

Caspase-1-dependent spatiality in triple-negative breast cancer and response to immunotherapy

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Supplementary information

Supplementary Figure 1-8

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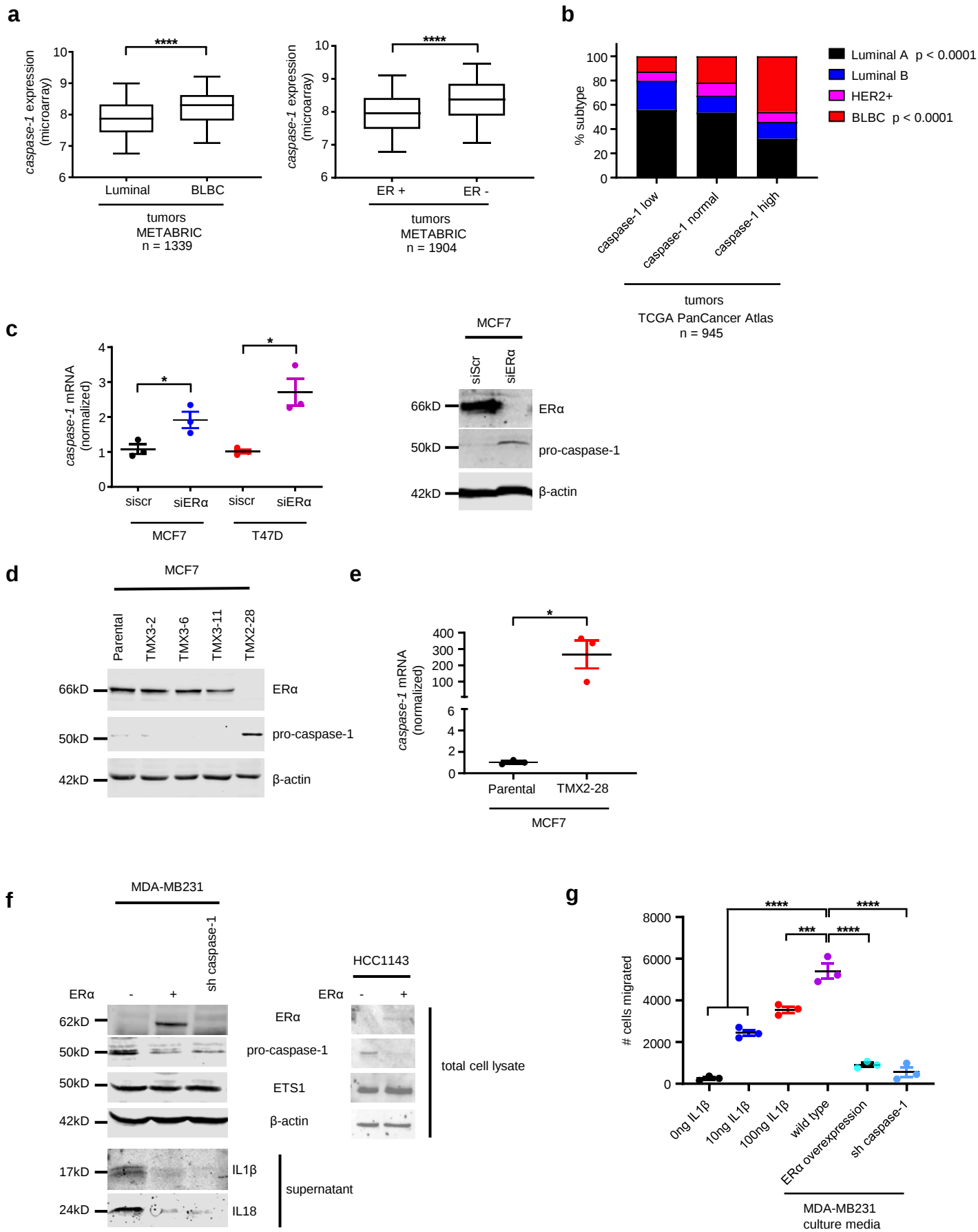


