

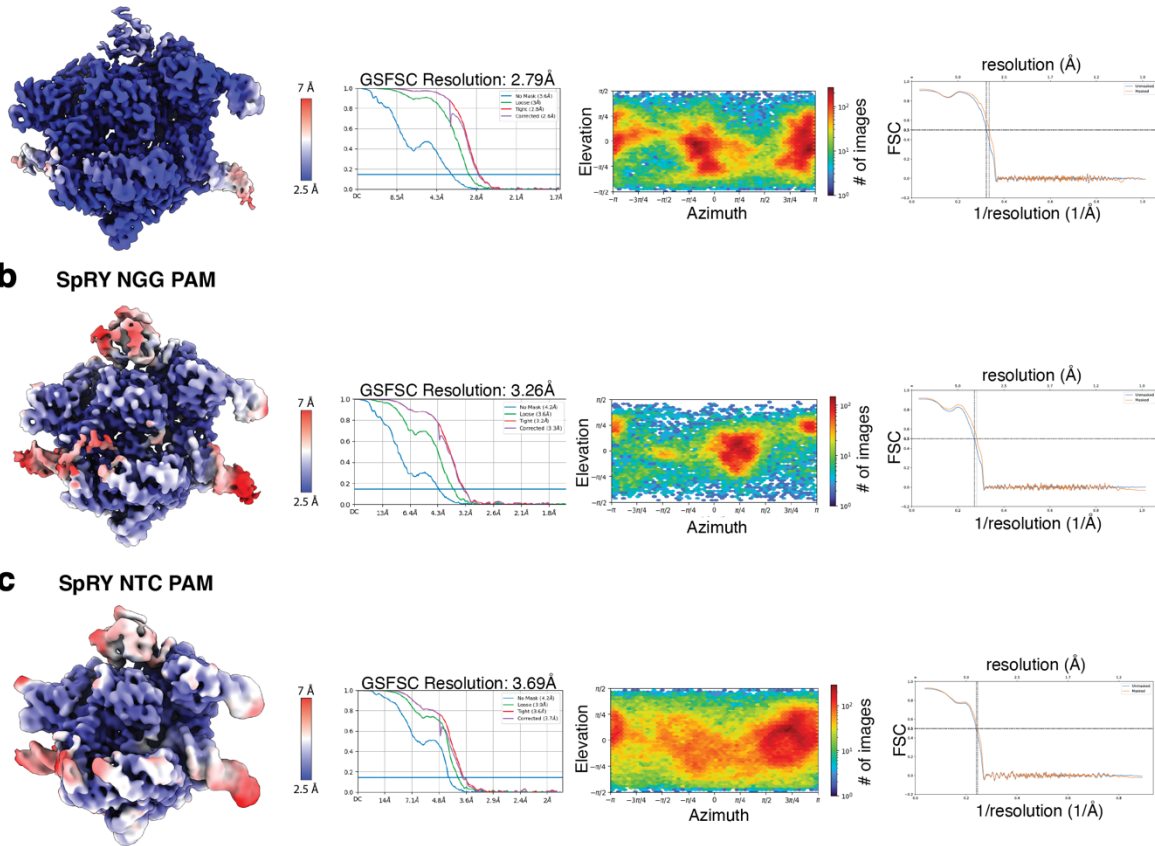
Supplementary Information

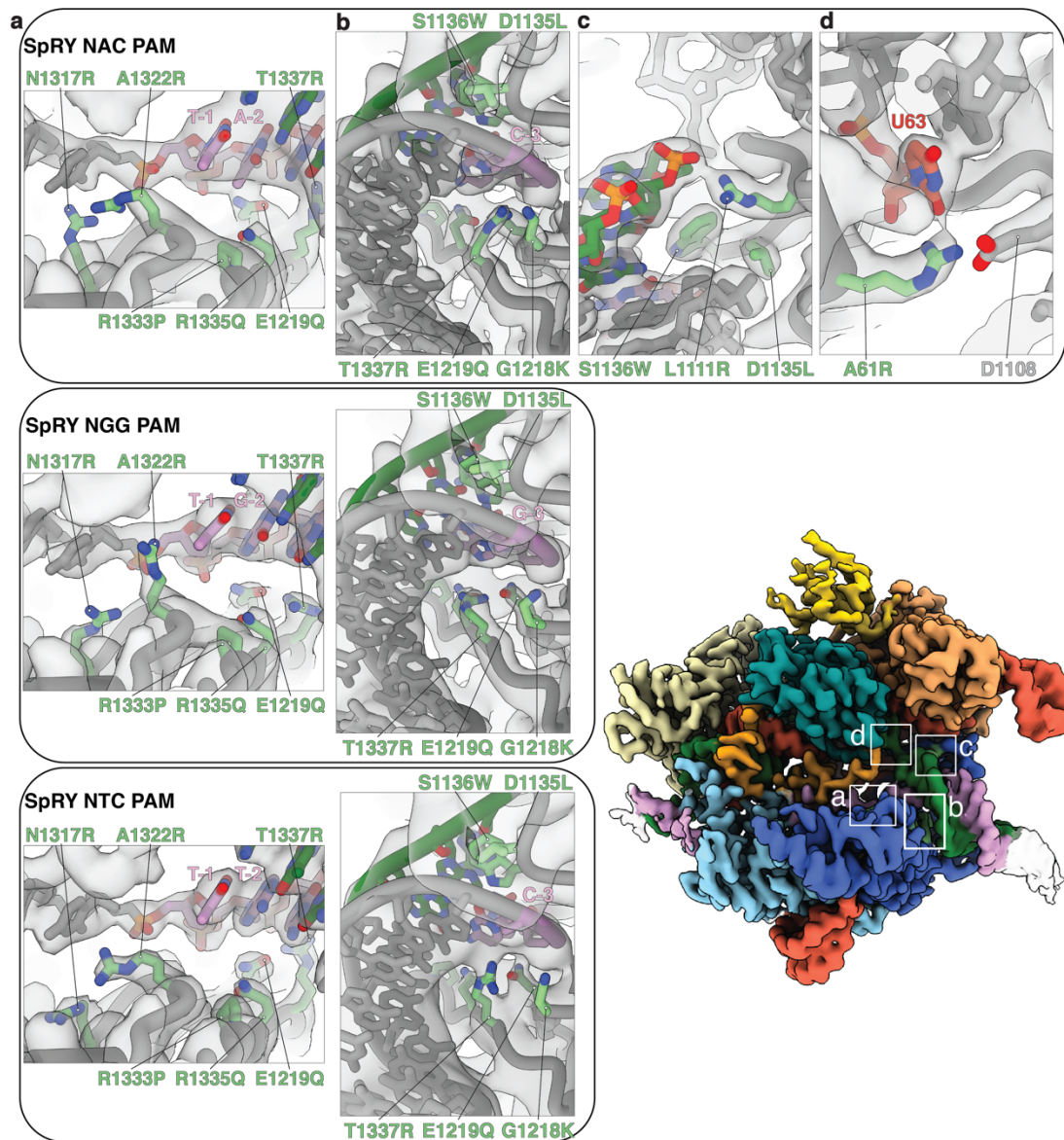
Unraveling the mechanisms of PAMless DNA interrogation by SpRY-Cas9

