J* = 8.0, 1.9 Hz, 2H), 3.32 (s, 6H), 1.50 (s, 18H); ¹³C NMR (101 MHz, CDCl₃) δ 154.23 (C), 149.79 (C), 144.66 (C), 131.24 (C), 124.97 (C), 124.64 (C), 117.18 (C), 81.42 (C), 37.20 (CH₃), 28.46 (CH₃).

3,6-bis(dimethylamino)-9H-fluoren-9-one (1c): (86%, orange solid) ¹H NMR (400 MHz, CDCl₃) δ 7.48 (d, *J* = 8.4 Hz, 2H), 6.75 (d, *J* = 2.2 Hz, 2H), 6.43 (dd, *J* = 8.4, 2.3 Hz, 2H), 3.09 (s, 12H); ¹³C NMR (101 MHz, CDCl₃) δ 191.76 (C), 154.59 (C), 146.19 (C), 125.34 (C), 124.47 (C), 110.38 (C), 103.19 (C), 40.72 (CH₃).

3,6-bis(diethylamino)-9H-fluoren-9-one (1d): 3,6-dibromo-9-fluorenone (338 mg, 1 mmol), Pd₂(dba)₃ (22.8 mg, 25 μmol, 0.025 eq), tBuONa (576 mg, 6 mmol, 6 eq), PtBu₃ (15 mg, 75 μmol, 10% wt in toluene, 0.075eq) and diethylamine (292 mg, 4 mmol, 4 eq) were used, affording **1d** as an orange solid (170 mg, 53%). ¹H NMR (400 MHz, CDCl₃) δ 7.48 (d, *J* = 8.4 Hz, 2H), 6.72 (d, *J* = 2.3 Hz, 2H), 6.43 (dd, *J* = 8.5, 2.3 Hz, 2H), 3.47 (q, *J* = 7.1 Hz, 8H), 1.24 (t, *J* = 7.1 Hz, 12H); ¹³C NMR (101 MHz, CDCl₃) δ 191.38 (C), 152.27 (C), 146.46 (C), 125.63 (C), 123.92 (C), 109.90 (C), 102.55 (C), 44.98 (CH₂), 13.01 (CH₃).

3,6-di(azetidin-1-yl)-9H-fluoren-9-one (1e): 3,6-dibromo-9-fluorenone (338 mg, 1 mmol), Pd₂(dba)₃ (22.8 mg, 25 μmol, 0.025 eq), tBuONa (576 mg, 6 mmol, 6 eq), PtBu₃ (15 mg, 75 μmol, 10% wt in toluene, 0.075eq) and azetidine (228 mg, 4 mmol, 4 eq) were used, affording **1e** as an orange solid (180 mg, 62%). ¹H NMR (400 MHz, CDCl₃) δ 7.45 (d, *J* = 8.0 Hz, 2H), 6.39 (s, 2H), 6.11 (d, *J* = 8.1 Hz, 2H), 4.01 (t, *J* = 7.1 Hz, 9H), 2.42 (p, *J* = 7.0 Hz, 5H); ¹³C NMR (101 MHz, CDCl₃) δ 155.43 (C), 145.78 (C), 125.21 (C), 124.89 (C), 109.04 (C), 102.06 (C), 51.81 (CH₂), 16.67 (CH₂).

3,6-bis(3,3-difluoroazetidin-1-yl)-9H-fluoren-9-one (1f): 3,6-dibromo-9-fluorenone (338 mg, 1 mmol), Pd₂(dba)₃ (22.8 mg, 25 μmol, 0.025 eq), tBuONa (576 mg, 6 mmol, 6 eq), PtBu₃ (15 mg, 75 μmol, 10% wt in toluene, 0.075eq) and 3,3-difluoroazetidine (372 mg, 4 mmol, 4 eq) were used, affording **1f** as an orange solid

(235 mg, 65%). ¹H NMR (400 MHz, CDCl₃) δ 7.50 (d, *J* = 8.1 Hz, 2H), 6.46 (d, *J* = 1.9 Hz, 2H), 6.23 (dd, *J* = 8.1, 2.0 Hz, 2H), 4.35 (t, *J* = 11.6 Hz, 8H); ¹³C NMR (101 MHz, CDCl₃) δ 191.34 (C), 153.52 (C), 145.73 (C), 126.51 (C), 125.63 (C), 115.68 (C), 111.08 (C), 103.63 (C), 63.23 (CH₂).

3,6-di(pyrrolidin-1-yl)-9H-fluoren-9-one (1g): 3,6-dibromo-9-fluorenone (338 mg, 1 mmol), Pd₂(dba)₃ (22.8 mg, 25 μmol, 0.025 eq), tBuONa (576 mg, 6 mmol, 6 eq), PtBu₃ (15 mg, 75 μmol, 10% wt in toluene, 0.075eq) and pyrrolidine (284 mg, 4 mmol, 4 eq) were used, affording **1g** as an orange solid (267 mg, 84%). ¹H NMR (400 MHz, CDCl₃) δ 7.48 (d, *J* = 8.3 Hz, 2H), 6.63 (d, *J* = 2.1 Hz, 2H), 6.30 (dd, *J* = 8.3, 2.2 Hz, 2H), 3.48 – 3.36 (m, 8H), 2.12 – 1.98 (m, 8H); ¹³C NMR (101 MHz, CDCl₃) δ 191.67 (C), 151.93 (C), 146.10 (C), 125.26 (C), 123.93 (C), 110.14 (C), 103.21 (C), 48.02 (CH₂), 25.54 (CH₂).

3,6-di(piperidin-1-yl)-9H-fluoren-9-one (1h): 3,6-dibromo-9-fluorenone (338 mg, 1 mmol), Pd₂(dba)₃ (22.8 mg, 25 μmol, 0.025 eq), tBuONa (576 mg, 6 mmol, 6 eq), PtBu₃ (15 mg, 75 μmol, 10% wt in toluene, 0.075eq) and piperidine (340 mg, 4 mmol, 4 eq) were used, affording **1h** as an orange solid (252 mg, 73%). ¹H NMR (400 MHz, CDCl₃) δ 7.49 (d, *J* = 8.4 Hz, 2H), 6.97 (s, 2H), 6.65 (dd, *J* = 8.4, 1.8 Hz, 2H), 3.45 – 3.33 (m, 8H), 1.75 – 1.61 (m, 12H); ¹³C NMR (101 MHz, CDCl₃) δ 191.44 (C), 155.81 (C), 146.03 (C), 125.67 (C), 125.39 (C), 113.16 (C), 106.06 (C), 49.32 (CH₂), 25.64 (CH₂), 24.49 (CH₂).

3,6-di(azepan-1-yl)-9H-fluoren-9-one (1i): 3,6-dibromo-9-fluorenone (338 mg, 1 mmol), Pd₂(dba)₃ (22.8 mg, 25 μmol, 0.025 eq), tBuONa (576 mg, 6 mmol, 6 eq), PtBu₃ (15 mg, 75 μmol, 10% wt in toluene, 0.075eq) and azepane (396 mg, 4 mmol, 4 eq) were used, affording **1i** as an orange solid (306 mg, 82%). ¹H NMR (400 MHz, CDCl₃) δ 7.47 (d, *J* = 8.4 Hz, 2H), 6.75 (d, *J* = 2.2 Hz, 2H), 6.45 (dd, *J* = 8.5, 2.3 Hz, 2H), 3.64 – 3.50 (m, 8H), 1.89 – 1.77 (m, 8H), 1.62 – 1.52 (m, 8H); ¹³C NMR (101 MHz, CDCl₃) δ 153.38 (C), 146.35 (C), 125.52 (C), 123.97 (C), 109.74 (C), 102.43 (C), 49.90 (CH₂), 27.73 (CH₂), 27.02 (CH₂).

3,6-bis(methyl(phenyl)amino)-9H-fluoren-9-one (1j): 3,6-dibromo-9-fluorenone (338 mg, 1 mmol), Pd₂(dba)₃ (22.8 mg, 25 μmol, 0.025 eq), tBuONa (576 mg, 6 mmol, 6 eq), PtBu₃ (15 mg, 75 μmol, 10% wt in toluene, 0.075eq) and N-methylaniline (428 mg, 4 mmol, 4 eq) were used, affording **1j** as an orange solid (273 mg, 70%). ¹H NMR (400 MHz, CDCl₃) δ 7.46 (d, *J* = 8.3 Hz, 2H), 7.43 – 7.37 (m, 4H), 7.25 – 7.18 (m, 6H), 6.83 (d, *J* = 2.1 Hz, 2H), 6.57 (dd, *J* = 8.3, 2.1 Hz, 2H), 3.40 (s, 6H); ¹³C NMR (101 MHz, CDCl₃) δ 191.47(C), 154.00(C), 147.78(C), 145.82(C), 130.02(C), 126.36(C), 125.87(C), 125.45(C), 125.26(C), 114.40(C), 106.80(C), 40.96(CH₃).

3,6-bis(diphenylamino)-9H-fluoren-9-one (1k): 3,6-dibromo-9-fluorenone (338 mg, 1 mmol), Pd₂(dba)₃ (22.8 mg, 25 μmol, 0.025 eq), tBuONa (576 mg, 6 mmol, 6 eq), PtBu₃ (15 mg, 75 μmol, 10% wt in toluene,

0.075eq) and diphenylamine (676 mg, 4 mmol, 4 eq) were used, affording **1k** as an orange solid (380 mg, 74%). ¹H NMR (400 MHz, CDCl₃) δ 7.48 (d, *J* = 8.2 Hz, 2H), 7.33 – 7.27 (m, 8H), 7.16 – 7.07 (m, 12H), 7.01 (d, *J* = 1.9 Hz, 2H), 6.77 (dd, *J* = 8.2, 2.0 Hz, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 191.01 (C), 153.53 (C), 146.82 (C), 145.34 (C), 129.70 (C), 128.60 (C), 125.71 (C), 125.27 (C), 124.52 (C), 121.40 (C), 113.45 (C).

3,6-di(indolin-1-yl)-9H-fluoren-9-one (1m): 3,6-dibromo-9-fluorenone (338 mg, 1 mmol), Pd₂(dba)₃ (22.8 mg, 25 μmol, 0.025 eq), tBuONa (576 mg, 6 mmol, 6 eq), PtBu₃ (15 mg, 75 μmol, 10% wt in toluene, 0.075eq) and indoline (476 mg, 4 mmol, 4 eq) were used, affording **1m** as a dark red solid (331 mg, 80%). ¹H NMR (400 MHz, CDCl₃) δ 7.63 (d, *J* = 8.2 Hz, 2H), 7.34 (d, *J* = 2.0 Hz, 2H), 7.31 (d, *J* = 8.0 Hz, 2H), 7.24 (d, *J* = 7.3 Hz, 2H), 7.17 (t, *J* = 7.7 Hz, 2H), 7.04 (dd, *J* = 8.2, 2.0 Hz, 2H), 6.87 (t, *J* = 7.4 Hz, 2H), 4.10 (t, *J* = 8.3 Hz, 4H), 3.19 (t, *J* = 8.3 Hz, 4H); ¹³C NMR (101 MHz, CDCl₃) δ 149.14 (C), 145.68 (C), 145.35 (C), 132.37 (C), 127.57 (C), 127.42 (C), 125.57 (C), 125.54 (C), 120.73 (C), 115.52 (C), 110.33 (C), 107.83 (C), 52.46 (CH₂), 28.28 (CH₂).

2) General procedure for preparation of AF dyes via nucleophilic addition of aryl lithium reagents

Method 1: The following procedure for AF1 is representative. A vial was charged with 2-bromo-1,3-dimethylbenzene (277.5 mg, 1.5 mmol, 1 eq), sealed, and vacuated/backfilled with nitrogen (3×). After dissolving the bromide in anhydrous THF (2 mL) and cooling the reaction to –78 °C, *n*-butyllithium (0.825 mL, 2 M in cyclohexane, 1.1 eq) was slowly added. The reaction was then stirred in the same temperature for further 5 min. A solution of 3,6-bis(*tert*-butyl carbamate)-9H-fluoren-9-one (**1a**; 205 mg, 0.5 mmol) in anhydrous THF (7 mL) was then added dropwise. The reaction mixture was warmed to room temperature and stirred for 30 min. It was subsequently quenched with water, extracted with EtOAc (2×). The combined organics were dried (Na₂SO₄), filtered, evaporated, and purified with silica gel chromatography (0–50% EtOAc/PE, linear gradient). The alcohol intermediate was then redissolved in CH₂Cl₂ (5 mL), and trifluoroacetic acid (1 mL) was added. The reaction was stirred at room temperature overnight. Toluene (3 mL) was added; the reaction mixture was concentrated to dryness and then triturated with Et₂O/PE (v/v, 1/4) to provide AF1 as a green solid (103 mg, 50% total, TFA salt). Analytical HPLC and NMR indicated that the material was >95% pure and did not require further purification.

AF1 (TFA salt): (50%, green solid) ¹H NMR (400 MHz, CDCl₃) δ 8.68 (s, 4H), 7.33 – 7.18 (m, 3H), 6.95 (s, 2H), 6.59 (d, *J* = 8.4 Hz, 2H), 6.18 (d, *J* = 8.4 Hz, 2H), 2.16 (s, 6H); ¹³C NMR (101 MHz, CDCl₃) δ 172.07 (C), 160.87 (C), 147.57 (C), 135.41 (C), 134.64 (C), 131.10 (C), 129.86 (C), 128.87 (C), 128.37 (C), 114.44

(C), 114.15 (C), 20.31 (CH₃); Maldi-Tof/Tof-MS calcd for C₂₁H₁₉N₂ [M]⁺ 299.1543, found 299.1694.

AF2 (TFA salt): Method 1 was applied. 2-bromo-1,3-dimethylbenzene (277.5 mg, 1.5 mmol, 1 eq), *n*-butyllithium (0.825 mL, 2 M in cyclohexane, 1.1 eq), 3,6-bis(*tert*-butyl methylcarbamate)-9H-fluoren-9-one (**1b**; 219 mg, 0.5 mmol) were used, affording AF2 as a green solid (163 mg, 74% total, TFA salt). ¹H NMR (400 MHz, CDCl₃) δ 9.11 (s, 2H), 7.26 – 7.21 (m, 1H), 7.19 (d, *J* = 1.2 Hz, 2H), 7.11 (d, *J* = 7.6 Hz, 2H), 6.61 (d, *J* = 8.7 Hz, 2H), 6.06 (dd, *J* = 8.7, 1.3 Hz, 2H), 3.09 (s, 6H), 2.17 (s, 6H); ¹³C NMR (101 MHz, CDCl₃) δ 171.59(C), 159.38(C), 146.59(C), 135.65(C), 133.93(C), 131.10(C), 129.95(C), 129.32(C), 127.88(C), 114.50(C), 109.90(C), 30.98(CH₃), 20.29(CH₃); Maldi-Tof/Tof-MS calcd for C₂₃H₂₃N₂ [M]⁺ 327.1856, found 327.1960.

Method 2: The following procedure for AF3 is representative. A vial was charged with 2-bromo-1,3-dimethylbenzene (277.5 mg, 1.5 mmol, 1 eq), sealed, and vacuated/backfilled with nitrogen (3×). After dissolving the bromide in anhydrous THF (2 mL) and cooling the reaction to −78 °C, *n*-butyllithium (0.825 mL, 2 M in cyclohexane, 1.1 eq) was slowly added. The reaction was then stirred in the same temperature for further 5 min. A solution of 3,6-bis(dimethylamino)-9H-fluoren-9-one (**1c**; 133 mg, 0.5 mmol) in anhydrous THF (7 mL) was then added dropwise. The reaction mixture was warmed to room temperature and stirred for 30 min. It was subsequently quenched with diluted perchloric acid (10 wt%, 1 mL), stirred for another 30 min until product precipitated from solution (147 mg, 65%). Analytical HPLC and NMR indicated that the material was >95% pure and did not require further purification.

AF3 (perchlorate salt): (65%, dark green solid) ¹H NMR (400 MHz, CDCl₃) δ 7.67 (d, *J* = 2.1 Hz, 2H), 7.24 (d, *J* = 7.5 Hz, 1H), 7.13 (d, *J* = 7.6 Hz, 2H), 6.58 (d, *J* = 8.9 Hz, 2H), 6.08 (dd, *J* = 8.9, 2.1 Hz, 2H), 3.44 (s, 12H), 2.22 (s, 6H); ¹³C NMR (101 MHz, CDCl₃) δ 158.36(C), 147.72(C), 135.39(C), 132.95(C), 129.20(C), 127.71(C), 113.62(C), 111.21(C), 42.17(CH₃), 20.09(CH₃); Maldi-Tof/Tof-MS calcd for C₂₅H₂₇N₂ [M]⁺ 355.2169, found 355.2269.

AF4 (perchlorate salt): Method 2 was applied. 2-bromo-1,3-dimethoxybenzene (325 mg, 1.5 mmol, 1 eq), *n*-butyllithium (0.825 mL, 2 M in cyclohexane, 1.1 eq) and 3,6-bis(dimethylamino)-9H-fluoren-9-one (**1c**; 133 mg, 0.5 mmol) were used, affording AF4 as a brown solid (216 mg, 89%, perchlorate salt). ¹H NMR (400 MHz, DMSO) δ 7.49 (t, *J* = 8.4 Hz, 1H), 7.44 (d, *J* = 2.1 Hz, 2H), 6.84 (d, *J* = 8.5 Hz, 2H), 6.78 (d, *J* = 9.0 Hz, 2H), 6.33 (dd, *J* = 9.0, 2.1 Hz, 2H), 3.73 (s, 6H), 3.34 (s, 12H); ¹³C NMR (101 MHz, DMSO) δ 158.53(C), 157.76(C), 147.30(C), 133.99(C), 133.58(C), 129.92(C), 112.23(C), 112.14(C), 105.49(C), 56.67(CH₃),

42.06(CH₃); Maldi-Tof/Tof-MS calcd for C₂₅H₂₇N₂O₂ [M]⁺ 387.2067, found 387.2234.

AF5 (perchlorate salt): Method 2 was applied. 2-bromo-1,3-dimethylbenzene (277.5 mg, 1.5 mmol, 1 eq), *n*-butyllithium (0.825 mL, 2 M in cyclohexane, 1.1 eq) and 3,6-bis(diethylamino)-9H-fluoren-9-one (**1d**; 161 mg, 0.5 mmol) were used, affording AF5 as a brown solid (217 mg, 85%, perchlorate salt). ¹H NMR (400 MHz, CDCl₃) δ 7.65 (s, 2H), 7.27 – 7.22 (m, 1H), 7.13 (d, *J* = 7.7 Hz, 2H), 6.58 (d, *J* = 8.9 Hz, 2H), 6.09 (s, 2H), 3.75 (d, *J* = 6.4 Hz, 8H), 2.23 (s, 6H), 1.37 (t, *J* = 7.1 Hz, 12H); ¹³C NMR (101 MHz, CDCl₃) δ 156.73(C), 148.18(C), 135.66(C), 133.13(C), 131.20(C), 129.32(C), 127.88(C), 113.40(C), 110.92(C), 47.19(CH₂), 20.33(CH₃), 13.97(CH₃); Maldi-Tof/Tof-MS calcd for C₂₉H₃₅N₂ [M]⁺ 411.2795, found 411.2924.

AF6 (perchlorate salt): Method 2 was applied. 2-bromo-1,3-dimethylbenzene (277.5 mg, 1.5 mmol, 1 eq), *n*-butyllithium (0.825 mL, 2 M in cyclohexane, 1.1 eq) and 3,6-di(azetidin-1-yl)-9H-fluoren-9-one (**1e**; 145 mg, 0.5 mmol) were used, affording AF6 as a dark green solid (220 mg, 92%, perchlorate salt). ¹H NMR (400 MHz, CDCl₃) δ 7.34 (s, 2H), 7.23 (d, *J* = 7.4 Hz, 1H), 7.12 (d, *J* = 7.6 Hz, 2H), 6.55 (d, *J* = 8.6 Hz, 2H), 5.78 (d, *J* = 8.6 Hz, 2H), 4.49 (t, *J* = 7.6 Hz, 8H), 2.60 – 2.47 (m, 4H), 2.19 (s, 6H); ¹³C NMR (101 MHz, DMSO) δ 156.66(C), 146.28(C), 135.50(C), 132.48(C), 129.81(C), 128.91(C), 128.42(C), 111.43(C), 110.71(C), 53.46(CH₂), 20.25(CH₃), 16.22(CH₂); Maldi-Tof/Tof-MS calcd for C₂₇H₂₇N₂ [M]⁺ 379.2169, found 379.2245.

AF7 (perchlorate salt): Method 2 was applied. 2-bromo-1,3-dimethylbenzene (277.5 mg, 1.5 mmol, 1 eq), *n*-butyllithium (0.825 mL, 2 M in cyclohexane, 1.1 eq) and 3,6-bis(3,3-difluoroazetidin-1-yl)-9H-fluoren-9-one (**1f**; 181 mg, 0.5 mmol) were used, affording AF7 as a dark green solid (240 mg, 87%, perchlorate salt). ¹H NMR (400 MHz, CDCl₃) δ 7.36 (d, *J* = 2.1 Hz, 2H), 7.26 (t, *J* = 7.6 Hz, 1H), 7.13 (d, *J* = 7.6 Hz, 2H), 6.63 (d, *J* = 8.6 Hz, 2H), 5.82 (dd, *J* = 8.6, 2.1 Hz, 2H), 4.87 – 4.68 (m, 8H), 2.20 (s, 6H); ¹³C NMR (101 MHz, CDCl₃) δ 178.35(C), 157.17(C), 148.14(C), 135.34(C), 134.53(C), 131.39(C), 130.43(C), 129.95(C), 128.14(C), 114.48(C), 113.34(C), 110.91(C), 64.40(CH₂), 20.29(CH₃); Maldi-Tof/Tof-MS calcd for C₂₇H₂₃F₄N₂ [M]⁺ 451.1792, found 451.2016.

AF8 (perchlorate salt): Method 2 was applied. 2-bromo-1,3-dimethylbenzene (277.5 mg, 1.5 mmol, 1 eq), *n*-butyllithium (0.825 mL, 2 M in cyclohexane, 1.1 eq) and 3,6-di(pyrrolidin-1-yl)-9H-fluoren-9-one (**1g**; 159 mg, 0.5 mmol) were used, affording AF8 as a brown solid (215 mg, 85%, perchlorate salt). ¹H NMR (400 MHz, CDCl₃) δ 7.56 (d, *J* = 1.8 Hz, 2H), 7.24 (dd, *J* = 8.1, 7.1 Hz, 1H), 7.12 (d, *J* = 7.7 Hz, 2H), 6.55 (d, *J* = 8.8 Hz, 2H), 5.97 (dd, *J* = 8.8, 1.7 Hz, 2H), 3.81 (s, 8H), 2.21 (s, 6H), 2.09 (t, *J* = 6.6 Hz, 8H); ¹³C NMR (101

MHz, CDCl₃) δ 172.57(C), 155.62(C), 147.49(C), 135.42(C), 132.66(C), 131.06(C), 129.52(C), 129.06(C), 127.65(C), 114.47(C), 111.73(C), 50.19(CH₃), 25.14(CH₂), 20.04(CH₂); Maldi-Tof/Tof-MS calcd for C₂₉H₃₁N₂ [M]⁺ 407.2482, found 407.2552.

AF9 (perchlorate salt): Method 2 was applied. 2-bromo-1,3-dimethylbenzene (277.5 mg, 1.5 mmol, 1 eq), *n*-butyllithium (0.825 mL, 2 M in cyclohexane, 1.1 eq) and 3,6-di(piperidin-1-yl)-9H-fluoren-9-one (**1h**; 173 mg, 0.5 mmol) were used, affording AF9 as a brown solid (216 mg, 81%, perchlorate salt). ¹H NMR (400 MHz, CDCl₃) δ 7.83 (s, 2H), 7.28 – 7.22 (m, 1H), 7.13 (d, *J* = 7.6 Hz, 2H), 6.62 (d, *J* = 8.9 Hz, 2H), 6.30 (s, 2H), 3.86 (s, 8H), 2.21 (s, 6H), 1.89 – 1.73 (m, 12H); ¹³C NMR (101 MHz, CDCl₃) δ 157.23(C), 148.06(C), 135.71(C), 133.09(C), 131.27(C), 129.61(C), 129.31(C), 127.89(C), 113.74(C), 111.38(C), 50.53(CH₂), 27.01(CH₂), 24.50(CH₂), 20.34(CH₃); Maldi-Tof/Tof-MS calcd for C₃₁H₃₅N₂ [M]⁺ 435.2795, found 435.3144.

AF10 (perchlorate salt): Method 2 was applied. 2-bromo-1,3-dimethylbenzene (277.5 mg, 1.5 mmol, 1 eq), *n*-butyllithium (0.825 mL, 2 M in cyclohexane, 1.1 eq) and 3,6-di(azepan-1-yl)-9H-fluoren-9-one (**1i**; 187 mg, 0.5 mmol) were used, affording AF10 as a brown solid (230 mg, 82%, perchlorate salt). ¹H NMR (400 MHz, DMSO) δ 7.63 (d, *J* = 2.2 Hz, 2H), 7.32 (dd, *J* = 8.3, 6.9 Hz, 1H), 7.23 (d, *J* = 7.7 Hz, 2H), 6.63 (d, *J* = 9.0 Hz, 2H), 6.43 (dd, *J* = 9.1, 2.2 Hz, 2H), 3.87 (t, *J* = 5.5 Hz, 8H), 2.17 (s, 6H), 1.80 (s, 9H), 1.55 (s, 8H); ¹³C NMR (101 MHz, DMSO) δ 170.44(C), 157.22(C), 147.57(C), 135.51(C), 133.26(C), 131.25(C), 129.88(C), 129.15(C), 128.42(C), 113.09(C), 112.47(C), 52.24(CH₃), 27.74(CH₂), 26.21(CH₂), 20.37(CH₂); Maldi-Tof/Tof-MS calcd for C₃₃H₃₉N₂ [M]⁺ 463.3108, found 463.3435.

AF11 (perchlorate salt): Method 2 was applied. 2-bromo-1,3-dimethoxybenzene (325 mg, 1.5 mmol, 1 eq), *n*-butyllithium (0.825 mL, 2 M in cyclohexane, 1.1 eq) and 3,6-di(azepan-1-yl)-9H-fluoren-9-one (**1i**; 187 mg, 0.5 mmol) were used, affording AF11 as a dark red solid (273 mg, 92%, perchlorate salt). ¹H NMR (400 MHz, CDCl₃) δ 7.43 (d, *J* = 2.2 Hz, 2H), 7.39 (t, *J* = 8.4 Hz, 1H), 6.72 (d, *J* = 9.0 Hz, 2H), 6.65 (d, *J* = 8.5 Hz, 2H), 6.10 (dd, *J* = 9.0, 2.2 Hz, 2H), 3.91 – 3.72 (m, 8H), 3.77 (s, 6H), 1.95 – 1.84 (m, 8H), 1.60 – 1.55 (m, 8H); ¹³C NMR (101 MHz, CDCl₃) δ 166.42(C), 158.50(C), 157.17(C), 148.24(C), 134.13(C), 132.63(C), 130.34(C), 112.11(C), 110.58(C), 109.75(C), 104.53(C), 56.24(CH₃), 52.21(CH₂), 27.90(CH₂), 26.61(CH₂); Maldi-Tof/Tof-MS calcd for C₃₃H₃₉N₂O₂ [M]⁺ 495.3006, found 495.3227.

AF12 (perchlorate salt): Method 2 was applied. 2-bromo-1,3-dimethylbenzene (277.5 mg, 1.5 mmol, 1 eq), *n*-butyllithium (0.825 mL, 2 M in cyclohexane, 1.1 eq) and 3,6-bis(methyl(phenyl)amino)-9H-fluoren-9-one (**1j**; 195 mg, 0.5 mmol) were used, affording AF12 as a brown solid (251 mg, 87%, perchlorate salt). ¹H NMR

(400 MHz, CDCl₃) δ 7.60 (d, J = 2.2 Hz, 2H), 7.50 (dd, J = 10.5, 4.8 Hz, 4H), 7.43 – 7.36 (m, 2H), 7.26 – 7.20 (m, 4H), 7.09 (d, J = 7.7 Hz, 2H), 6.98 (s, 1H), 6.42 (d, J = 8.9 Hz, 2H), 5.88 – 5.85 (m, 2H), 3.83 (s, 6H), 2.22 (s, 6H); ¹³C NMR (101 MHz, CDCl₃) δ 175.57(C), 158.99(C), 148.63(C), 144.83(C), 135.45(C), 133.12(C), 131.07(C), 130.47(C), 129.62(C), 128.76(C), 128.23(C), 127.97(C), 126.73(C), 114.20(C), 114.10(C), 43.23(CH₃), 20.35(CH₃); Maldi-Tof/Tof-MS calcd for C₃₅H₃₁N₂ [M]⁺ 479.2482, found 479.2677.

AF13 (perchlorate salt): Method 2 was applied. 2-bromo-1,3-dimethylbenzene (277.5 mg, 1.5 mmol, 1 eq), *n*-butyllithium (0.825 mL, 2 M in cyclohexane, 1.1 eq) and 3,6-bis(diphenylamino)-9H-fluoren-9-one (**1k**; 257 mg, 0.5 mmol) were used, affording AF13 as a purple solid (327 mg, 93%, perchlorate salt). ¹H NMR (400 MHz, CDCl₃) δ 7.45 (t, J = 7.7 Hz, 8H), 7.34 (t, J = 7.4 Hz, 4H), 7.26 – 7.22 (m, 9H), 7.13 (d, J = 7.6 Hz, 2H), 6.75 (d, J = 2.0 Hz, 2H), 6.63 (d, J = 8.8 Hz, 2H), 6.22 (dd, J = 8.8, 1.8 Hz, 2H), 2.28 (s, 6H); ¹³C NMR (101 MHz, CDCl₃) δ 175.99(C), 158.30(C), 147.34(C), 143.64(C), 135.63(C), 134.39(C), 133.64(C), 130.47(C), 130.08(C), 128.70(C), 128.17(C), 127.62(C), 118.48(C), 116.80(C), 20.60(CH₃); Maldi-Tof/Tof-MS calcd for C₄₅H₃₅N₂ [M]⁺ 603.2795, found 603.3048.

AF14 (perchlorate salt): Method 2 was applied. 2-bromo-1,3-dimethylbenzene (277.5 mg, 1.5 mmol, 1 eq), *n*-butyllithium (0.825 mL, 2 M in cyclohexane, 1.1 eq) and 3,6-di(indolin-1-yl)-9H-fluoren-9-one (**1m**; 207 mg, 0.5 mmol) were used, affording AF14 as a purple solid (247 mg, 82%, perchlorate salt). ¹H NMR (400 MHz, CDCl₃) δ 7.87 (s, 2H), 7.43 (d, J = 8.1 Hz, 2H), 7.35 – 7.26 (m, 5H), 7.19 – 7.09 (m, 4H), 6.91 (d, J = 8.9 Hz, 2H), 6.71 (d, J = 8.8 Hz, 2H), 4.67 (t, J = 7.5 Hz, 4H), 3.30 (t, J = 7.4 Hz, 4H), 2.27 (s, 6H); ¹³C NMR (101 MHz, CDCl₃) δ 172.23(C), 153.17(C), 147.80(C), 142.61(C), 136.93(C), 135.75(C), 132.87(C), 132.50(C), 129.66(C), 128.29(C), 128.07(C), 126.50(C), 125.93(C), 117.30(C), 116.73(C), 114.39(C), 54.74(CH₃), 28.63(CH₂), 20.49(CH₂); Maldi-Tof/Tof-MS calcd for C₃₇H₃₁N₂ [M]⁺ 503.2482, found 503.2802.

3) Procedure for preparation of AF derivatives

Preparation of AF2Ac: AF2 (33 mg, 0.075 mmol, 1 eq) was added into a round bottom flask, and dissolved with dried DCM (5 mL). Triethylamine (38 mg, 0.375 mmol, 5 eq) was then added and the solution was stirred rigorously. Then acetyl chloride (30 mg, 0.375 mol, 5 eq) was added and stirred for further 10 min. It was subsequently concentrated to dryness, washed with PE and EA (1:1, v/v, 10 mL totally) and filtrated (3 times) to afford AF2Ac as a yellow solid (19 mg, 69%). ¹H NMR (400 MHz, CDCl₃) δ 7.45 (s, 1H), 7.23 (d, J = 7.8 Hz, 1H), 7.15 (d, J = 7.5 Hz, 2H), 7.03 (dd, J = 7.8, 1.7 Hz, 1H), 6.86 (d, J = 7.9 Hz, 1H), 6.74 (s, 1H),

6.50 (d, $J = 9.7$ Hz, 1H), 5.88 (d, $J = 9.1$ Hz, 1H), 3.30 (s, 3H), 2.70 (s, 3H), 2.12 (d, $J = 6.5$ Hz, 6H), 1.94 (s, 3H). Maldi-Tof/Tof-MS calcd for $C_{25}H_{24}N_2O$ $[M+H]^+$ 369.1967, found 368.9492.

Preparation of AF2B: (4-(4,4,5,5-Tetramethyl-1,3,2-dioxaborolan-2-yl)phenyl)methanol (936 mg, 4 mmol) was dissolved with dry THF (30 mL) in a round bottom flask. The solution was stirred in ice bath, added with triethylamine (606 mg, 6 mmol) and triphosgen (594 mg, 2 mmol), and then further stirred for 4 h. The reaction mixture was filtered off through a celite pad, concentrated to dryness to afford chloridate intermediate as yellowish oil. AF2 (44 mg, 0.1 mmol, 1 eq) was dissolved with dry DCM (5 mL) in a round bottom flask. The solution was stirred in ice bath, then added with DIPEA (52 mg, 0.4 mmol, 4eq) and stirred for 5 min. A solution of chloridate intermediate (45 mg, 0.15 mmol, 1.5 eq) in 2 mL DCM was added into the reaction mixture and further stirred for 30 min. The reaction mixture was diluted with 30 mL DCM and washed with 0.1 M HCl, and concentrated to dryness. Et₂O (10 mL) was then added and the solution was filtered to collect the precipitate as crude product. The crude product was purified with HPLC (MeOH/H₂O, 0.1% TFA was added) to afford AF2B as a dark green solid (10 mg, 20%). ¹H NMR (400 MHz, CDCl₃) δ 8.03 – 7.75 (m, 2H), 7.47 – 7.29 (m, 3H), 7.21 – 7.06 (m, 2H), 6.91 (dd, $J = 48.3, 8.6$ Hz, 2H), 6.80 – 6.57 (m, 3H), 6.42 (d, $J = 9.0$ Hz, 1H), 5.24 (s, 2H), 3.47 (s, 3H), 3.29 (s, 3H), 2.17 (s, 6H). HRMS (ESI) calcd for $C_{31}H_{29}BN_2O_4$ $[M+H]^+$ 505.2299, found 505.2292.

Preparation of intermediate 2: A round bottom flask was dried, sealed and vacuated/backfilled with nitrogen (3 $\times$), then charged with 2,5-dibromo-1,3-dimethylbenzene (5.28 g, 20 mmol, 1 eq) and dry THF (60 mL). After cooling the solution to -78 °C, *n*-butyllithium (10 mL, 2 M in cyclohexane, 1 eq) was slowly added and the reaction was stirred vigorously for 2 h. A solution of bis(tert-butoxycarbonyl)oxide (4.37 g, 20 mmol, 1 eq) was dissolved in dry THF (15 mL), added into reaction mixture dropwise, and further stirred for 2 h. The reaction was quenched with water and extracted with DCM (2 $\times$). The combined organics were dried (Na₂SO₄), filtered, evaporated, and purified with silica gel chromatography (100% PE) to afford intermediate 2 as a colorless oil (1.94 g, 34%).

Preparation of AF3-COOH: Method 2 was applied to afford AF3-COOH *tert*-butyl ester as an intermediate. intermediate 2 (427 mg, 1.5 mmol, 1 eq), *n*-butyllithium (0.825 mL, 2 M in cyclohexane, 1.1 eq) and 3,6-bis(dimethylamino)-9H-fluoren-9-one (**1c**; 133 mg, 0.5 mmol) were used, affording AF3-COOH *tert*-butyl ester as a brown solid. Then the intermediate was dissolved with DCM/TFA (4:1, v/v, 10 mL totally) and stirred under room temperature for 12 h. Toluene (10 mL) was added and the reaction mixture was concentrated to dryness, further dried under vacuum pump overnight to afford AF3-COOH as a brown solid

(161 mg, 63%, TFA salt). ¹H NMR (400 MHz, DMSO) δ 7.79 (s, 2H), 7.62 (d, J = 1.7 Hz, 2H), 6.65 (d, J = 9.0 Hz, 2H), 6.37 (dd, J = 9.0, 1.7 Hz, 2H), 3.38 (s, 12H), 2.21 (s, 6H). ¹³C NMR (101 MHz, DMSO) δ 169.03 (C), 167.70 (C), 158.07 (C), 146.97 (C), 136.31 (C), 135.88 (C), 132.79 (C), 131.96 (C), 129.14 (C), 128.78 (C), 113.62 (C), 113.04 (C), 42.31 (CH₃), 20.20 (CH₃). Maldi-Tof/Tof-MS calcd for C₂₆H₂₇N₂O₂ [M]⁺ 399.2067, found 399.2165.

Preparation of AF2-COOH: Method 1 was applied. Intermediate **2** (427 mg, 1.5 mmol, 1 eq), *n*-butyllithium (0.825 mL, 2 M in cyclohexane, 1.1 eq), 3,6-bis(*tert*-butyl methylcarbamate)-9H-fluoren-9-one (**1b**; 219 mg, 0.5 mmol) were used, affording AF2-COOH as a green solid (121 mg, 50% total, TFA salt). ¹H NMR (400 MHz, DMSO) δ 9.29 (s, 2H), 7.78 (s, 2H), 7.20 (s, 2H), 6.65 (d, J = 7.8 Hz, 2H), 6.23 (d, J = 8.5 Hz, 2H), 3.07 (s, 6H), 2.22 (s, 6H); ¹³C NMR (101 MHz, DMSO) δ 169.18 (C), 167.68 (C), 159.42 (C), 147.03 (C), 136.32 (C), 135.82 (C), 133.45 (C), 131.95 (C), 129.09 (C), 128.77 (C), 31.17 (CH₃), 20.20 (CH₃); Maldi-Tof/Tof-MS calcd for C₂₄H₂₃N₂O₂ [M]⁺ 371.1754, found 371.1864.

Preparation of intermediate 3: 2-bromo-5-iodo-*m*-xylene (3.11 g, 10 mmol, 1 eq) was dissolved with diisopropylamine (50 mL) in a round bottom flask, stirred rigorously under room temperature. [PdCl₂(PPh₃)₂] (210 mg, 0.3 mmol, 0.03 eq), CuI (114 mg, 0.6 mmol, 0.06 eq) and ethynyltrimethylsilane (1.18 g, 12 mmol, 1.2 eq) were then added and further stirred for 4 h. The reaction mixture was diluted with EA (200 mL) and washed with water (3×). The organic phase was dried (Na₂SO₄), filtered, evaporated, and purified with silica gel chromatography (100% PE) to afford intermediate **3** as a yellow oil (2.67 g, 95%).

Preparation of intermediate 4: Intermediate **3** (1.40 g, 5 mmol, 1 eq) and K₂CO₃ (1.38 g, 2 eq) were added into a round bottom flask, then MeOH (20 mL) was added, and stirred under room temperature for 12 h. The reaction mixture was diluted with water (200 mL), extracted with Et₂O (3×). The combined organics were dried (Na₂SO₄), filtered and evaporated to afford intermediate **4** as a yellow oil (0.91 g, 87%).

Dextran-N₃ was prepared according to the reported procedure. All characterizations of intermediate **5** and **6** can be found in reference.³

Preparation of intermediate 5: 3-bromopropan-1-amine hydrobromide (3.28 g, 15 mmol, 1 eq) and NaN₃ (3.32 g, 51 mmol, 3.4 eq) were added into a round bottom flask, dissolved with water (25 mL), stirred, and heated to reflux for 15 h. The reaction mixture was cooled to room temperature, Et₂O (25 mL) was added and cooled to 0 °C, then KOH (4 g) was added. After KOH was totally dissolved, another 25 mL Et₂O was added and the organic phase was then separated. The water phase was further extracted with Et₂O (2×). The combined organics were dried (Na₂SO₄), filtered and evaporated carefully to afford intermediate **5** as a yellow

liquid (608 mg, 40%).

Preparation of intermediate 6: Intermediate **5** (1.8 g, 18 mmol, 1 eq) was added into a round bottom flask, dissolved with DCM (60 mL) and stirred under room temperature, then a solution of bis(4-nitrophenyl) carbonate (5.4 g, 18 mmol, 1 eq) in DCM (40 mL) was added dropwise in 10 min, and further stirred for 4 h. A solution of NaOH (792 mg, 19.8 mmol, 1.1 eq) in water (60 mL) was added and stirred, then organic phase was separated, dried (Na₂SO₄), filtered, evaporated and purified with silica gel chromatography (PE/EA, 3/1, v/v) to afford intermediate **6** as a white solid (2.57 g, 54%).

Preparation of Dextran-N₃: High vacuum dried Dextran-40000 (2 g, 0.05 mmol, 1 eq) and LiCl (2 g) was dissolved with dry DMF in a dried round bottom flask under N₂ atmosphere and stirred. DIPEA (2 mL) and intermediate **6** (172 mg, 0.65 mmol, 13 eq) was then added. The reaction mixture was heated to 70 °C and further stirred for 24 h, then cooled down and concentrated to dryness, dissolved with water and dialyzed against water for 3 days with 3× per day exchange of dialysate for fresh water. The water solution was finally lyophilized to afford Dextran-N₃ as a white solid (1.76 g, 85%). The amount of N₃ group in each Dextran-N₃ molecule was quantified to be 11.97 based on ¹H-NMR.

Preparation of AF3-Alkyne: Method 2 was applied. intermediate **4** (313 mg, 1.5 mmol, 1 eq), *n*-butyllithium (1.575 mL, 2 M in cyclohexane, 2.1 eq, added in excess of 1 equivalent to neutralize the alkynyl-H in intermediate **4**) and 3,6-bis(dimethylamino)-9H-fluoren-9-one (**1c**; 133 mg, 0.5 mmol) were used, affording AF3-Alkyne as a brown solid (119 mg, 50%, perchlorate salt). ¹H NMR (400 MHz, CDCl₃) δ 7.64 (s, 2H), 7.27 (s, 2H), 6.54 (d, J = 8.9 Hz, 2H), 6.08 (d, J = 8.9 Hz, 2H), 3.44 (s, 12H), 3.11 (s, 1H), 2.19 (s, 6H); ¹³C NMR (101 MHz, CDCl₃) δ 172.04 (C), 158.60 (C), 147.82 (C), 136.04 (C), 132.92 (C), 131.93 (C), 131.46 (C), 129.59 (C), 123.17 (C), 113.94 (C), 111.47 (C), 83.23 (C), 78.26 (CH), 42.40 (CH₃), 20.12 (CH₃); Maldi-Tof/Tof-MS calcd for C₂₇H₂₇N₂ [M]⁺ 379.2169, found 379.2277.