Fig. S1. Caspase-1 expression is ER α -dependant. **a** mRNA expression of caspase-1 in human breast tumor samples (METABRIC dataset^{1,2}) grouped either by subtype (1140 luminal and 199 BLBC samples) or by ER α expression (1459 ER+ and 445 ER- samples) (mean with 95% CIs, two-tailed, unpaired t-test with equal variances **** : $p < 0.0001$). **b** Proportion of breast cancer subtypes in caspase-1 low, normal, and high subgroups of human breast cancer (TCGA PanCancer Atlas dataset^{1,2}) divided by a cutoff z-score of +/-1. P-values represent the significance level of a positive association between caspase-1 high tumors and the basal-like subtype, and a negative association between caspase-1 high tumors and the luminal A subtype (Fisher's exact test). **c** Caspase-1 mRNA level in ER positive MCF7 and T47D cell lines with siRNA transient KD of ER α , quantified by real time quantitative polymerase chain reaction (RTqPCR) ($n = 3$ biologically independent experiments, mean with SEM, two-tailed, unpaired t-test with equal variances, MCF7 siscr vs siER α : * : $p = 0.0367$; T47D siscr vs siER α : * : $p = 0.0125$) and associated immunoblot of pro-caspase-1 protein in the MCF7 cells . β -actin is included as a loading control. **d** Immunoblot of ER α and pro-caspase-1 in parental MCF7 and several tamoxifen-resistant MCF7 cell lines. **e** Caspase-1 mRNA levels in tamoxifen-resistant TMX2-28 cells compared to parental MCF7 ($n = 3$ independent biologically replicates, mean with SEM, two-tailed, unpaired t-test with equal variances, * : $p = 0.0351$). **f** Immunoblot of ER α , pro-caspase-1, and ETS1 from wild type MDA-MB231 or HCC1143 TNBC cells, cells overexpressing ER α , and cells with shRNA KD of caspase-1 and associated immunoblot of IL1 β and IL18 in MDA-MB231 cell culture supernatant. **g** Transwell migration assay of THP-1 macrophages after the addition of increasing concentrations of recombinant IL1 β , or conditioned media from wild type MDA-MB231 cells, cells overexpressing ER α , and cells with shRNA KD of caspase-1 ($n = 3$ biologically independent experiments, mean with SEM, one-way ANOVA with Dunnett's multiple comparisons test, *** : $p = 0.0003$, **** : $p < 0.0001$). $n = 3$ biologically independent experiments for immunoblots and representative images are shown. Source data are provided as a Source Data file.