Grace N. Hibshman, Jack P. K. Bravo, Matthew M. Hooper, Tyler L. Dangerfield, Hongshan Zhang, Ilya J. Finkelstein, Kenneth A. Johnson, David W. Taylor

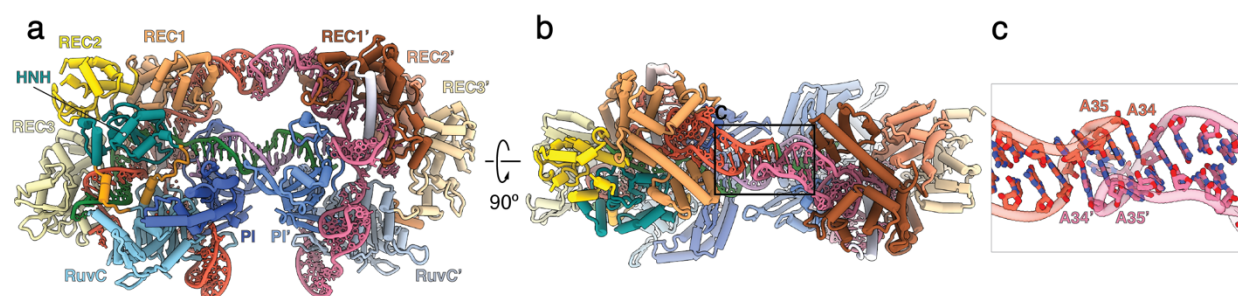
Supplementary Fig. 1. Cryo-EM data analysis of SpRY **a**, NAC PAM DNA **b**, NGG PAM DNA, and **c**, NTC PAM DNA.