Preparation of AF3-Dextran: Dextran-N₃ (59 mg, 0.0014 mmol, 1 eq), AF3-Alkyne (8 mg, 0.0168 mmol, 12 eq), copper(II) sulfate pentahydrate (4.2 g, 0.0168 mmol, 12 eq) and DMSO (5 mL) was added in a dried round bottom flask under N₂ atmosphere and stirred under room temperature, then sodium ascorbate (3.9 mg, 0.0192 mmol, 14 eq) was added and the reaction mixture was further stirred for 24 h. The solution was dialyzed against water (3 days, 3× water exchange per day) and lyophilized to afford AF3-Dextran as a brown solid (60 mg, 81%). The amount of AF3 in each AF3-Dextran molecule was measured to be 4.7 according to the absorption spectra (Using the absorption coefficient of AF3 as a reference, the peak absorbance of an AF3-Dextran PBS solution was measured at 0.367, which spectrally correlated to 17.7 μM AF3. Then 50 mL

of this solution (containing 0.885 μmol AF3) underwent lyophilized and was then weighed, registering a mass of 8.1 mg. Considering the molecular weight of AF3-Dextran as 43000 Da and the amount of AF3-Dextran was 0.188 μmol , then the average number of AF3 groups per molecule can be calculated to be $0.885/0.184=4.7$).

Supplementary Tables

Supplementary Table 1 | Molecular weights, absorption/emission wavelengths and wavelength-to-molecular-weight ratios of AF dyes and the fluorescent dyes in Figure 1.

Dye	Molecular weight (g/mol)	λ_{abs} (nm)	λ_{em} (nm)	Wavelength-to-molecular-weight ratio ($\lambda_{\text{abs}}/\text{Mw}$)	Dye	Molecular weight (g/mol)	λ_{abs} (nm)	λ_{em} (nm)	Wavelength-to-molecular-weight ratio ($\lambda_{\text{abs}}/\text{Mw}$)
AF1	299	800	976	2.676	18	848	868	562	1.509
AF2	327	847	933	2.590	19	636	658	646	0.985
AF3	355	959	1014	2.701	20	688	701	534	1.288
AF4	387	983	1046	2.540	21	910	1060	747	1.218
AF5	411	966	1015	2.350	22	625	698	600	1.042
AF6	379	962	1014	2.538	23	602	618	412	1.461
AF7	451	946	995	2.098	24	688	715	557	1.235
AF8	407	968	1017	2.378	25	742	764	521	1.424
AF9	435	966	1026	2.221	26	775	831	751	1.032
AF10	463	973	1026	2.102	27	981	1032	547	1.793
AF11	495	994	1045	2.008	28	1046	1080	742	1.410
AF12	479	965	1029	2.015	29	1080	1130	610	1.770
AF13	603	1045	1196	1.733	30	650	684	511	1.272
AF14	503	1101	1187	2.189	31	746	766	537	1.389
1	851	907	546	1.559	32	862	908	563	1.531
2	701	816	446	1.572	33	1026	1045	664	1.545
3	641	659	309	2.074	34	1072	1103	1019	1.052
4	646	660	399	1.619	35	700	743	439	1.595
5	721	740	531	1.358	36	630	807	603	1.045
6	700	728	391	1.790	37	666	736	495	1.345
7	732	745	362	2.022	38	1014	1070	652	1.555
8	654	744	474	1.380	39	932	980	509	1.831
9	817	853	424	1.927	40	709	975	680	1.043
10	880	960	659	1.335	41	879	1120	845	1.040
11	880	921	711	1.238	42	763	1065	800	0.954
12	665	683	284	2.342	43	932	1230	894	1.043
13	621	679	440	1.411	44	750	1055	969	0.774
14	596	740	299	1.993	45	627	633	314	1.997
15	668	770	324	2.062	46	698	705	514	1.358
16	686	710	770	0.891	47	745	750	714	1.043
17	723	738	552	1.310					

Supplementary Table 2 | Photophysical properties of AF3 in different solvents.

Solvent	λ_{abs} (nm)	ϵ ($\text{M}^{-1} \text{cm}^{-1}$)	λ_{em} (nm)	Stokes shift (nm)	Quantum yield ($10^{-2} \%$)
DCM	959	23052	1014	55	1.1
Toluene	946	27288	993	47	2.0
Dioxane	940	23765	997	57	1.4
DMF	949	18773	1015	66	0.9
DMSO	951	19618	1022	71	1.1
EtOH	950	20368	1006	55	0.7
PBS	942	20764	1022	80	0.4

Supplementary Table 3 | Crystal data and structure refinements for AF3.

Compound	AF3
CCDC No.	2263552
Formula	$\text{C}_{13}\text{H}_{14.50}\text{Cl}_{1.50}\text{NO}_2$
Formula wt.	269.93
T (K)	173
Wavelength ($\text{\AA}$)	1.34138 $\text{\AA}$
Crystal size	0.23×0.16×0.03
Crystal system	Monoclinic
Space group	$P2_1/n$
a/ $\text{\AA}$	11.2051 (4)
b/ $\text{\AA}$	8.7168 (3)
c/ $\text{\AA}$	25.7043 (10)
α (deg.)	90
β (deg.)	91.984 (2)
γ (deg.)	90
Volume/ $\text{\AA}^3$	2509.10 (16)
Z	8
Density (calcd.)	1.429
μ (mm^{-1})	2.388
θ (deg.)	2.99 to 59.40
F(000)	1128
Completeness	1.000
Goodness-of-fit	1.045
final R indices ($I > 2\sigma(I)$)	0.0428/0.1158
R indices (all data)	0.0495/0.1220
largest diff. peak and hole ($\text{e \AA}^{-3}$)	0.491/-0.515

Supplementary Table 4 | MCI indexes (in electrons) for AF3 in the S₀, S₁, and T₁ states computed at various levels.

Methods	S ₀	S ₁	T ₁
LC-BLYP*	0.1025	0.1030 (Δ SCF) 0.1051/0.1039 (TD/TDA) ^a	0.1126 (UDFT) ^b 0.1092 (TDA) ^a
B3LYP	0.0654	0.0670 (TD)	0.0726 (UDFT) ^b
CASSCF(2,2)	0.0159	0.0183	0.0224

^athe Tamm–Dancoff approximation (TDA) scheme of TD-DFT is shown to reliably predict triplet excited state versus traditional TD-DFT. ^bunrestricted density functional theory (UDFT).

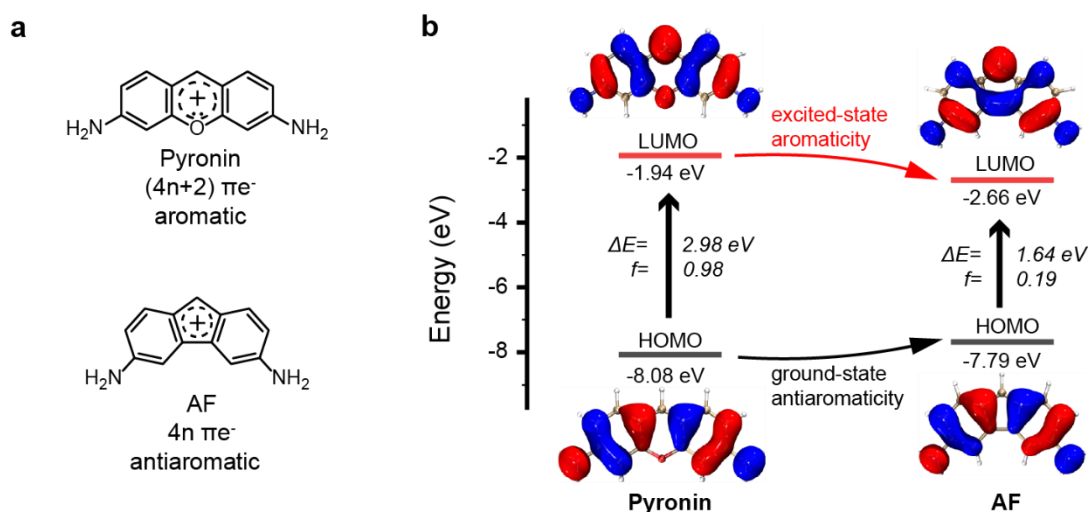
Supplementary Table 5 | Molecular weight, net charge and cLog D of modified and unmodified AF dyes. cLog D was calculated using ChemAxon software.

Name	Molecular weight (Da)	Net charge (pH = 7)	cLogD (pH = 7)
AF2	327	+1	1.92
AF2-COOH	370	0	-2.27
AF3	355	+1	3.30
AF3-COOH	399	0	-0.89
AF3-Dextran	~43300	—	—

Supplementary Table 6 | Calculated NICS(1)_{zz} values (ppm) based on the central five-membered ring of AF3 using three methods including B3LYP, LC-BLYP* and CASSCF(2,2) with a 6-31G(d) basis set. Note that NICS(1)_{zz} was defined as the zz component of the NICS value located 1 angstrom perpendicular to the plane above the five-membered ring. The CASSCF calculations were performed using Dalton 2022 software.^{41,42}

State	B3LYP	LC-BLYP*	CASSCF(2,2)
S ₀	34.42	37.33	32.07
S ₁	-23.87	-25.30	-66.04

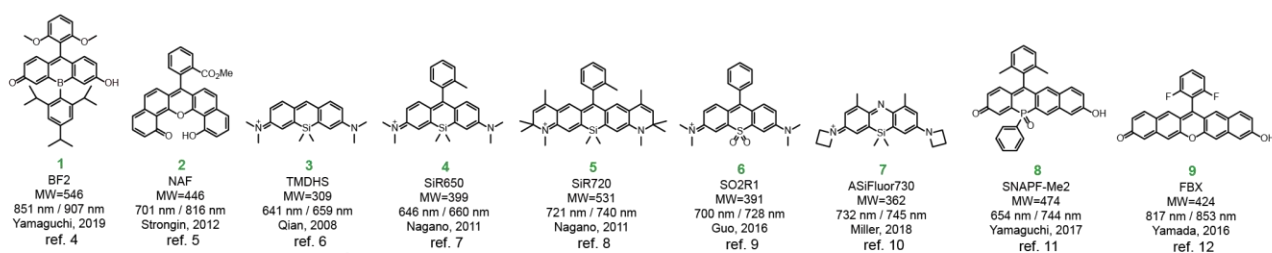
Supplementary Figures



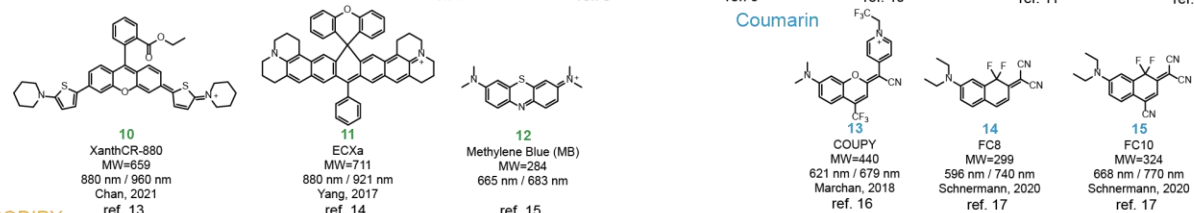
Supplementary Figure 1 | Schematic illustration of aromatic and antiaromatic molecular

skeleton. a, Schematically shows the structure of AF skeleton with a $4n\pi$ -electron five-membered ring as an antiaromatic core and corresponding pyronin skeleton with a $(4n+2)\pi$ -electron aromatic core. **b**, HOMO/LUMO energy level and ΔE comparison of AF and pyronin skeleton shows the gap narrowing effect of antiaromatic skeleton introduction. The calculation was conducted using (TD)DFT at the LC-BLYP*/def-TZVP with DCM in PCM solvent model.

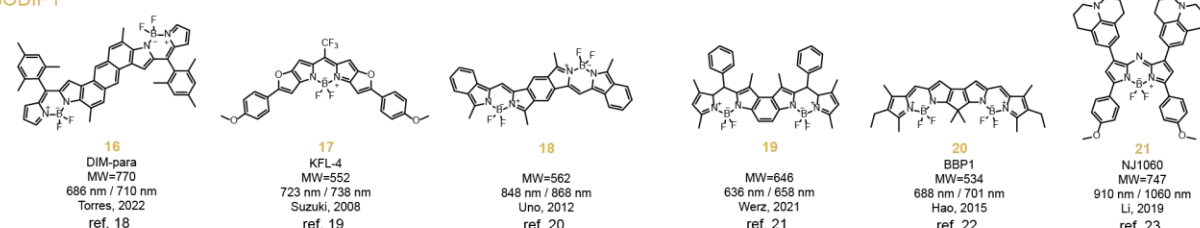
Xanthene



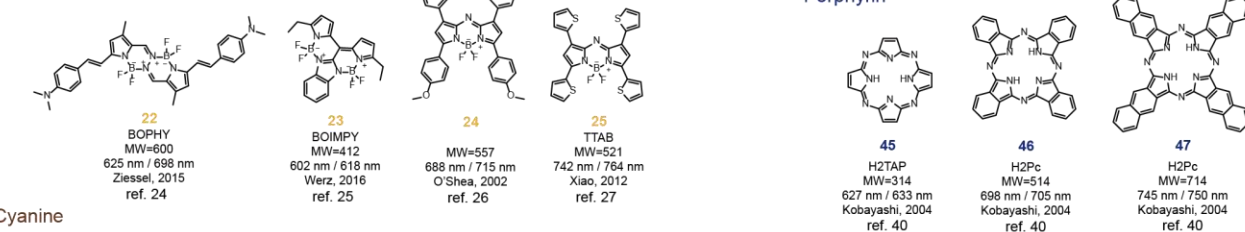
Coumarin



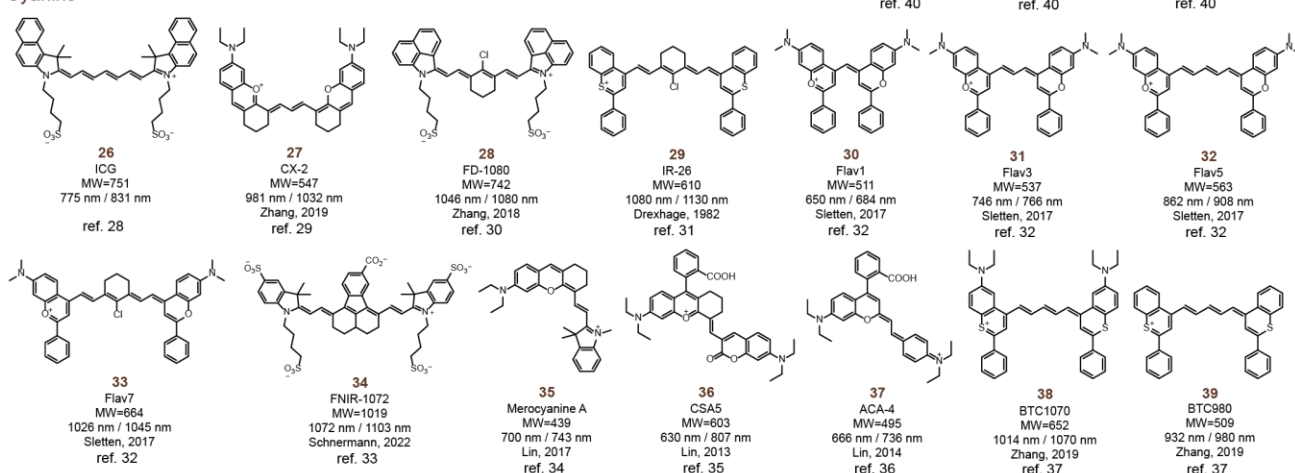
BODIPY



Porphyrin



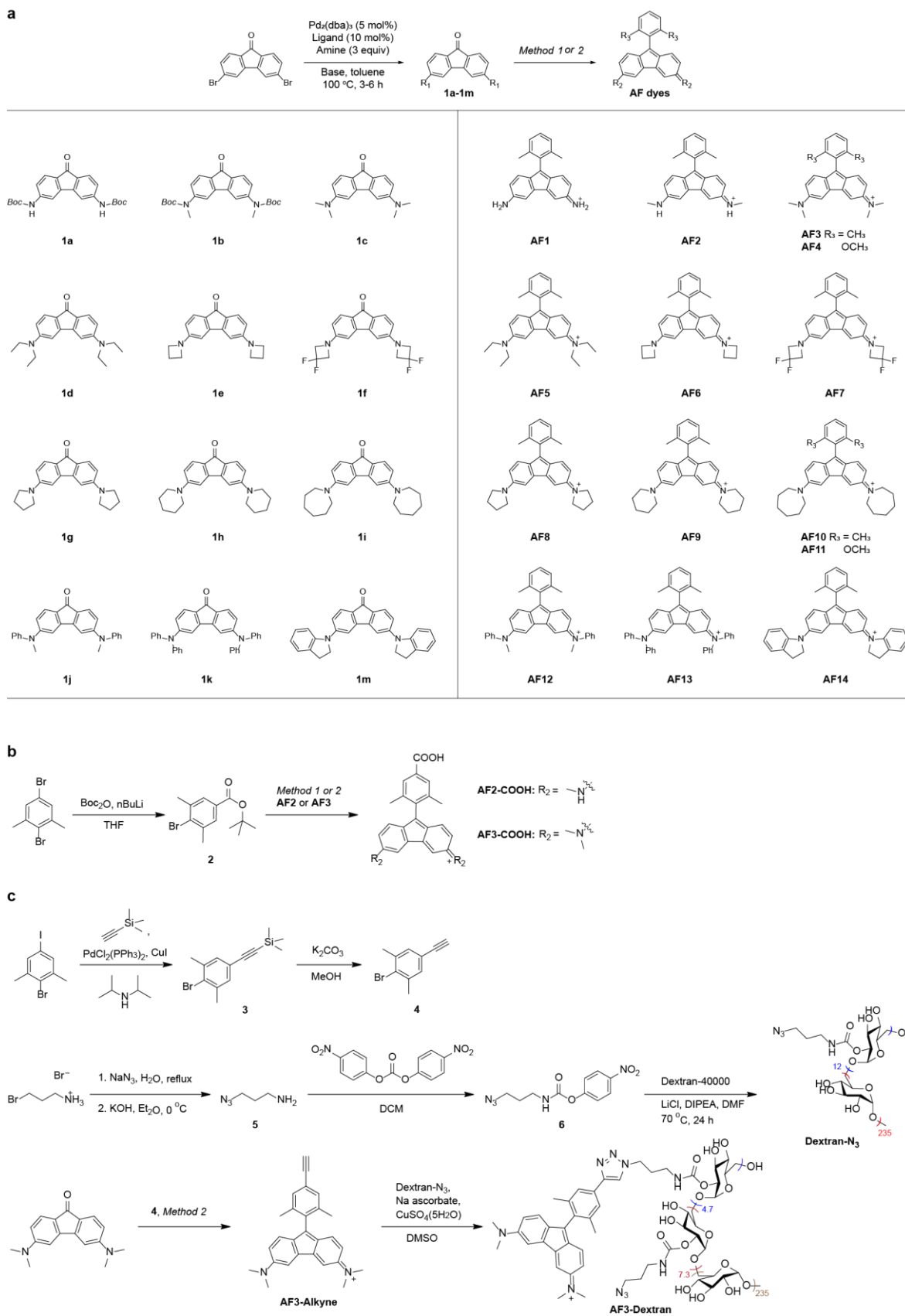
Cyanine



D-A-D

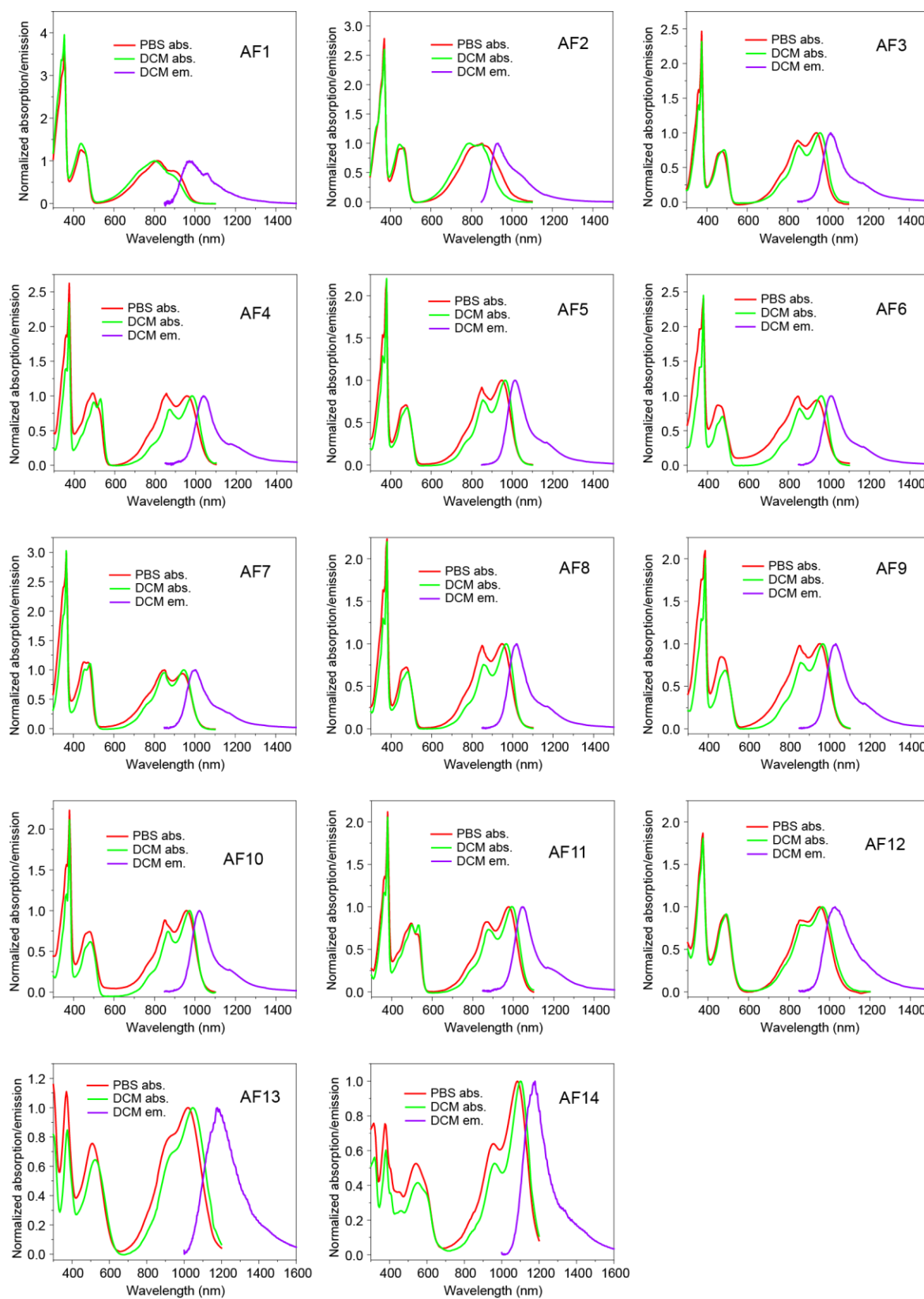


Supplementary Figure 2 | Molecular structures and absorption/emission wavelength of compared fluorescent dyes in Figure 1.



Supplementary Figure 3 | Synthetic route of AF dyes and derivatives. a, Synthetic route and corresponding structure of AF dyes. **b**, Synthetic route of AF2-COOH and AF3-COOH. **c**, Synthetic

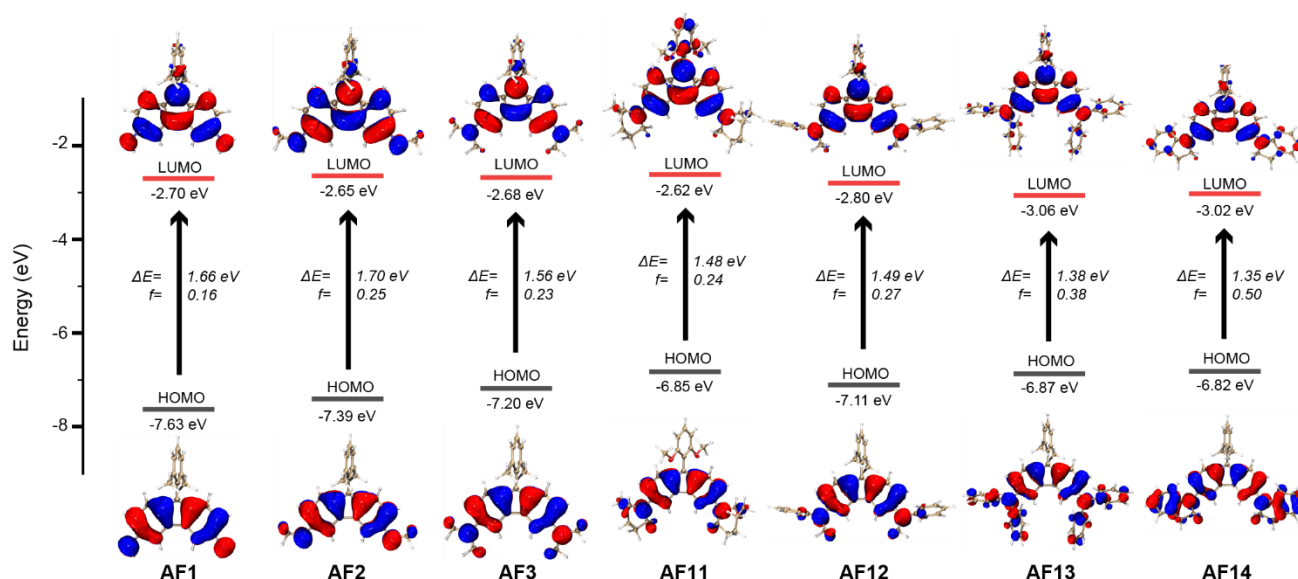
route of AF3-Dextran.



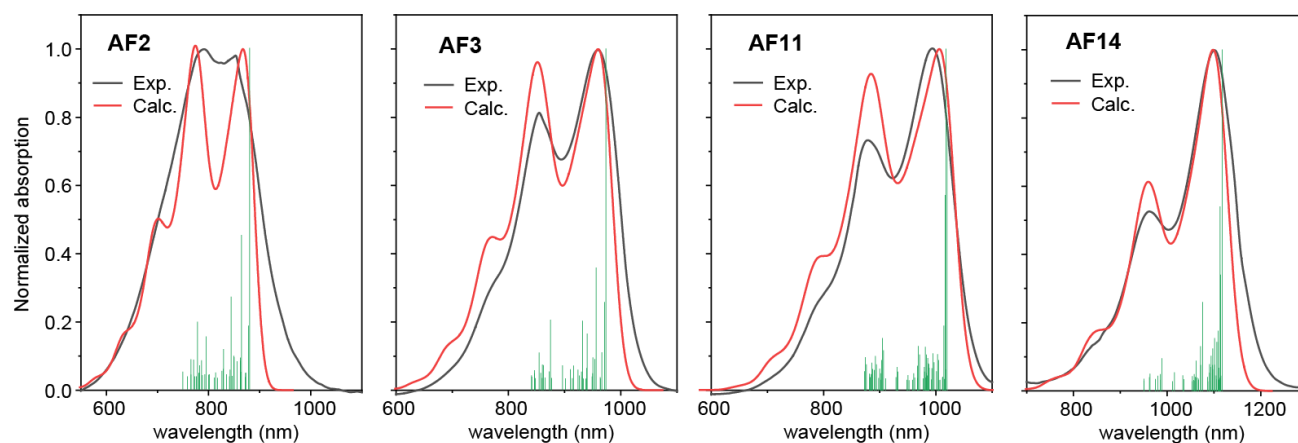
Supplementary Figure 4 | Normalized absorption spectra and emission spectra of AF1-AF14.

Absorption spectra were collected in 1×PBS 7.4 solution and DCM, emission spectra were collected

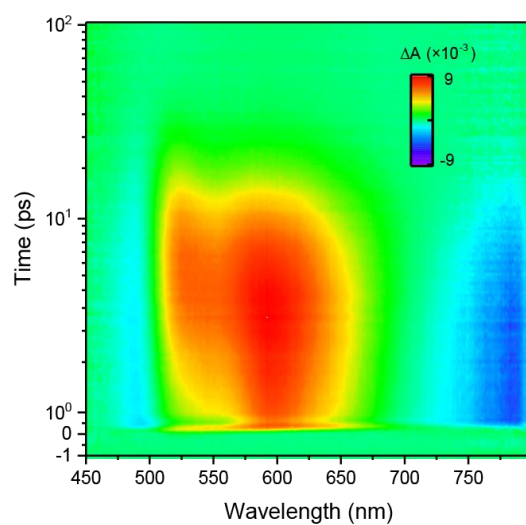
in DCM under 808 nm excitation.



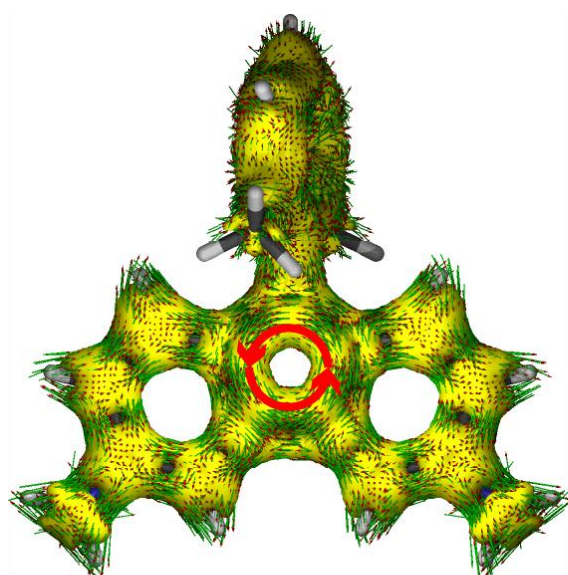
Supplementary Figure 5 | Comparison of the HOMO and LUMO energy levels, S_0 - S_1 excitation energies and oscillator strengths for AF1, AF2, AF3, AF11, AF12, AF13 and AF14 based on TDDFT calculations at the LC-BLYP*/def-TZVP with DCM in PCM solvent model.



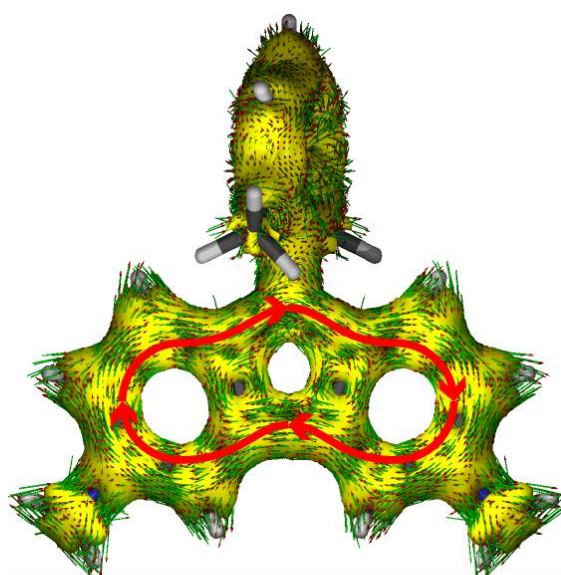
Supplementary Figure 6 | Vibration resolved absorption spectra (red) and experimental spectra (black) of AF dyes. Green sticks show the normalized strength of each mainly contributed vibration resolved transition. Gaussian broadening was applied for the calculated spectra and half width at half maximum (HWHM) was set to 225 cm^{-1} . The calculated results were shifted (AF3: +20 nm; AF14: +10 nm) to fit the experimental results better.



Supplementary Figure 7 | Femtosecond transient absorption contour maps of AF3 under 375 nm pump excitation.



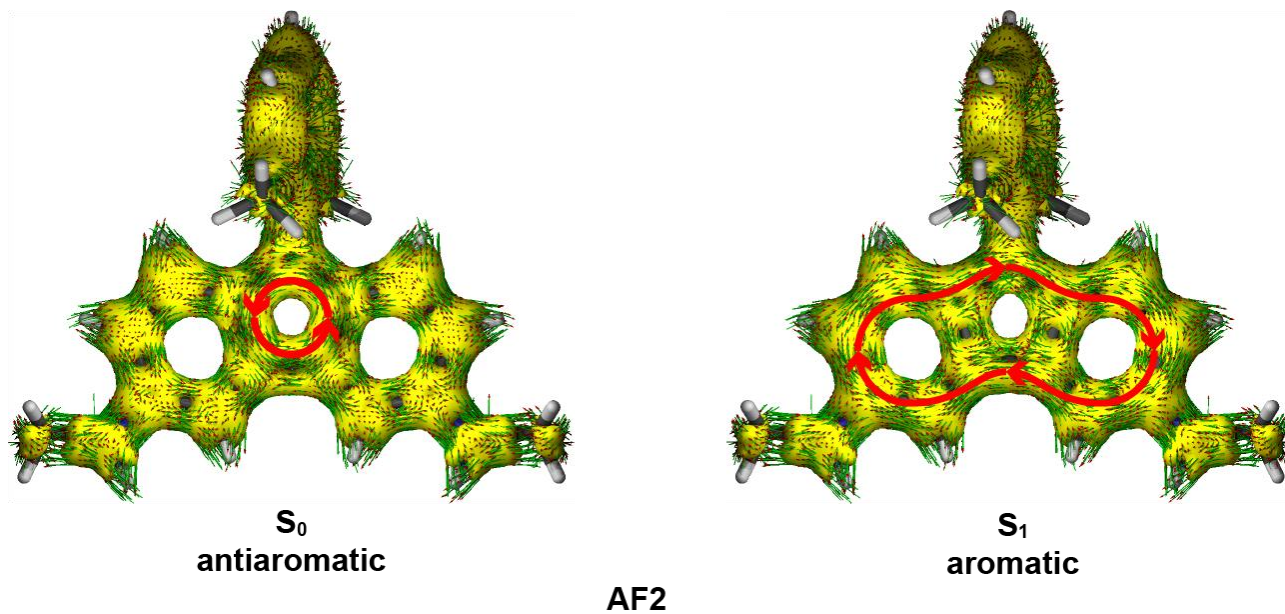
S₀
antiaromatic



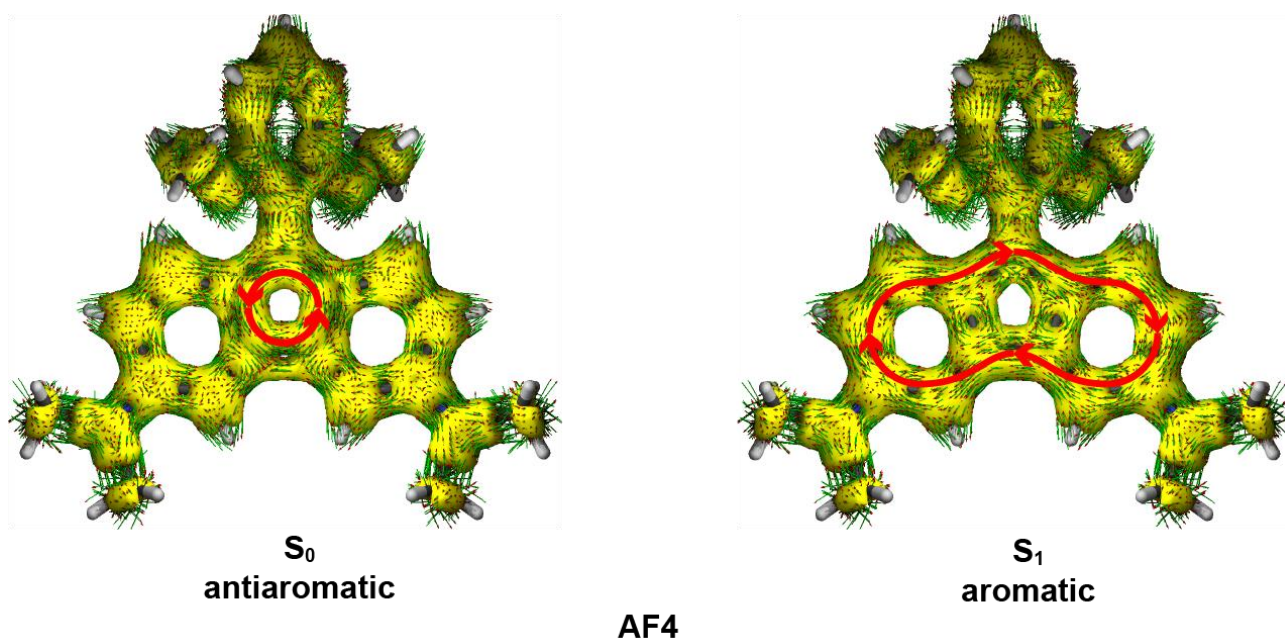
S₁
aromatic

AF1

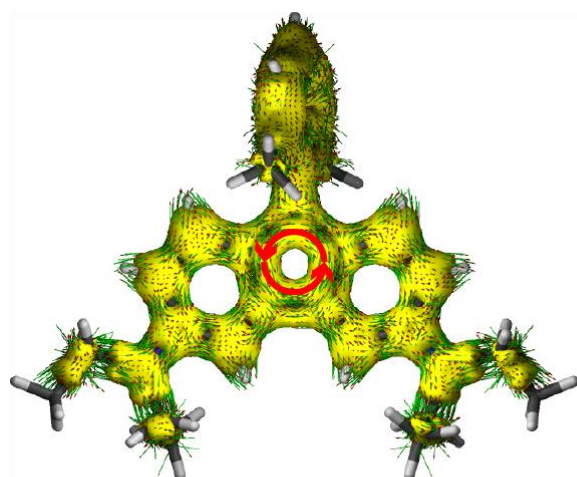
Supplementary Figure 8 | Theoretical calculated AICD diagram of AF1 in S₀ and S₁ state.



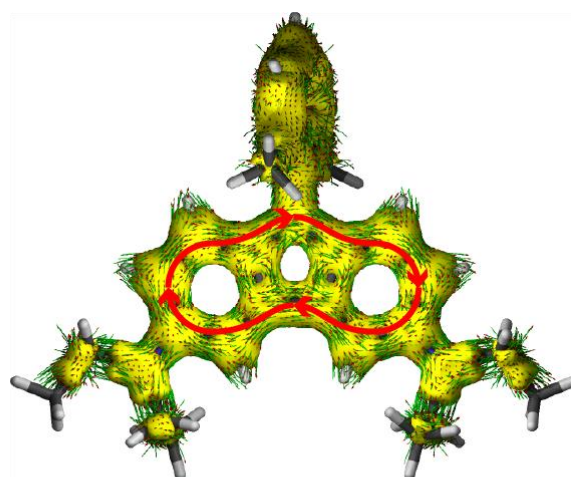
Supplementary Figure 9 | Theoretical calculated AICD diagram of AF2 in S₀ and S₁ state.



Supplementary Figure 10 | Theoretical calculated AICD diagram of AF4 in S₀ and S₁ state.



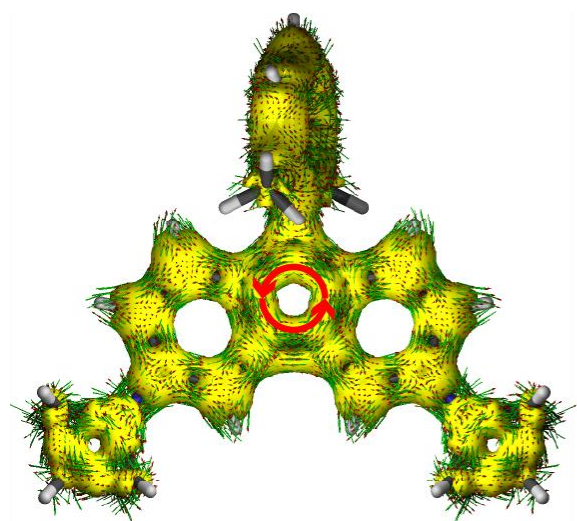
S_0
antiaromatic



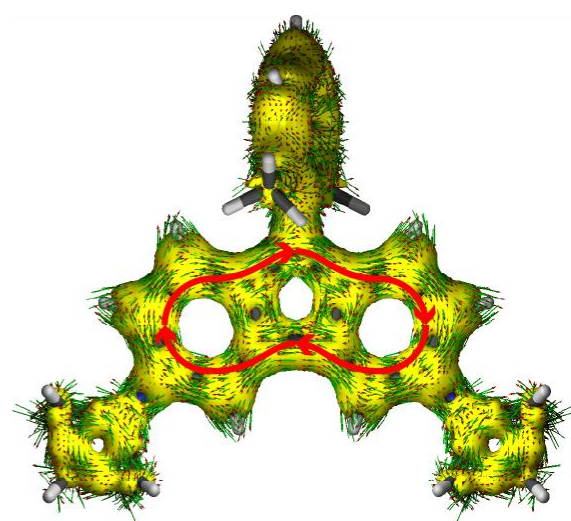
S_1
aromatic

AF5

Supplementary Figure 11 | Theoretical calculated AICD diagram of AF5 in S_0 and S_1 state.