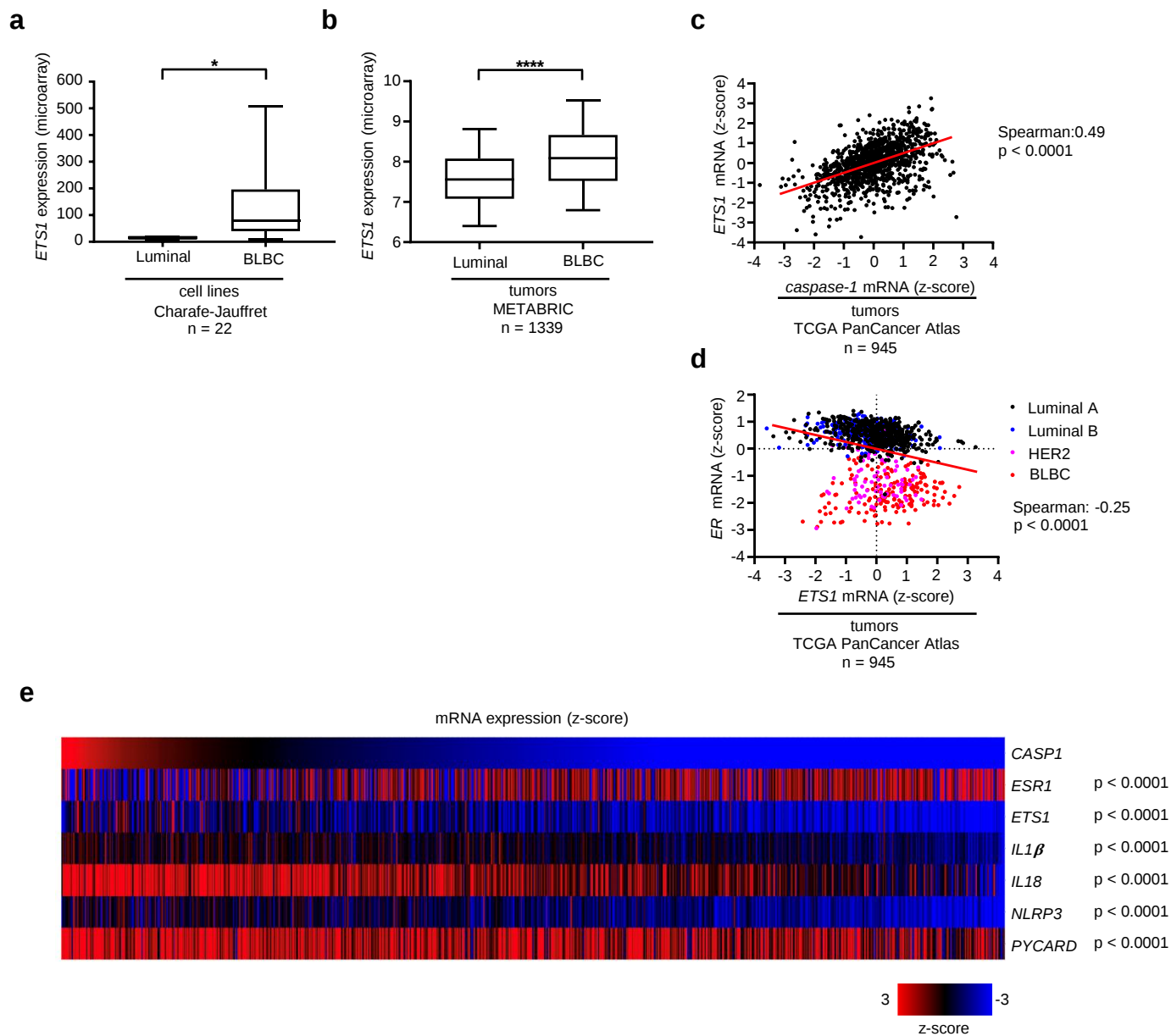
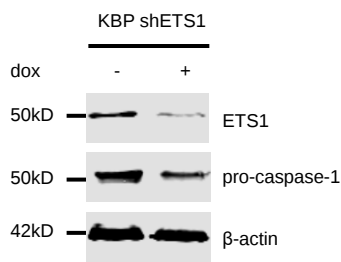
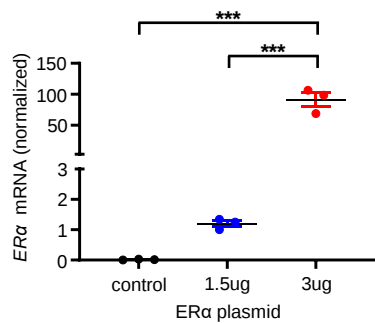


Fig. S2. ETS1 is associated with breast cancer subtype and caspase-1 expression. **a** mRNA expression level of ETS1 in 22 human breast cancer cell lines (13 Luminal and 9 BLBC cell lines) (Charafe-Jauffret dataset³), quantified by microarray, classified into two groups based on subtype (luminal vs BLBC), mean 95% CI, two-tailed, unpaired Welch's t-test with unequal variances, * : $p = 0.0376$. **b** mRNA expression level of ETS1 in 1339 human breast cancer samples (1140 luminal and 199 BLBC samples) (METABRIC dataset^{1,2}) quantified by microarray, classified into two groups based on subtype (luminal vs BLBC) (mean 95% CI, two-tailed, unpaired t-test with equal variances, **** : $p < 0.0001$). Association between ETS1 and caspase-1 mRNA expression (**c**), and between ETS1 and ER mRNA expression (**d**) in 945 human breast cancer samples (TCGA PanCancer Atlas dataset^{1,2}) quantified by RNAseq. **e** Heatmap expression levels of caspase-1 (*CASP1*), ER (*ESR1*), *ETS1*, *IL1 β* , *IL18*, and inflammasome components *NLRP3* and *ASC* (*PYCARD*) in human breast tumors from the TCGA PanCancer Atlas dataset^{1,2}. P-values represent positive Spearman correlations between caspase-1 and other listed genes, with the exception of *ER* (negative Spearman correlation). Source data are provided as a Source Data file.