C SpRY NTC PAM

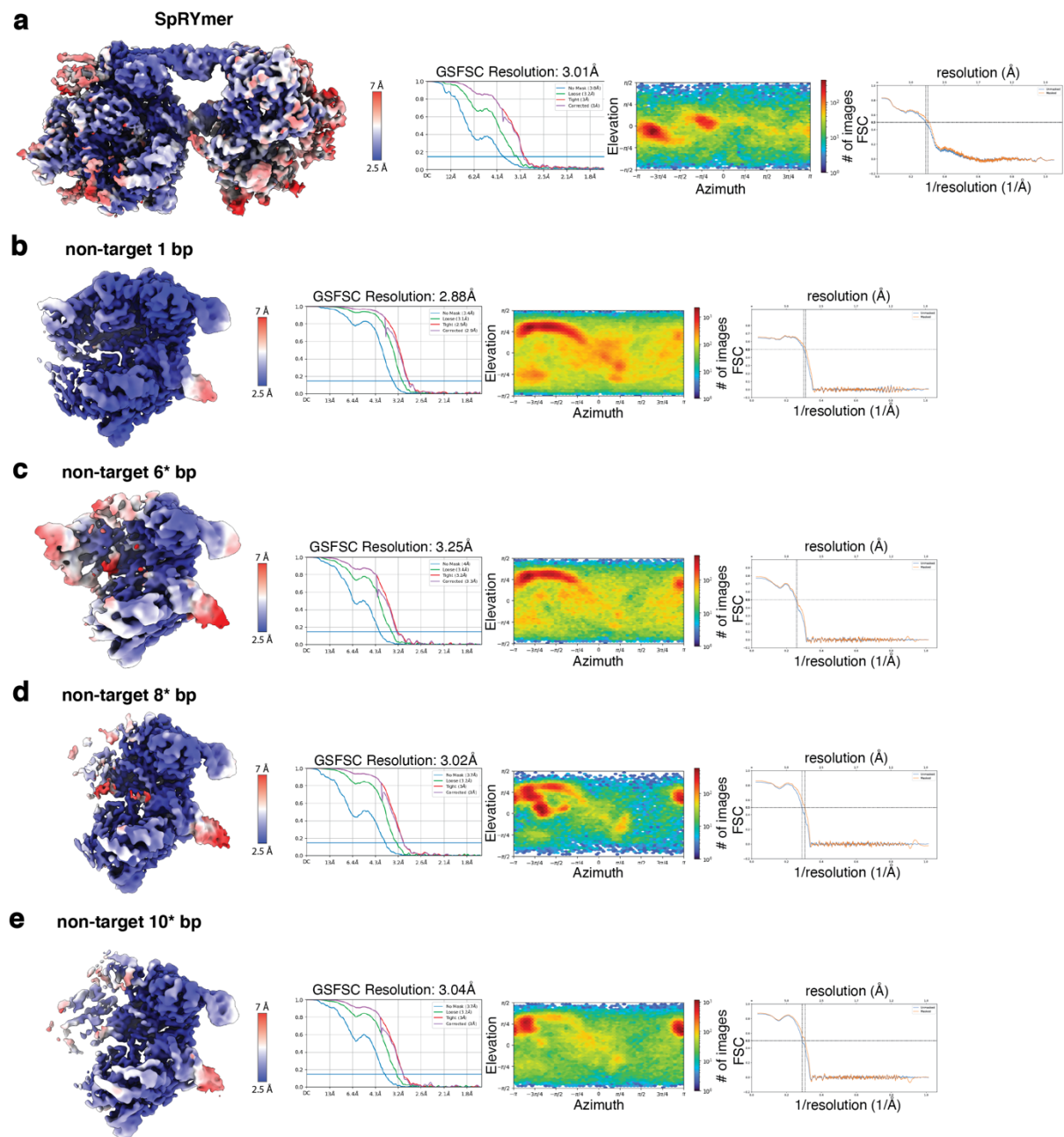




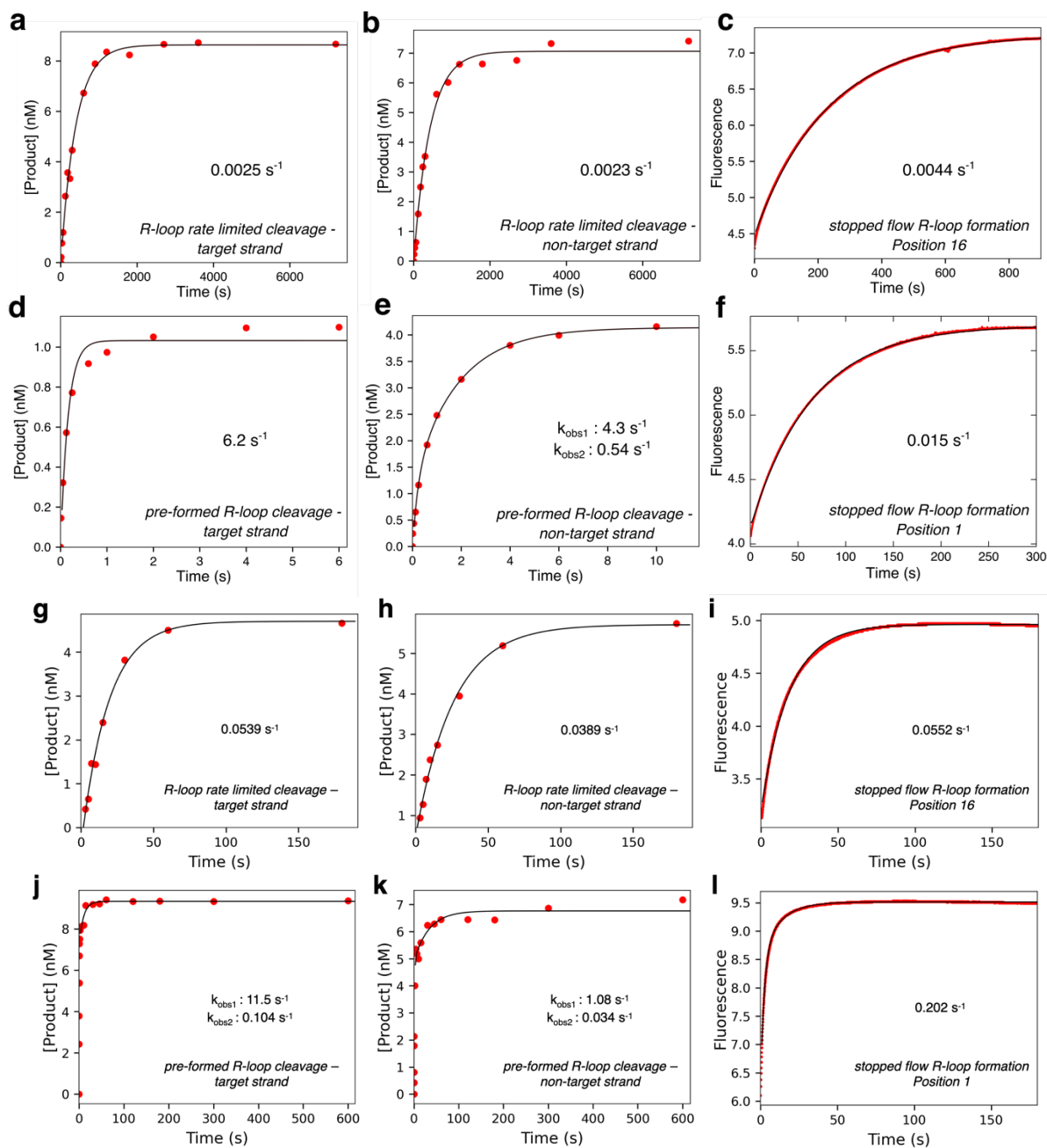
Supplementary Fig. 2. Detailed view of SpRY mutations and the rotamers adopted by different PAM sequences.



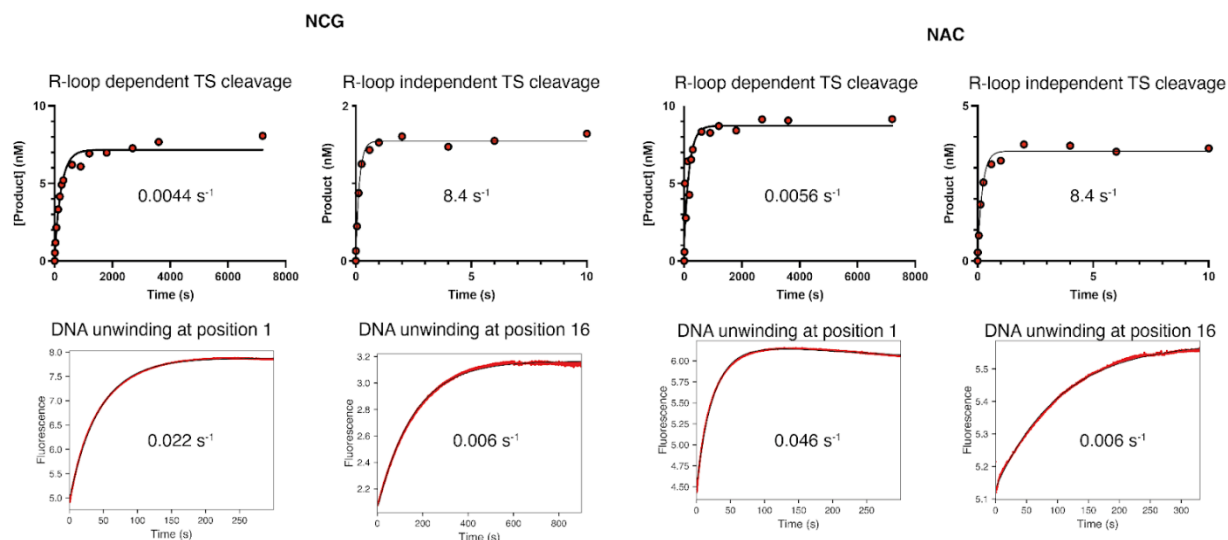
Supplementary Fig. 3. SpRYmer stem loop interaction. **a**, Overview of the SpRYmer atomic model with domains labeled. **b**, Top-down view of the SpRYmer atomic model. **c**, Detailed view of the stem loop kissing interaction.