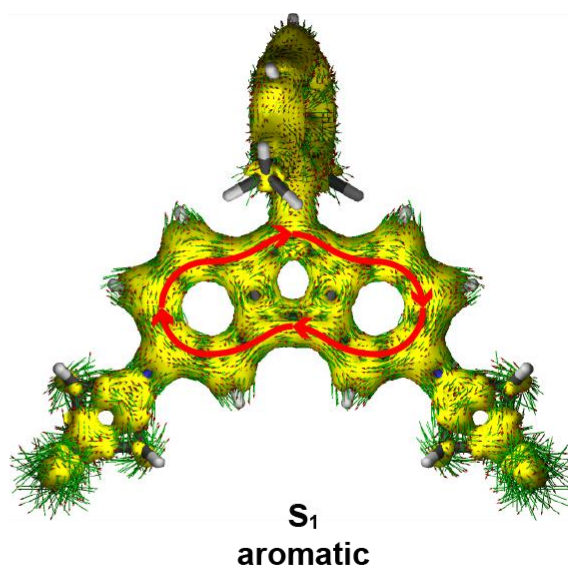
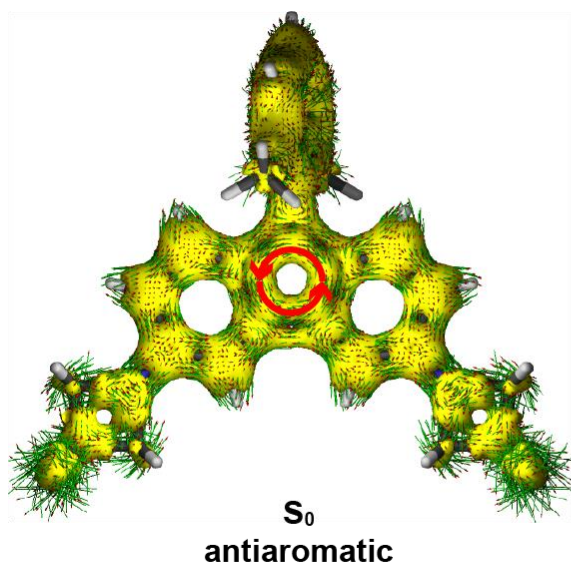
S_0
antiaromatic



S_1
aromatic

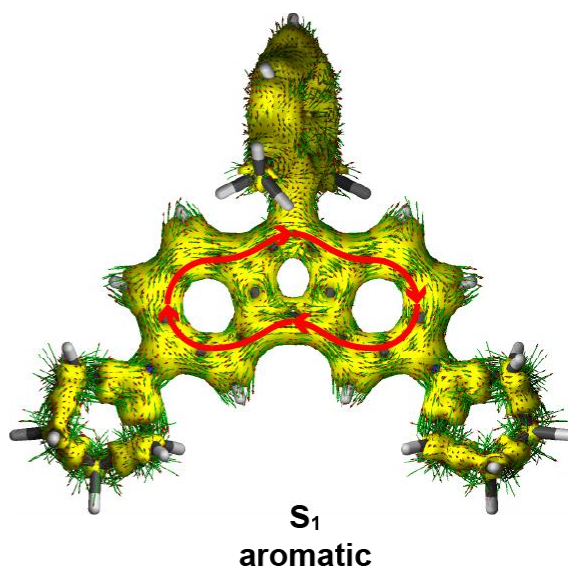
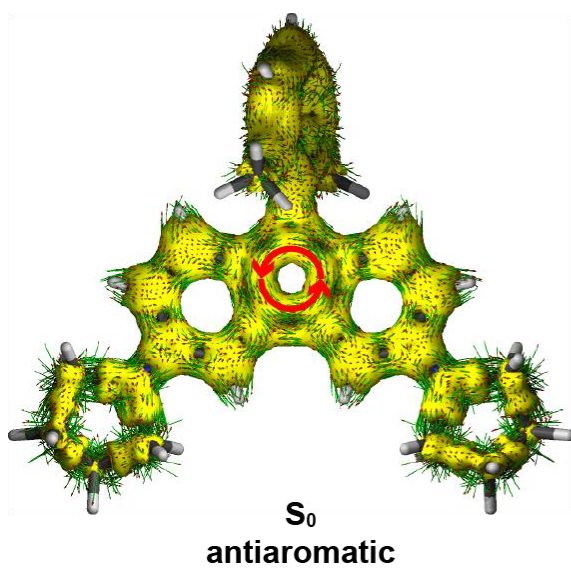
AF6

Supplementary Figure 12 | Theoretical calculated AICD diagram of AF6 in S_0 and S_1 state.



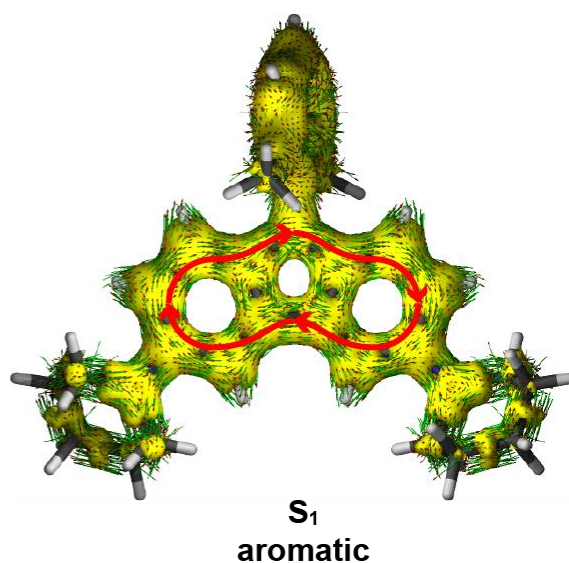
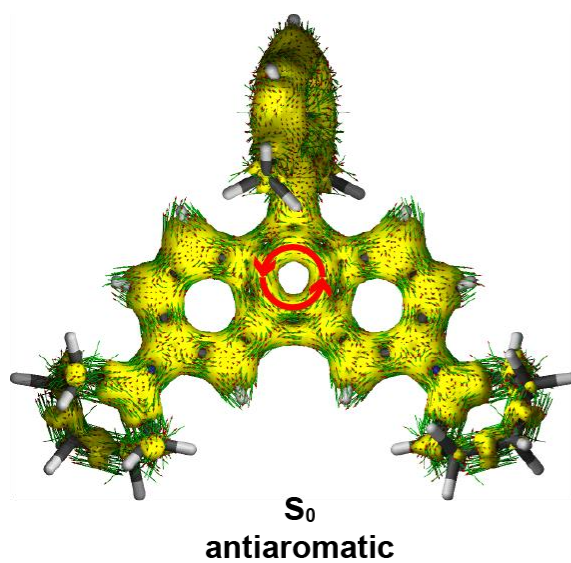
AF7

Supplementary Figure 13 | Theoretical calculated AICD diagram of AF7 in S₀ and S₁ state.



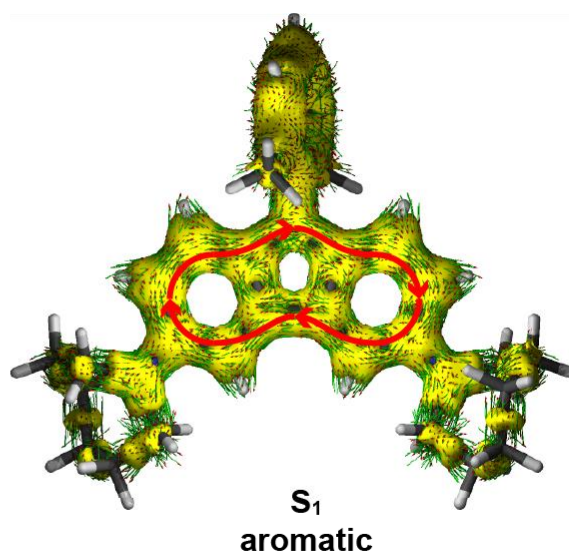
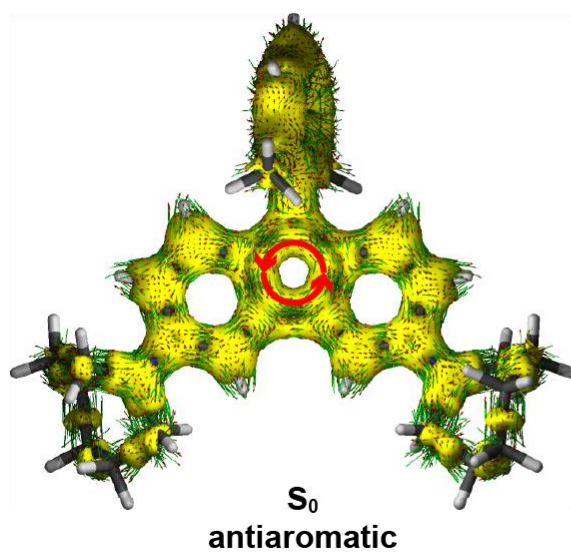
AF8

Supplementary Figure 14 | Theoretical calculated AICD diagram of AF8 in S₀ and S₁ state.



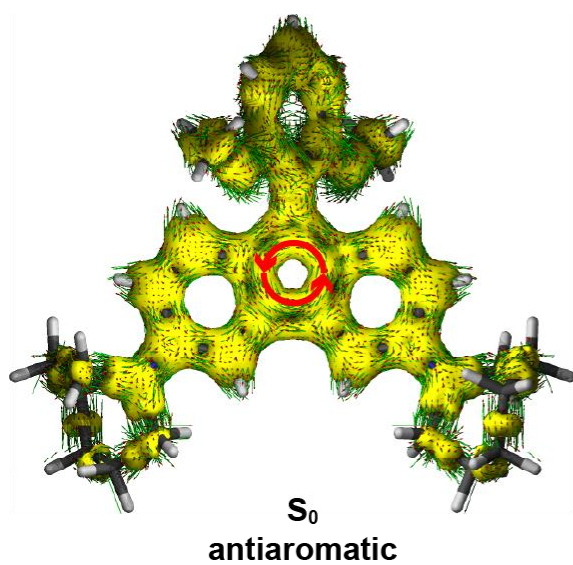
AF9

Supplementary Figure 15 | Theoretical calculated AICD diagram of AF9 in S₀ and S₁ state.

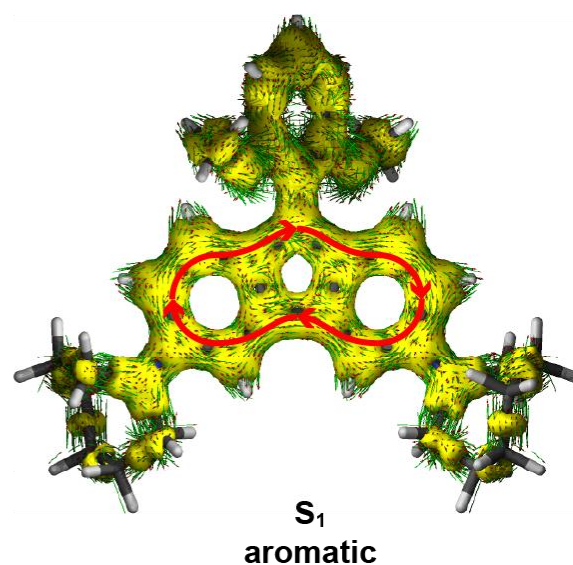


AF10

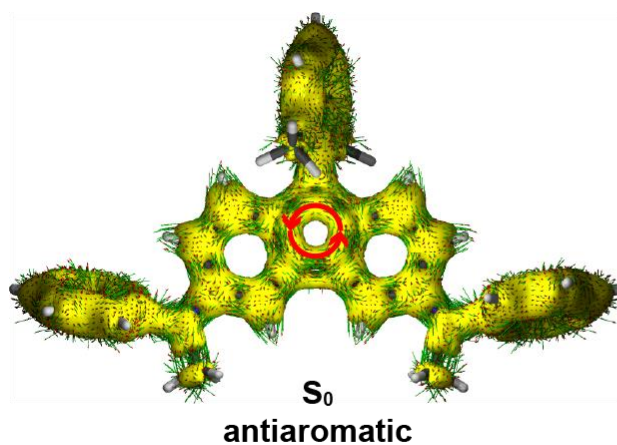
Supplementary Figure 16 | Theoretical calculated AICD diagram of AF10 in S₀ and S₁ state.



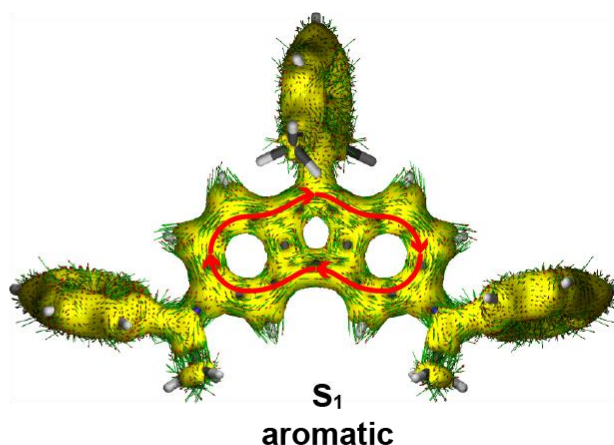
AF11



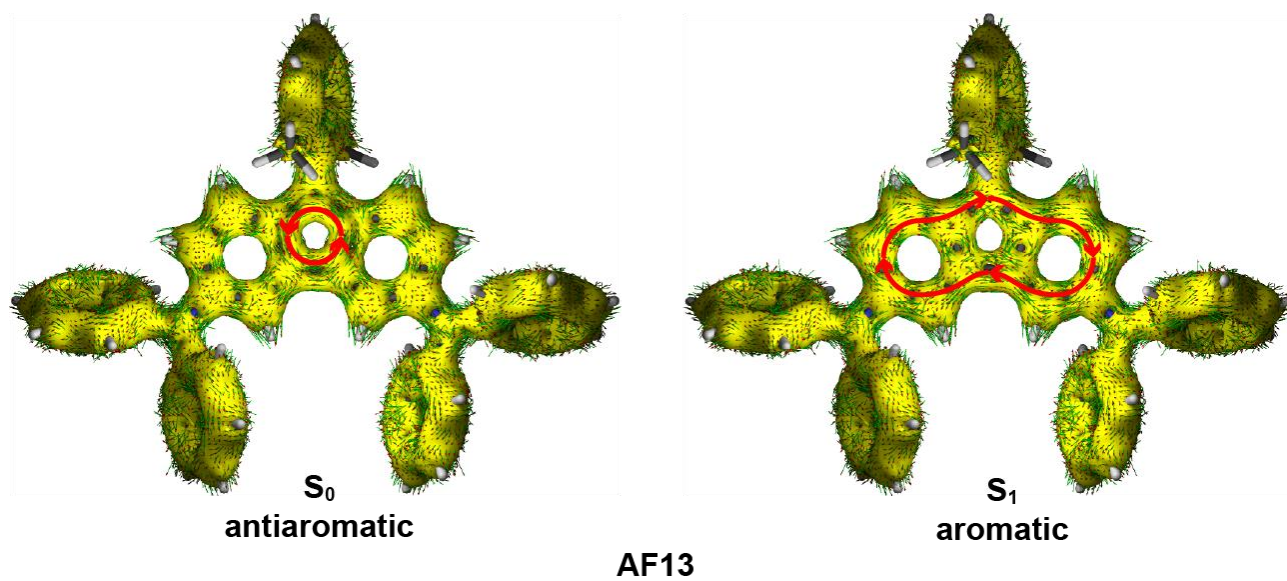
Supplementary Figure 17 | Theoretical calculated AICD diagram of AF11 in S₀ and S₁ state.



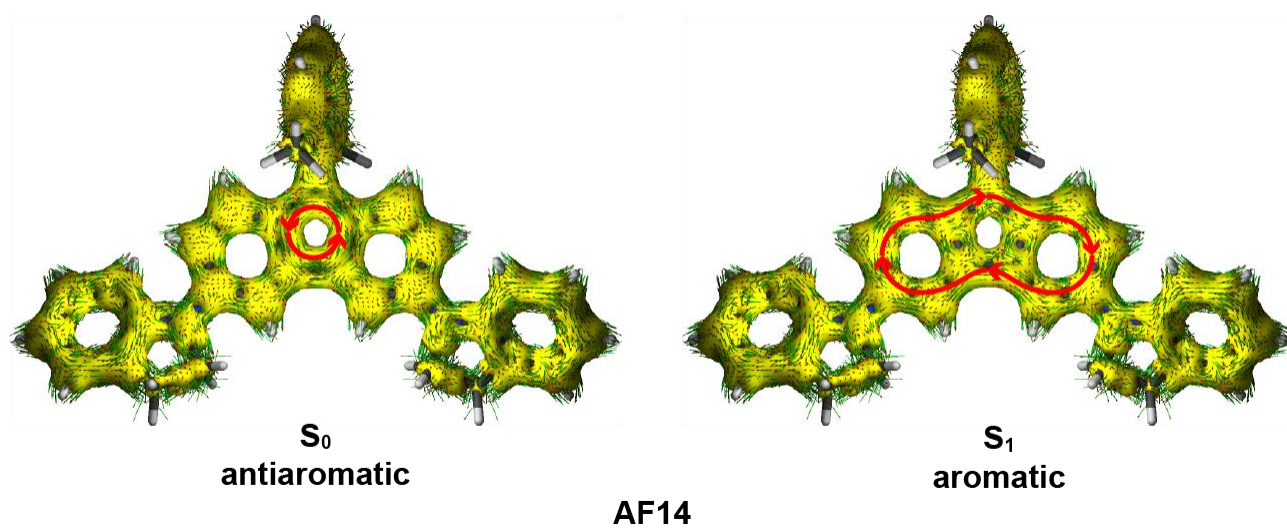
AF12



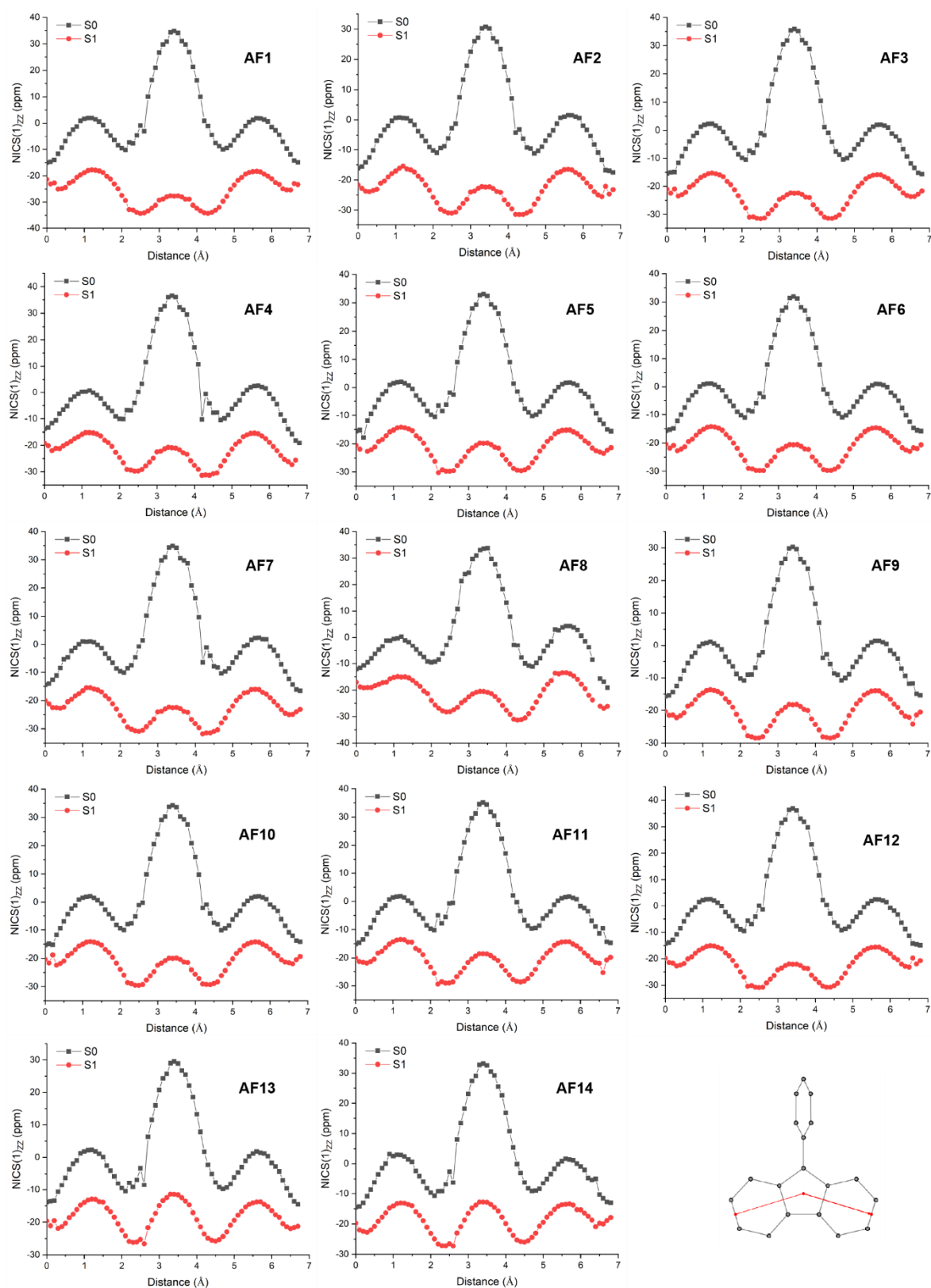
Supplementary Figure 18 | Theoretical calculated AICD diagram of AF12 in S₀ and S₁ state.



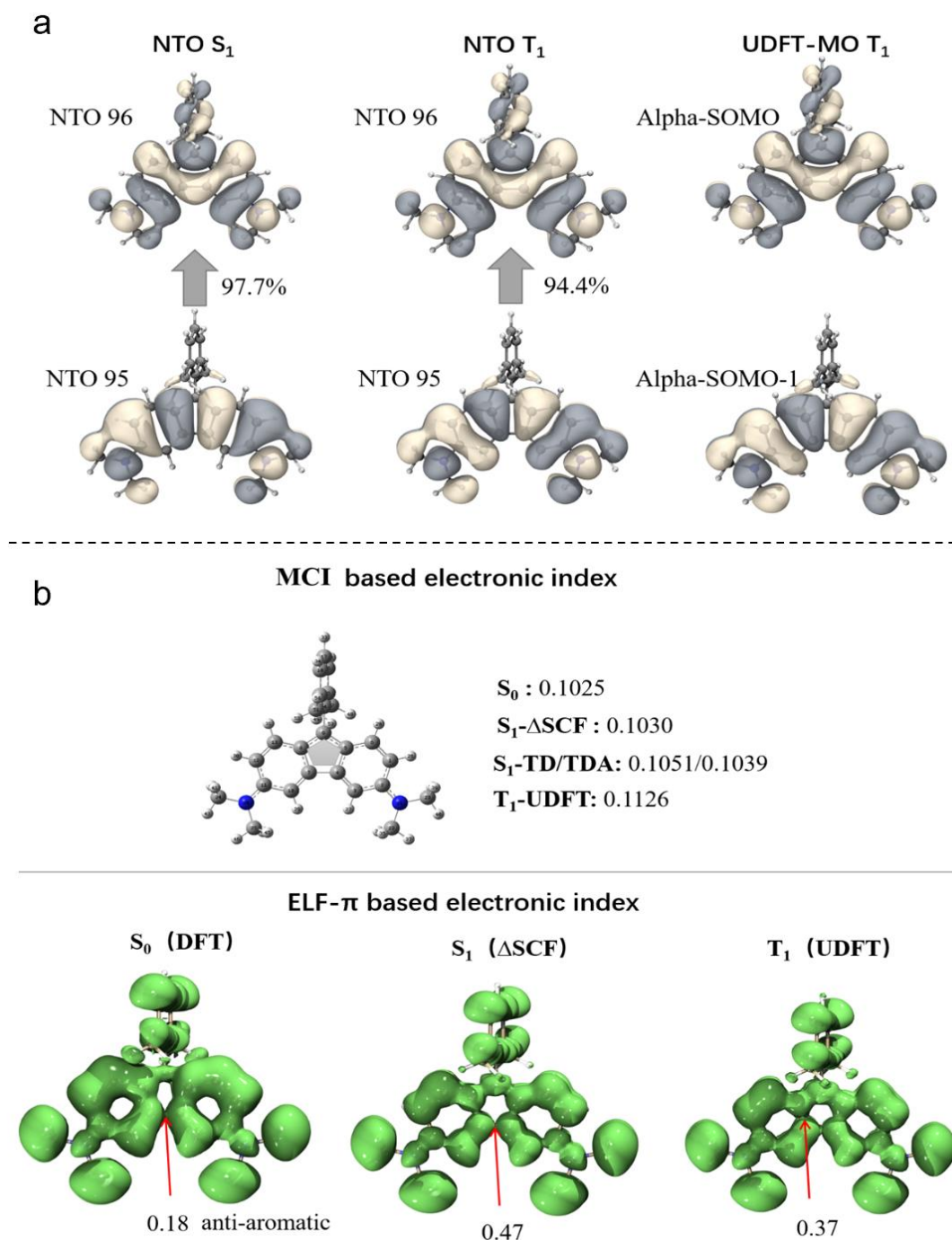
Supplementary Figure 19 | Theoretical calculated AICD diagram of AF13 in S_0 and S_1 state.



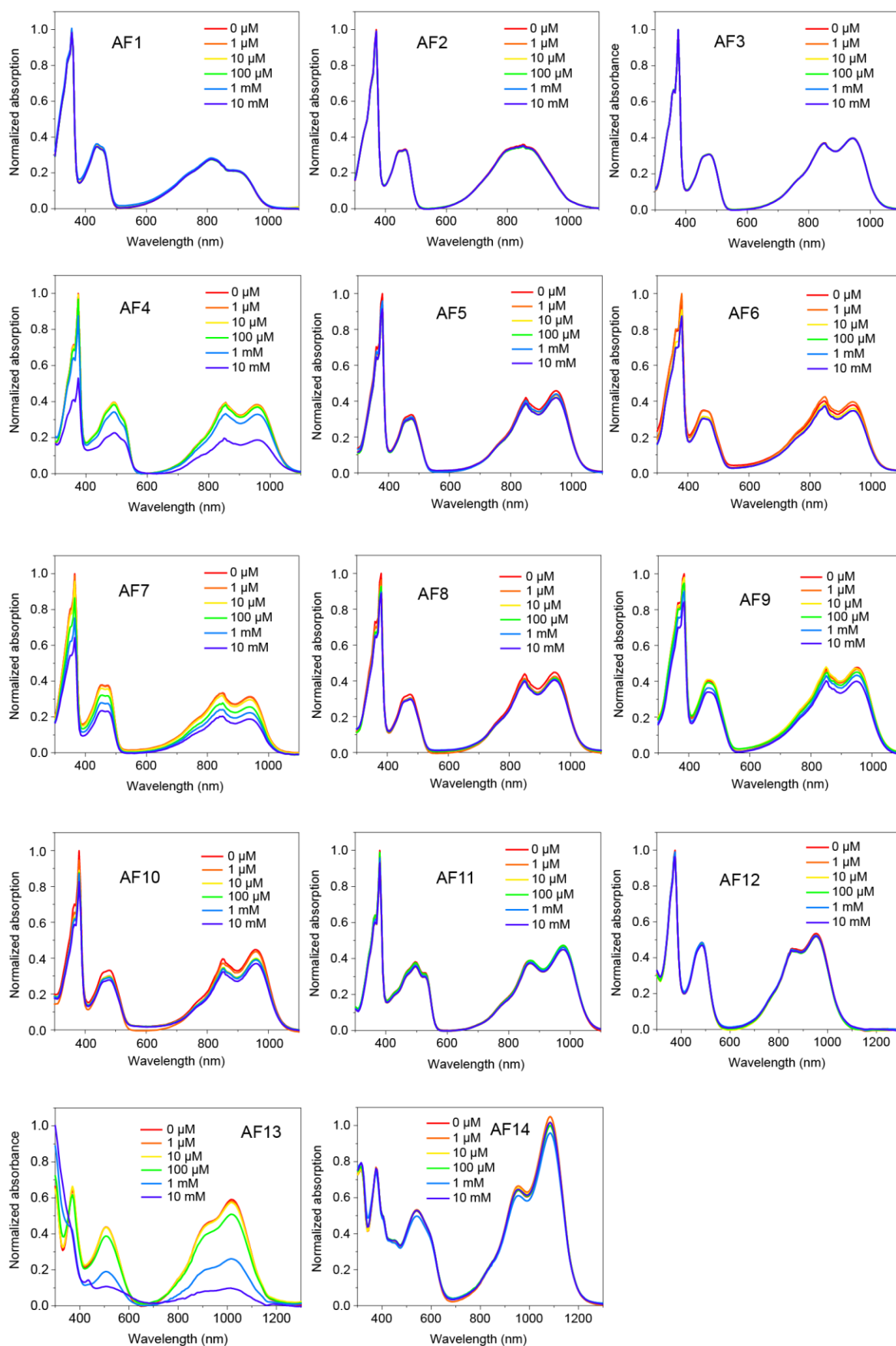
Supplementary Figure 20 | Theoretical calculated AICD diagram of AF14 in S_0 and S_1 state.



Supplementary Figure 21 | NICS-XY scans of AF dyes in both S₀ (black) and S₁ (red) state. All the scans are performed at a height of 1 Å. The red line in the bottom right image shows the scanning path with a scanning interval of 0.1 Å.

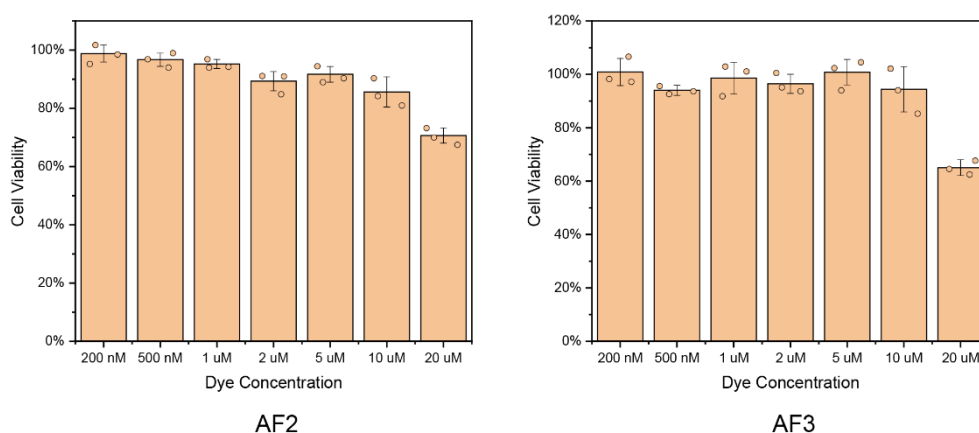


Supplementary Figure 22 | Calculated electronic indexes for (anti-)aromaticity characterization. **a**, The analysis of the natural transition orbitals (NTOs) using TDA-DFT and the electron configurations based on singly occupied molecular orbitals (SOMOs) using UDFT. **b**, Calculated electronic indexes based on multicenter index (MCI) and electron localization function- π (ELF- π) used for characterizing aromaticity in the five-member ring of AF3 and detailed definition of formula can be found in Lu et al. *J. Comput. Chem.*, 2012, 33, 580 and the manual of Multiwfn code.⁴³

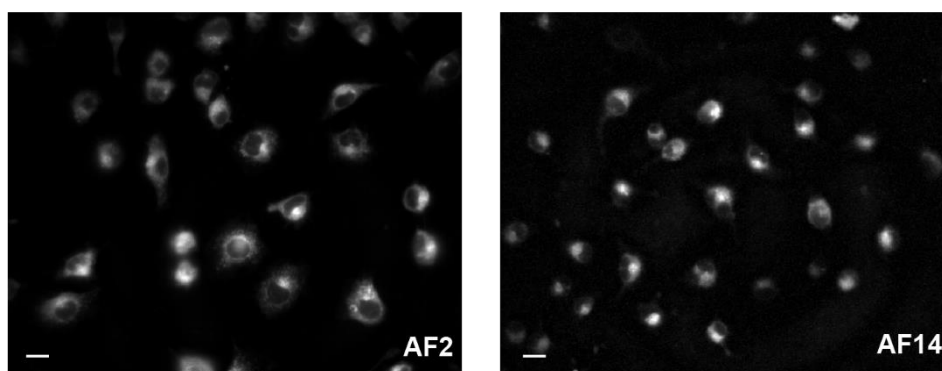


Supplementary Figure 23 | Dose-response curves of all AF dyes as a function of GSH concentration. Note: According to the dose-response curves, we can obtain the dissociation

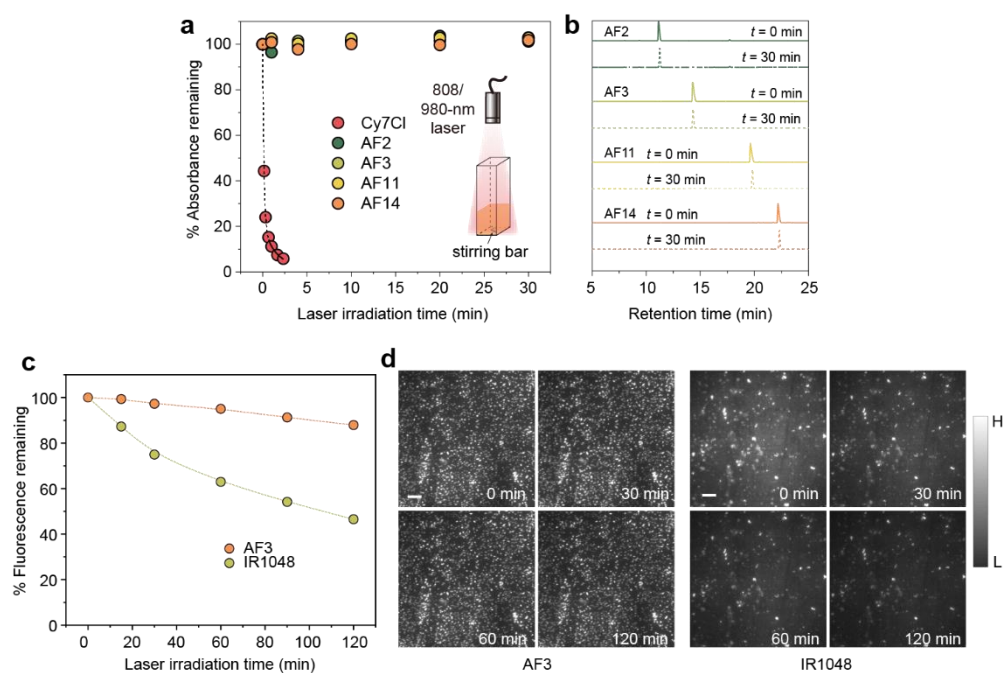
constant⁴⁴ ($K_{d, \text{GSH}}$), where the absorbance of the dissociated form is reduced to half of the maximum. Most of AF dyes have $K_{d, \text{GSH}}$ values greater than 10 mM. For *o*-dimethyl substituted AF7 and AF13, the decline of $K_{d, \text{GSH}}$ could be attributed to the insufficient electron donating effect of N-substituents. Besides, $K_{d, \text{GSH}}$ is also slightly reduced at AF4, which could be attributed to the insufficient steric protection effect of *o*-dimethoxyl groups compared to *o*-dimethyl groups. As the cellular concentration of GSH is in the range of 1-10 mM, these AF dyes are stable for use in biological applications.



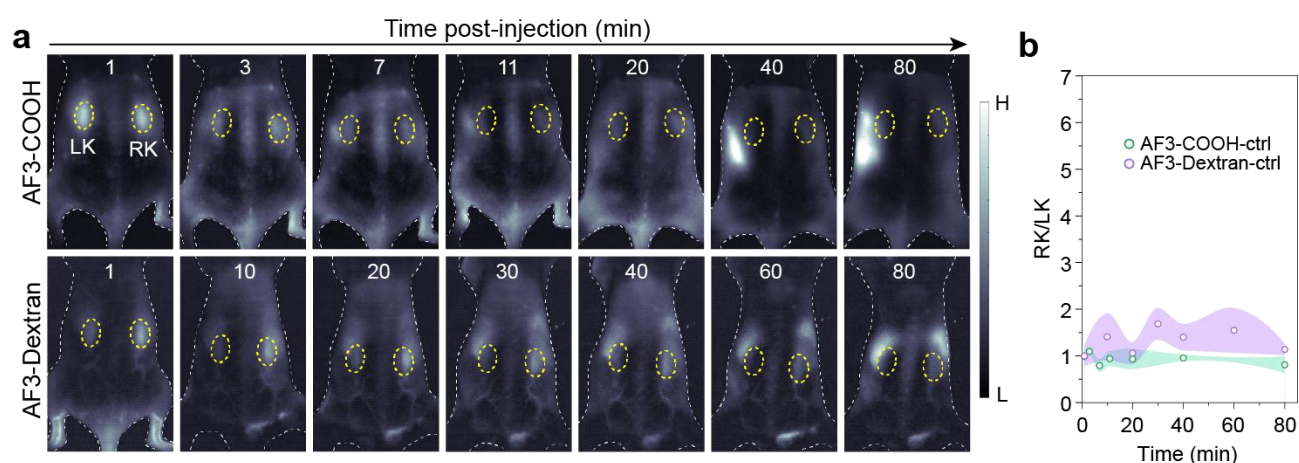
Supplementary Figure 24 | Cell viability results of AF2 and AF3 in 4T1 cells. The bars represent mean $\pm$ s.d. derived from $n = 3$ independent groups.



Supplementary Figure 25 | Fluorescence images of AF dye-stained endothelial cells (ECs). Dye concentration: 1 μM in 1 $\times$ PBS solution. Staining time: 10 min. Fluorescence images were captured under 808 nm excitation and 1000 LP filter. Scale bar, 10 μm .

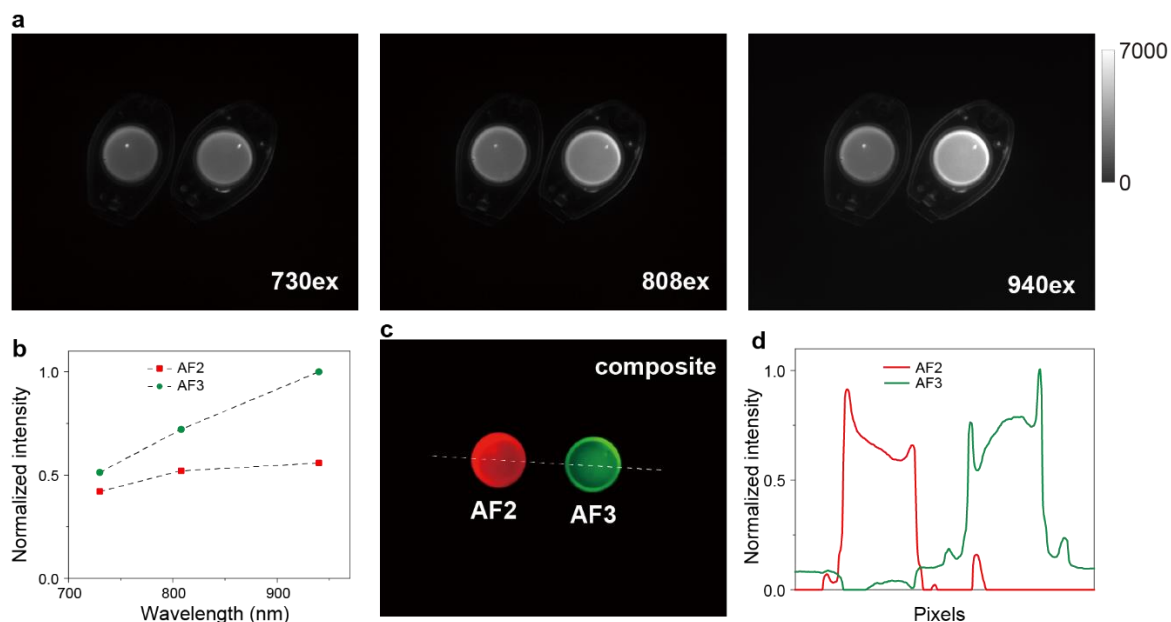


Supplementary Figure 26 | In vitro photostability characterization. **a**, Photostability comparison of AF2, AF3, AF11, Cy7-Cl (808 nm ex), and AF14 (980 nm ex) in aqueous solution. The absorbance of AF2, AF3, AF11, Cy7-Cl at 808 nm and AF14 at 980 nm were set to 0.5 and laser power density was 1.6 W/cm². **b**, HPLC chromatogram of AF dyes before and after irradiation. **c-d**, NIR-II fluorescence microscopic images of dyes exposed to air after certain time irradiation under 808 nm (AF3) and 980 nm (IR1048) laser excitation (**d**), and fluorescence intensity statistic results (**c**). 40×/0.95 objective was used. Laser power density: 200 kW cm⁻². Samples were prepared using microscope slides by dropping DCM solution of each dye and slowly evaporating. Scale bar, 10 μm.



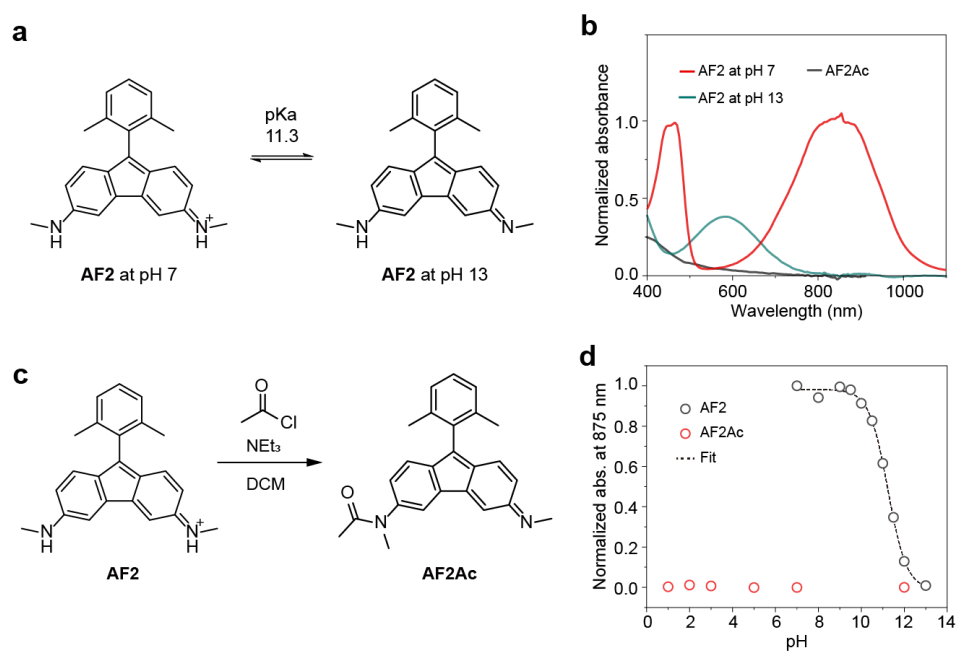
Supplementary Figure 27 | In vivo NIR-II fluorescent imaging of control group mice without renal ischemia-reperfusion treatment. **a**, In vivo NIR-II fluorescent images of control group mice without renal ischemia-reperfusion treatment. The yellow circles indicate the kidney position. **b**,

Time-dependent evolution of the fluorescence intensity ratio of right-to-left kidney (RK/LK) for images in **a**.