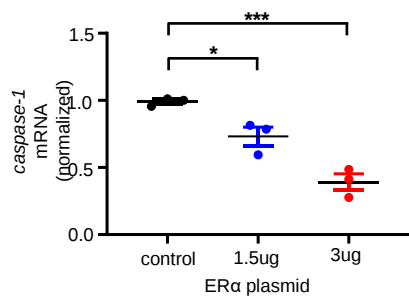
a



b



c



d

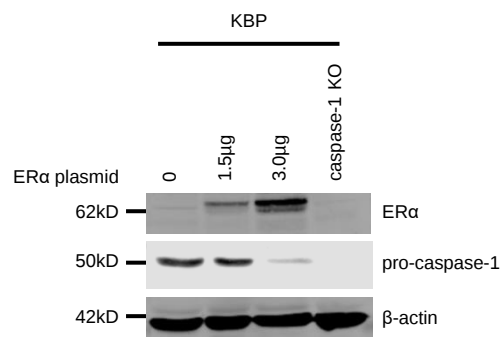


Fig. S3. ER α and caspase-1 are inversely related in murine TNBC. **a** Immunoblot of ETS1 and pro-caspase-1 in lysates from mouse KBP TNBC cells containing a doxycycline-inducible shRNA targeting ETS1, treated without (-) and with (+) doxycycline (dox). ER α (control vs 3 μ g: *** : p = 0.0002; 1.5 μ vs 3 μ g: *** : p = 0.0002) **(b)** and caspase-1 (* : p = 0.034; *** : p = 0.006) **(c)** mRNA levels in KBP cells transiently transfected with serial amounts (0 μ g, 1.5 μ g, 3 μ g) of a mouse ER α -expressing plasmid. **d** Immunoblot of ER α and pro-caspase-1 in KBP cells transiently transfected with ER α -expressing plasmid (n = 3 biologically independent experiments, including immunoblots where representative images are shown; mean with SEM, one-way ANOVA with Dunnett's multiple comparisons test). Source data are provided as a Source Data file.