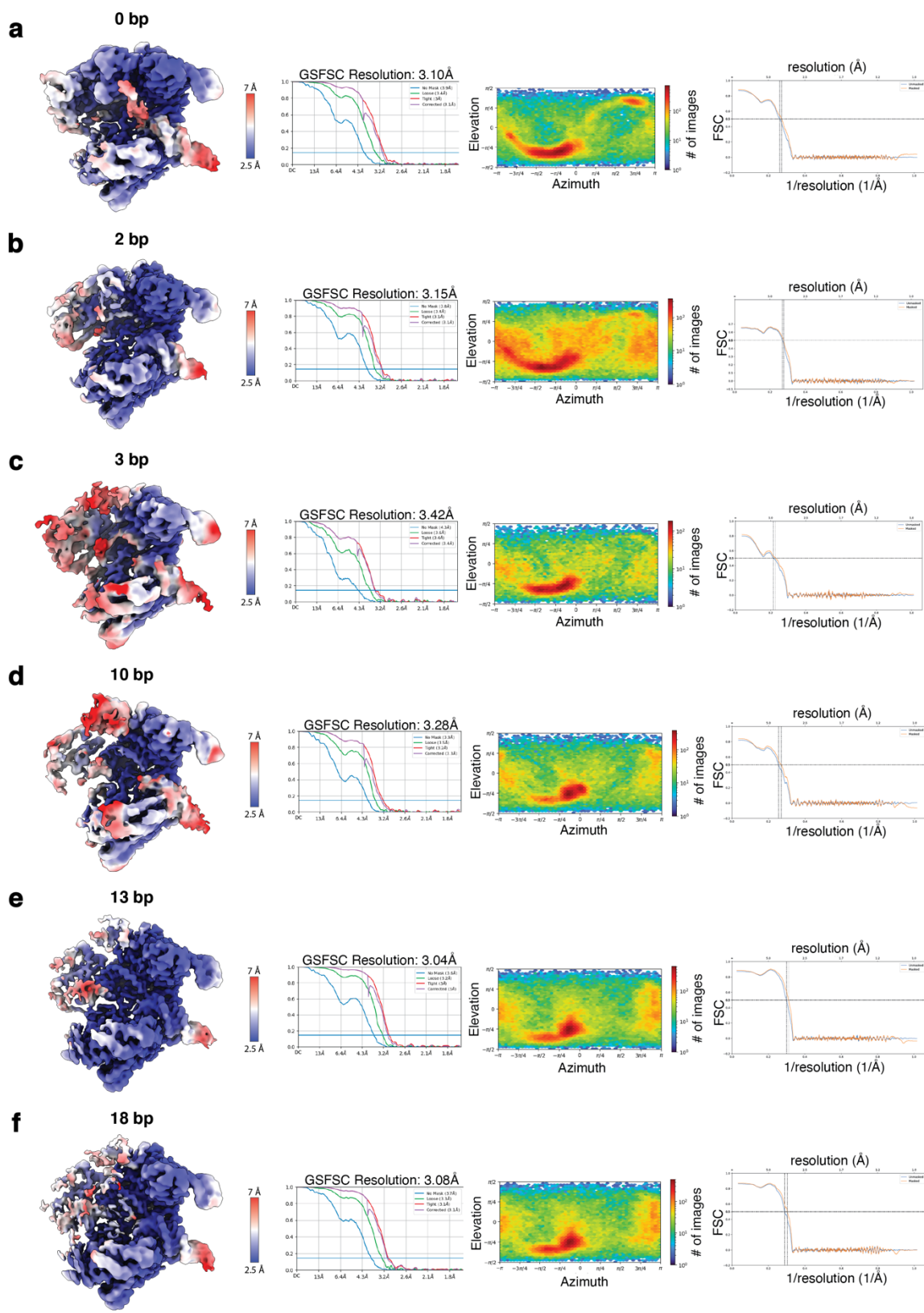
Supplementary Fig. 4. Cryo-EM data analysis of the **a**, SpRYmer **b**, SpRY bound to off-target DNA with 1 bp **c**, 6 bp **d**, 8 bp **d**, and **e**, 10 bp of R-loop formed. *These structures contain mismatches.