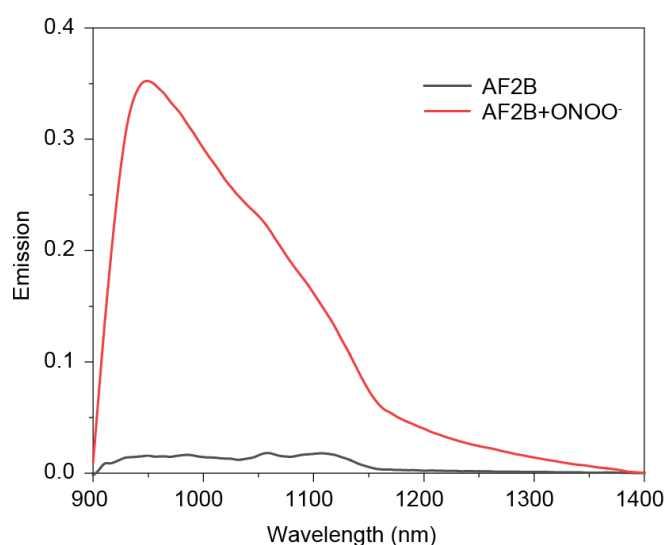
Supplementary Figure 28 | In-vitro fluorescence multispectral unmixing result of AF2 and AF3.

a, Fluorescence image of AF2 and AF3 samples under 730 nm, 808 nm and 940 nm excitation; **b**, Acquired excitation spectra of AF2 and AF3; **c**, Multispectral unmixing result of AF2 and AF3; **d**, normalized signal intensity of AF2/AF3 channel on the dotted line region in **c**.

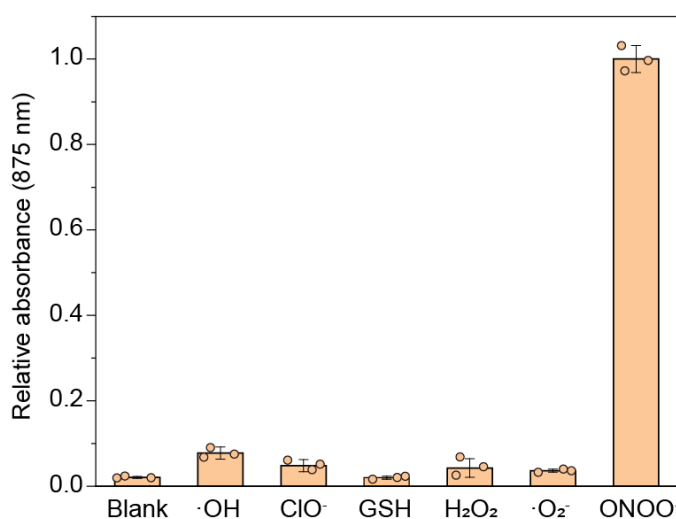


Supplementary Figure 29 | pH-response characteristics of AF2 and AF2Ac. a, Acid-base

equilibrium of AF2. **b**, Normalized absorption spectra of AF2 at pH 7 and 13, as well as AF2Ac at pH 7 (7:3 v/v PBS:MeCN). The 590 nm absorption peak of AF2 at pH 13 proved that the reaction product between AF2 and base was the deprotonated form shown in **a**, rather than an OH nucleophilic addition form at the 9- position, because this will result in the lose of conjugation and no visible absorbance will be observed. **c**, Chemical structure and synthetic route of AF2Ac. **d**, Absorbance at 875 nm of AF2 and AF2Ac versus pH values. Boltzmann curve fitting (dotted line) gave the pKa value of ~11.3.

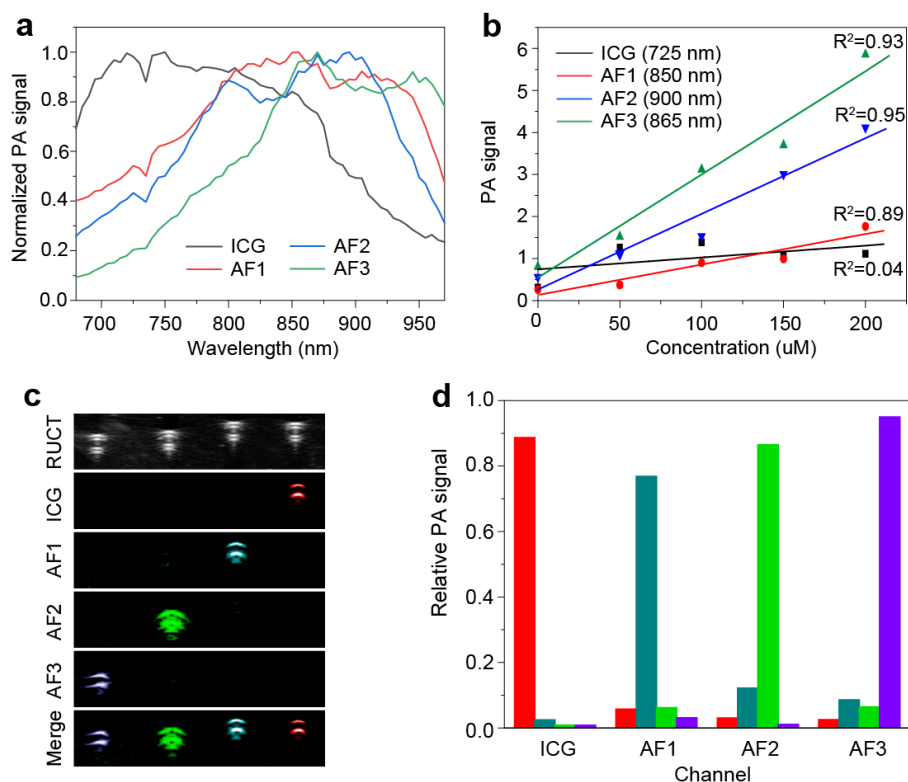


Supplementary Figure 30 | Absorption spectra of AF2B (50 μ M) after incubation with ONOO⁻ (25 μ M).

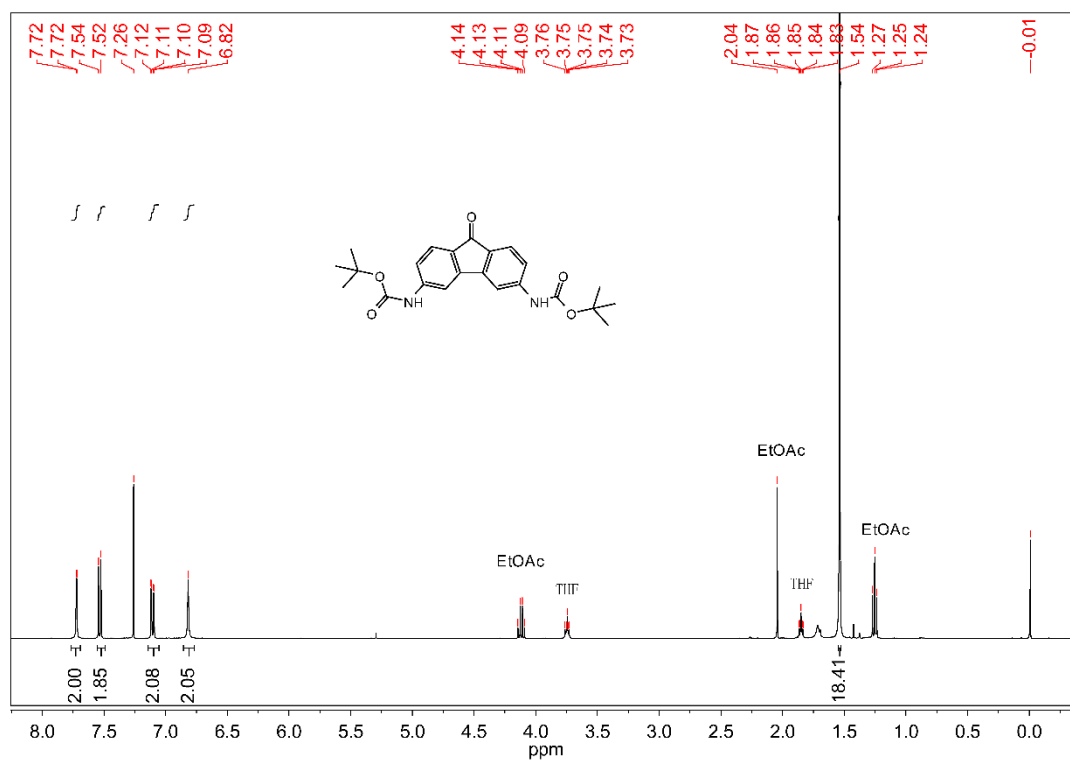


Supplementary Figure 31 | Selectivity of AF2B over ClO⁻ (50 eq.), H₂O₂ (50 eq.), •OH (20 eq.),

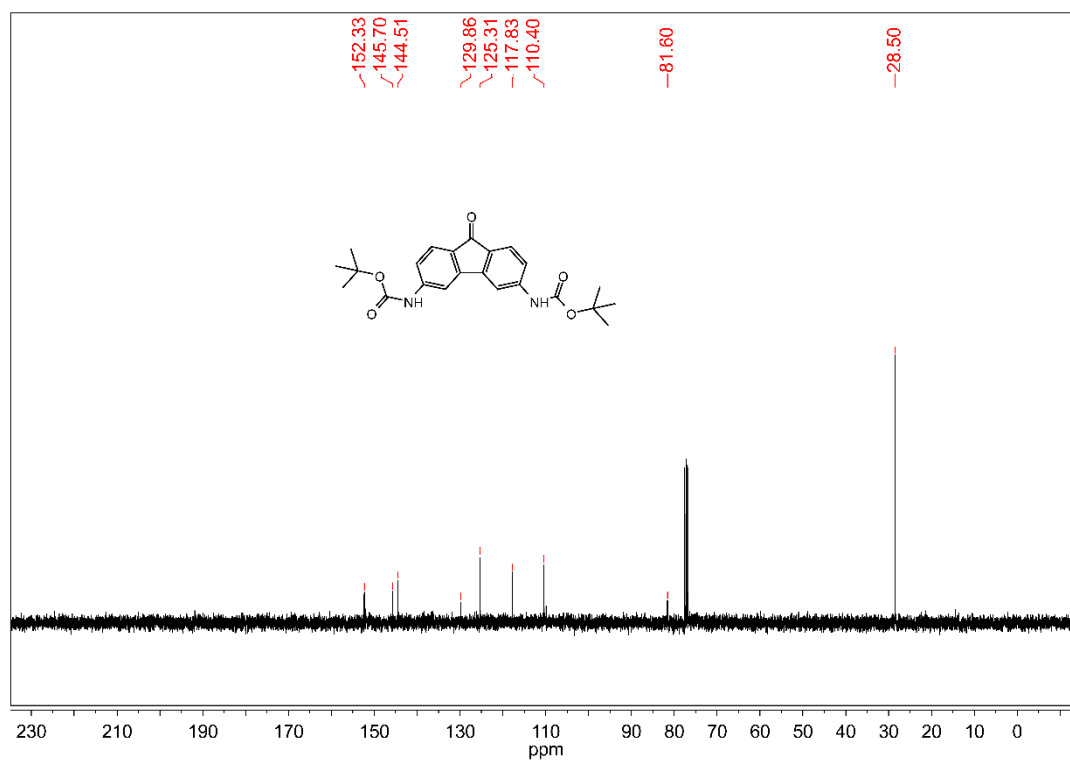
•O₂⁻ (50 eq.), GSH (50 eq.) and ONOO⁻ (0.5 eq.). The bars represent mean ± s.d. derived from n = 3 independent groups.



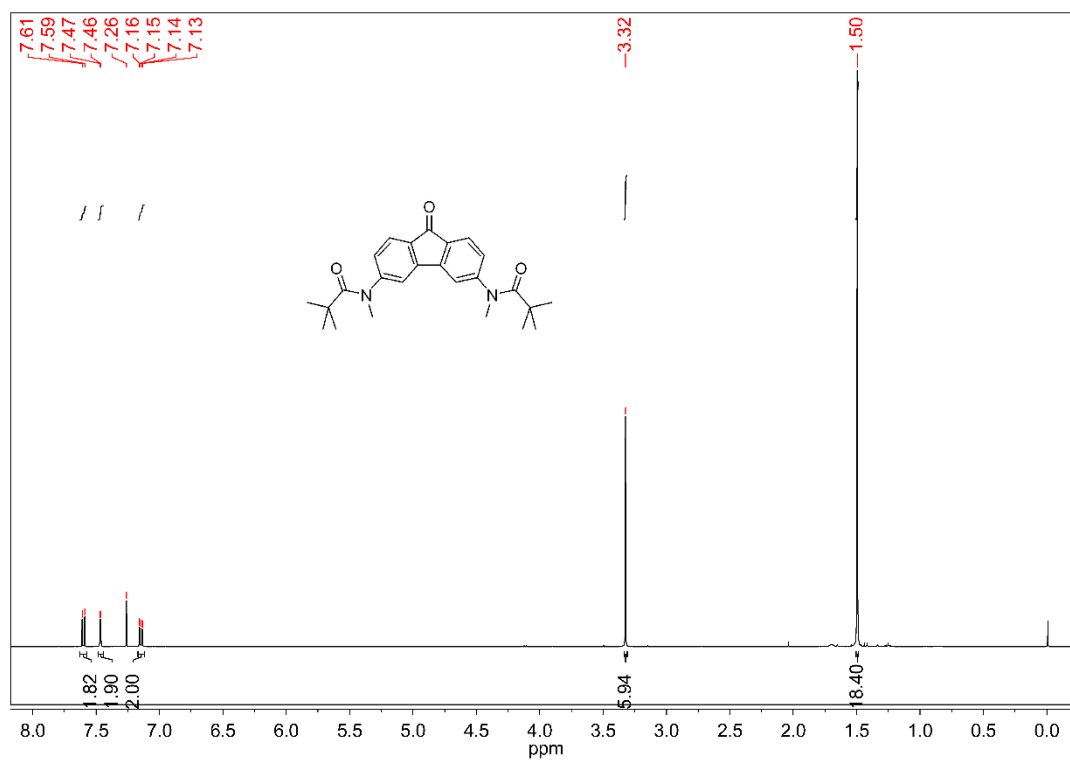
Supplementary Figure 32 | in-vitro photoacoustic (PA) spectra and multispectral unmixing of ICG, AF1, AF2, and AF3. **a**, Normalized PA spectra of ICG, AF1, AF2, and AF3 (200 μM, 1×PBS solution). **b**, Linear fit results of PA signal of ICG, AF1, AF2, and AF3 at certain wavelength. **c-d**, Multispectral PA unmixing results of ICG, AF1, AF2, and AF3.



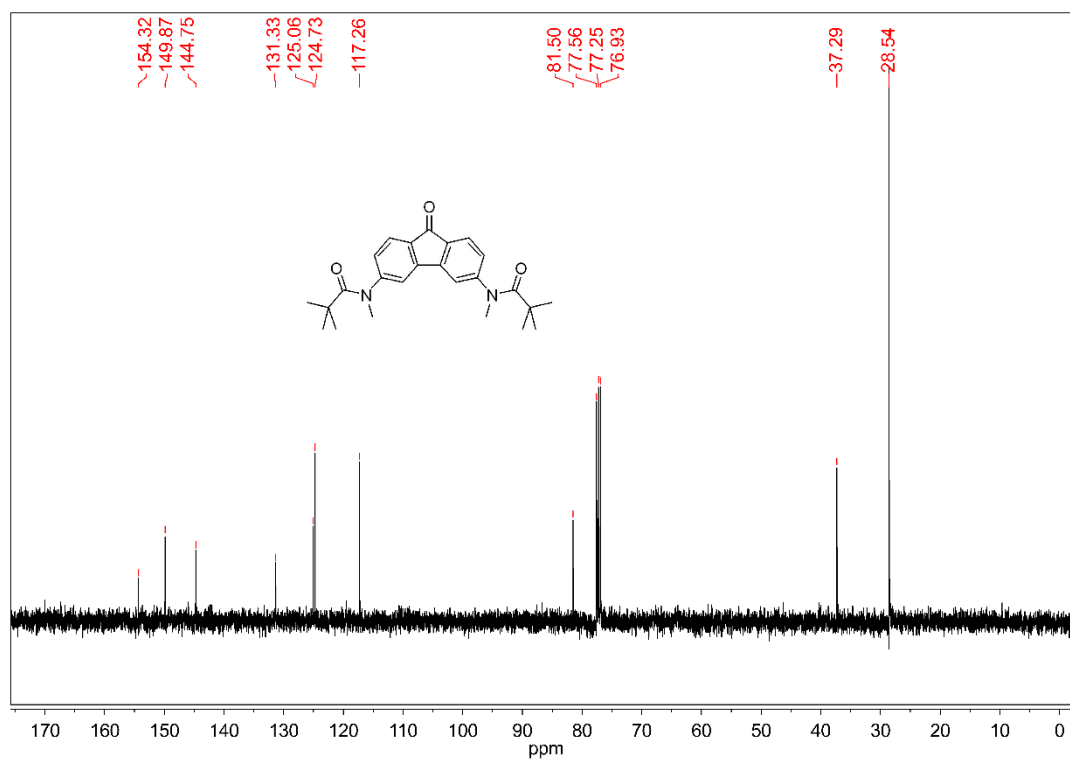
Supplementary Figure 33 | ¹H-NMR of 1a.



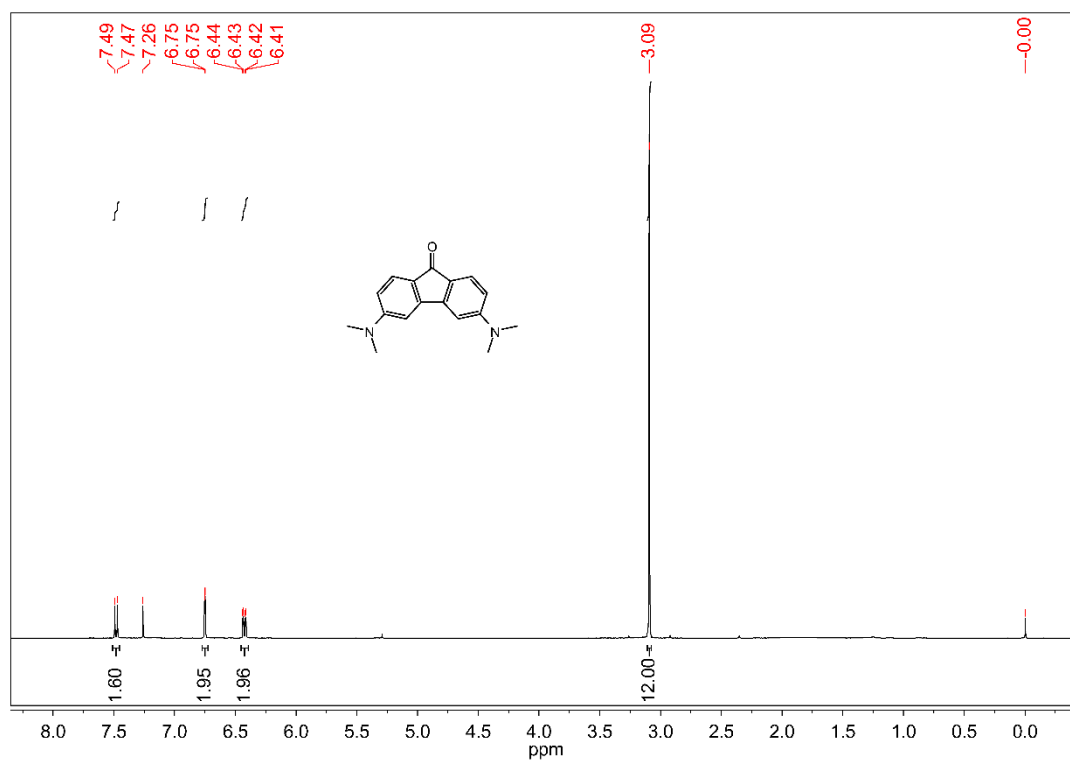
Supplementary Figure 34 | ¹³C-NMR of 1a.



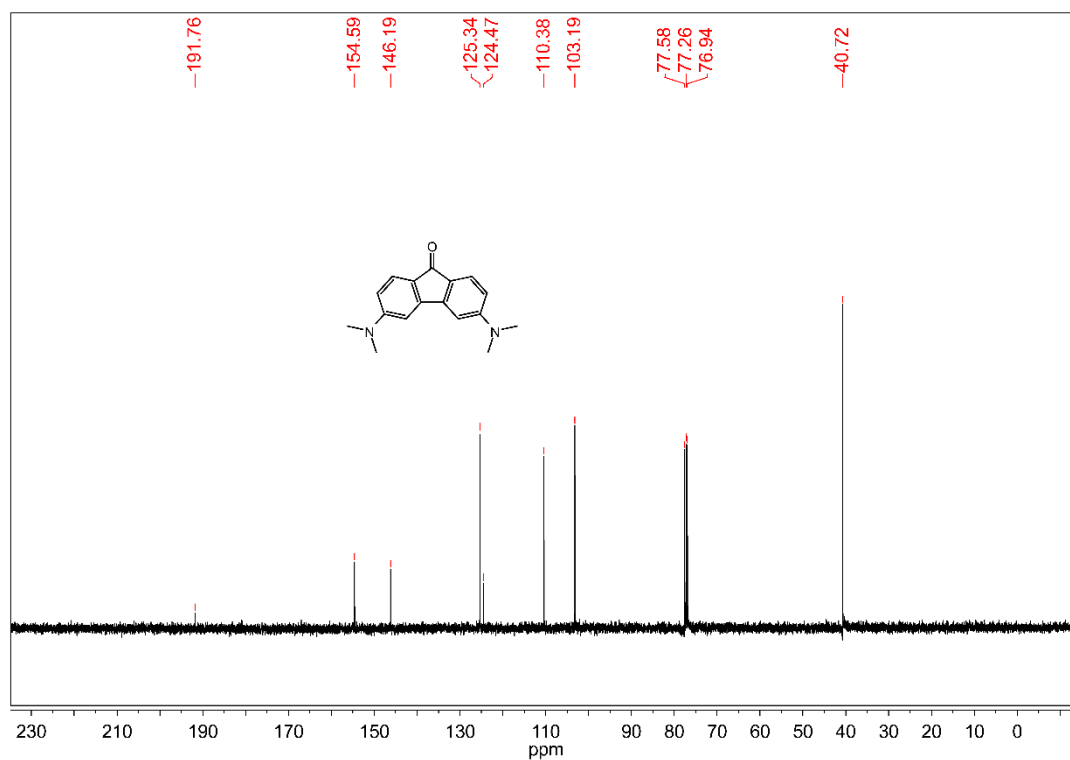
Supplementary Figure 35 | ¹H-NMR of 1b.



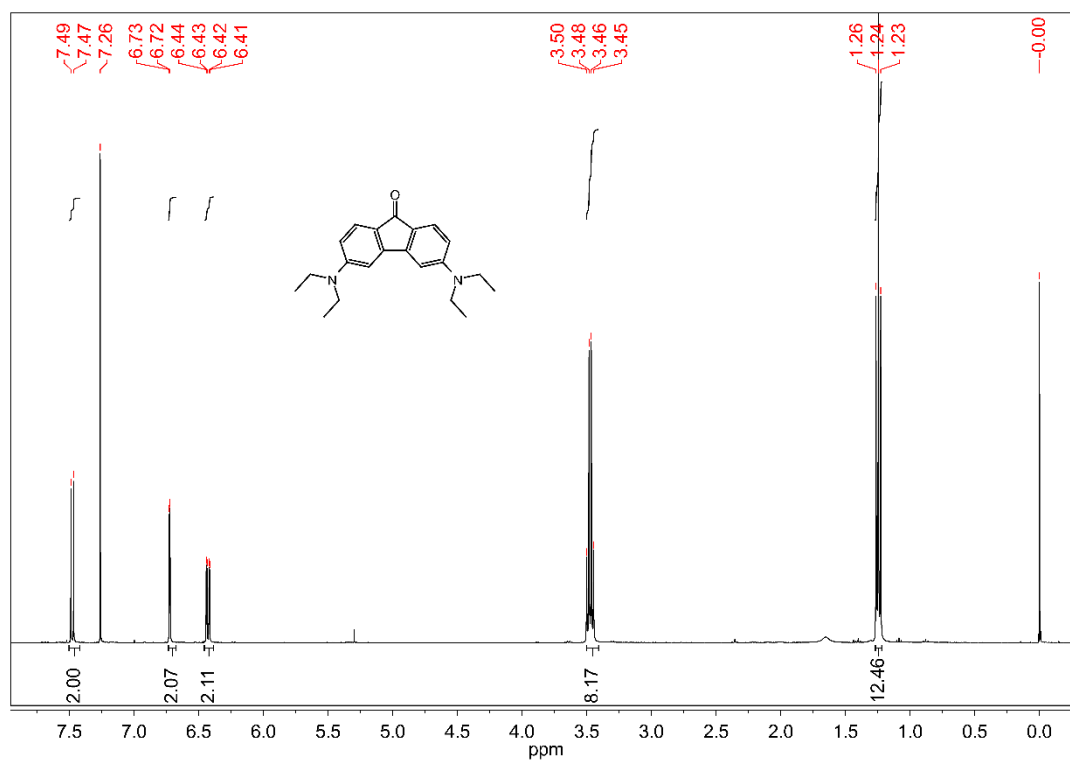
Supplementary Figure 36 | ¹³C-NMR of 1b.



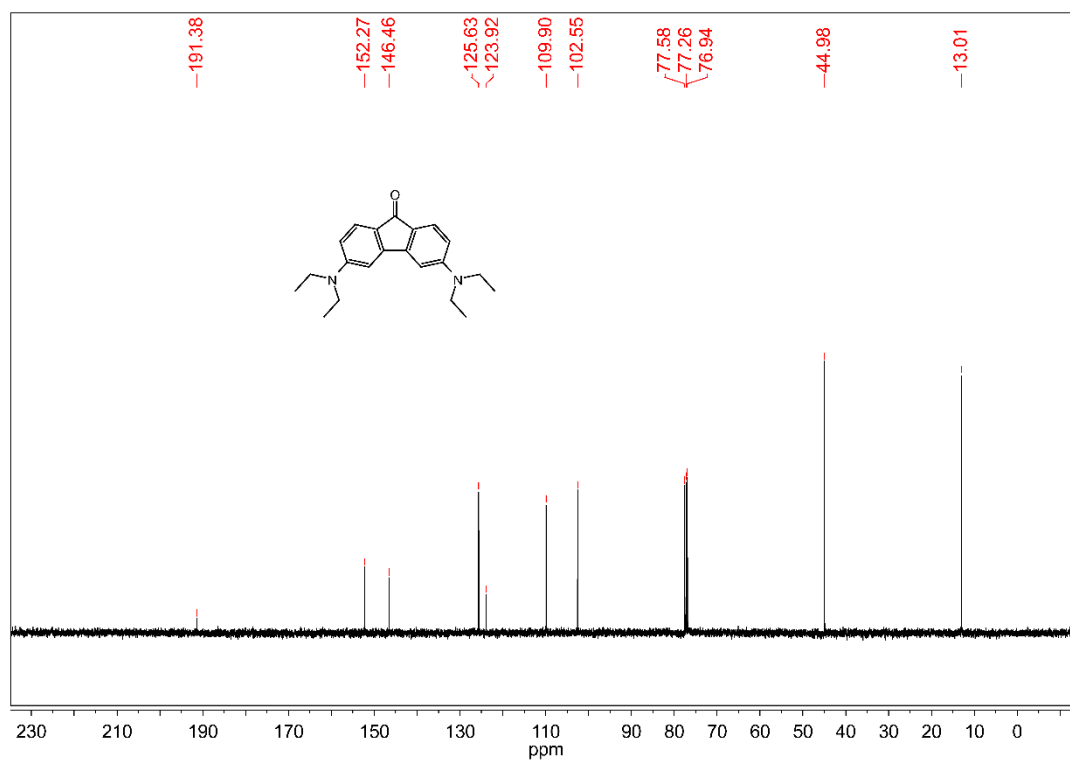
Supplementary Figure 37 | ¹H-NMR of 1c.



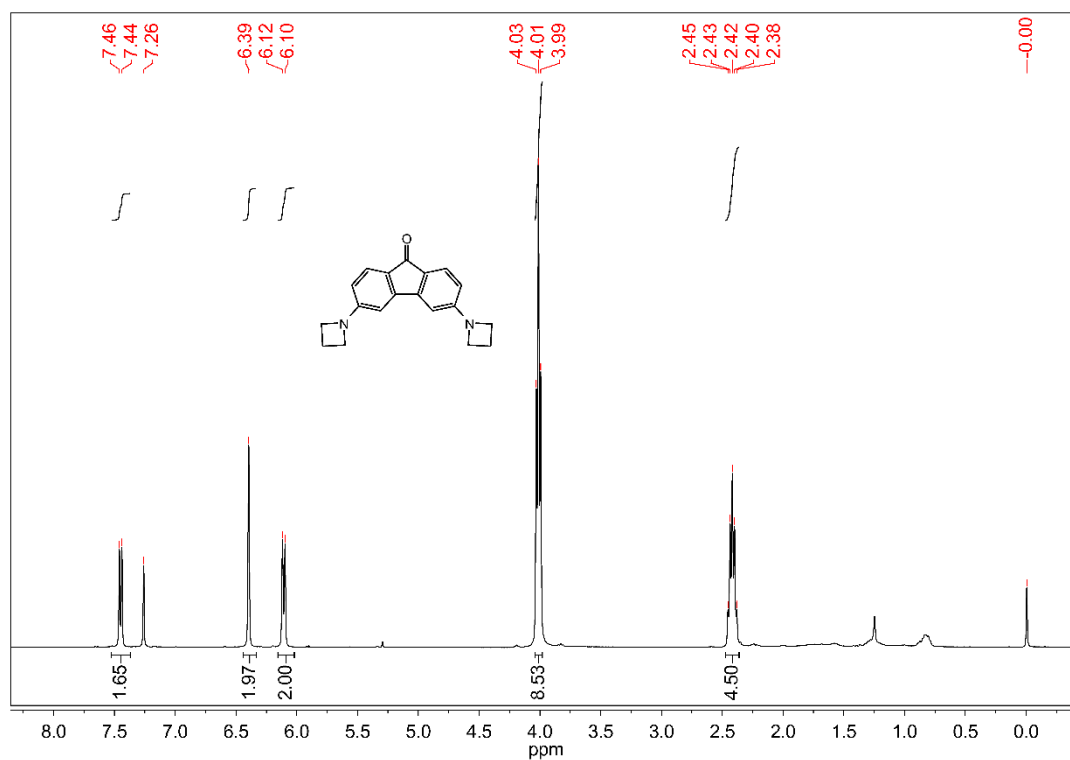
Supplementary Figure 38 | ¹³C-NMR of 1c.



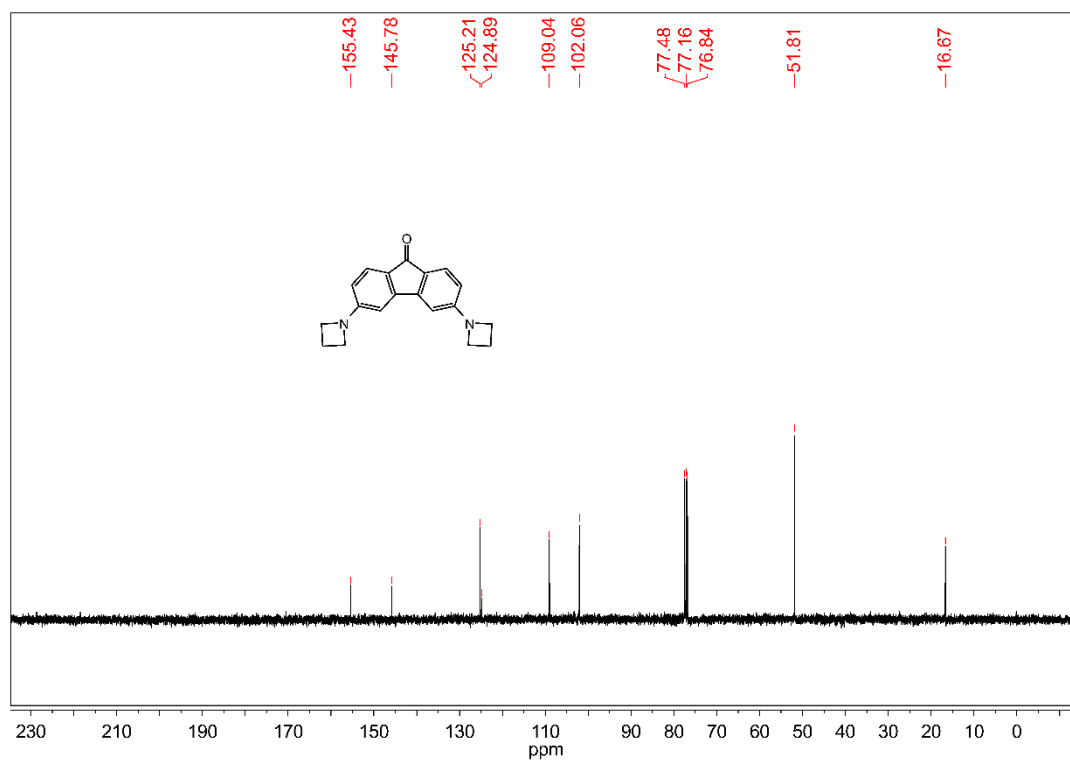
Supplementary Figure 39 | ¹H-NMR of 1d.



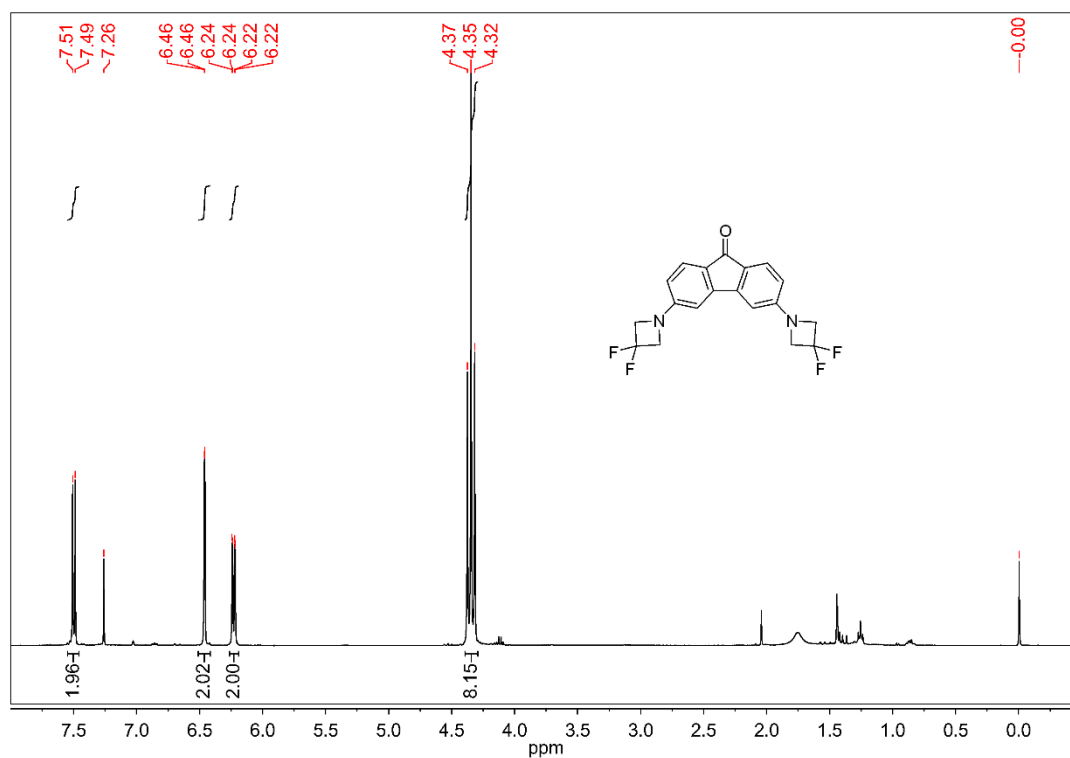
Supplementary Figure 40 | ¹³C-NMR of 1d.



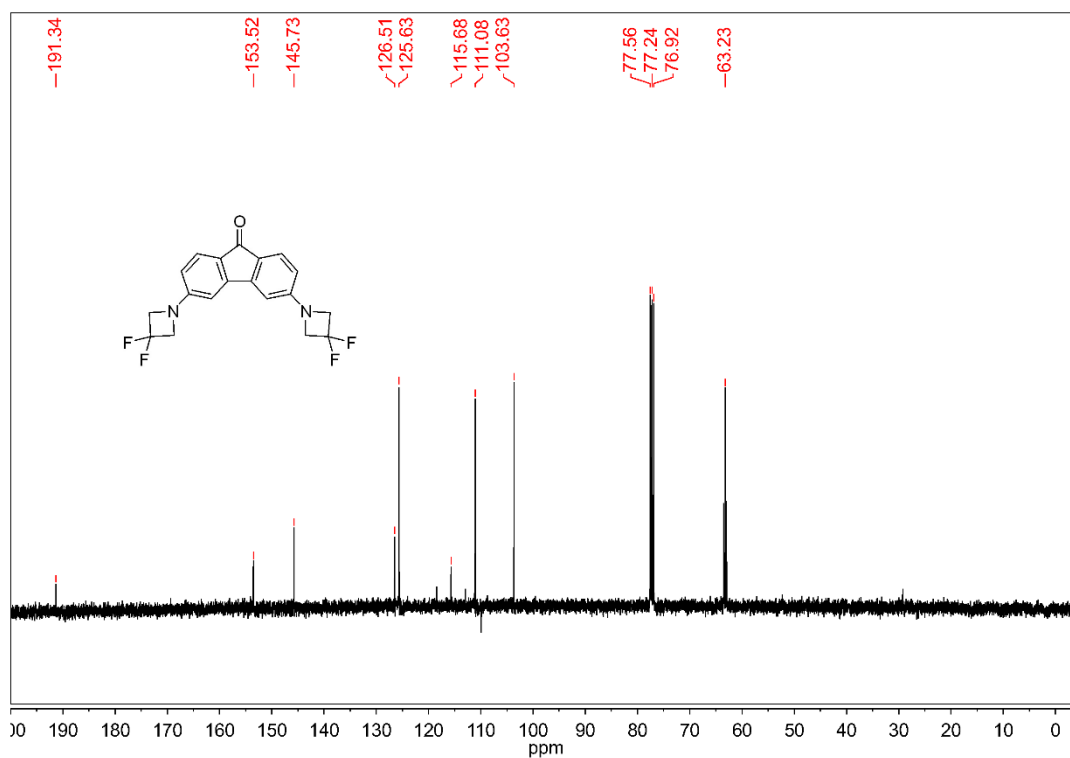
Supplementary Figure 41 | ¹H-NMR of 1e.



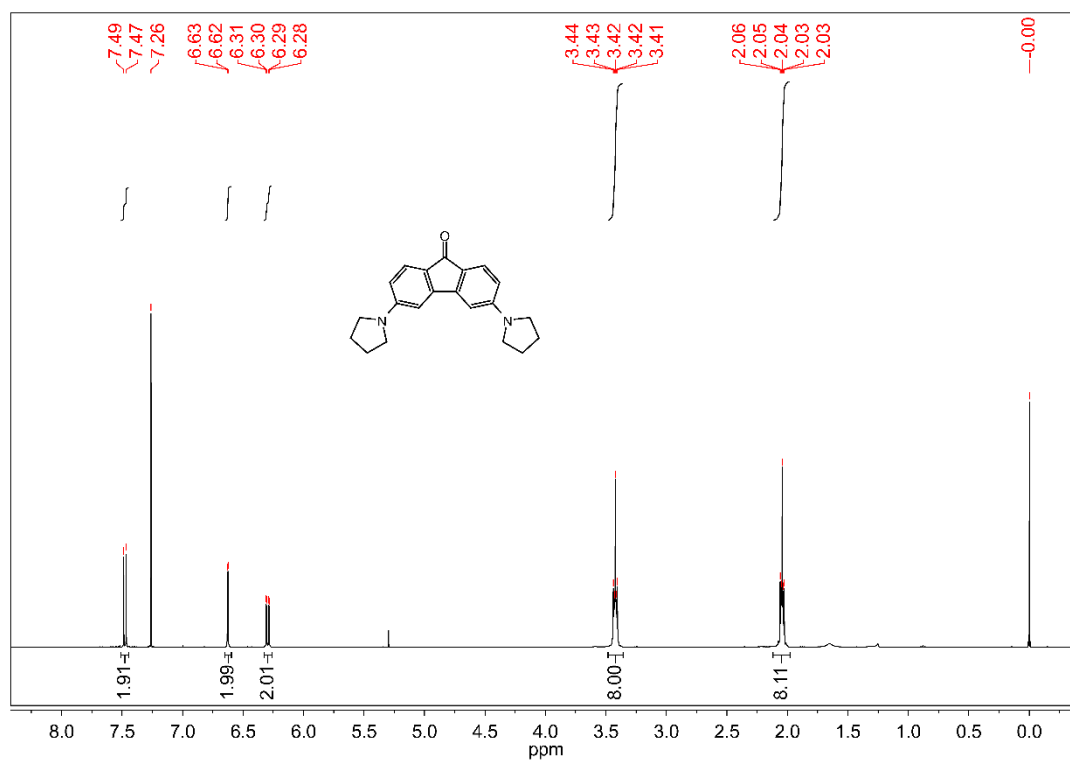
Supplementary Figure 42 | ¹³C-NMR of 1e.