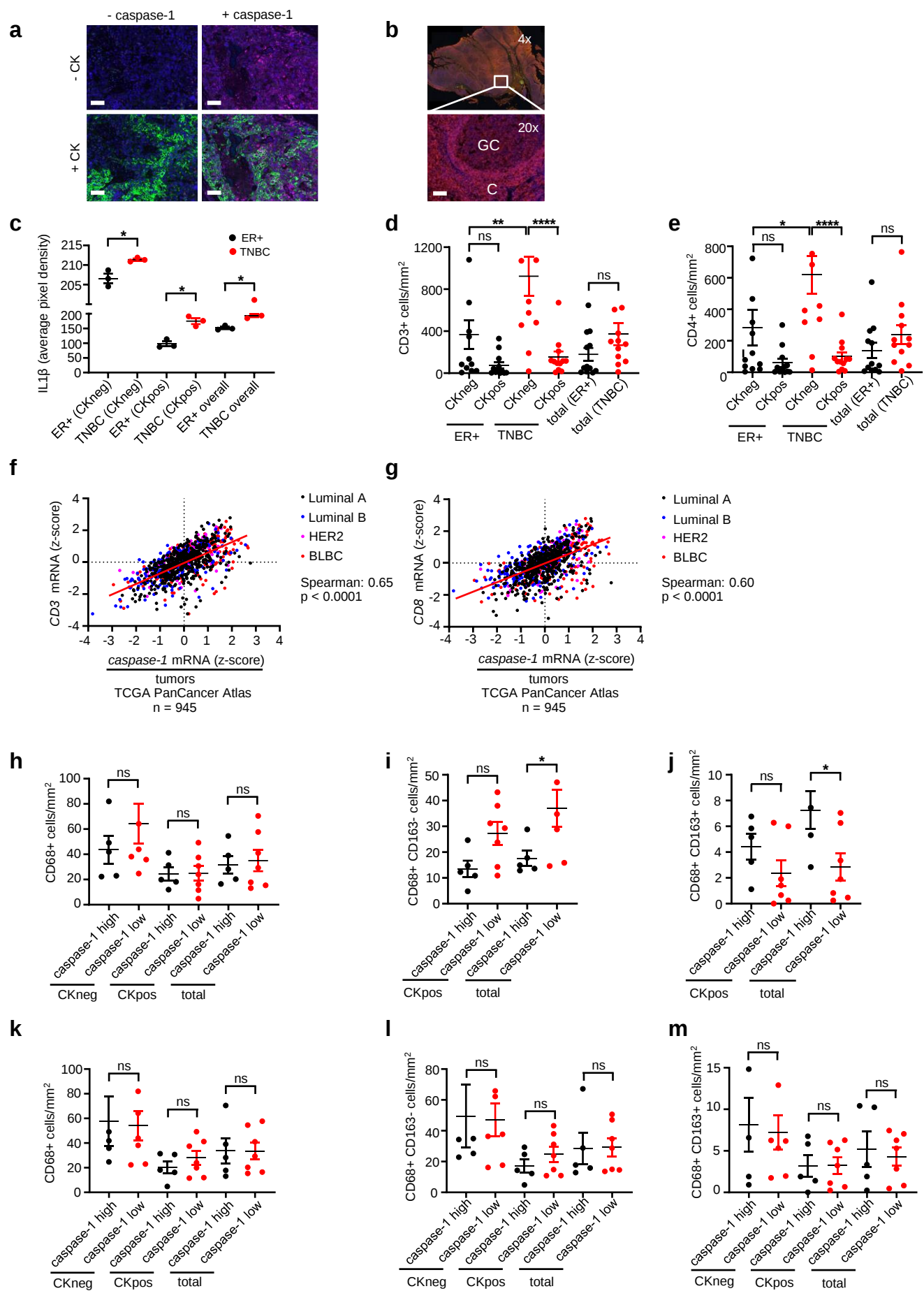


Fig. S4. Caspase-1 is associated with spatial immunophenotype in breast cancer. a

Multiplex IHC of TNBC probed with secondary antibody only (negative control, -caspase-1) or with anti-caspase-1 and secondary antibodies (+caspase-1) (DAPI: blue, CK: green, caspase-1: magenta; scalebar corresponds to 100 μ m). **b** IHC of human palatine tonsil (4X) with high magnification view of a lymph nodule (20X, scalebar corresponds to 100 μ m) confirming absence of germinal center (GC) staining and positive staining of the corona (C)⁴. **c** Multiplex IHC quantification of IL1 β in the stromal compartment (CKneg), intratumoral compartment (CKpos), or both (overall) in ER+ and TNBC tumor samples (n = 3 ER+, n = 3 TNBC; ER+ (CKneg) vs TNBC (CKneg) * : p = 0.0181; ER+ (CKpos) vs TNBC (CKpos) * : p = 0.0275; ER+ overall vs TNBC overall * : p = 0.0273) . Multiplex IHC quantification of CD3+ lymphocytes (** : p = 0.009; **** : p < 0.0001) (**d**) and CD4+ T-cells (* : p = 0.0315; **** : p < 0.0001) (**e**) in ER positive (ER+) and TNBC, in the stromal (CKneg) or intratumoral (CKpos) compartments, or overall (total) (n = 12 ER+, n = 12 TNBC). Association between CD3 and caspase-1 mRNA expression (**f**), and between CD8 and caspase-1 mRNA expression (**g**) in 945 human breast cancer samples (TCGA PanCancer Atlas dataset^{1,2}) quantified by RNAseq. Quantification of CD68+ macrophages (n = 7) (**h, k**), CD68+ CD163- M1-like macrophages (* : p = 0.0286, n = 7) (**i, l**) and CD68+ CD163+ M2-like macrophages (* : p = 0.0311, n = 7) (**j, m**) within the CKneg and CKpos compartments, or overall (total). Caspase-1 high and low groups were categorized according to intratumoral (**h - j**) or stromal (**k - m**) caspase-1 quantification, divided by the mean caspase-1 value in those compartments. Mean with SEM, one-way ANOVA with Bonferroni's multiple comparisons test. ns = not significant. Source data are provided as a Source Data file.