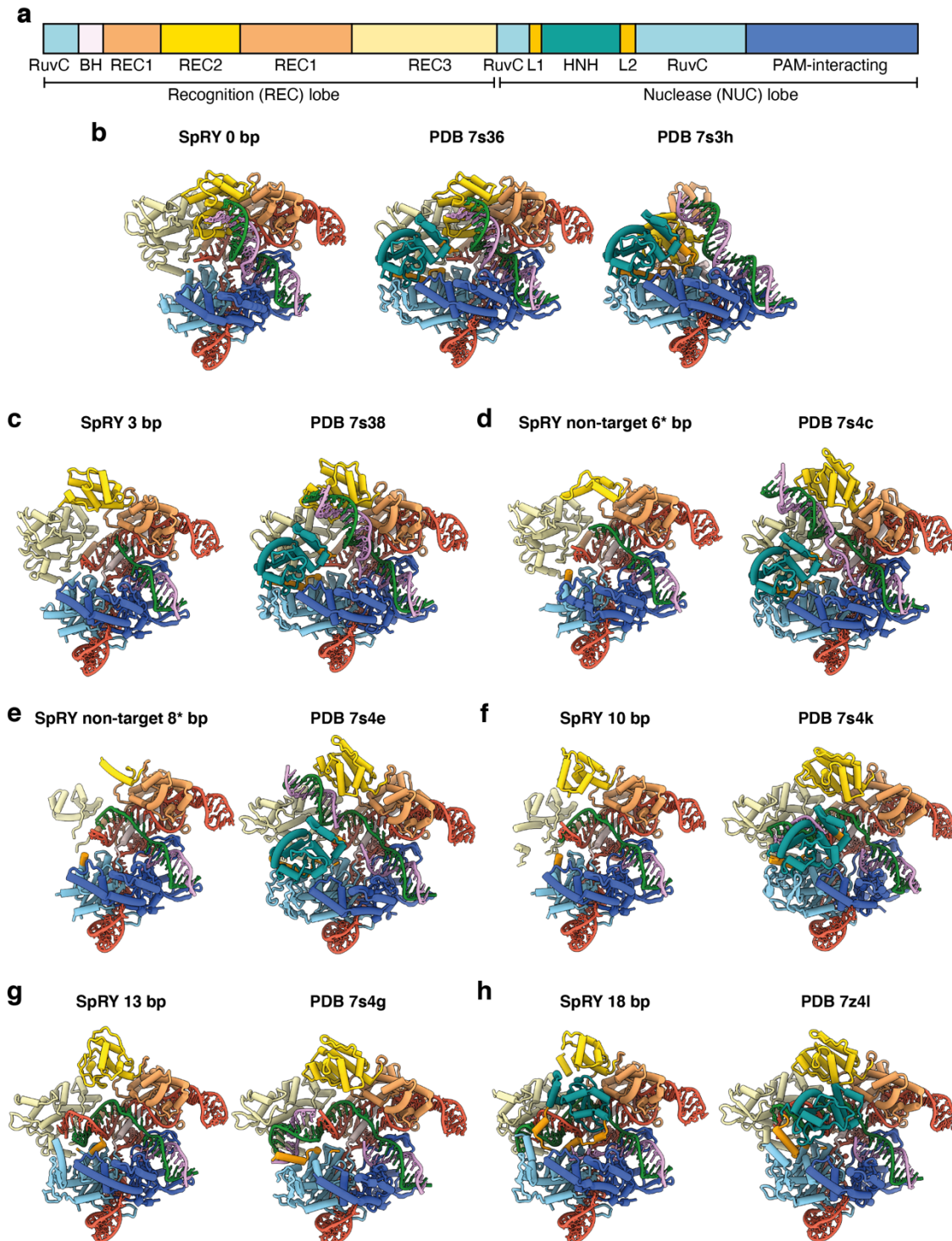
Supplementary Fig. 5. NGG PAM DNA cleavage kinetics for **a-f**, SpRY, and **g-l**, SpG.



Supplementary Fig. 6. DNA cleavage and unwinding kinetics for NGC and NAC PAM substrates. The low amplitude of product for the Mg^{2+} -initiated reactions may be due to association of SpRY at partially complementary, off-target sites, thus reducing the effective enzyme concentration.

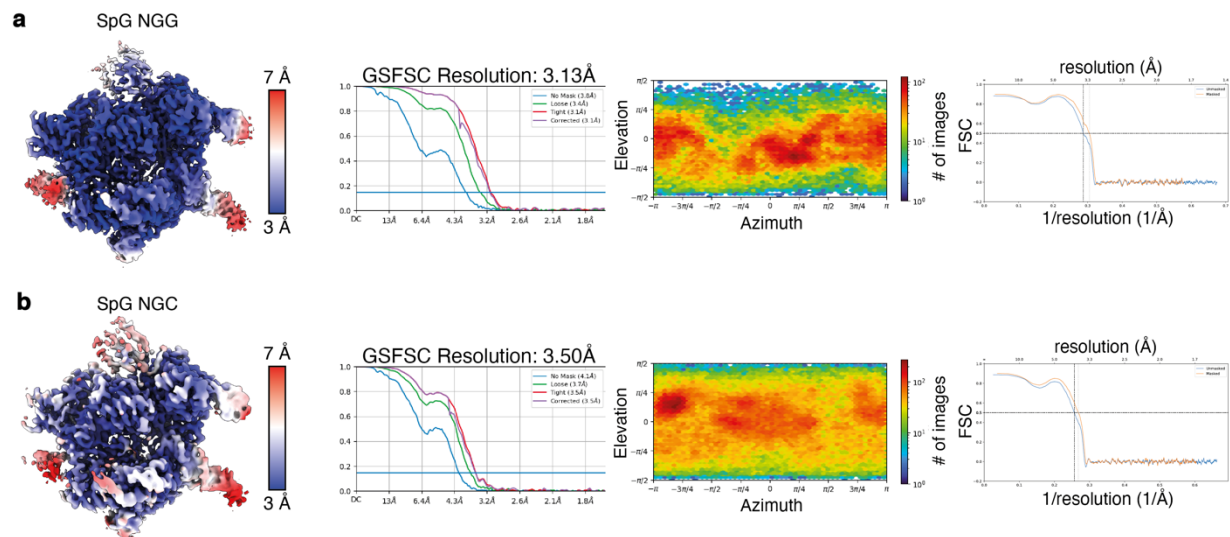


Supplementary Fig. 7. Cryo-EM data analysis of SpRY **a**, 0 bp **b**, 2 bp **c**, 3 bp **d**, 10 bp **e**, 13 bp **f**, 18 bp, and **g**, 20 bp (product state) R-loop intermediates.