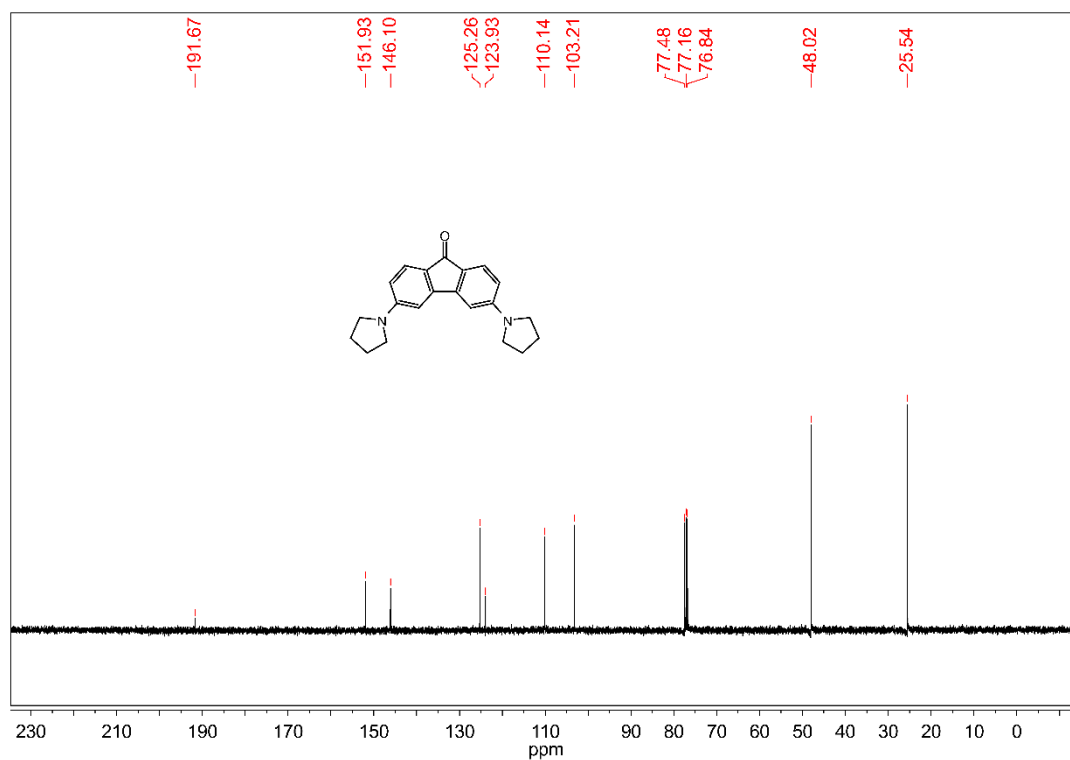
Supplementary Figure 43 | ¹H-NMR of 1f.



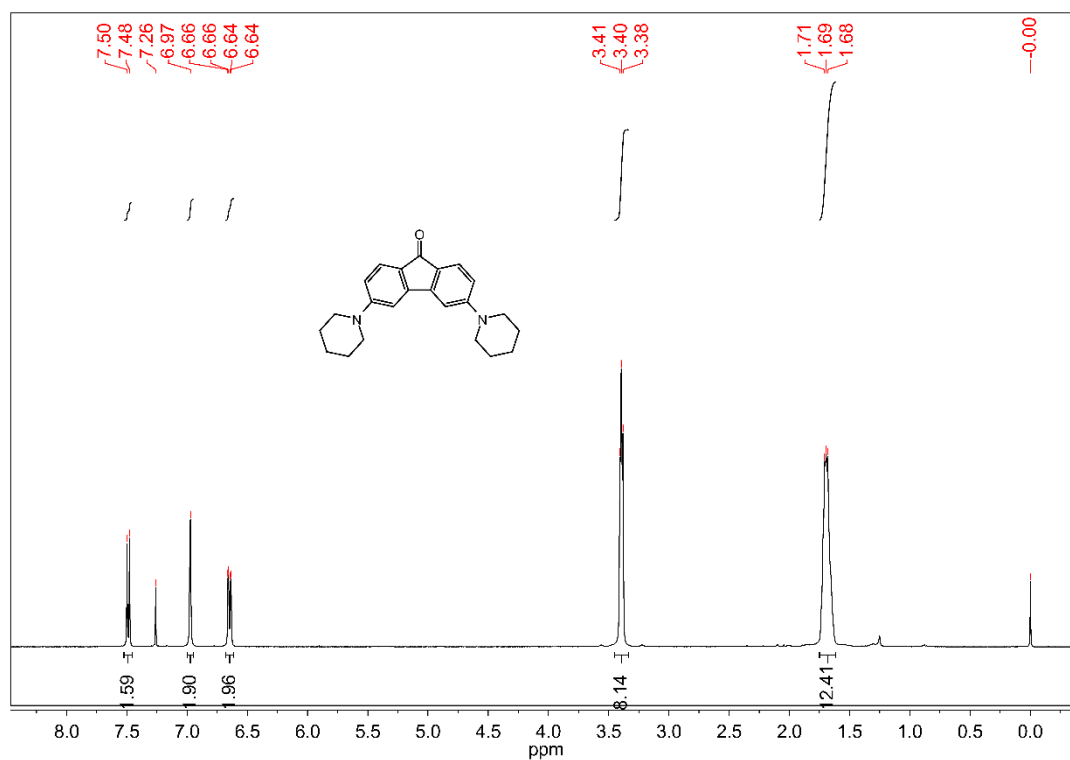
Supplementary Figure 44 | ¹³C-NMR of 1f.



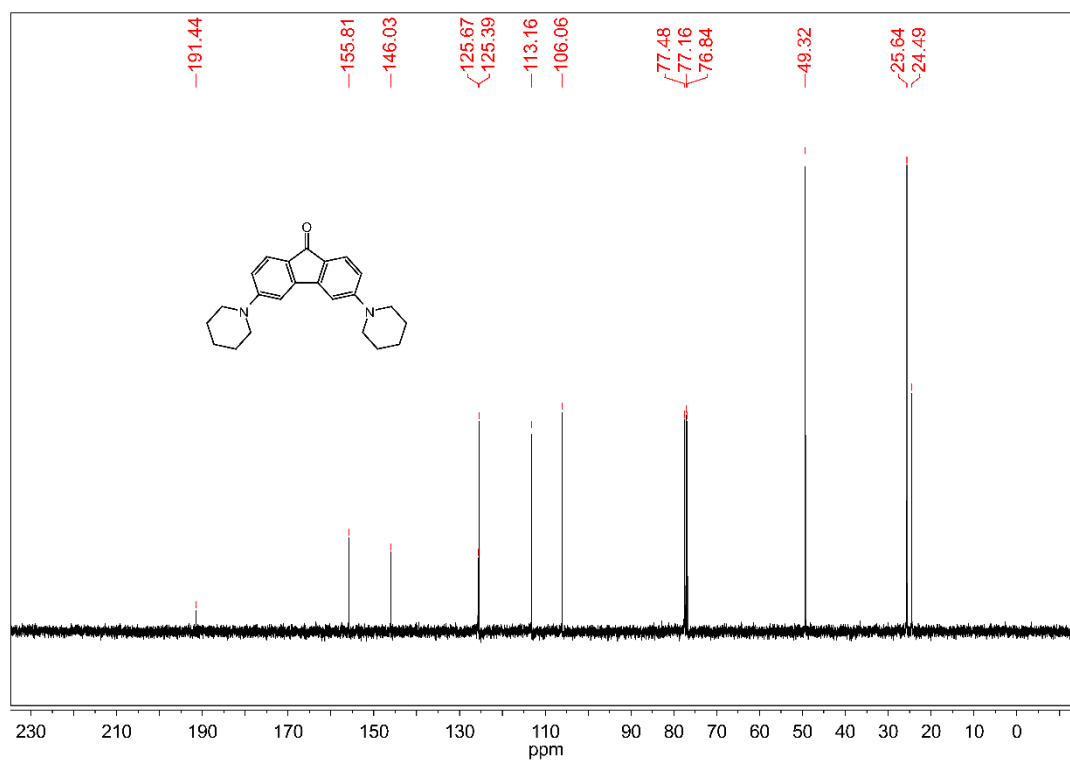
Supplementary Figure 45 | ¹H-NMR of 1g.



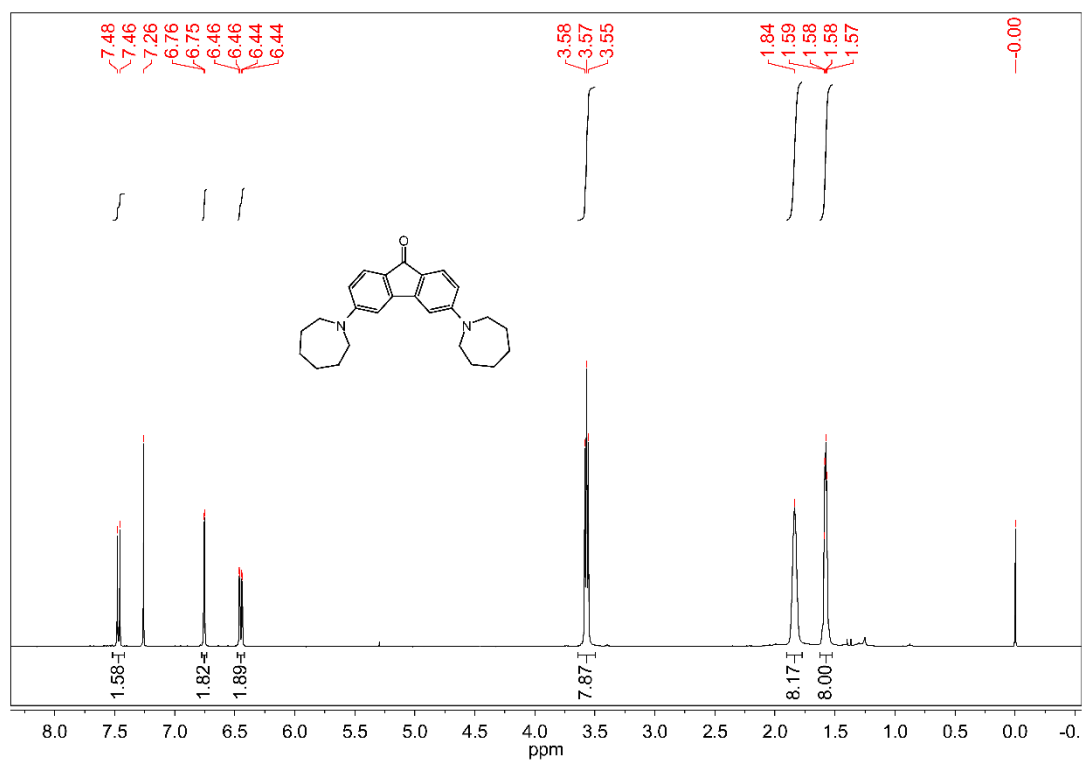
Supplementary Figure 46 | ¹³C-NMR of 1g.



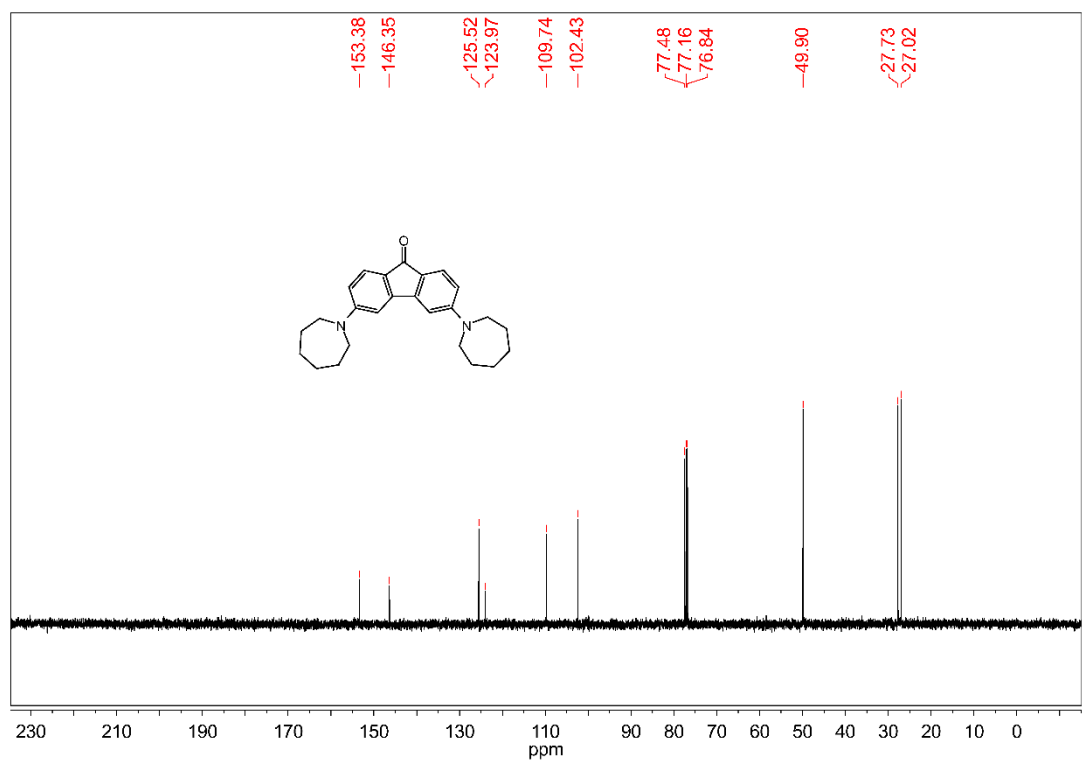
Supplementary Figure 47 | ^1H -NMR of 1h.



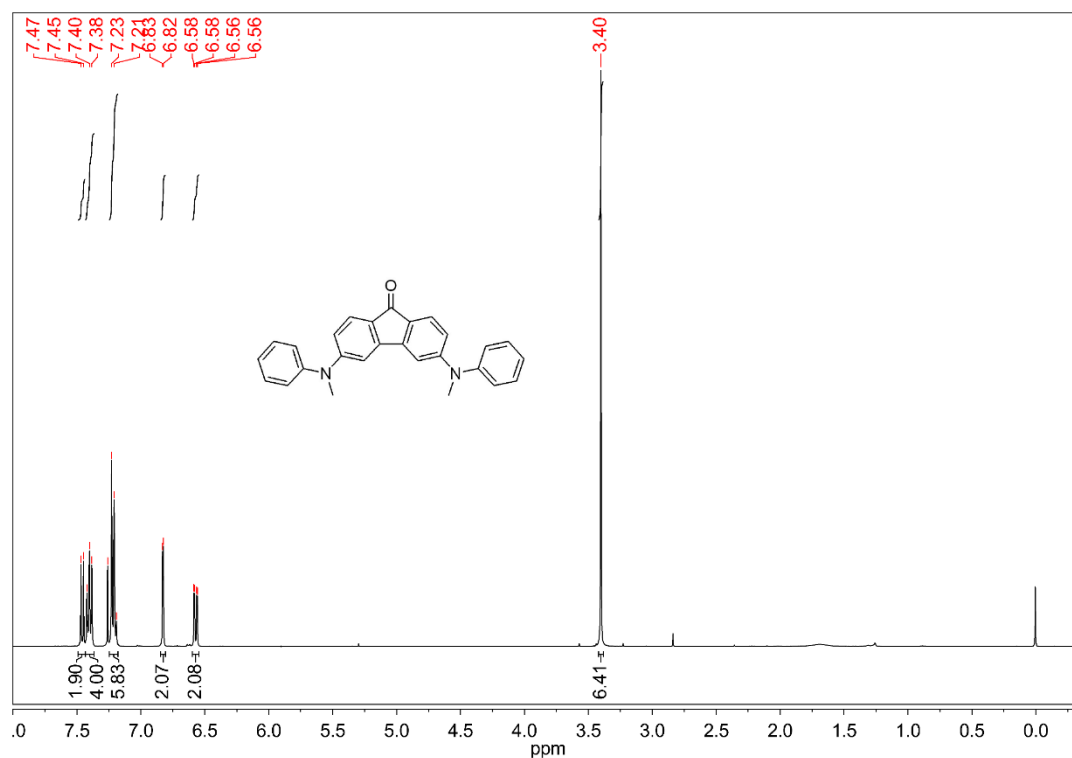
Supplementary Figure 48 | ^{13}C -NMR of 1h.



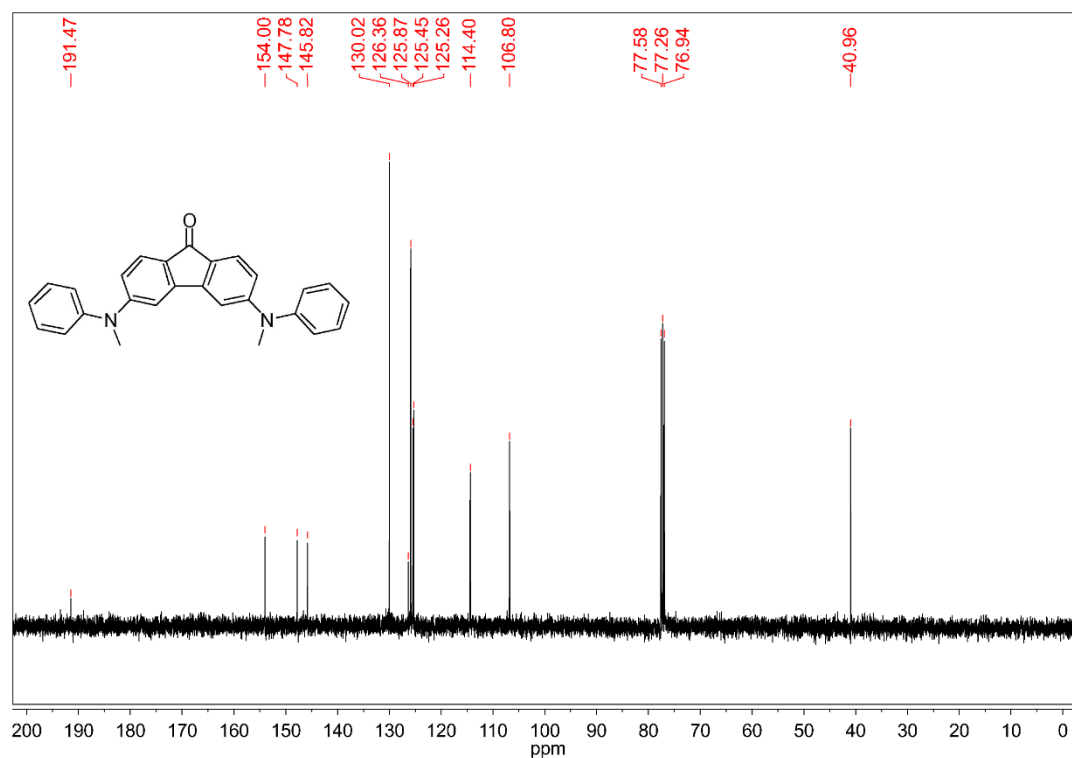
Supplementary Figure 49 | ¹H-NMR of 1i.



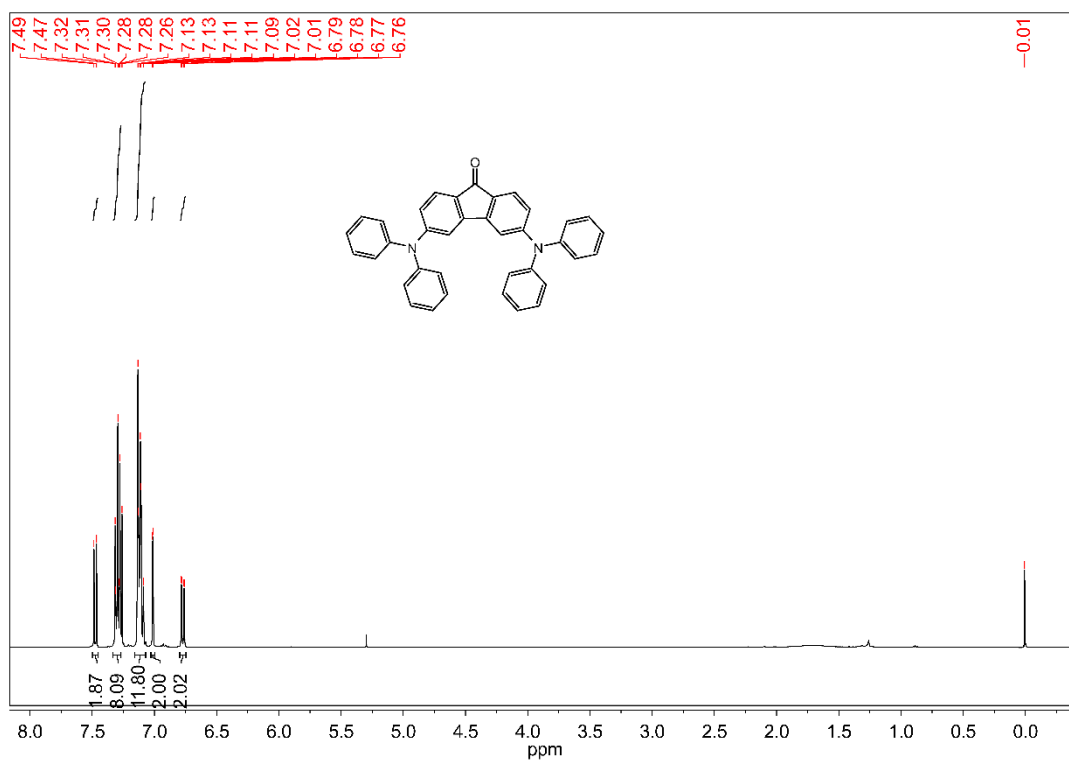
Supplementary Figure 50 | ¹³C-NMR of 1i.



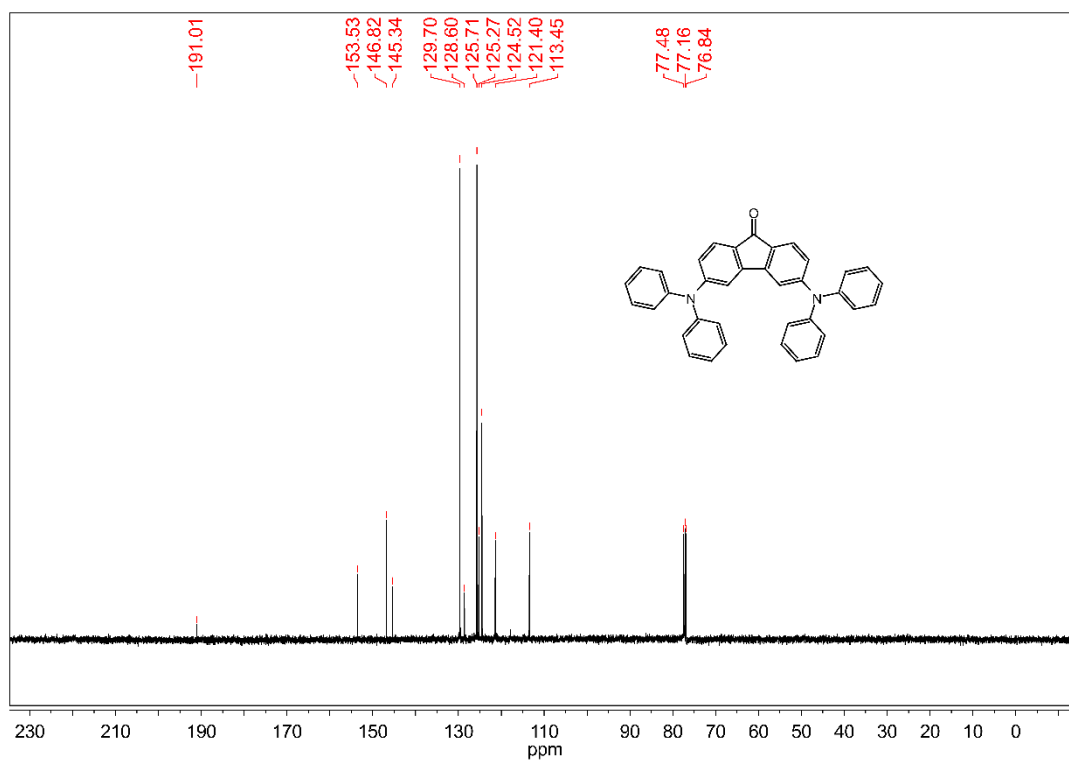
Supplementary Figure 51 | ¹H-NMR of 1j.



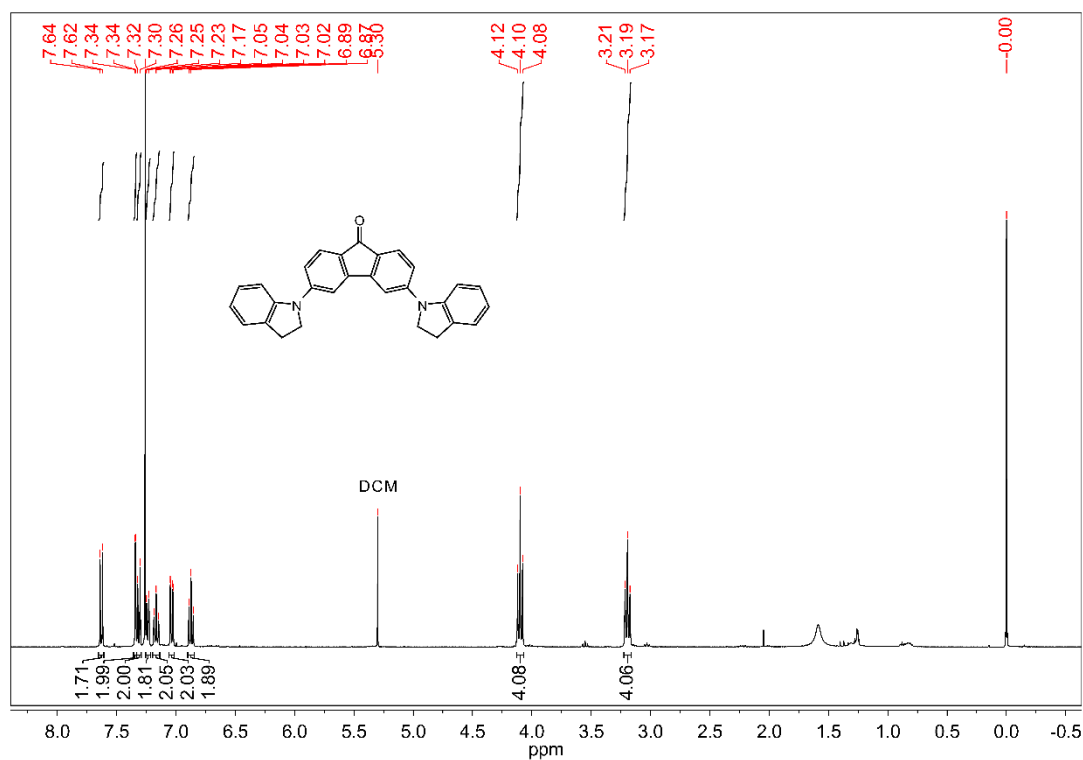
Supplementary Figure 52 | ¹³C-NMR of 1j.



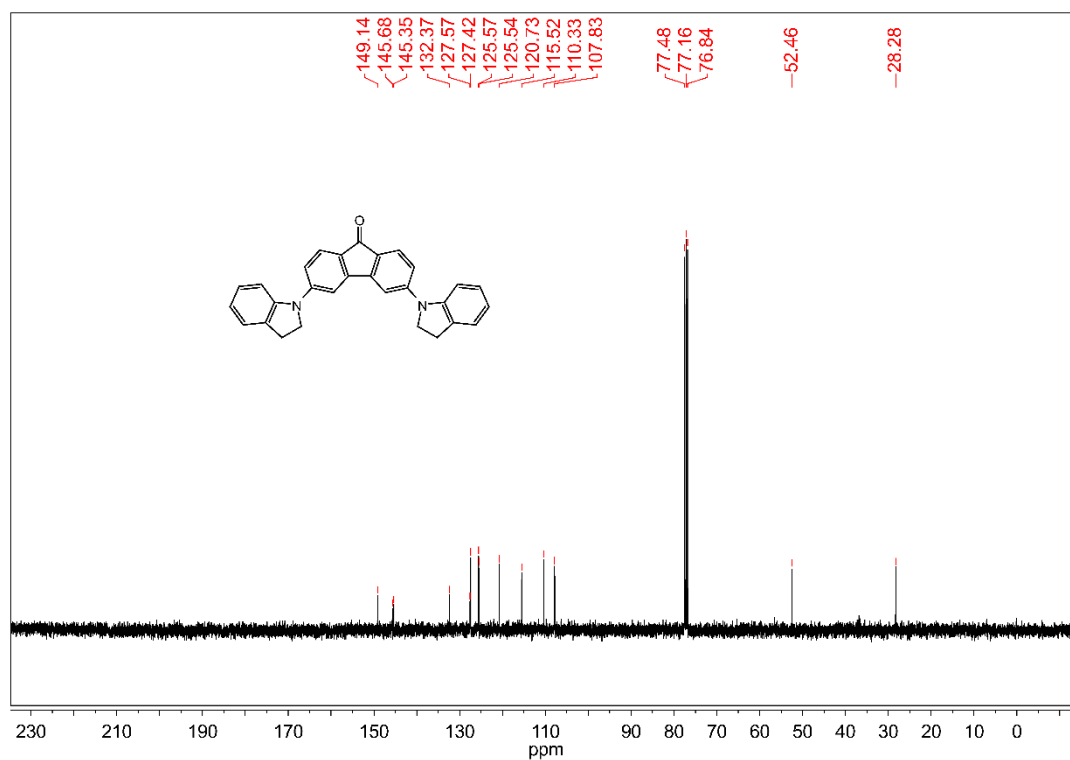
Supplementary Figure 53 | ¹H-NMR of 1k.



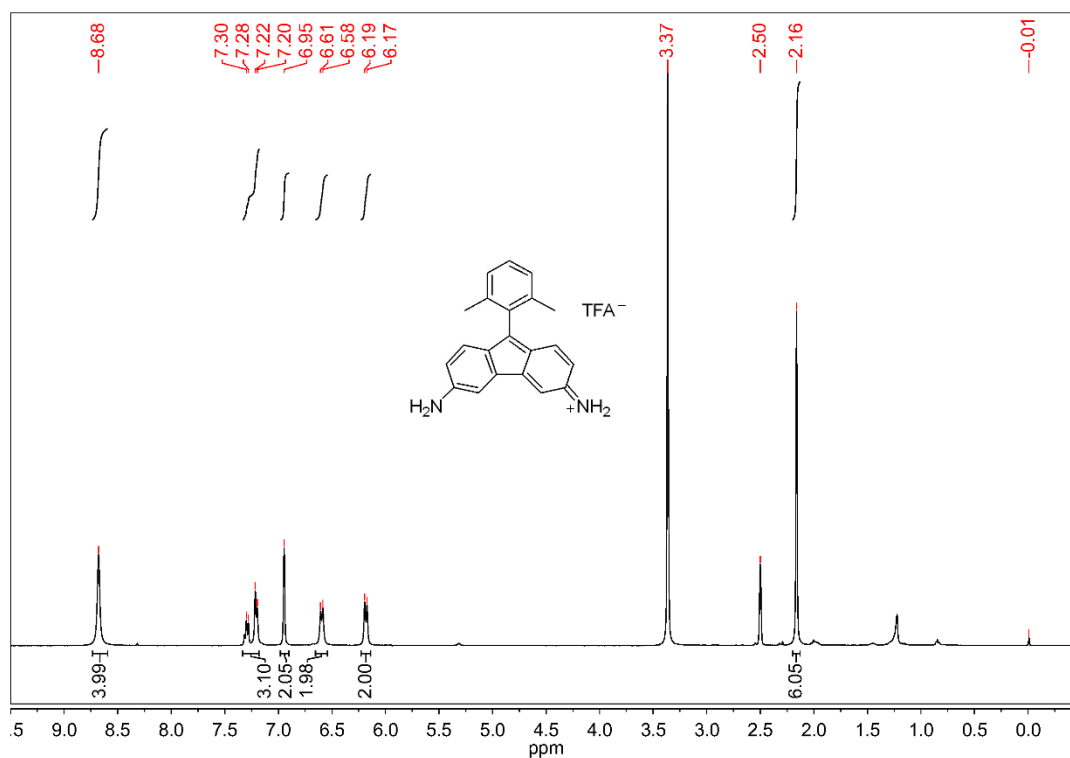
Supplementary Figure 54 | ¹³C-NMR of 1k.



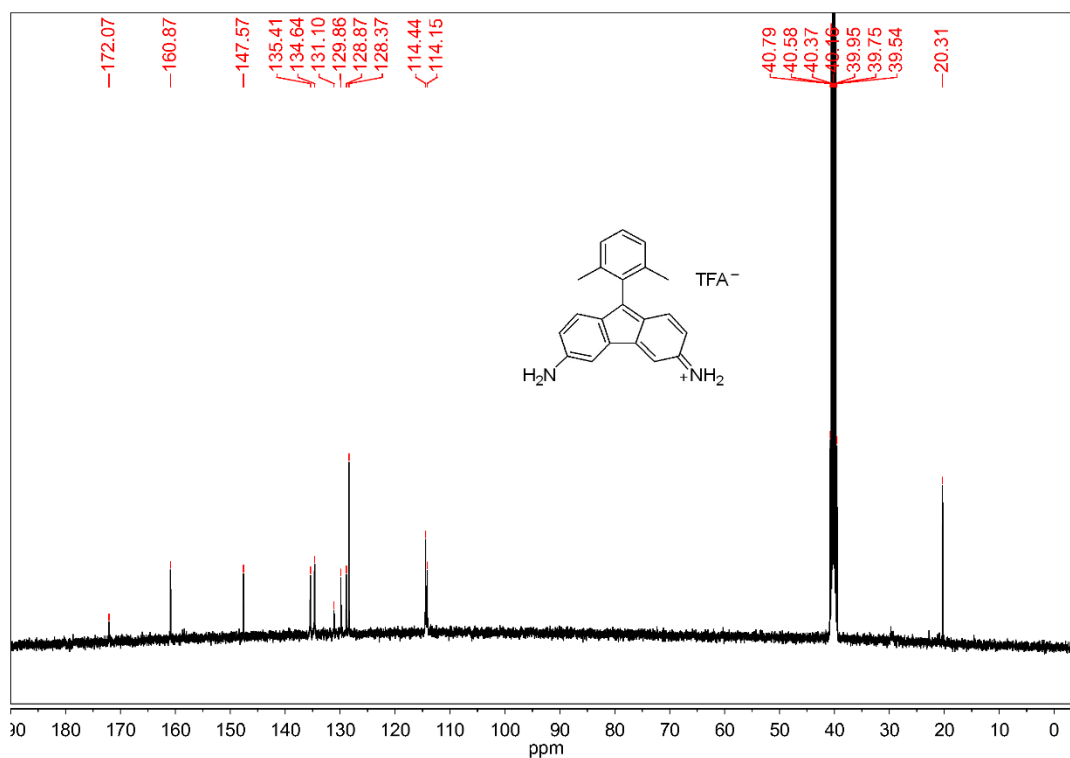
Supplementary Figure 55 | ¹H-NMR of 1m.



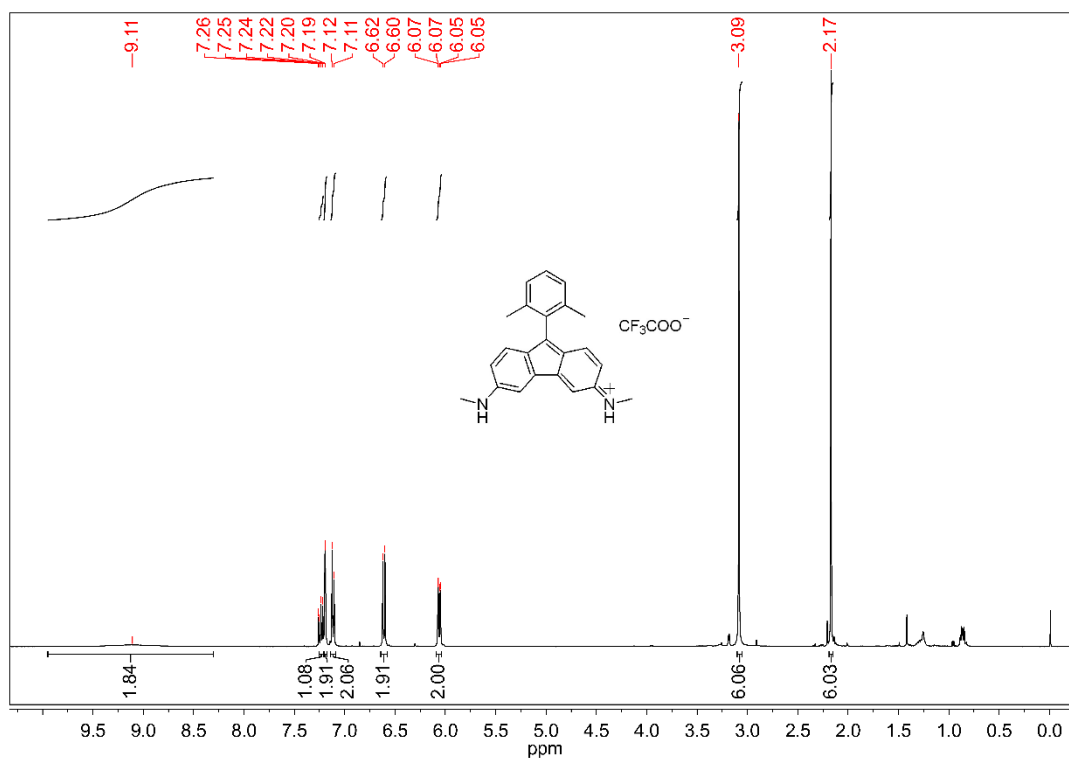
Supplementary Figure 56 | ¹³C-NMR of 1m.



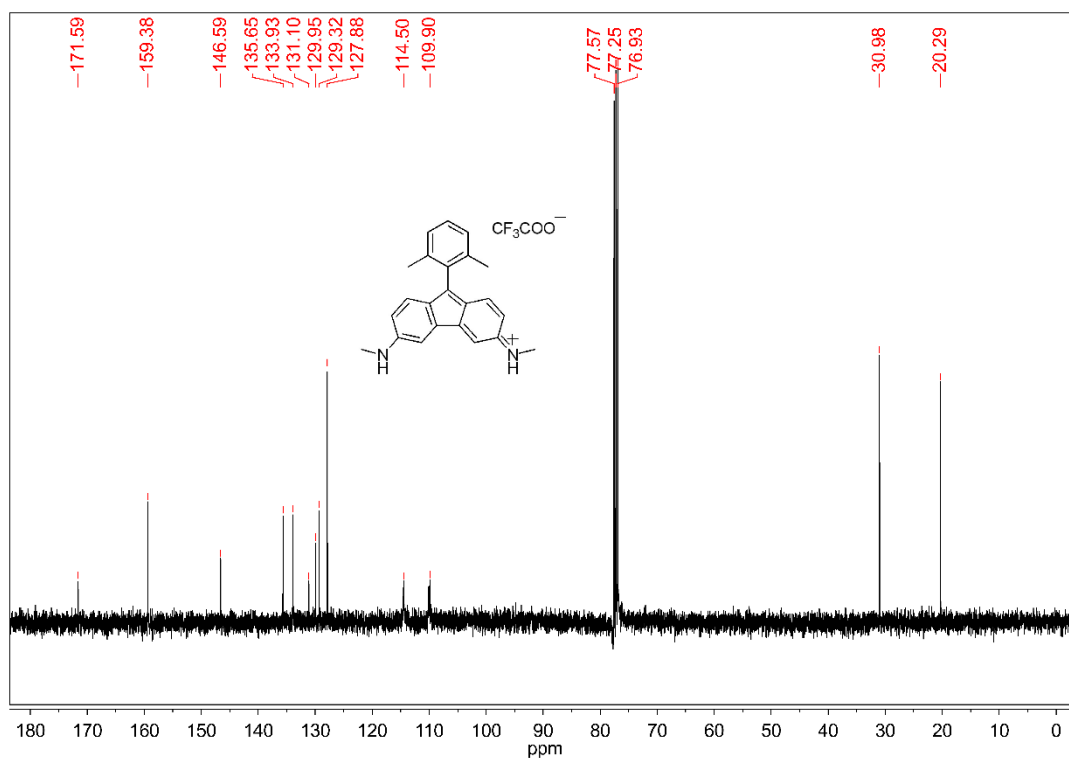
Supplementary Figure 57 | ¹H-NMR of AF2.



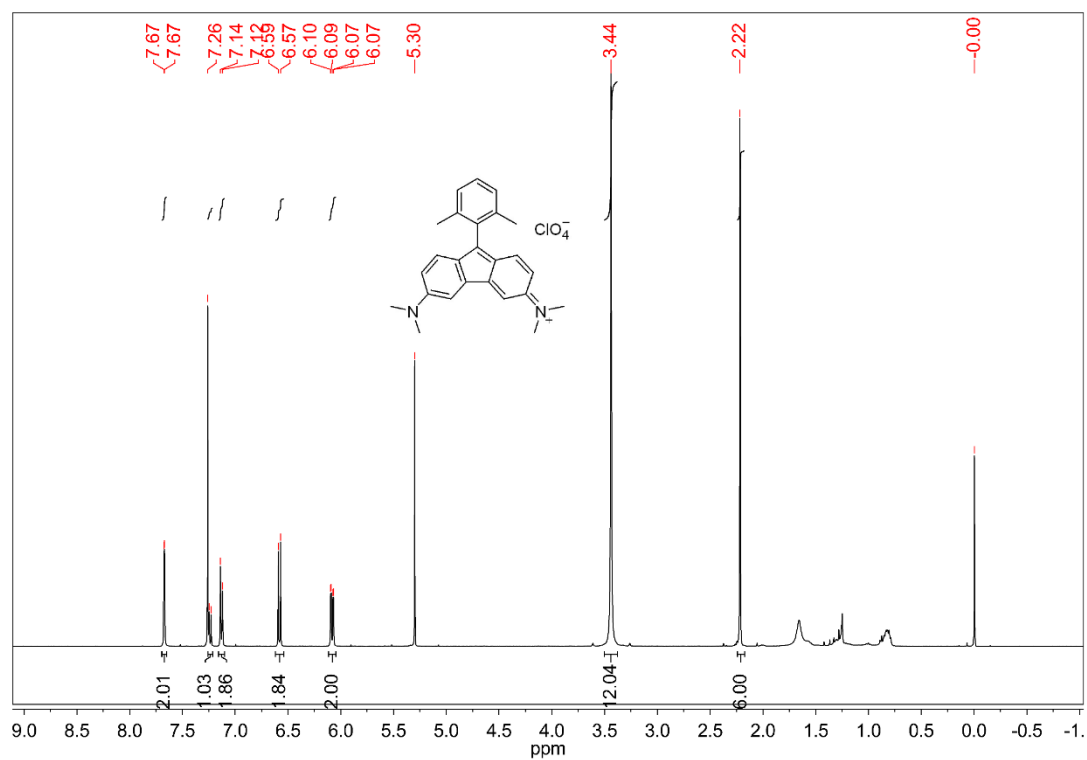
Supplementary Figure 58 | ¹³C-NMR of AF2.