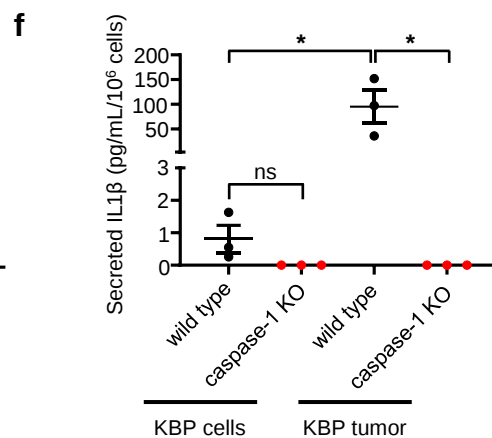
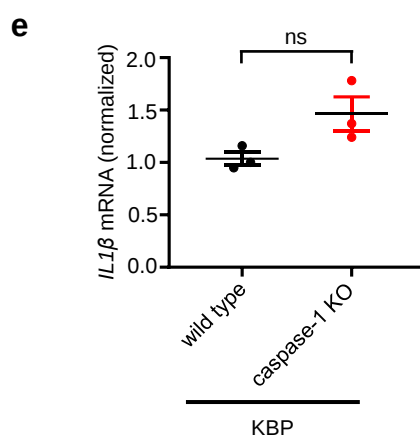
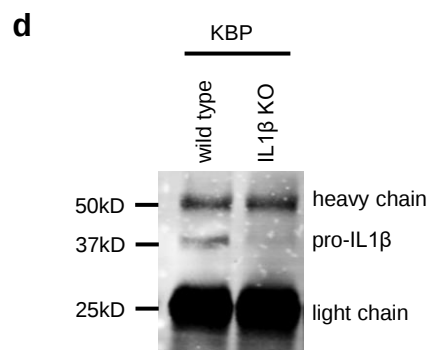
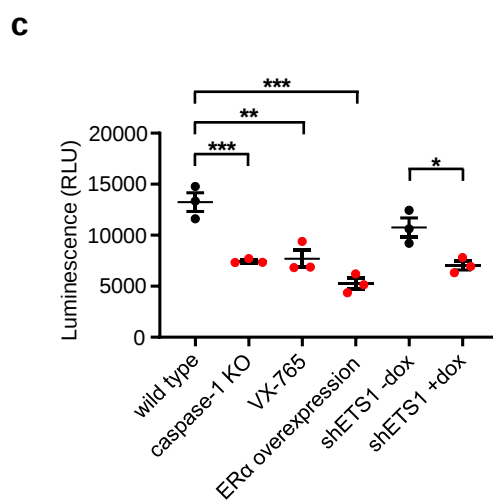
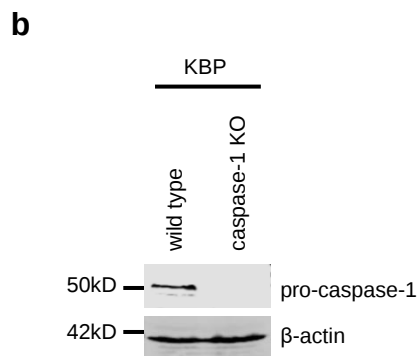
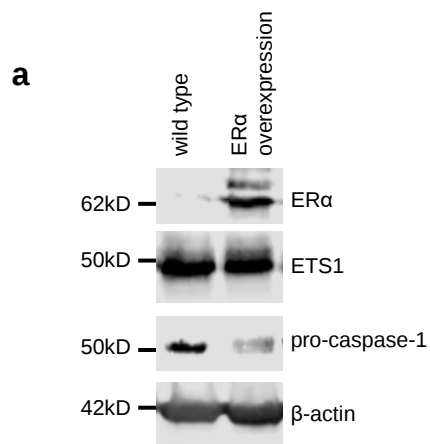


Fig. S5. Caspase-1 production is ER α -, and ETS1-dependent, and is required for IL1 β secretion in murine KBP cells. **a** Immunoblot of ER α , ETS1, and pro-caspase-1 in wild type or ER α -overexpressing KBP cells. **b** Immunoblot of pro-caspase-1 in wild type and CRISPR-mediated caspase-1 KO KBP cells. β -actin is included as a loading control. **c** Analysis of caspase-1 activity (Caspase-1 Glo Inflammasome assay) in wild type, VX-765 (Belnacasan®) caspase-1 antagonist-treated, ER α -overexpressing and doxycycline-inducible shETS1 KBP cells. Mean with SEM, one-way ANOVA with Bonferroni's multiple comparisons test. * : $p = 0.0393$, ** : $p = 0.0017$, wild type vs caspase-1 KO: *** : $p = 0.0009$; wild type vs ER α overexpression: *** : $p = 0.0006$, $n = 3$. **d** Immunoprecipitation (anti-IL1 β) immunoblot of IL1 β in wild type and IL1 β KO KBP cells. **e** IL1 β mRNA levels detected by RTqPCR in wild type and caspase-1 KO KBP cells ($n = 3$ biologically independent experiments, mean with SEM, two-tailed, unpaired t-test with equal variances). **f** Secreted IL1 β levels detected by ELISA in wild type and caspase-1 KO KBP cells in culture and harvested from KBP orthotopic tumors. One-way ANOVA with Bonferroni's multiple comparisons test for ELISA, ns = not significant, KBP cells vs tumor: * : $p = 0.0245$; KBP wild type vs caspase-1 KO tumor: * : $p = 0.0233$; $n = 3$. $n = 3$ biologically independent experiments for all immunoblots and representative images are shown. Source data are provided as a Source Data file.