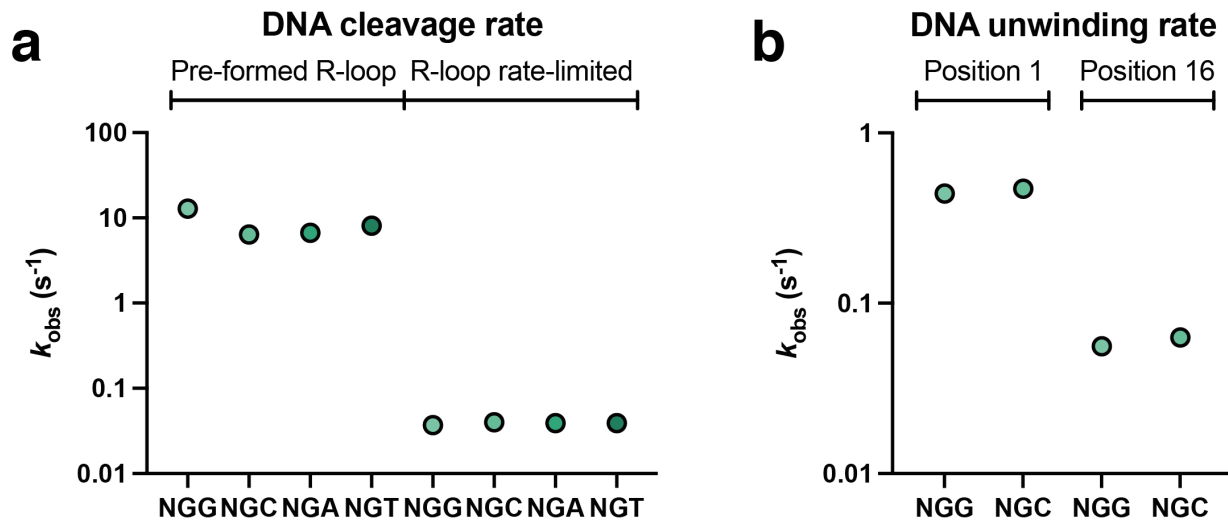


Supplementary Fig. 8. **a**, Comparison of SpRY 0 bp R-loop intermediate with previously determined Cas9 0 bp structures (PDB 7s36, 7s3h). **b**, Comparison of SpRY 3 bp R-loop intermediate with previously determined Cas9 3 bp structure (PDB 7s38). **c**, Comparison of SpRY 6* bp off-target DNA structure with previously determined Cas9 6 bp structure (PDB 7s4c). **d**, Comparison of SpRY 8* bp off-target DNA structure with previously determined Cas9 8 bp

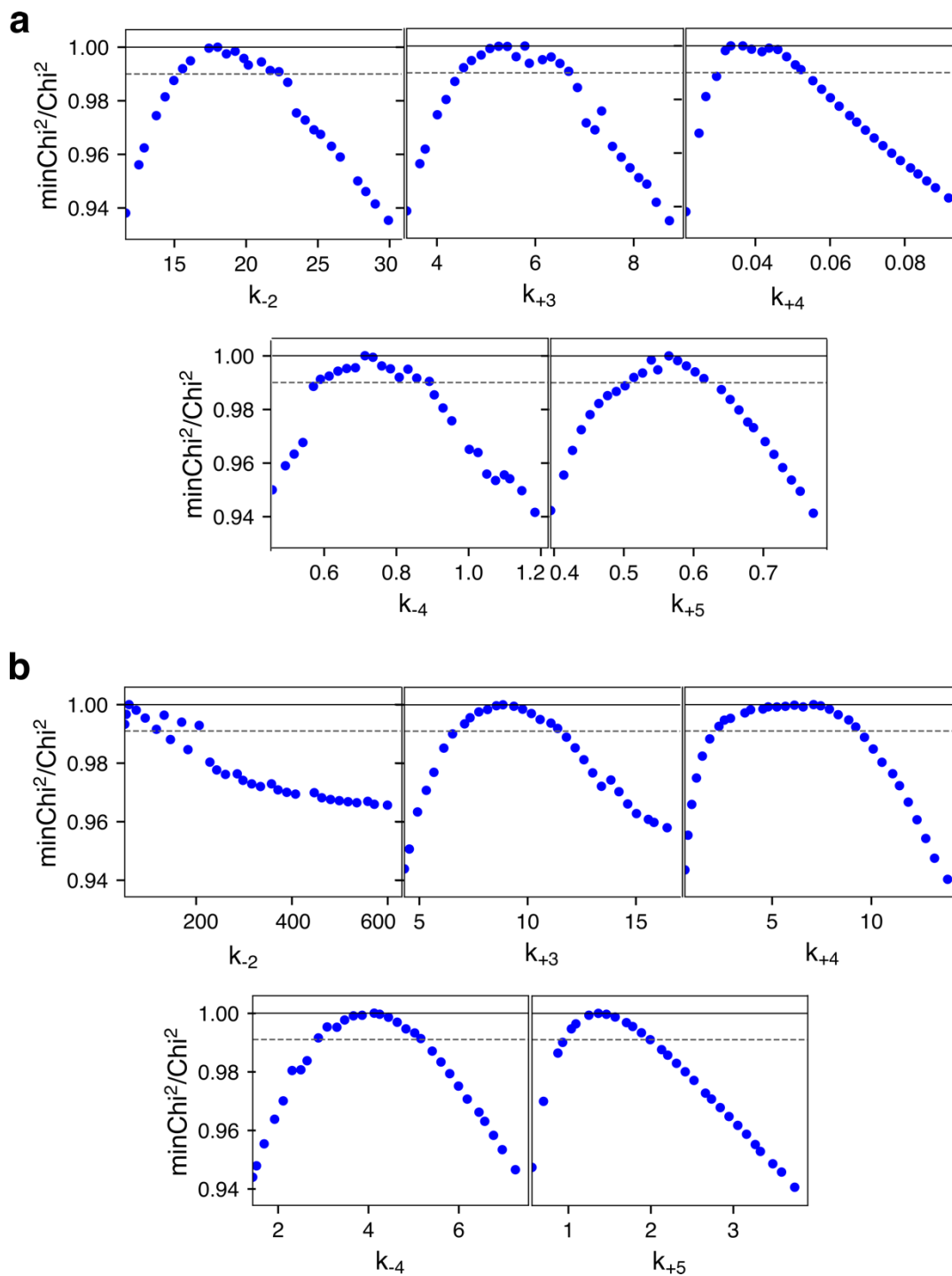
structure (PDB 7s4e). **e**, Comparison of SpRY 10 bp R-loop intermediate with previously determined Cas9 10 bp structure (PDB 7s4k). **f**, Comparison of SpRY 13 bp R-loop intermediate with previously determined Cas9 12 bp structure (PDB 7s4g). **g**, Comparison of SpRY 18 bp R-loop intermediate with previously determined Cas9 18 bp structure (PDB 7z4l).