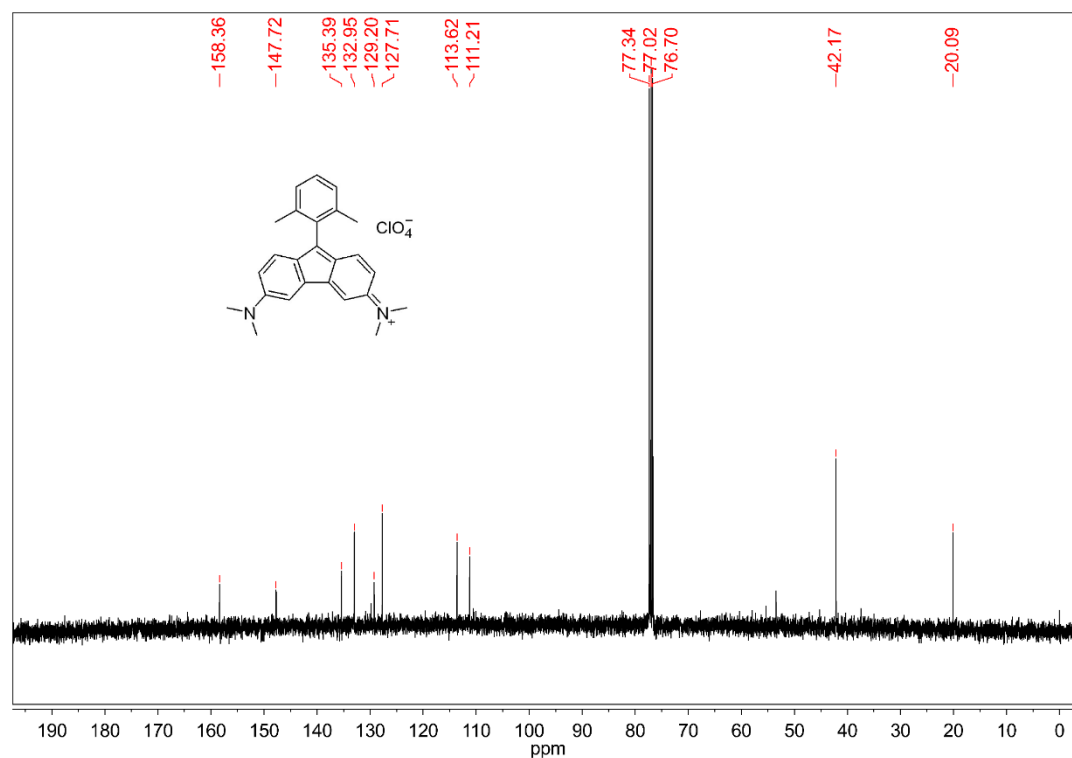
Supplementary Figure 59 | ¹H-NMR of AF2.



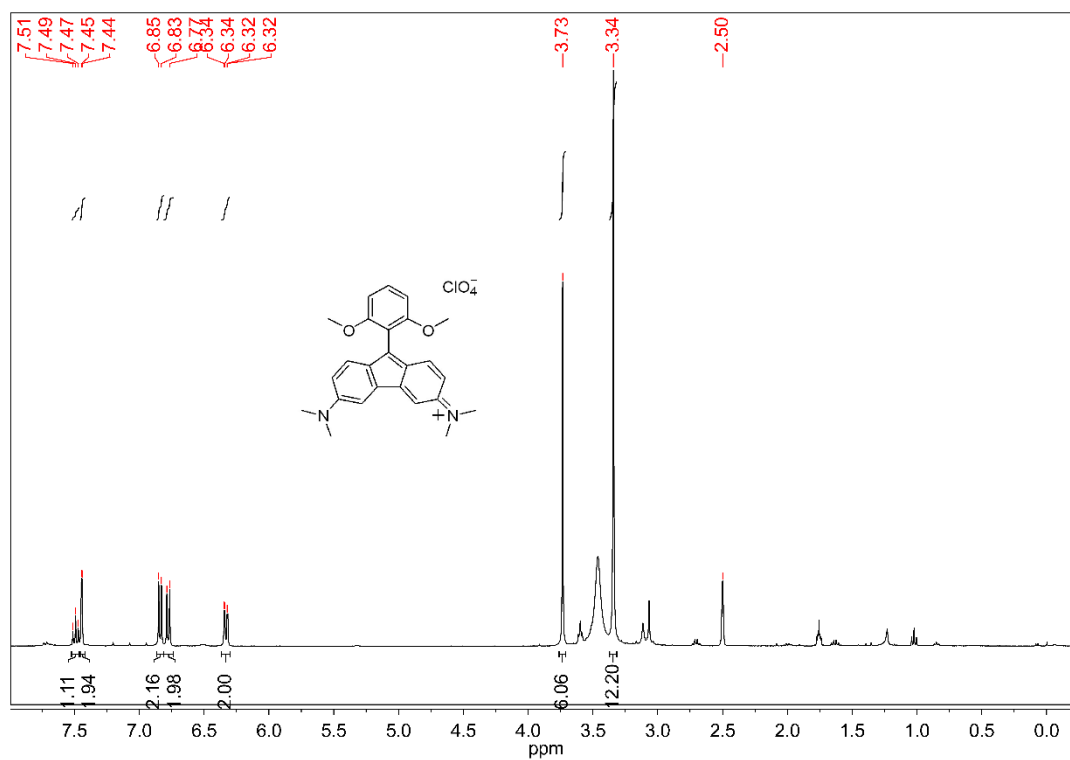
Supplementary Figure 60 | ¹³C-NMR of AF2.



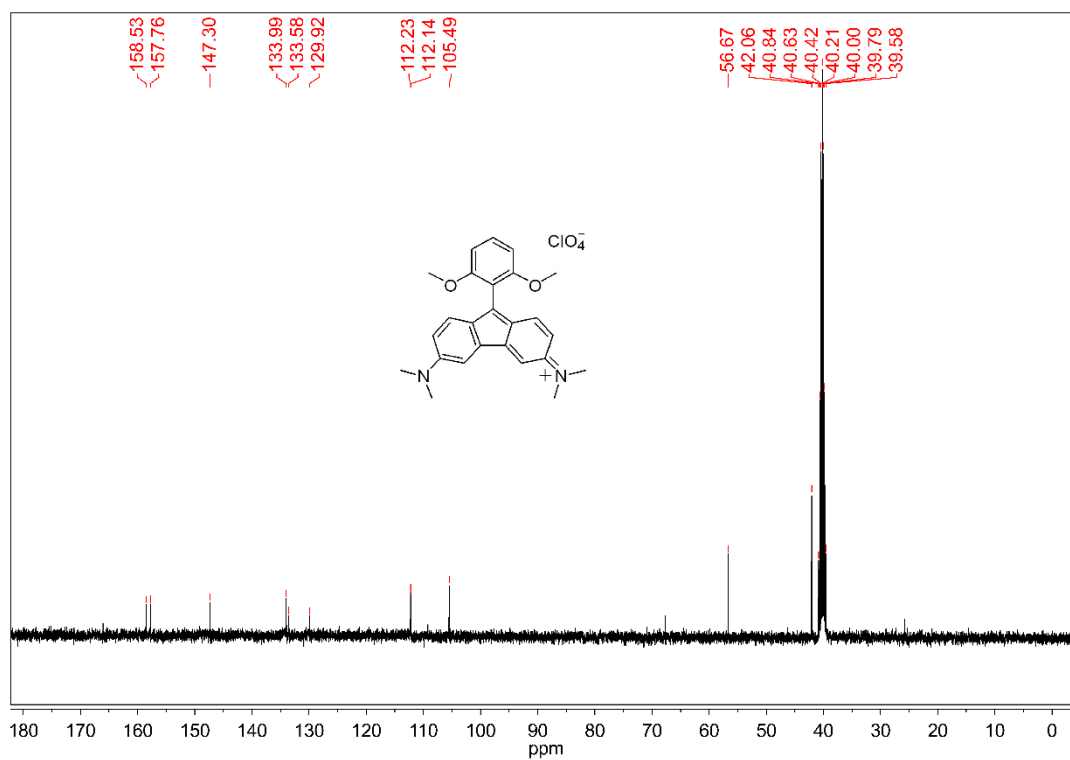
Supplementary Figure 61 | ¹H-NMR of AF3.



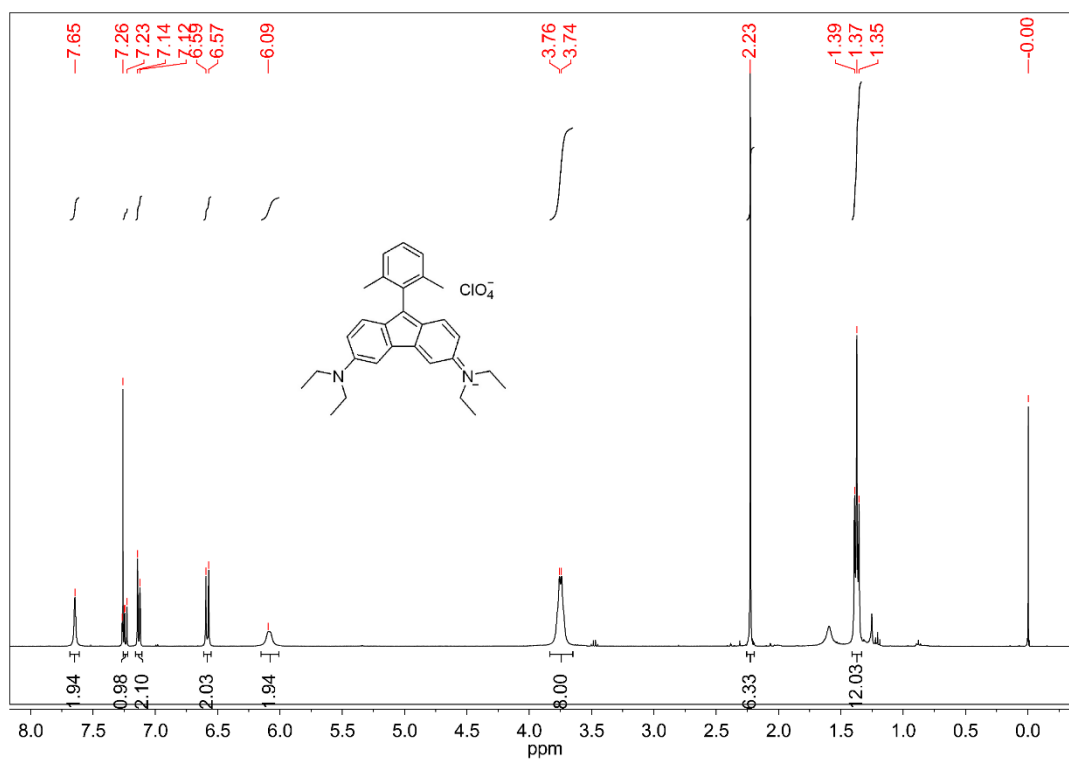
Supplementary Figure 62 | ¹³C-NMR of AF3.



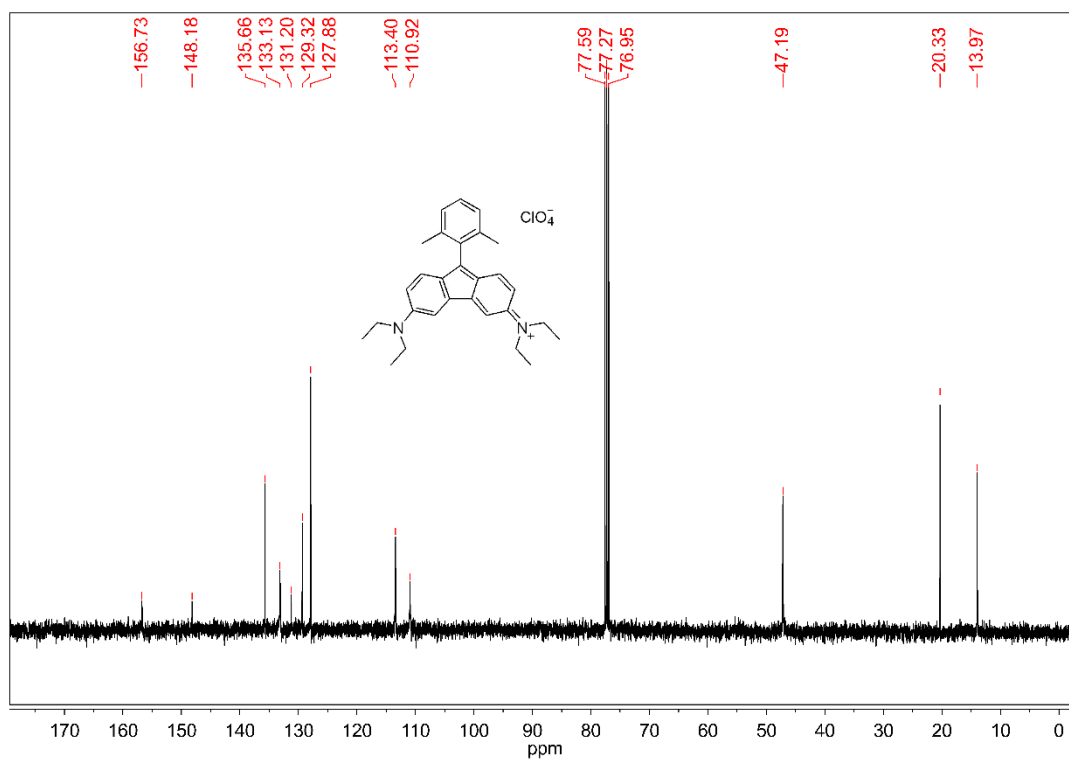
Supplementary Figure 63 | ¹H-NMR of AF4.



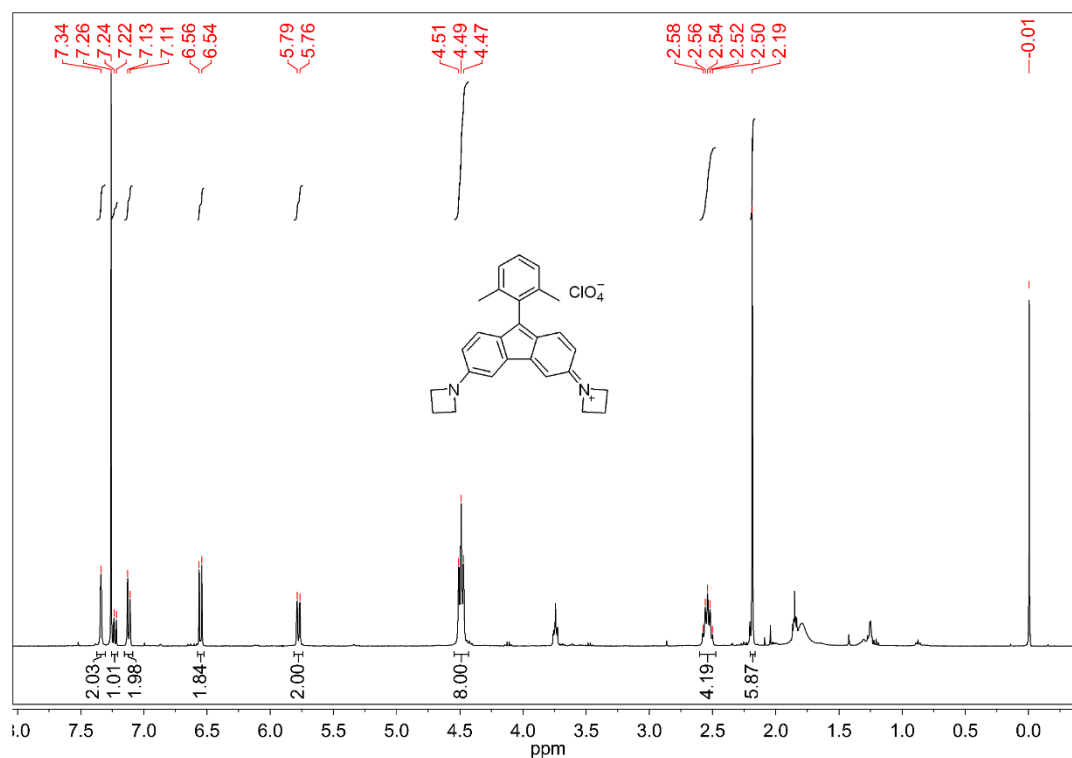
Supplementary Figure 64 | ¹³C-NMR of AF4.



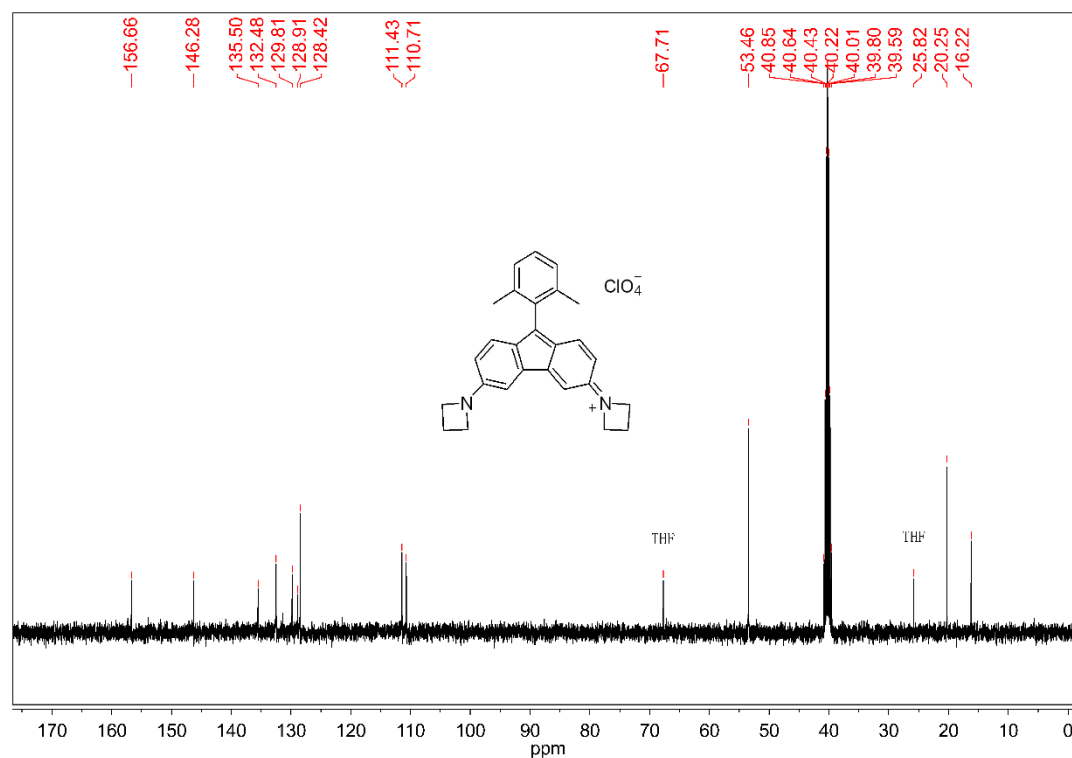
Supplementary Figure 65 | ¹H-NMR of AF5.



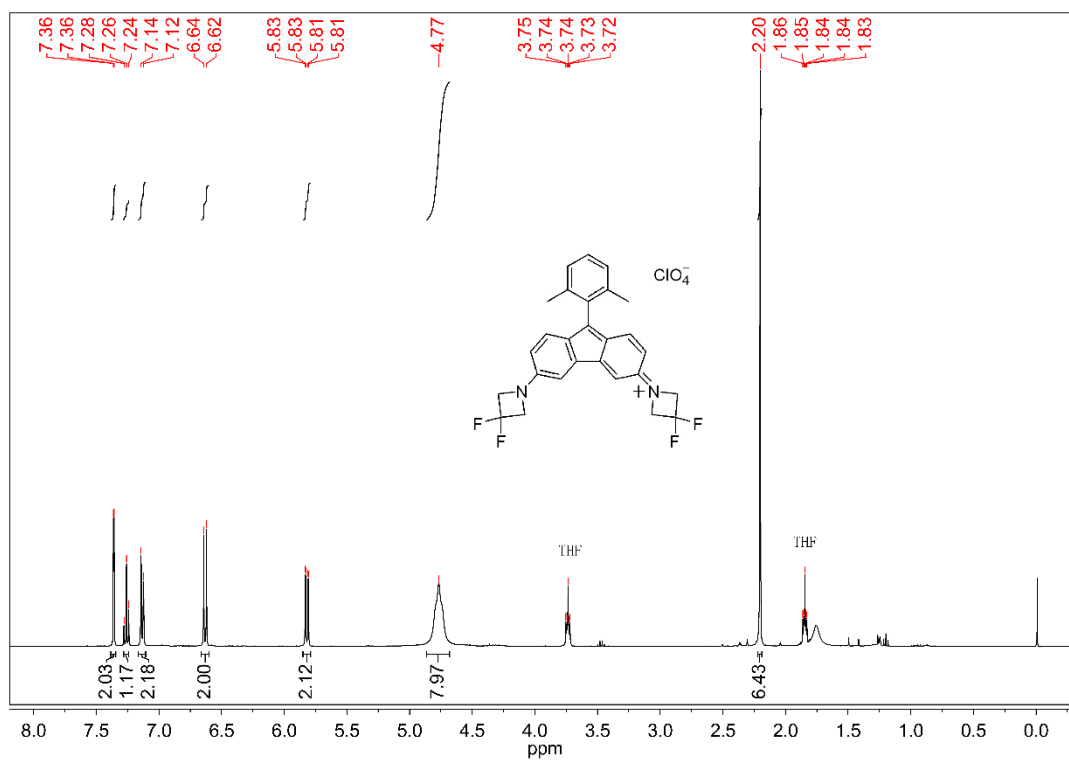
Supplementary Figure 66 | ¹³C-NMR of AF5.



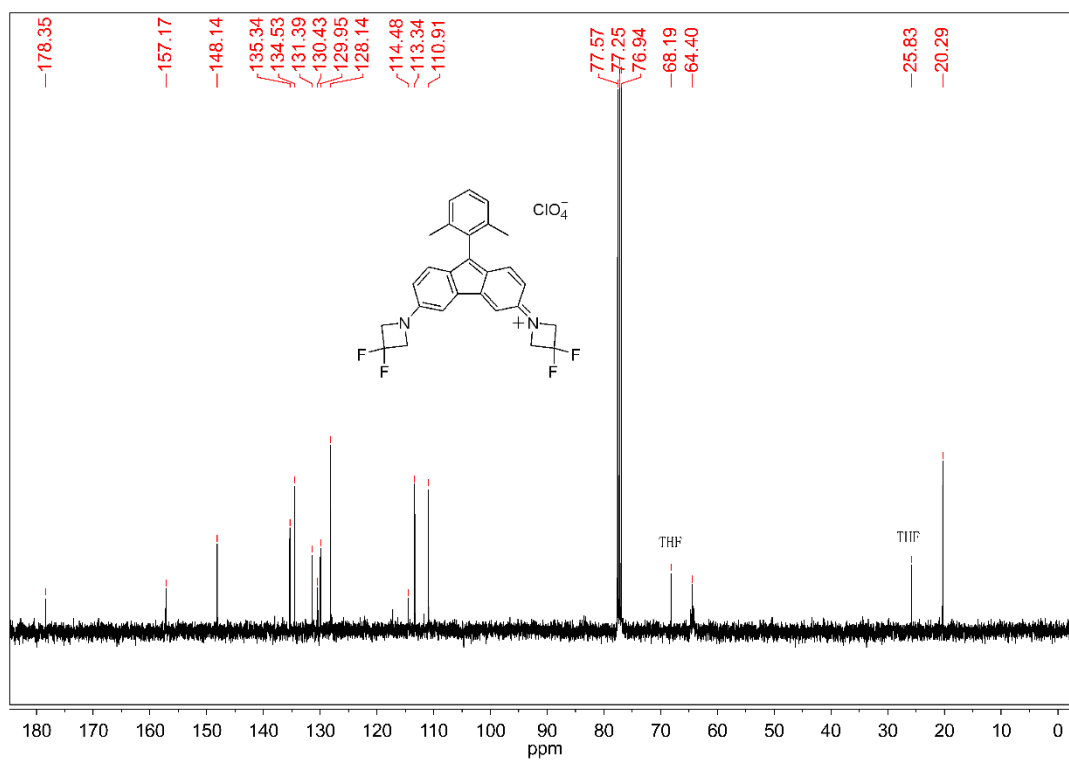
Supplementary Figure 67 | ¹H-NMR of AF6.



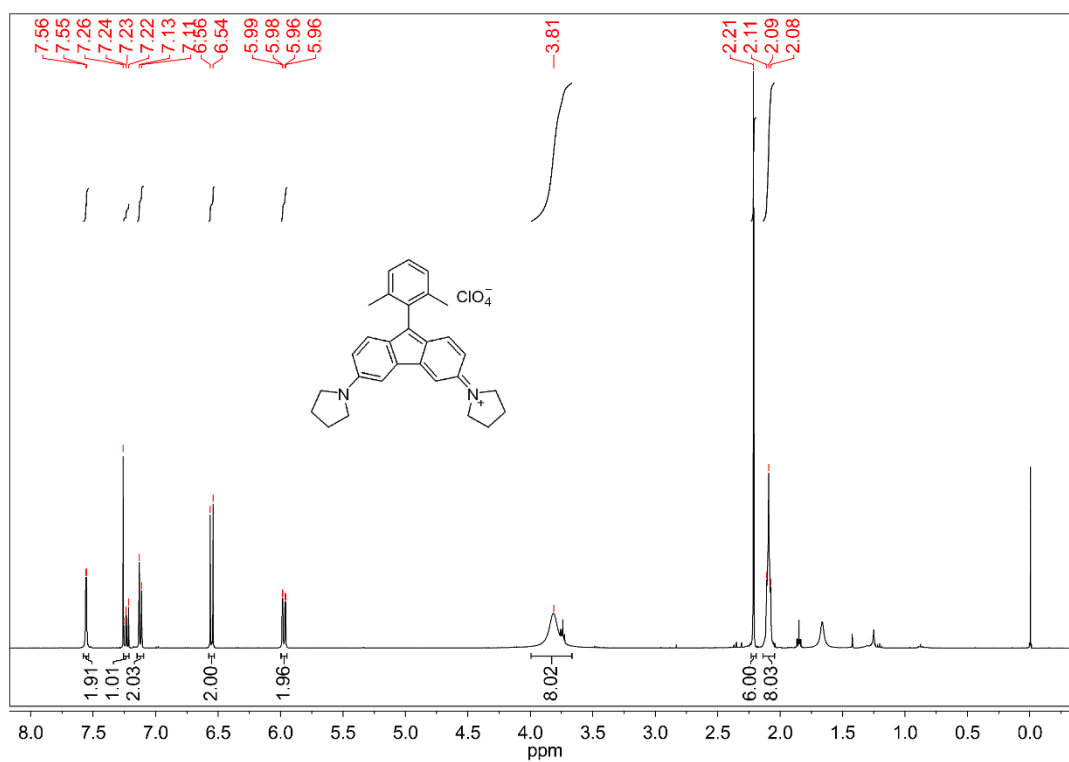
Supplementary Figure 68 | ¹³C-NMR of AF6.



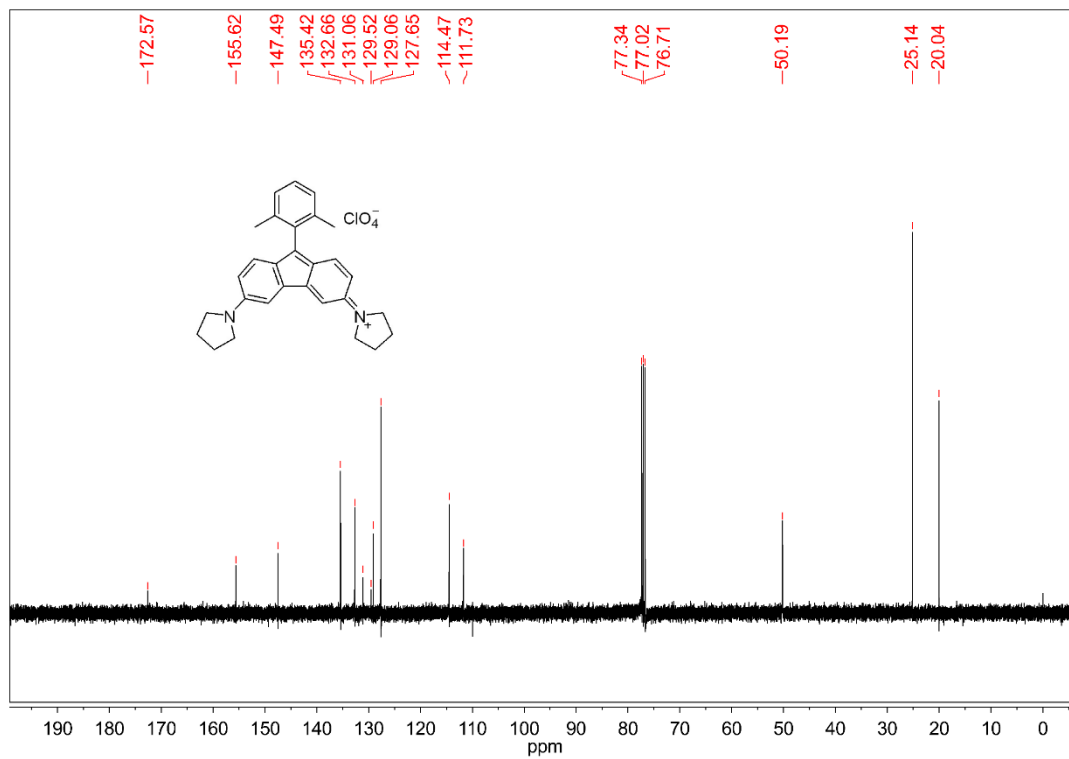
Supplementary Figure 69 | ¹H-NMR of AF7.



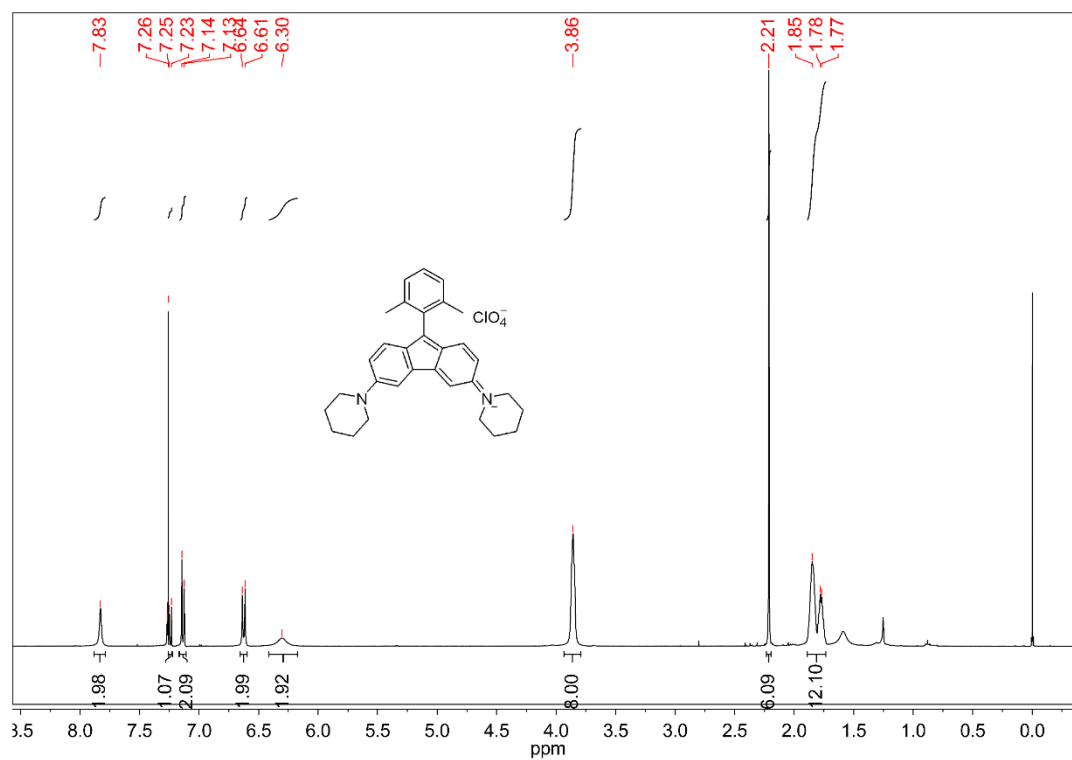
Supplementary Figure 70 | ¹³C-NMR of AF7.



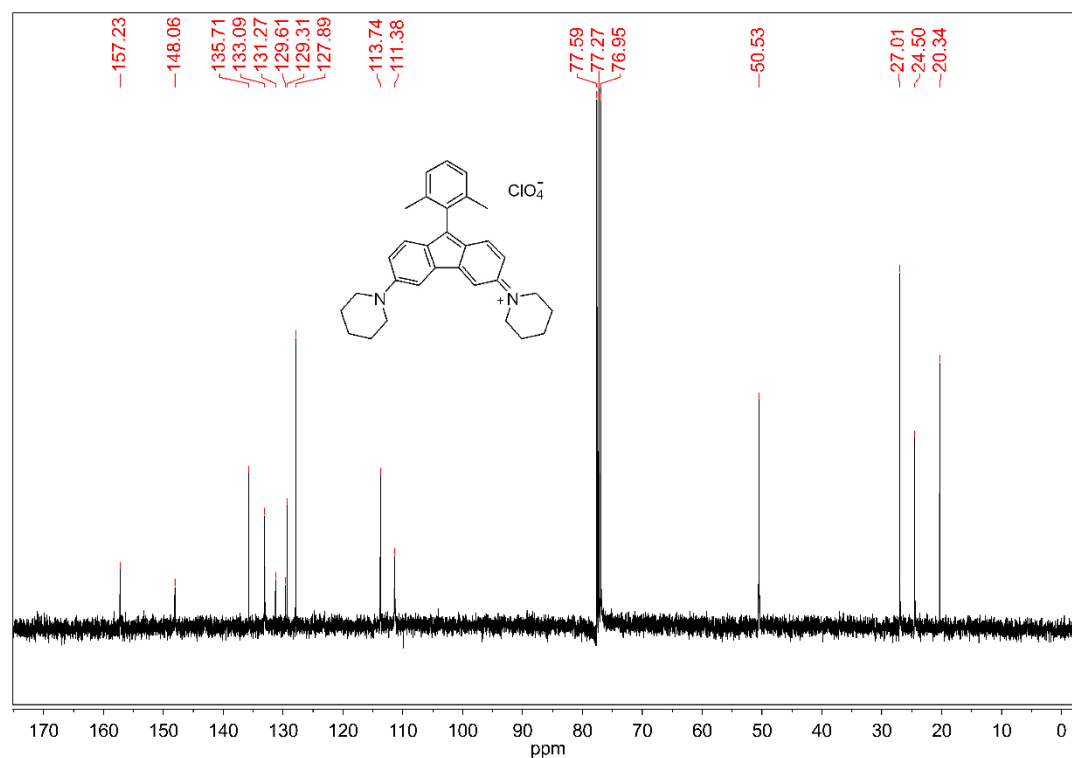
Supplementary Figure 71 | ¹H-NMR of AF8.



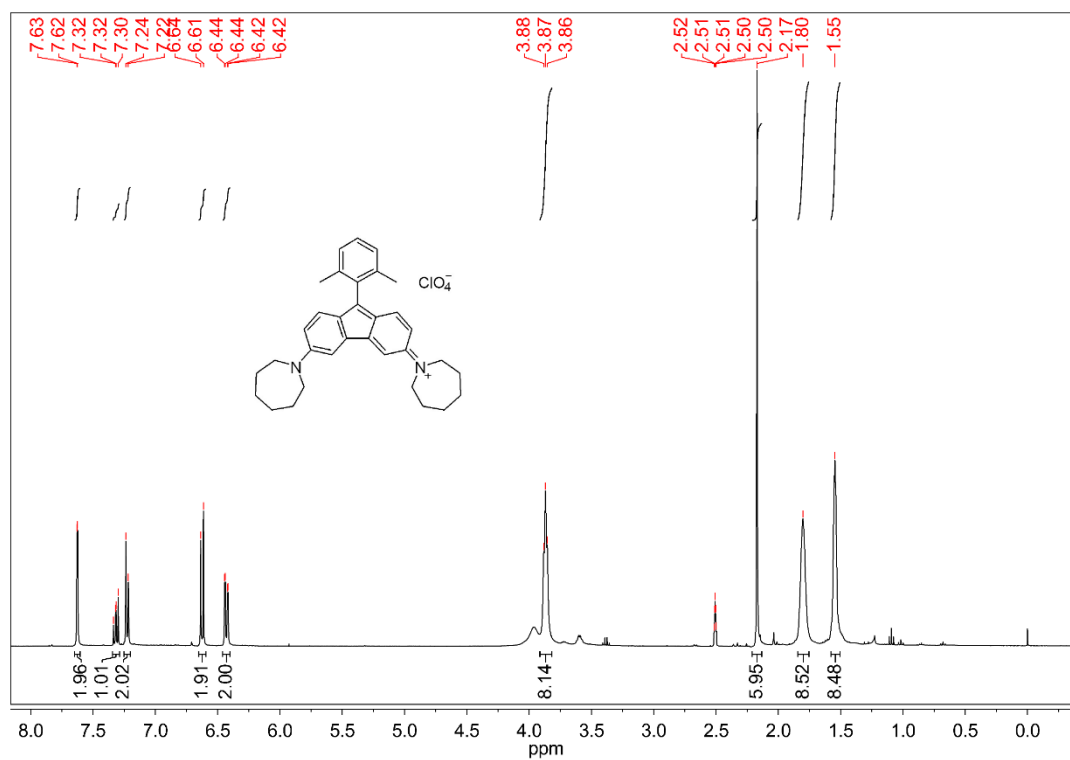
Supplementary Figure 72 | ¹³C-NMR of AF8.



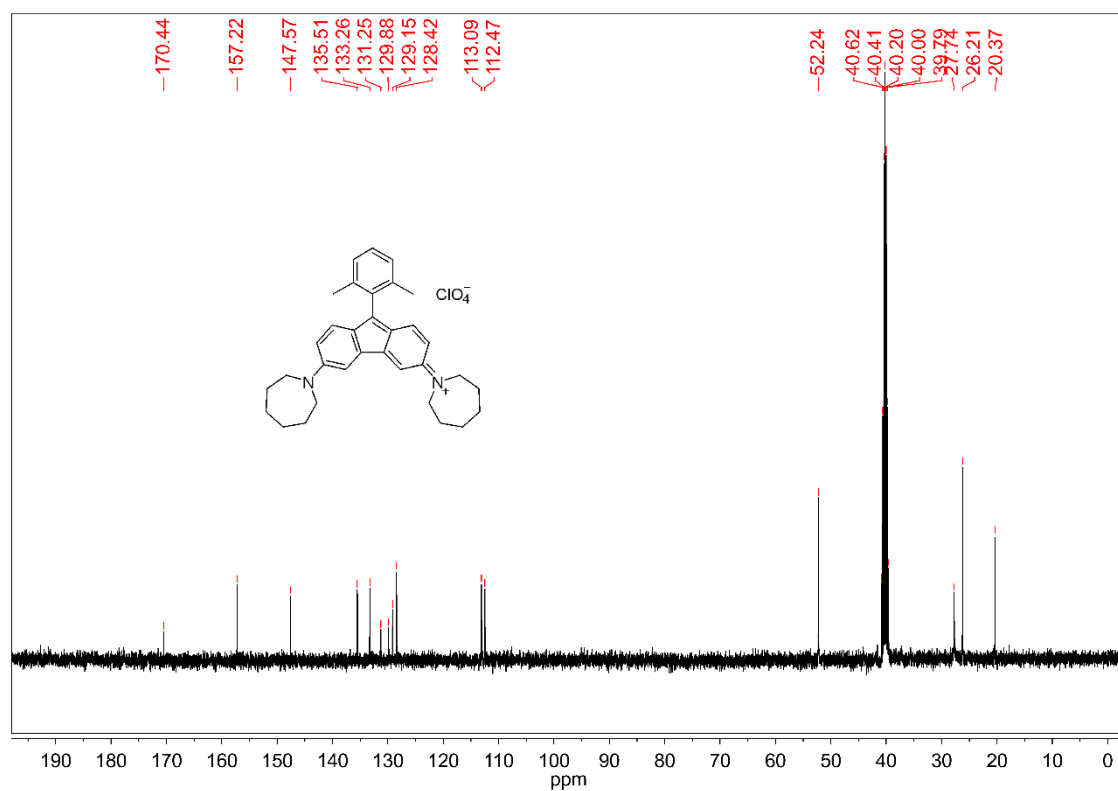
Supplementary Figure 73 | ¹H-NMR of AF9.



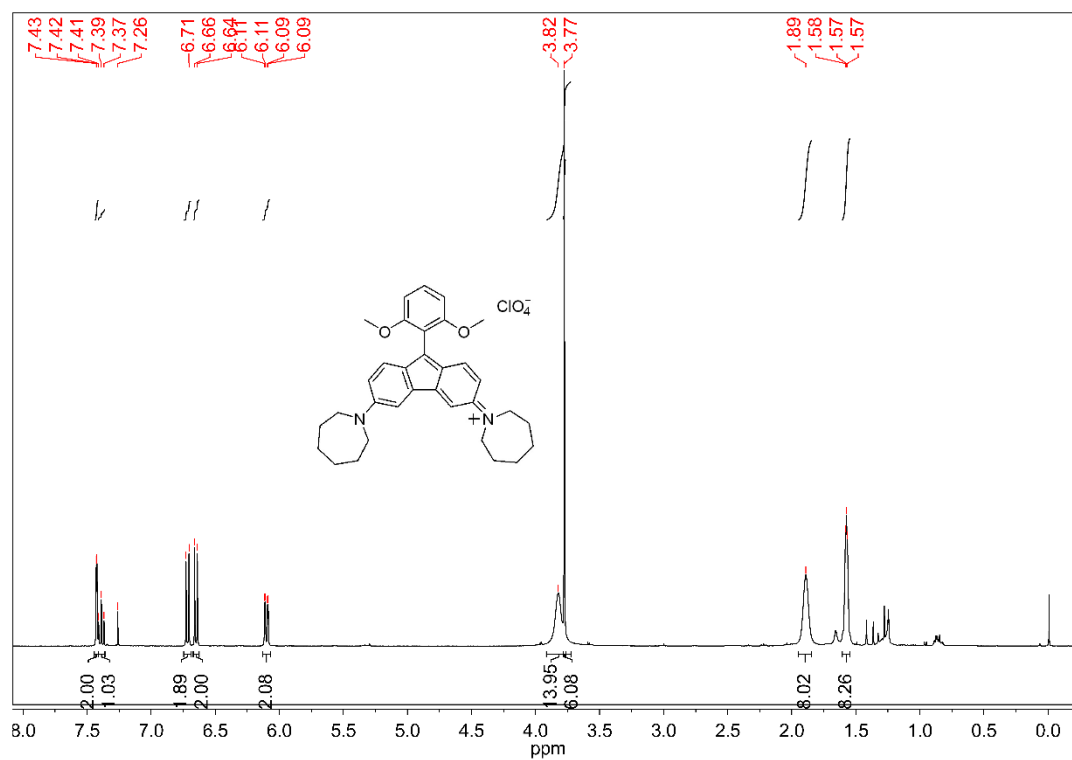
Supplementary Figure 74 | ¹³C-NMR of AF9.