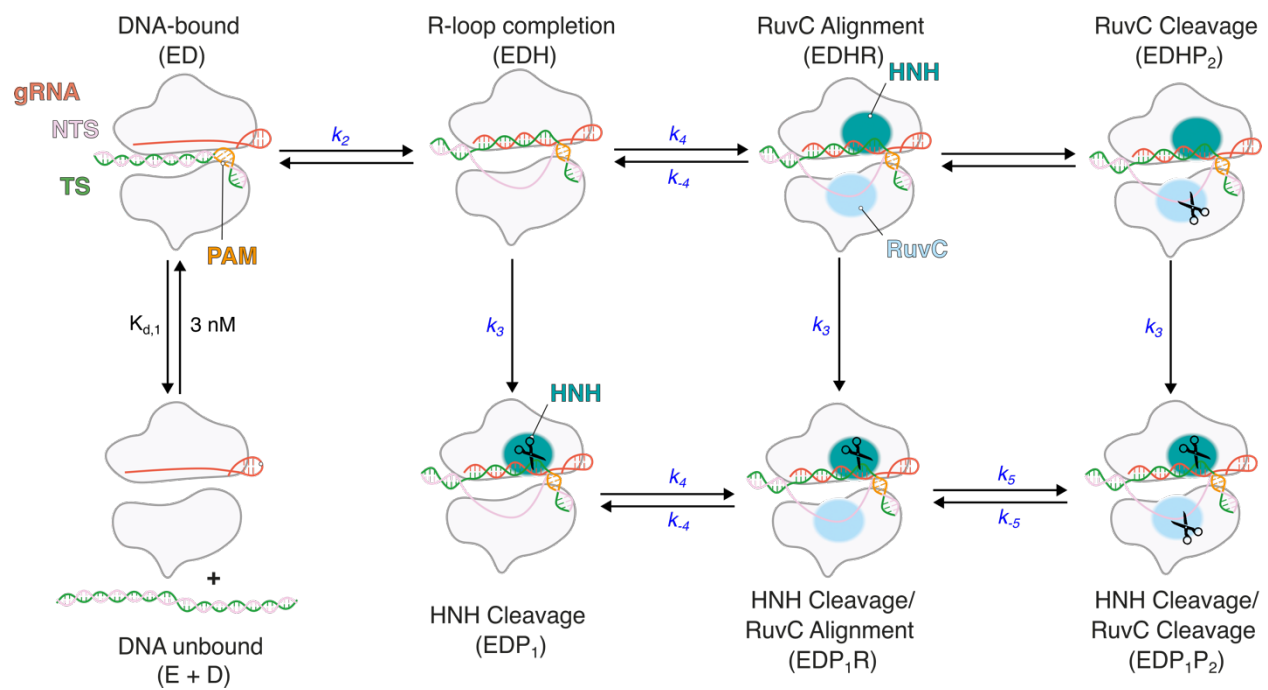
Supplementary Fig. 9. Cryo-EM data analysis of SpG bound to **a**, NGG PAM DNA, and **b**, NGC PAM DNA.



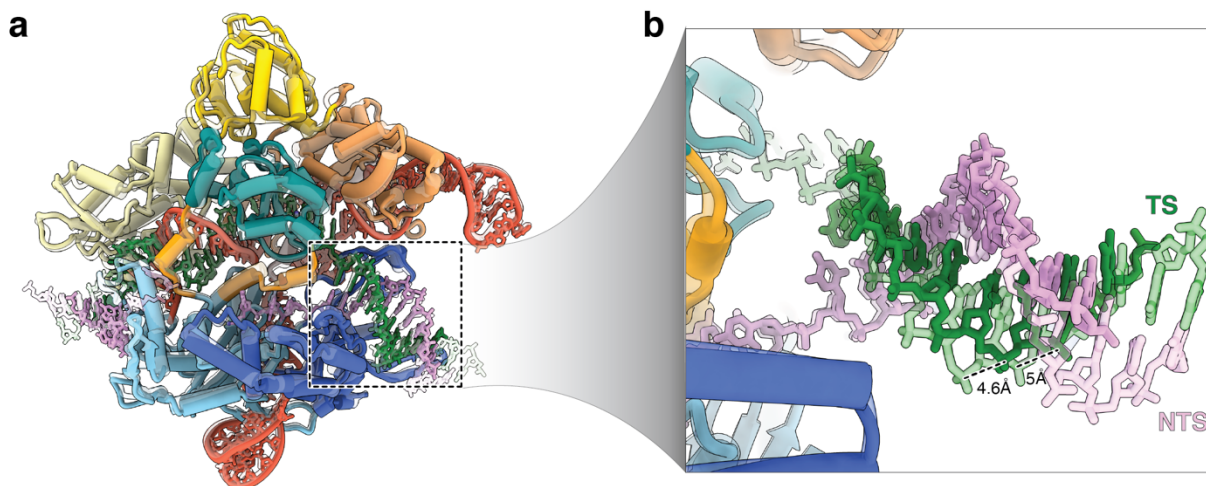
Supplementary Fig. 10. Observed rates for DNA cleavage and DNA unwinding for SpG on DNA substrates with various PAM sequences.



Supplementary Fig. 11. Confidence contours from global data fitting for **a**, SpRY, and **b**, SpG. Contours show the change in χ^2 as a function of values for individual rate. The dashed line indicates the χ^2 threshold corresponding to the 95% confidence interval used for reporting upper and lower limits on parameters listed in **Supplementary Table 3**.