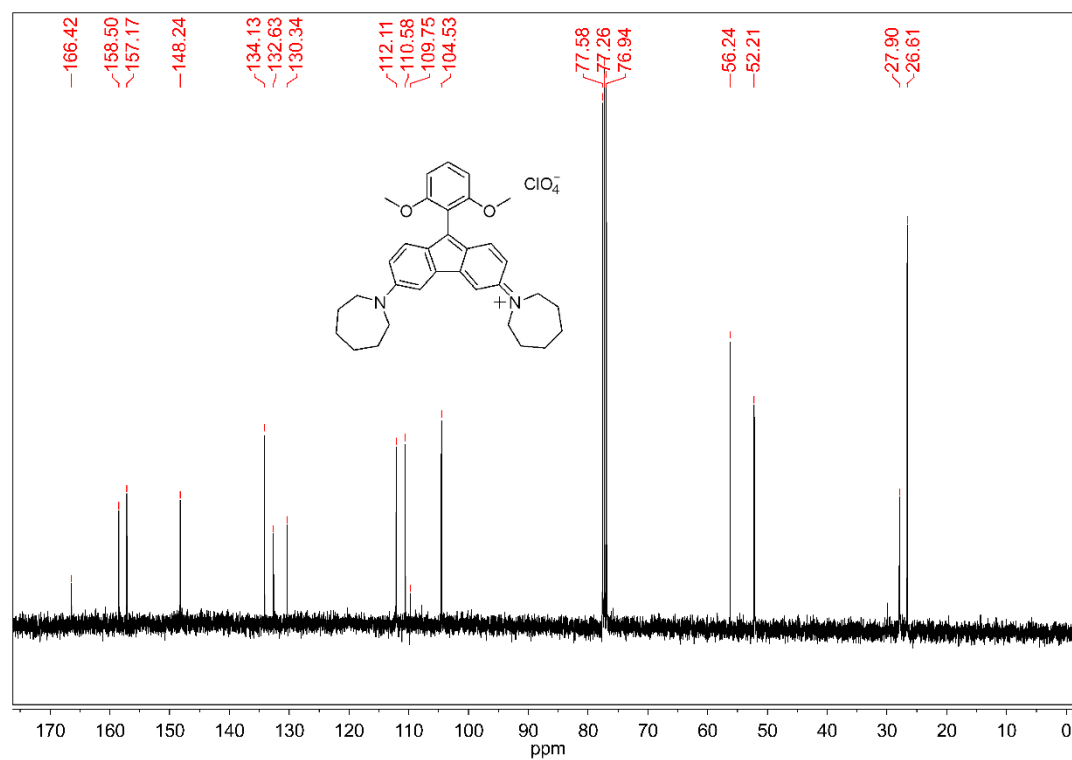
Supplementary Figure 75 | ¹H-NMR of AF10.



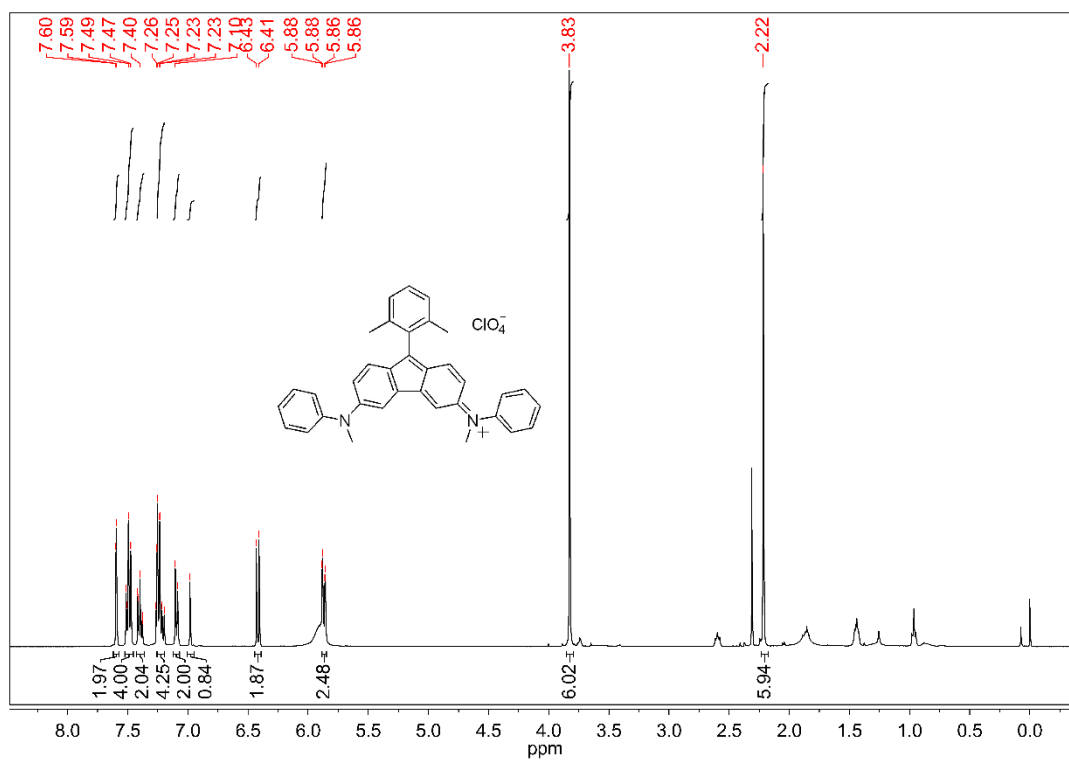
Supplementary Figure 76 | ¹³C-NMR of AF10.



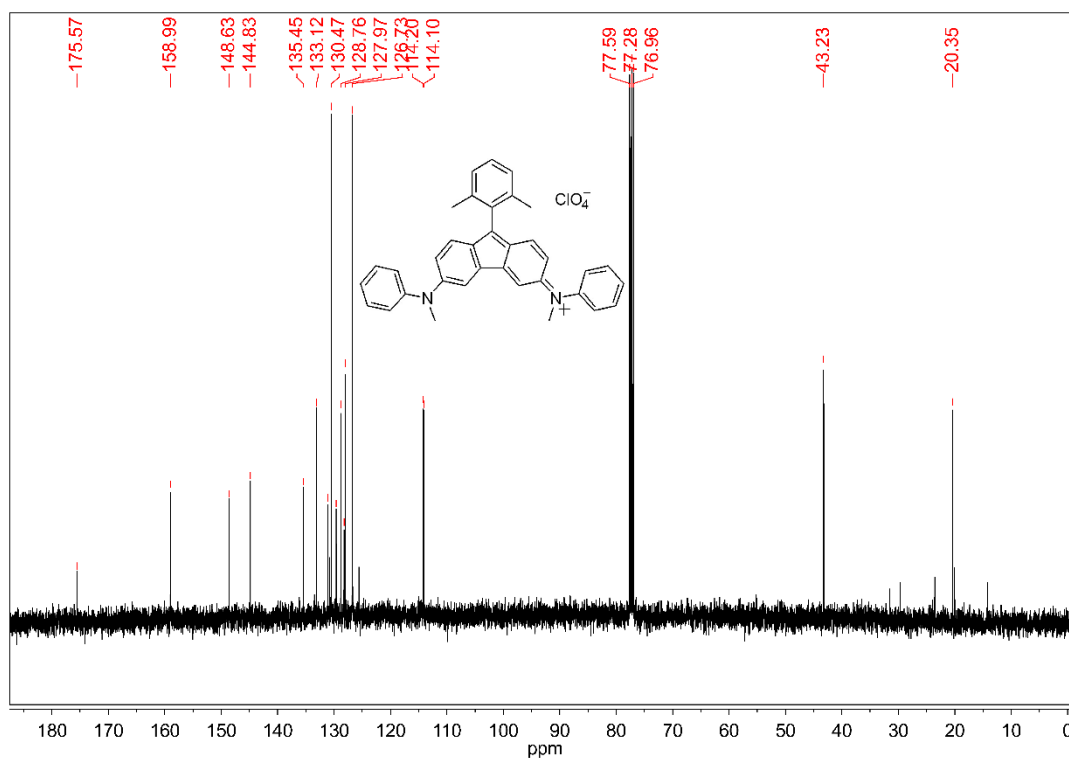
Supplementary Figure 77 | ¹H-NMR of AF11.



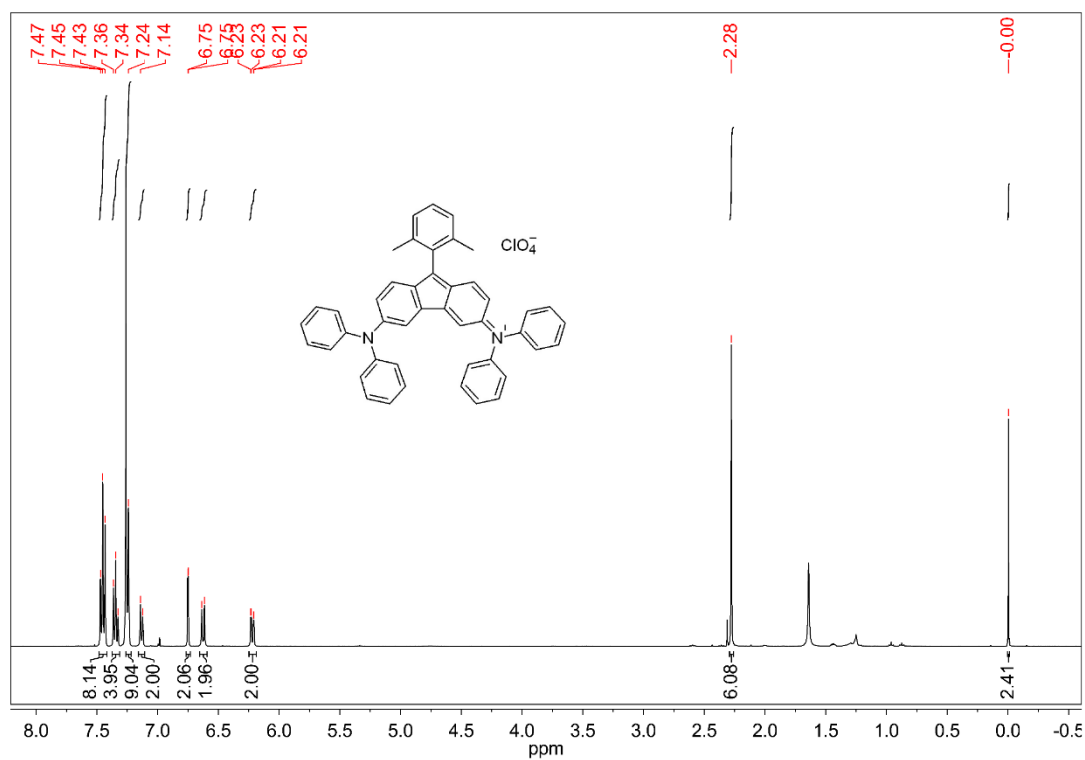
Supplementary Figure 78 | ¹³C-NMR of AF11.



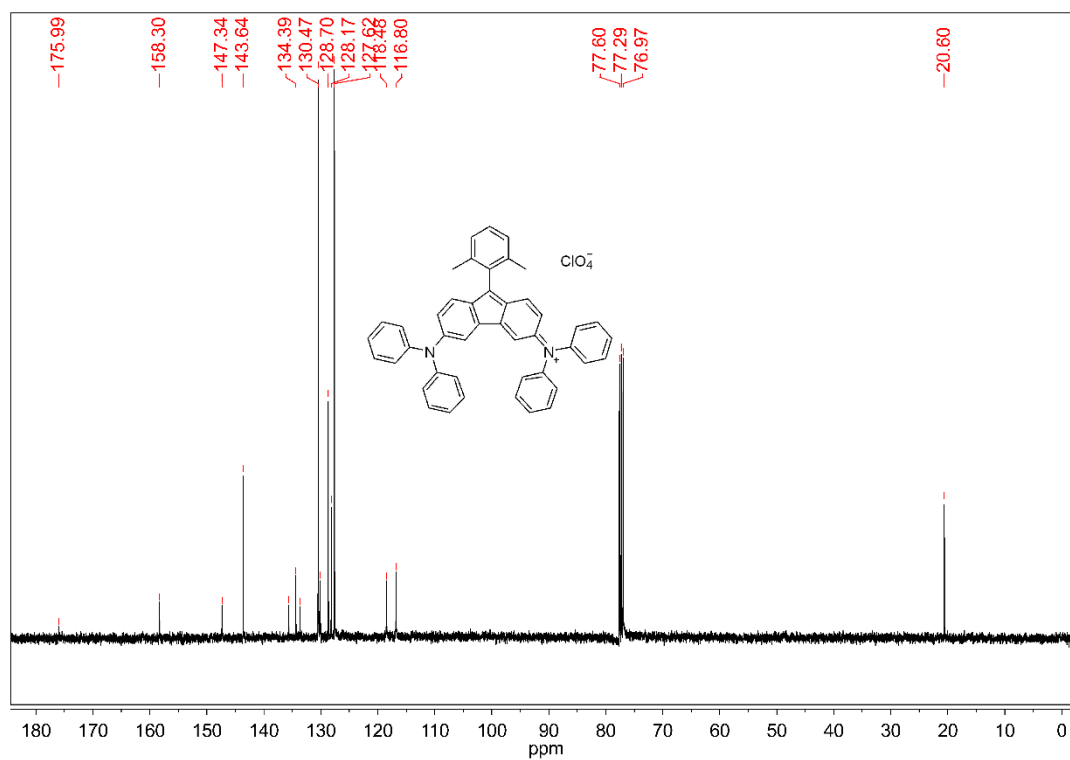
Supplementary Figure 79 | ¹H-NMR of AF12.



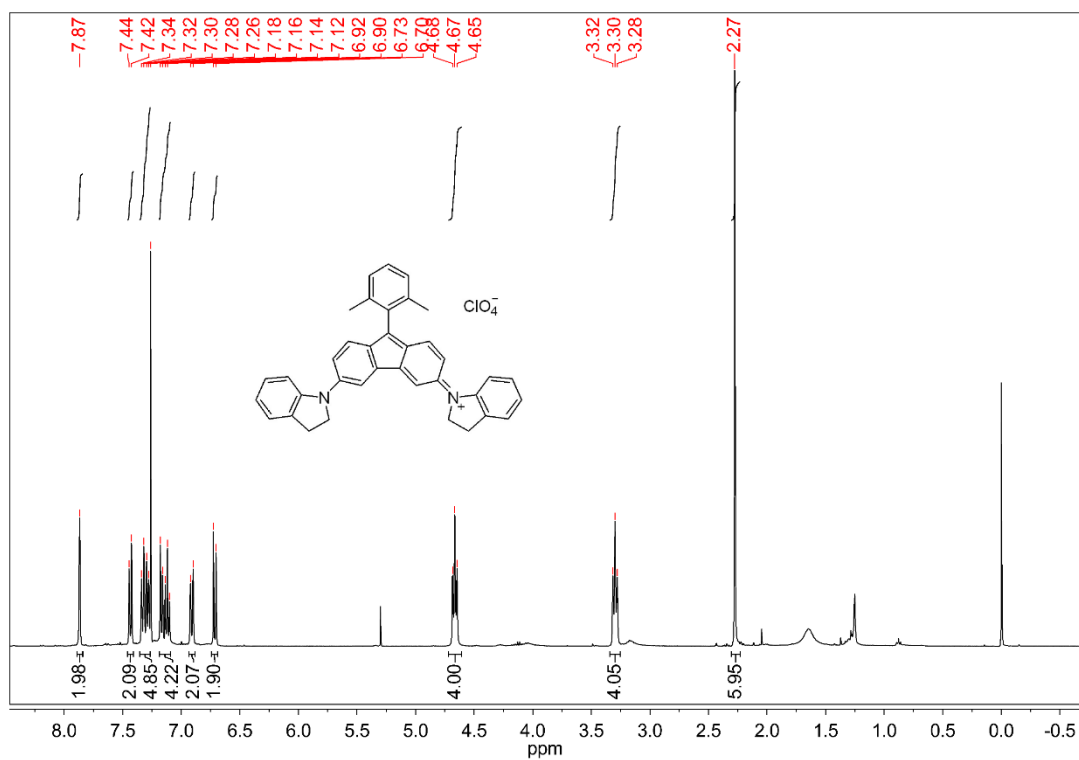
Supplementary Figure 80 | ¹³C-NMR of AF12.



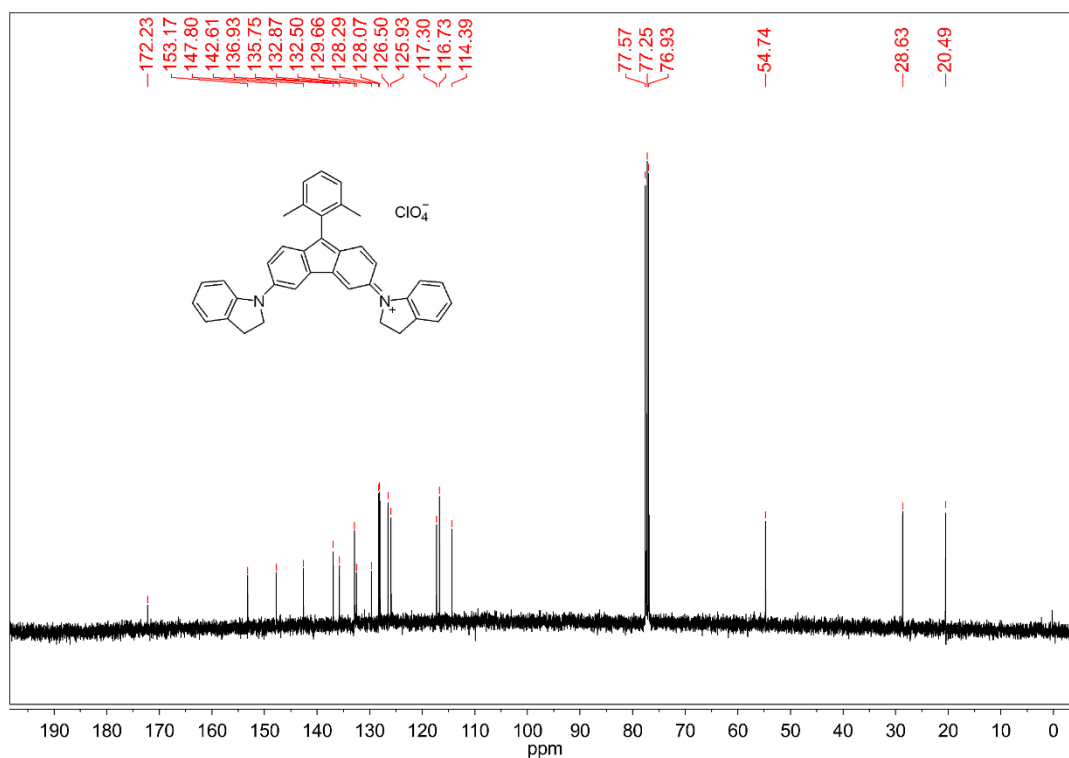
Supplementary Figure 81 | ¹H-NMR of AF13.



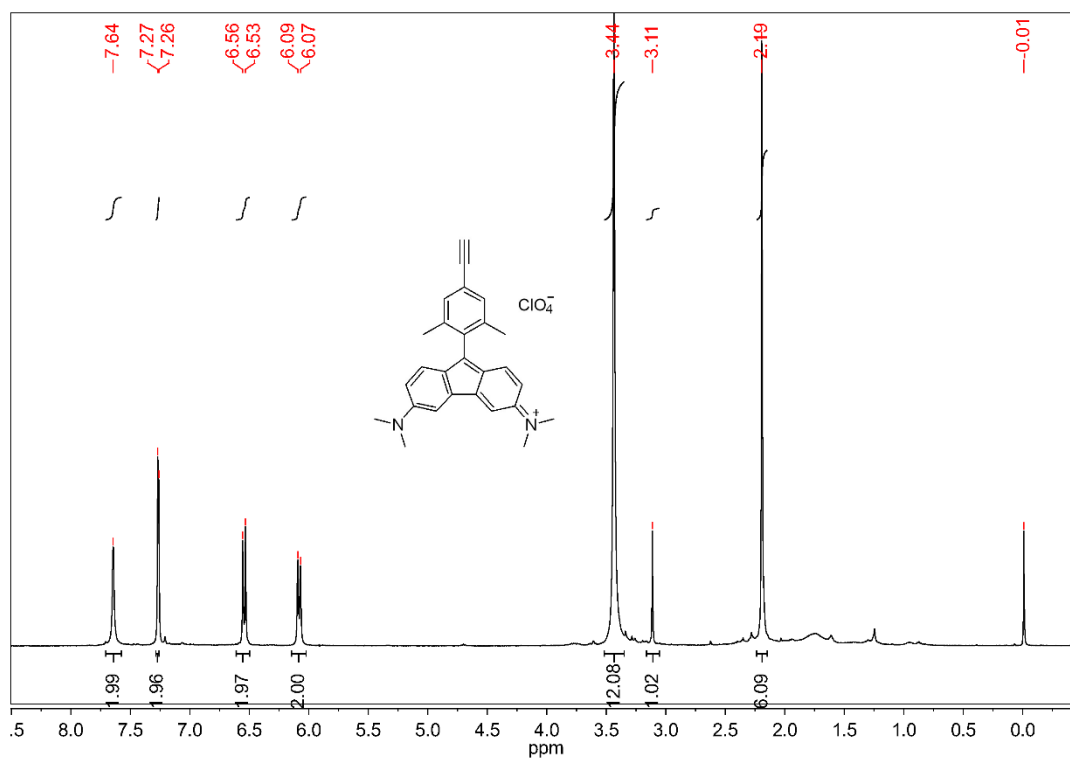
Supplementary Figure 82 | ¹³C-NMR of AF13.



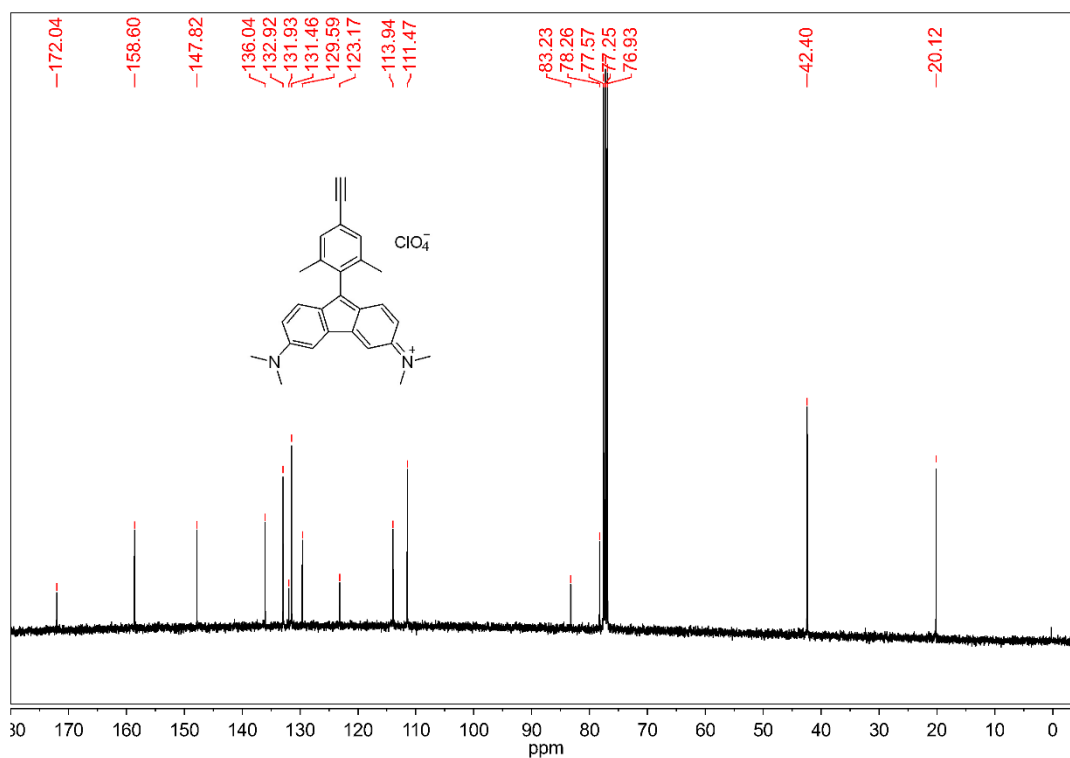
Supplementary Figure 83 | ¹H-NMR of AF14.



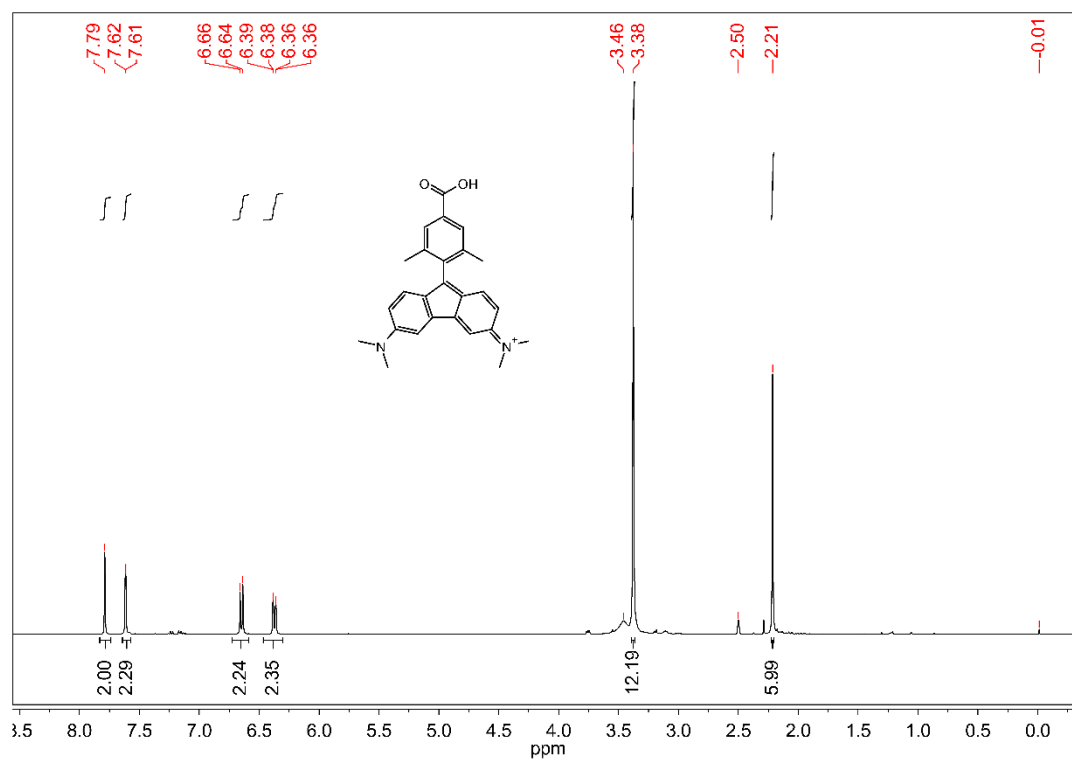
Supplementary Figure 84 | ¹³C-NMR of AF14.



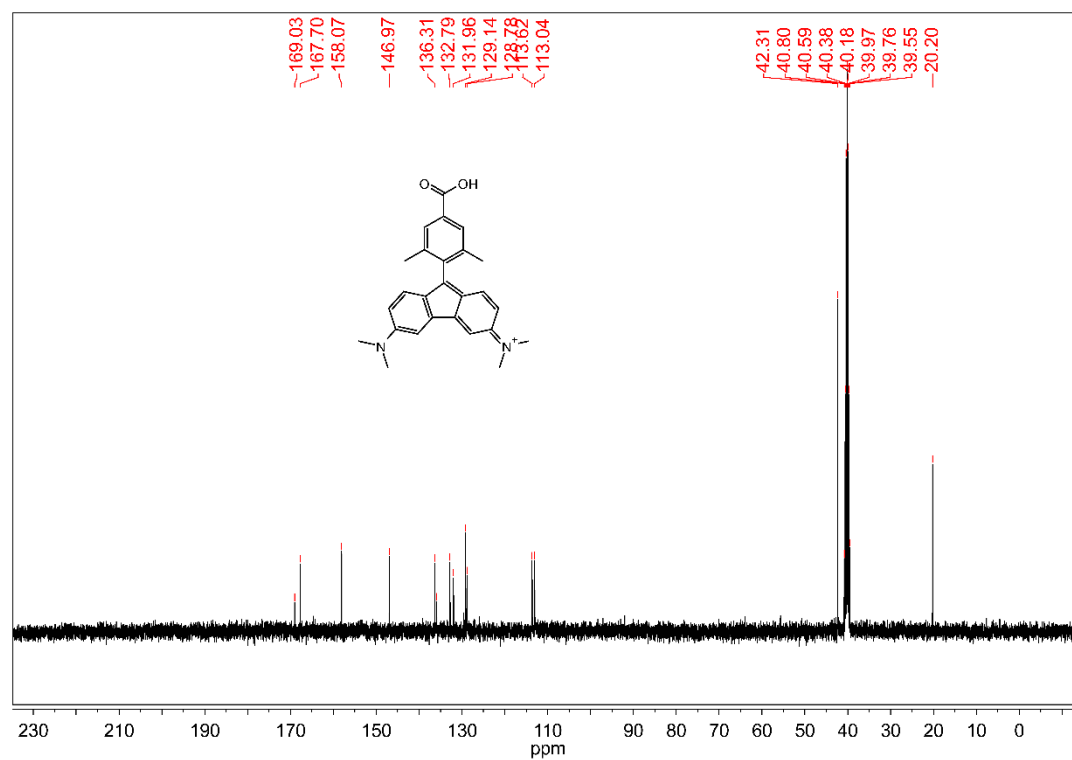
Supplementary Figure 85 | ¹H-NMR of AF3-Alkyne.



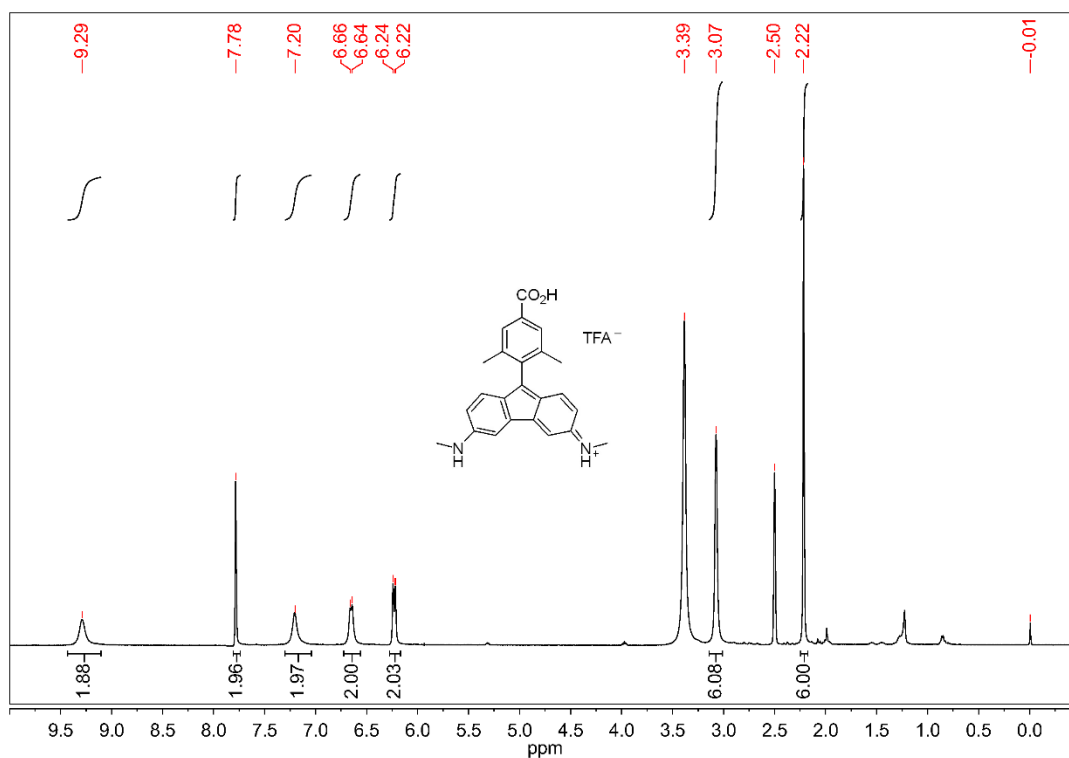
Supplementary Figure 86 | ¹³C-NMR of AF3-Alkyne.



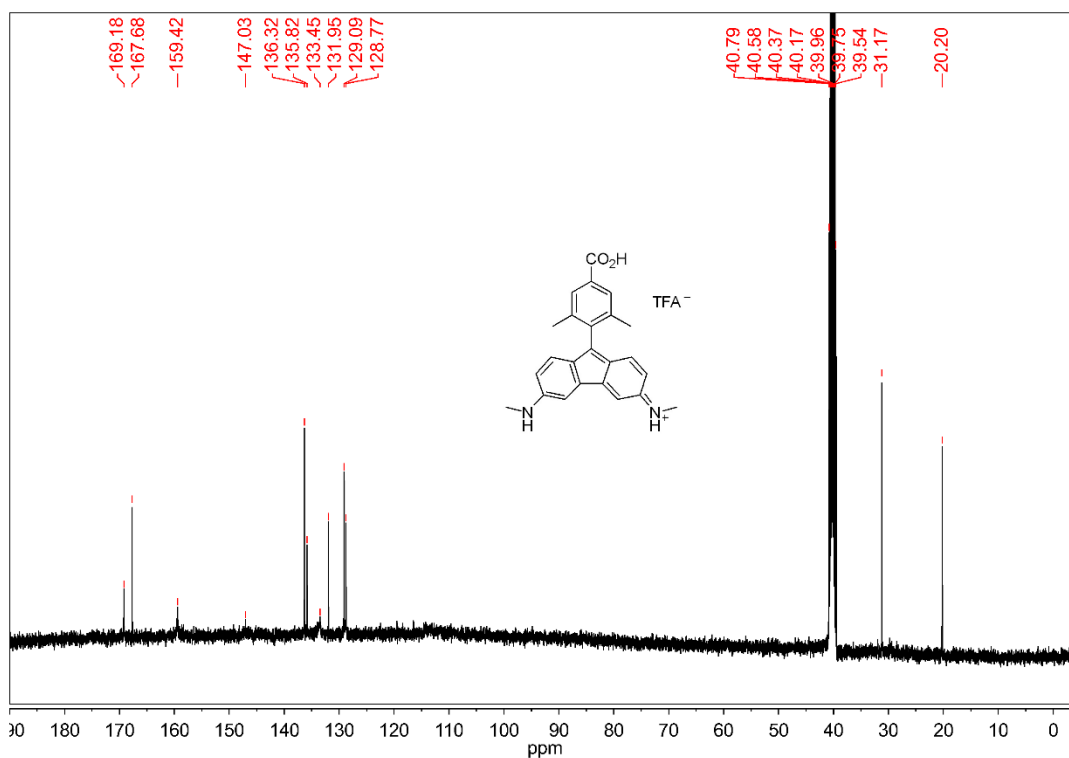
Supplementary Figure 87 | ¹H-NMR of AF3-COOH.



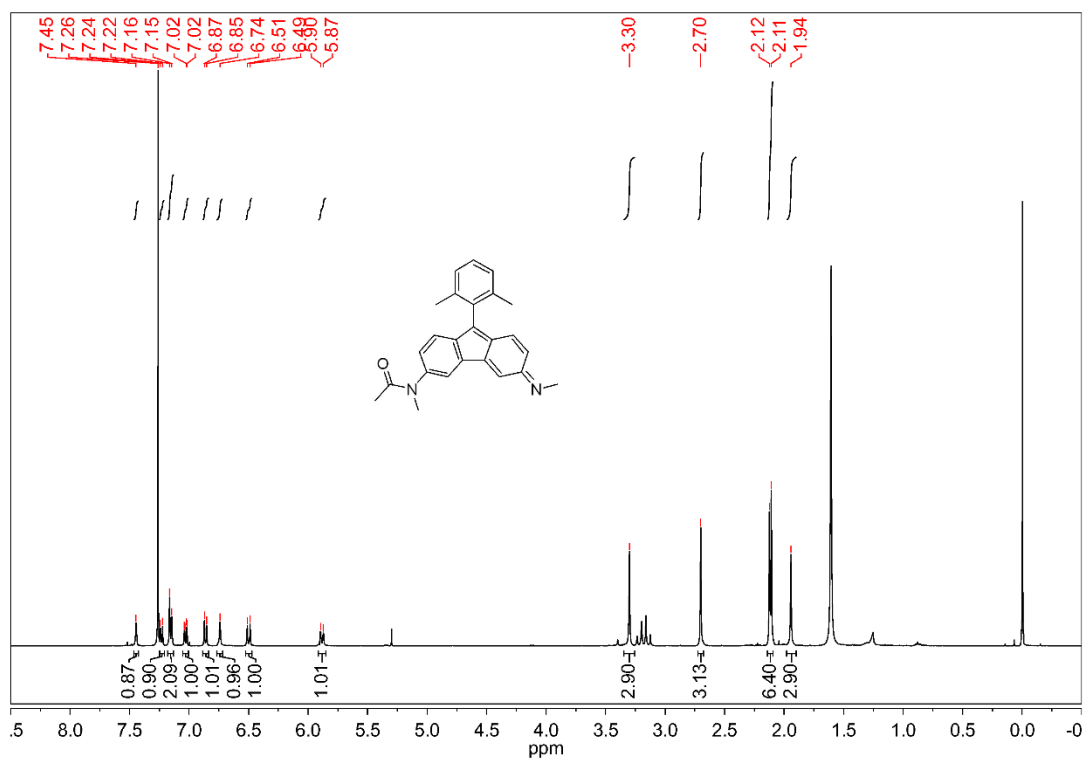
Supplementary Figure 88 | ¹³C-NMR of AF3-COOH.



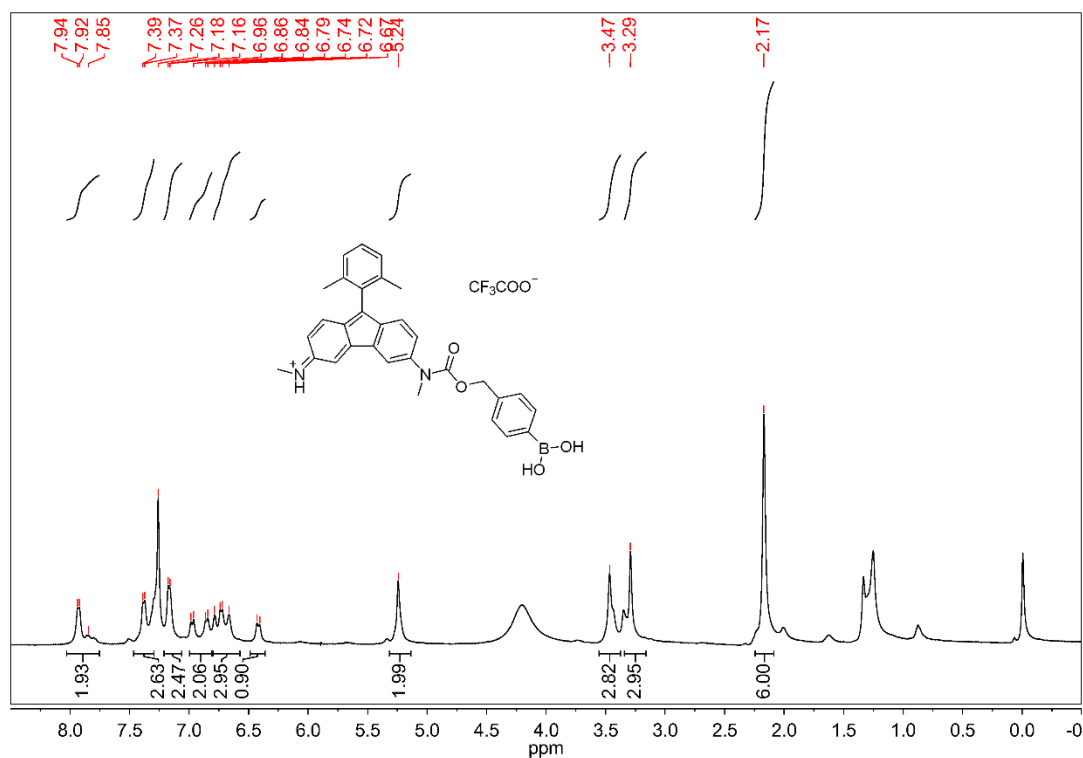
Supplementary Figure 89 | ¹H-NMR of AF2-COOH.