Supplementary Fig. 12. Full kinetic scheme used for global data fitting.



Supplementary Fig. 13. **a**, Overlay of SpRY bound to the NAC PAM DNA substrate and Cas9 (PDB 7s4x). **b**, Close up view of the PAM region of the structures shown in **a**. Cas9 is partially transparent, and SpRY is opaque.

Supplementary Tables

Name	Sequence (5'-3')	Source
TS NGG PAM	/6-FAM/agc tga cgt ttg tac tcc agc gtc tca tct tta tgc gtc agc aga gat ttc tgc t	IDT
NTS NGG PAM	agc aga aat ctc tgc tga cgc ata aag atg aga cgc TGG agt aca aac gtc agc t	IDT
TS NAC PAM	/6-FAM/agctgacgtttgtactGTAgcgtctcatctttatgcgtcagcagagatttctgct	IDT
NTS NAC PAM	agcagaaatctctgctgacgcataaagatgagacgctACagtacaaacgtcagct	IDT
TS NTC PAM	/6-FAM/agctgacgtttgtactGAagcgtctcatctttatgcgtcagcagagatttctgct	IDT
NTS NTC PAM	agcagaaatctctgctgacgcataaagatgagacgctTCagtacaaacgtcagct	IDT
TS NGC PAM	/6-FAM/agctgacgtttgtactGCagcgtctcatctttatgcgtcagcagagatttctgct	IDT
NTS NGC PAM	agcagaaatctctgctgacgcataaagatgagacgctGCagtacaaacgtcagct	IDT
NTS NGG PAM Hot	/6-FAM/agc aga aat ctc tgc tga cgc ata aag atg aga cgc TGG agt aca aac gtc agc t	IDT
Scrambled TS	/56-FAM/agctgacgtttgtactccac*g*c*a*gagtagaaatacgagagcagagatttctgct	IDT
Scrambled NTS	agcagaaatctctgctctgctgatttct*a*c*t*c*t*g*cgtggagtacaaacgtcagct	IDT
TS NGG cold	agc tga cgt ttg tac tcc agc gtc tca tct tta tgc gtc agc aga gat ttc tgc t	IDT
TS NAC cold	agctgacgtttgtactGTAgcgtctcatctttatgcgtcagcagagatttctgct	IDT
TS NGG cold	agctgacgtttgtactGAagcgtctcatctttatgcgtcagcagagatttctgct	IDT
NGG_NTS_tCo_16	agc aga aat ctc tgc tga cg[tCo] ata aag atg aga cgc tgg agt aca aac gtc agc t	Biosynthesis
NGC_NTS_tCo_16	agc aga aat ctc tgc tga cg[tCo] ata aag atg aga cgc tgc agt aca aac gtc agc t	Biosynthesis
NAC_NTS_tCo_16	agc aga aat ctc tgc tga cg[tCo] ata aag atg aga cgc tac agt aca aac gtc agc t	Biosynthesis
NGG_NTS_tCo_1	agc aga aat ctc tgc tga cgc ata aag atg aga cg[tCo] tgg agt aca aac gtc agc t	Biosynthesis
NGC_NTS_tCo_1	agc aga aat ctc tgc tga cgc ata aag atg aga cg[tCo] tgc agt aca aac gtc agc t	Biosynthesis
NAC_NTS_tCo_1	agc aga aat ctc tgc tga cgc ata aag atg aga cg[tCo] tac agt aca aac gtc agc t	Biosynthesis

Supplementary Table 1. List of DNA substrates used in this study. *Corresponds to phosphorothioate-modified DNA.

Supplementary Table 2. Cryo-EM data processing and modeling.

Parameter	Best fit		
	SpRY	SpG	Cas9
k_1	$100 \mu\text{M s}^{-1} *$	$100 \mu\text{M s}^{-1} *$	$100 \mu\text{M}^{-1}\text{s}^{-1}*$
k_{-1}	$0.3 \text{ s}^{-1} *$	$0.3 \text{ s}^{-1} *$	$0.3 \text{ s}^{-1} *$
k_2	$[> 0.02 \text{ s}^{-1}]$	$[< 0.3 \text{ s}^{-1}]$	2.5
k_{-2}	$16.1 \text{ s}^{-1} (12.9, 20.1)$	$59.9 \text{ s}^{-1} (44.2, 206)$	1.2 s^{-1}
k_3	$5.36 \text{ s}^{-1} (4.29, 6.72)$	$8.9 \text{ s}^{-1} (7.1, 11.4)$	5.6 s^{-1}
k_4	$0.038 \text{ s}^{-1} (0.024, 0.048)$	$7.1 \text{ s}^{-1} (2.3, 53.8)$	1.8 s^{-1}
k_{-4}	$0.65 \text{ s}^{-1} (0.51, 0.77)$	$4.1 \text{ s}^{-1} (2.8, 5.2)$	2.3 s^{-1}
k_5	$0.55 \text{ s}^{-1} (0.48, 0.63)$	$1.36 \text{ s}^{-1} (0.86, 2.0)$	4.4 s^{-1}
k_{-5}	$\{ 0.0004 \text{ s}^{-1} \}$	$\{ 0.0004 \text{ s}^{-1} \}$	0.0004 s^{-1}

Supplementary Table 3. Rate constants derived from fitting the data in **Fig. 4c-g** and **Fig 6e-i** to the model shown in **Supplementary Figure 12** are summarized. Values for SpRY and SpG in parentheses are lower and upper limits based on a 95% confidence interval from the confidence contour analysis. For SpRY and SpG, since k_2 was not well defined, it was locked at a lower limit of 0.02 s^{-1} and an upper limit of 0.3 s^{-1} for SpRY and SpG, respectively, for confidence contour calculations (locked rate constants are shown in brackets). Rates for Cas9 are taken from (26). The * indicates that rates for DNA binding and dissociation were not defined by the data but were locked at these rate constants to give a K_d of 3 nM. Values for k_{-5} in curly brackets were set at small values that did not change the results of the fitting to satisfy thermodynamic constraints and to allow generation of a free energy profile (20).