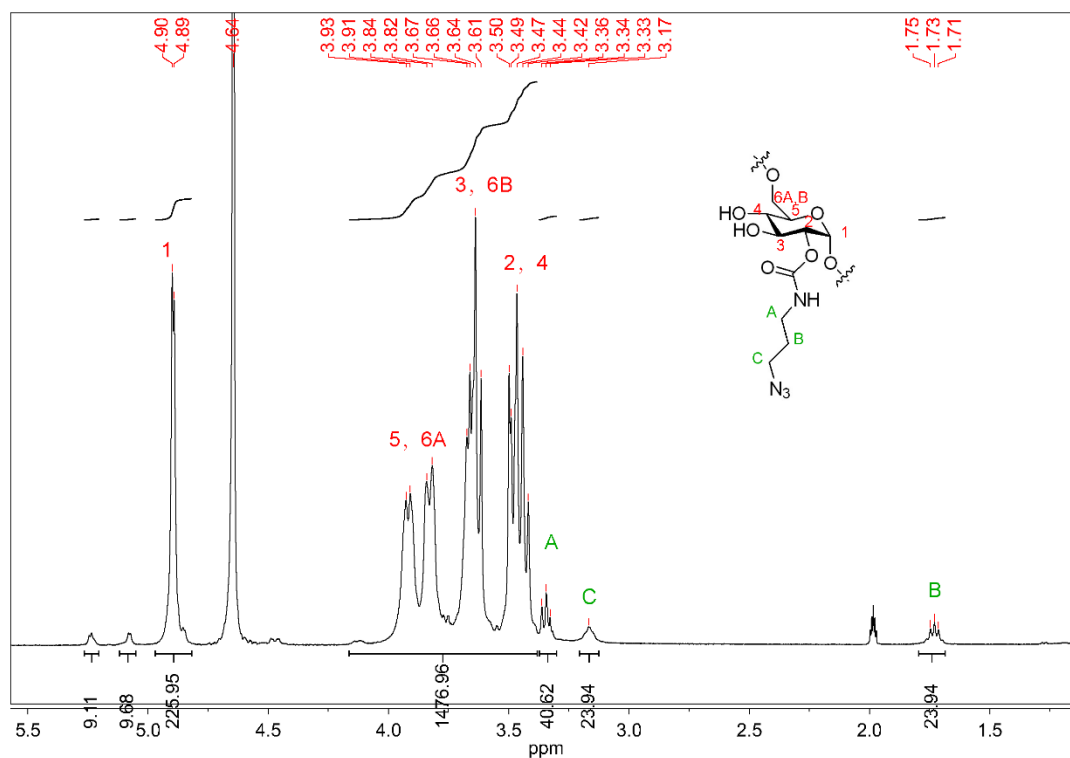
Supplementary Figure 90 | ¹³C-NMR of AF2-COOH.



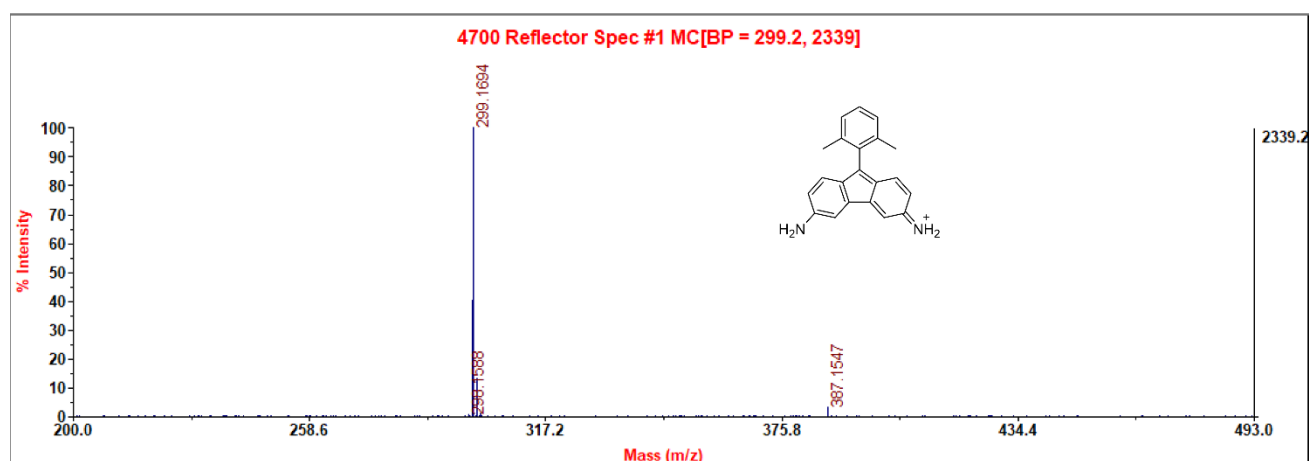
Supplementary Figure 91 | ¹H-NMR of AF2Ac.



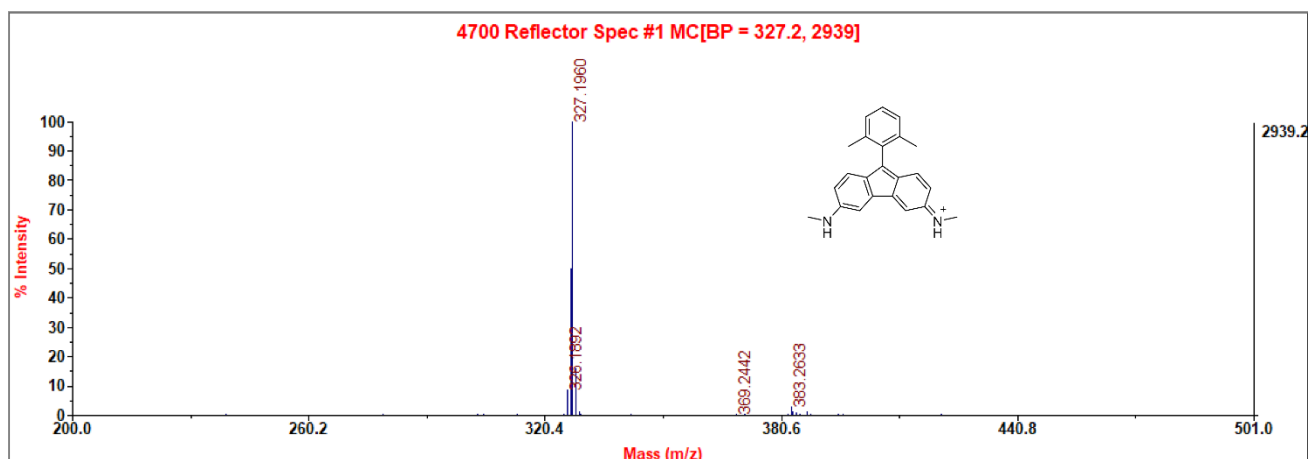
Supplementary Figure 92 | ¹H-NMR of AF2B.



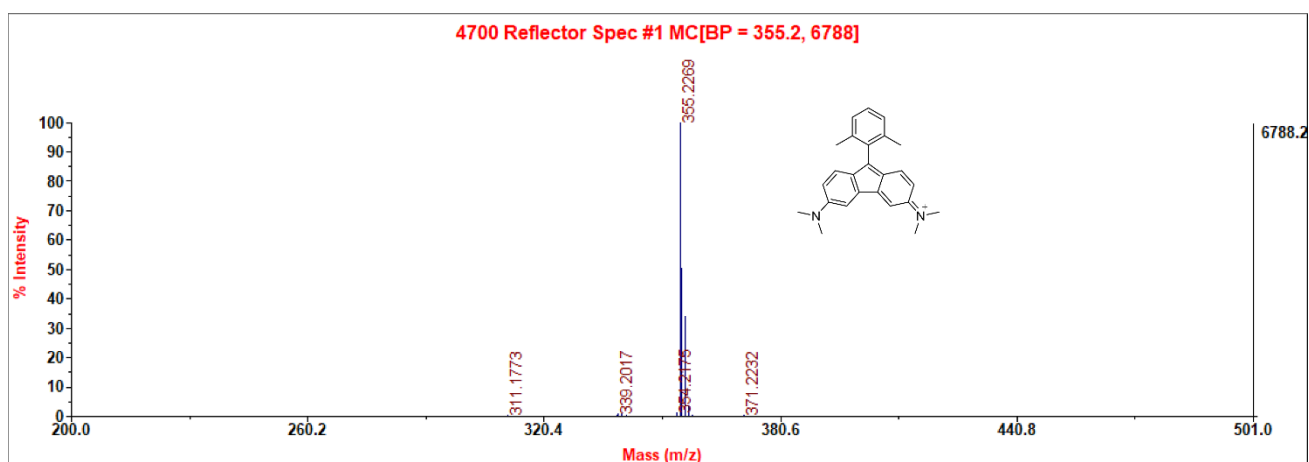
Supplementary Figure 93 | ^1H -NMR of Dextran- N_3 . The total integration of all dextran C-H signals was set to 1726.13 (7 protons $\times$ 246.59 glucose units per 40000 Da dextran). The integral of signal (green B) was used to determine the average number of azidopropyl linker groups per dextran-40000 ($23.94/2=11.97$)



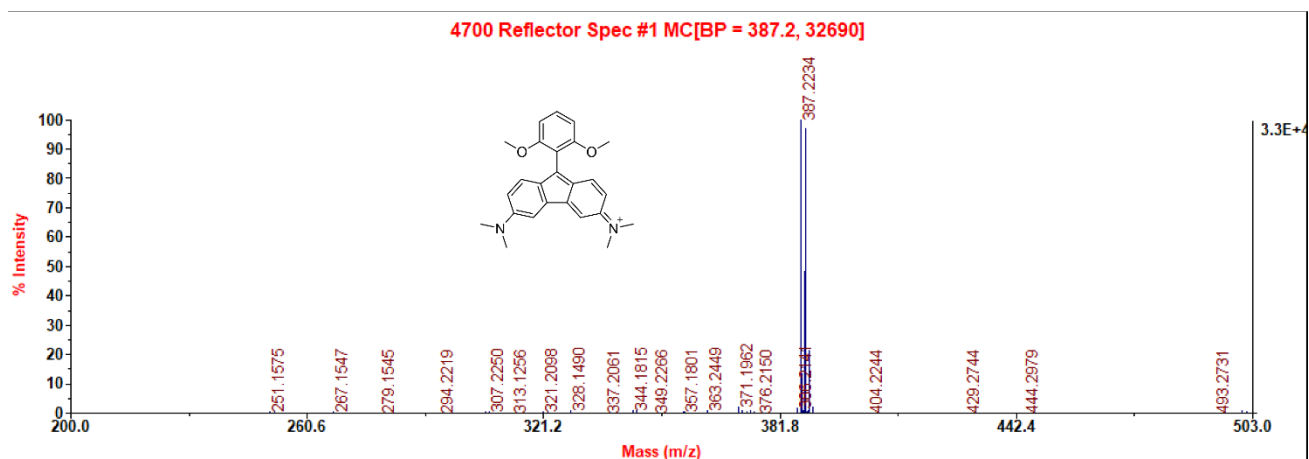
Supplementary Figure 94 | MALDI-TOF MS spectra of AF1.



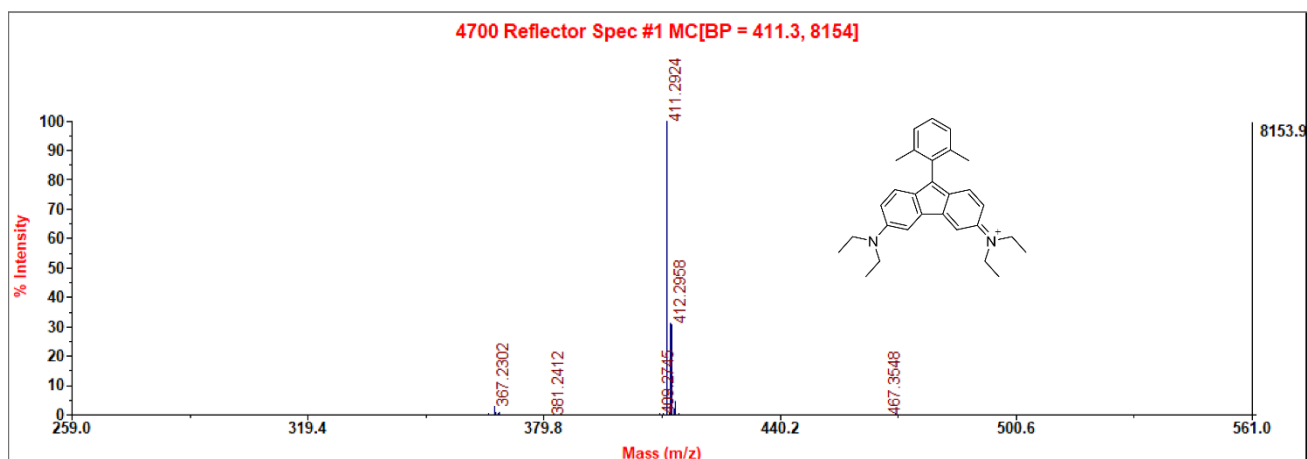
Supplementary Figure 95 | MALDI-TOF MS spectra of AF2.



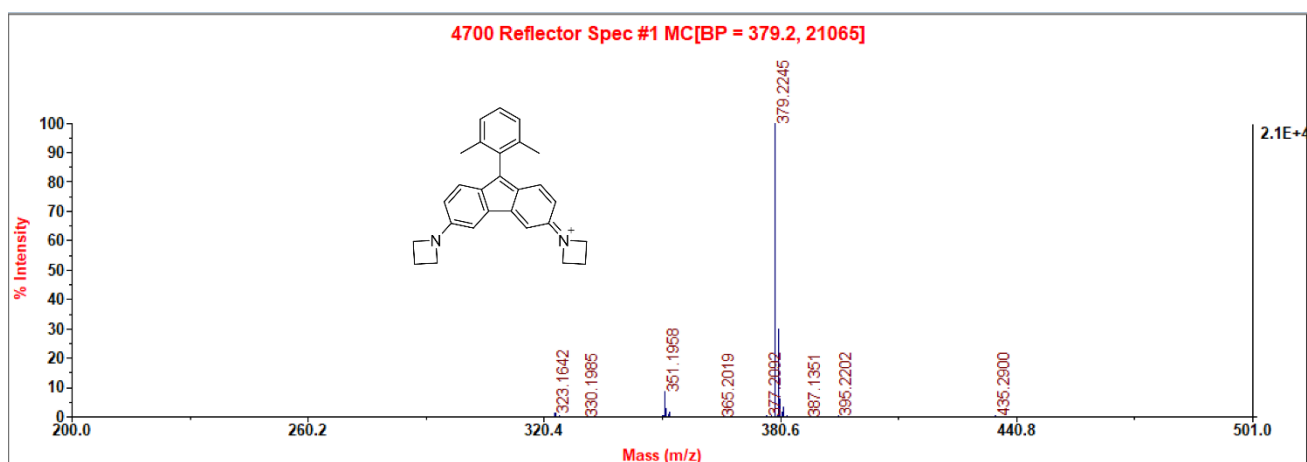
Supplementary Figure 96 | MALDI-TOF MS spectra of AF3.



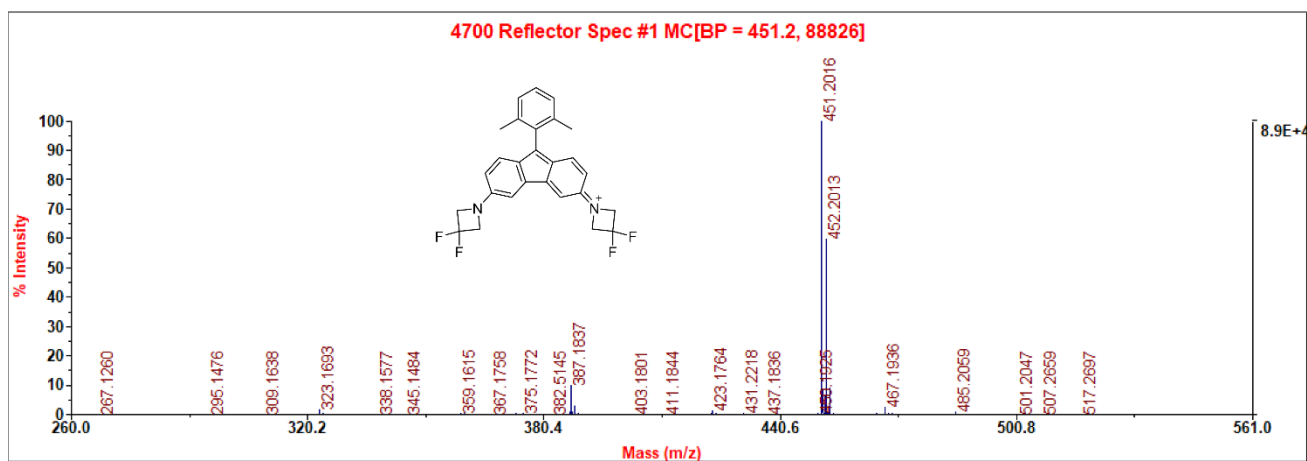
Supplementary Figure 97 | MALDI-TOF MS spectra of AF4.



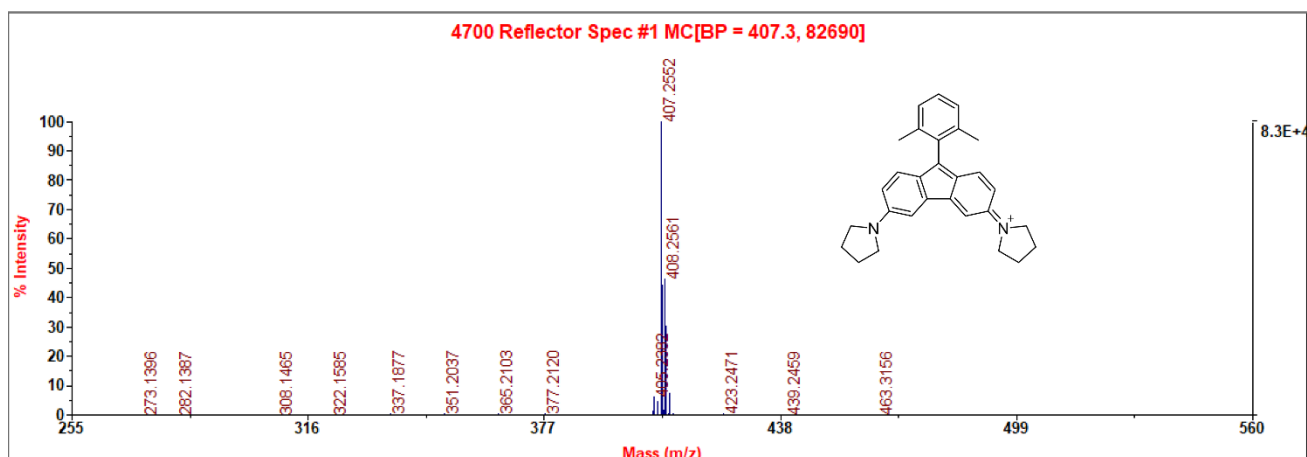
Supplementary Figure 98 | MALDI-TOF MS spectra of AF5.



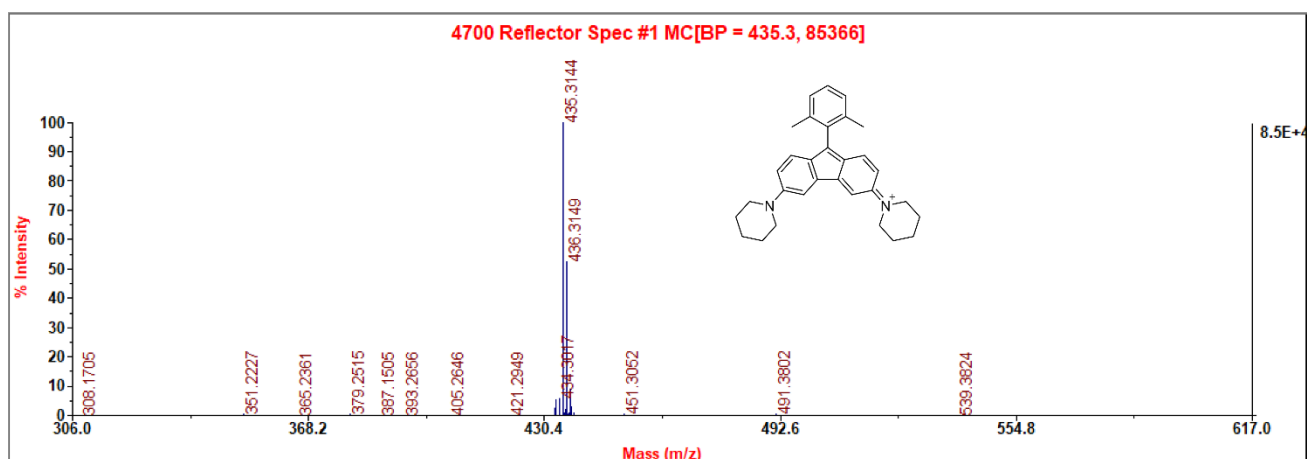
Supplementary Figure 99 | MALDI-TOF MS spectra of AF6.



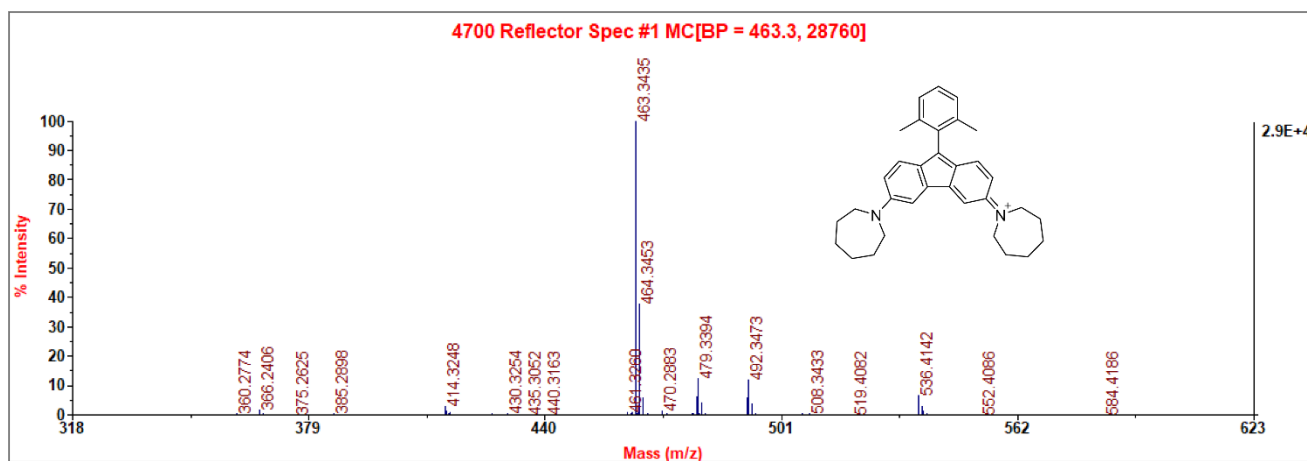
Supplementary Figure 100 | MALDI-TOF MS spectra of AF7.



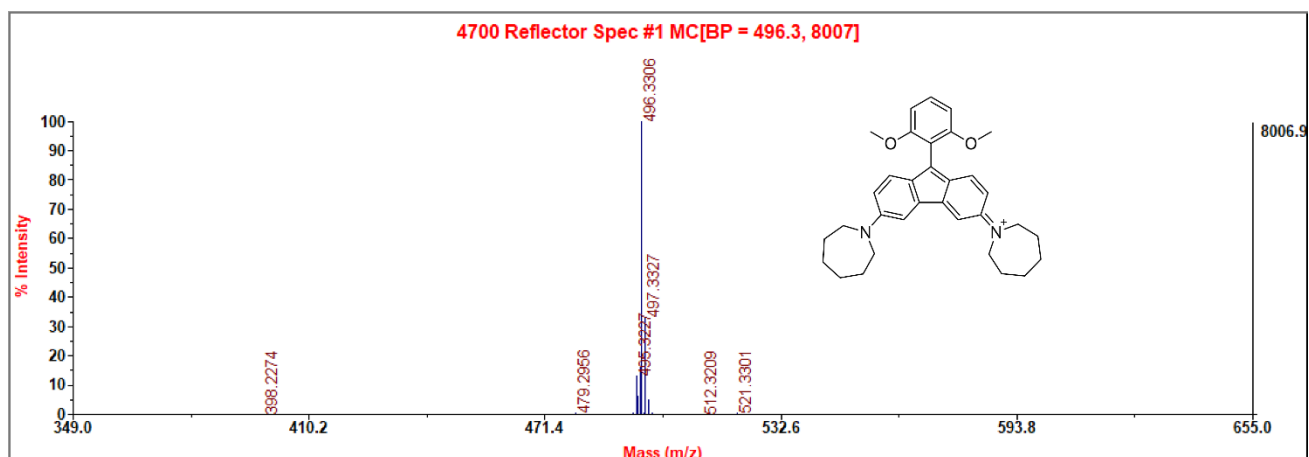
Supplementary Figure 101 | MALDI-TOF MS spectra of AF8.



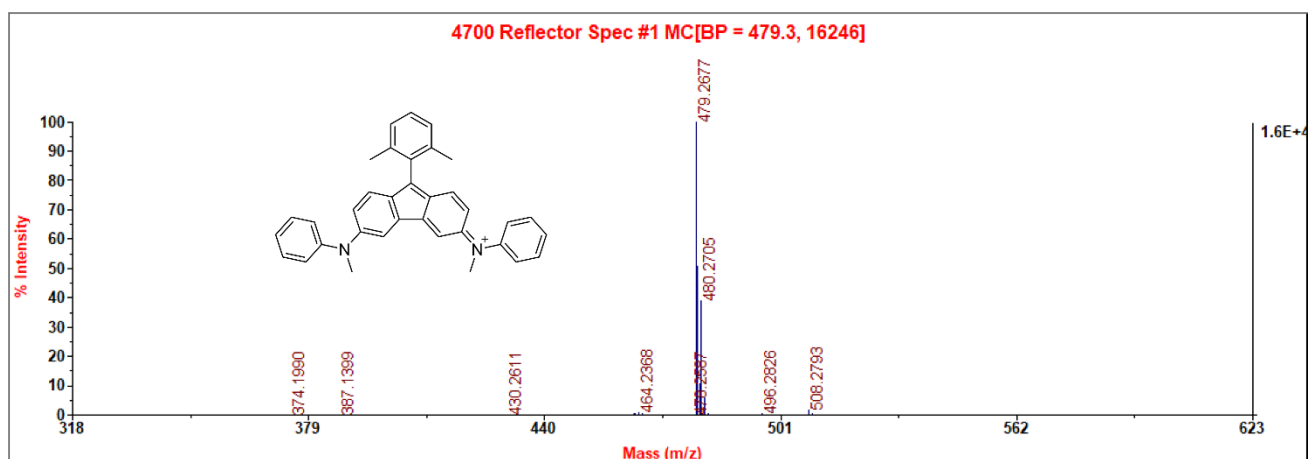
Supplementary Figure 102 | MALDI-TOF MS spectra of AF9.



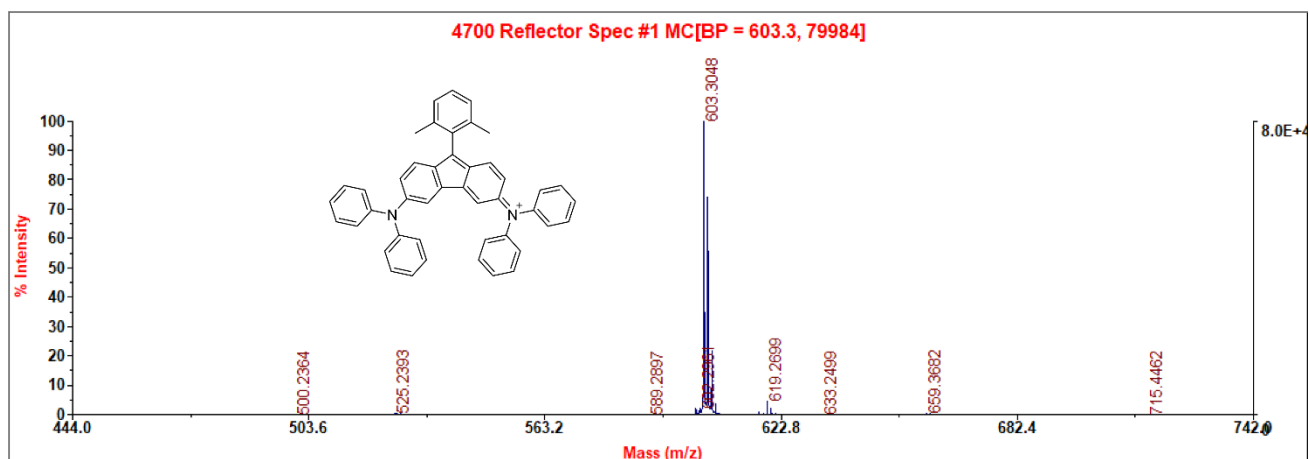
Supplementary Figure 103 | MALDI-TOF MS spectra of AF10.



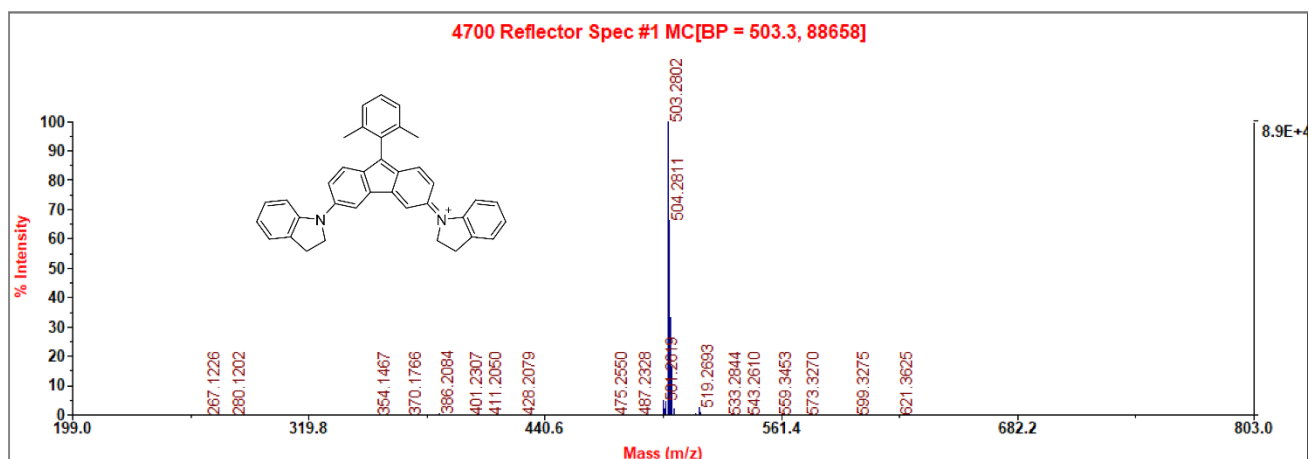
Supplementary Figure 104 | MALDI-TOF MS spectra of AF11.



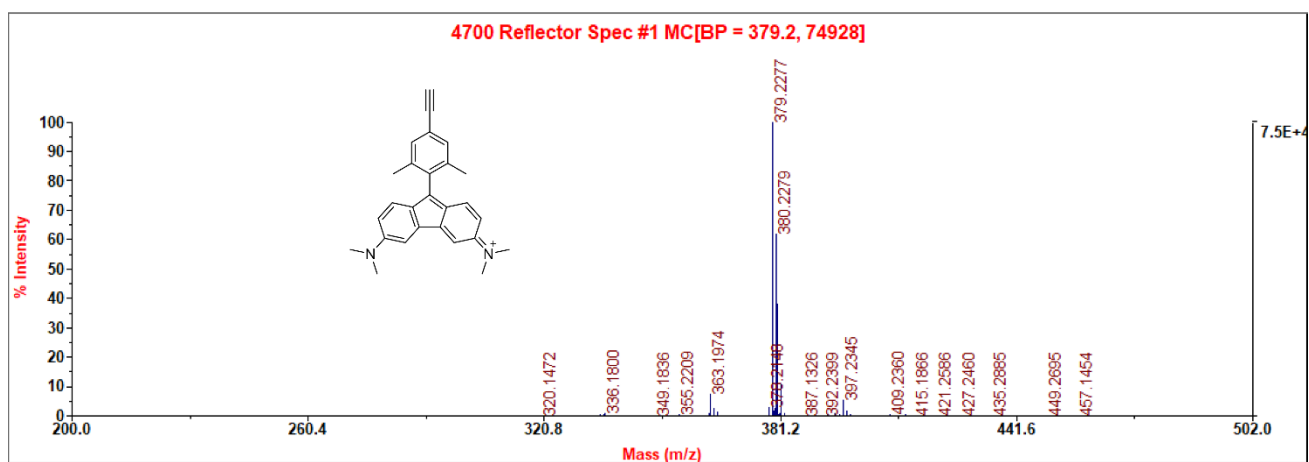
Supplementary Figure 105 | MALDI-TOF MS spectra of AF12.



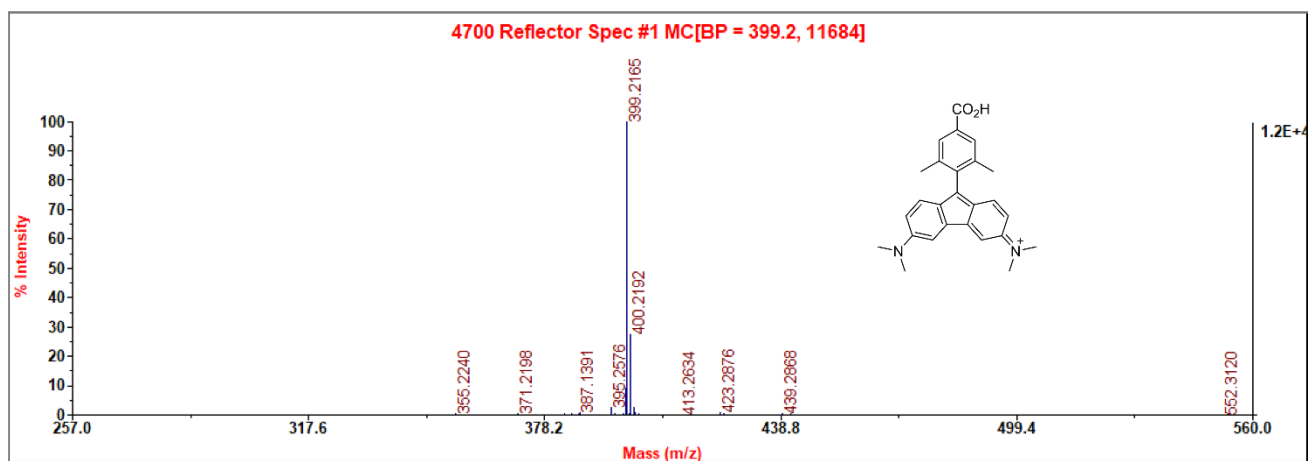
Supplementary Figure 106 | MALDI-TOF MS spectra of AF13.



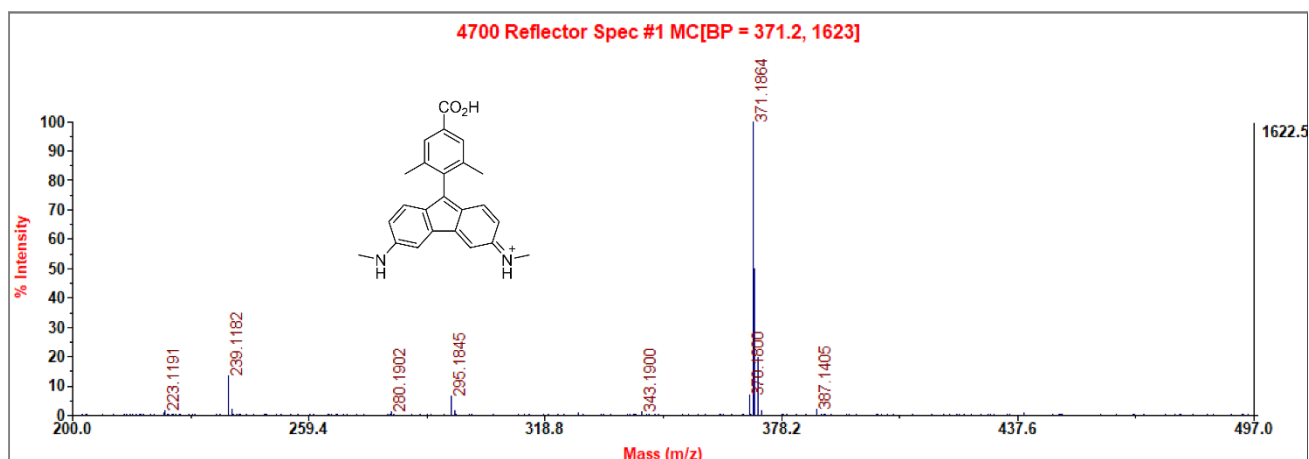
Supplementary Figure 107 | MALDI-TOF MS spectra of AF14.



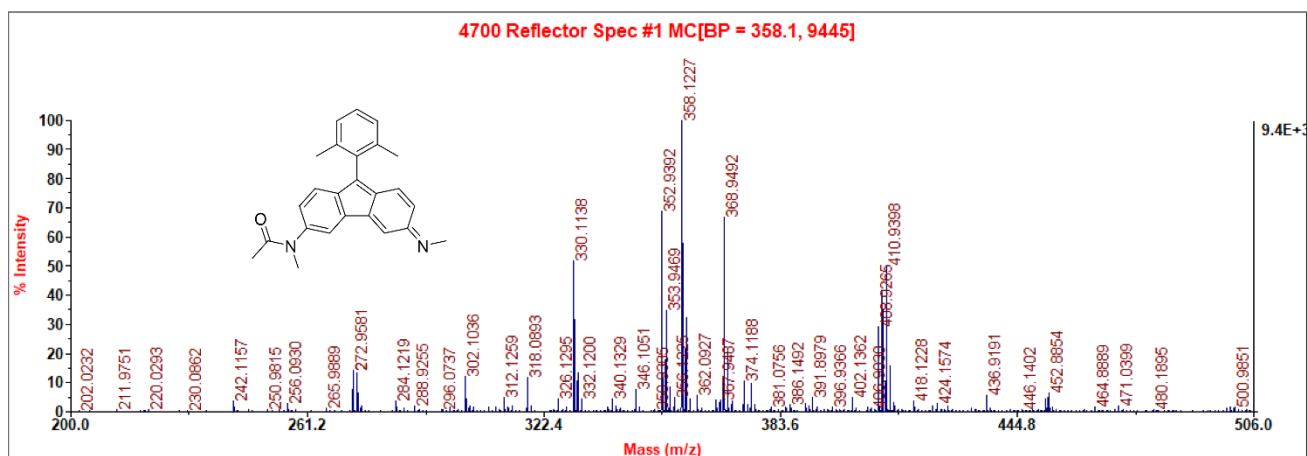
Supplementary Figure 108 | MALDI-TOF MS spectra of AF3-Alkyne.



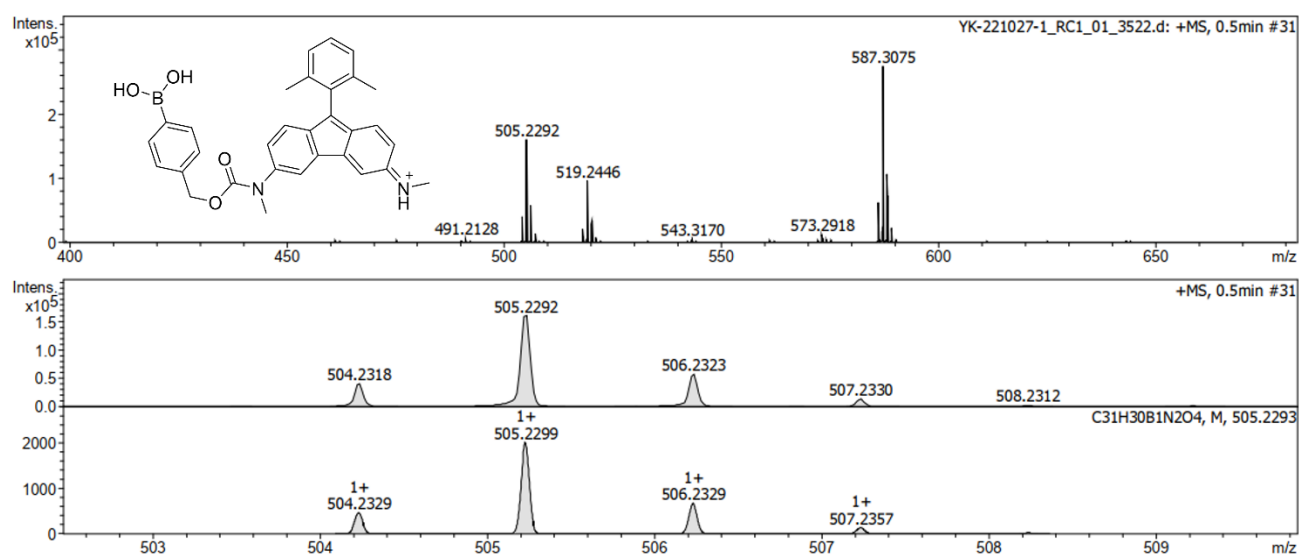
Supplementary Figure 109 | MALDI-TOF MS spectra of AF3-COOH.



Supplementary Figure 110 | MALDI-TOF MS spectra of AF2-COOH.



Supplementary Figure 111 | MALDI-TOF MS spectra of AF2Ac.



Supplementary Figure 112 | HRMS spectra of AF2B.

Supplementary Notes

Supplementary note for Supplementary Figure 22

For MCI,^{45,46} the larger the MCI value, the stronger the aromaticity. Considering the fact that the typical TDDFT underestimates MCI values for S_1 state, the MCI values for T_1 state possessing similar excited state characters vs. S_1 state (Figure S21a) are obtained using unrestricted KS-DFT method. It can be seen that the MCI values are in the order of T_1 -UDFT (0.1126) > S_1 -TD/TDA (0.1051/0.1039) > S_1 - Δ SCF (0.1030) > S_0 (0.1025). In addition, the MCI values for both S_0 and S_1 states are also computed at the CASSCF(2,2)/6-31G(d) level. All the calculated MCI values of T_1 states are larger than that of S_0 state, suggesting more aromatic character of excited state. Last but not least, it should be noted that the MCI is not a good choice to determine the antiaromaticity since the antiaromatic systems typically possessing very small MCI that is indistinguishable from those with weak aromaticity.⁴³

For ELF- π index,⁴⁷ the space enclosed by the higher ELF values makes it easier for electrons to delocalize within this space and simultaneously more difficult to populate outside of this space. The position indicated by the red arrow is called a bifurcation point, on which larger ELF- π values suggest the more aromatic. According to the definition of ELF- π value,⁴⁸ the molecules with ELF- π of 0.17~0.35 correspond to an antiaromatic character, and ELF- π of > 0.70 to aromatic character. Thus, it can be seen that the ELF- π value of 0.18 for the S_0 state of AF3 indicates its antiaromaticity character. And the ELF- π value for the S_1 state is calculated to be 0.47 using the Δ SCF method. Note that usage of ELF- π to analyze the triplet state Baird-aromaticity is proved by Ottosson et al.,⁴⁹ herein the ELF- π value for T_1 state is calculated to be 0.37 using the unrestricted KS-DFT method. Unfortunately, these obtained ELF- π values for excited states are significantly smaller than the reference value of 0.70, suggesting not aromatic characters at all.

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