Fig. S6. Inhibition of caspase-1 production mimics IL1 β knockout and reduces macrophage infiltrates in mouse models of TNBC. Growth curves, tumor weights, flow cytometric quantification of macrophages (F4/80 and CD11b double positive cells as a proportion of CD45 positive immune cells) and IHC staining/quantification of macrophages in the following TNBC mouse models: **a** Wild type and ER α -overexpressing KBP allografts (tumor weight: ** : $p = 0.0086$; F4/80+ CD11b+ in CD45+ cells: ** : $p = 0.0033$; F4/80+ in total cells: ** : $p = 0.0012$; **** : $p < 0.0001$; $n = 3$). **b** Representative FACS plots demonstrating the gating strategy for F4/80+ CD11b+ cells in CD45+ cells, in wild type and ER α -overexpressing KBP allografts. **c** KBP shETS1 tumors in mice fed regular (-dox) or doxycycline-containing (+dox) diet at tumor onset (tumor weight: * : $p = 0.016$; F4/80+ CD11b+ in CD45+ cells: * : $p = 0.0498$; *** : $p = 0.0005$; **** : $p < 0.0001$; $n = 3$). **d** Wild type and caspase-1 knockout (KO) KBP tumors (F4/80+ CD11b+ in CD45+ cells: * : $p = 0.024$; F4/80+ in total cells: * : $p = 0.0156$; *** : $p = 0.0009$; **** : $p < 0.0001$, $n = 4$). **e** Wild type and IL1 β KO KBP tumors (* : $p = 0.0122$; ** : $p = 0.0011$; *** : $p = 0.0002$; **** : $p < 0.0001$, $n = 8$). **f** Wild type KBP tumors treated with vehicle control or VX-765 (* : $p = 0.0414$; tumor weight: ** : $p = 0.0075$; F4/80+ CD11b+ in CD45+ cells: ** : $p = 0.0042$; **** : $p < 0.0001$, $n = 4$). **g** Growth curve, tumor weight, flow cytometric quantification of macrophages (CD14+ HLA-DR+, CD163+ M2-like) and T cells (CD8+ and GrB+) in humanized mice bearing MDA-MB231 xenografts, treated with vehicle control or VX-765 (CD14+ HLADR+ in CD45+ cells: * : $p = 0.0134$; %GrB+ in CD8+ cells: * : $p = 0.0227$, $n = 6$). Mean with SEM, two-way ANOVA with Bonferroni's multiple comparisons test for growth curves, two-tailed, unpaired t-test with equal variances for tumor weights and macrophage quantification. Scalebar corresponds to 50 μ m. Source data are provided as a Source Data file.

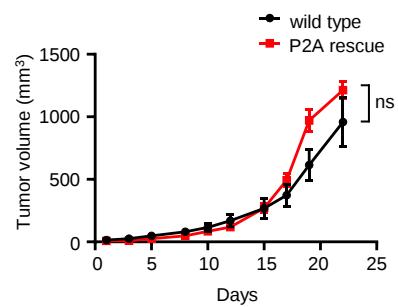
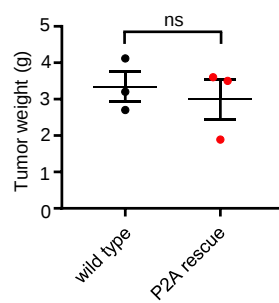
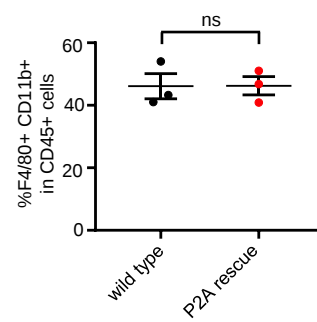
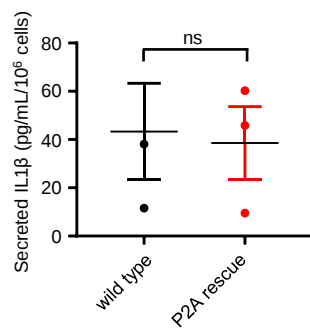
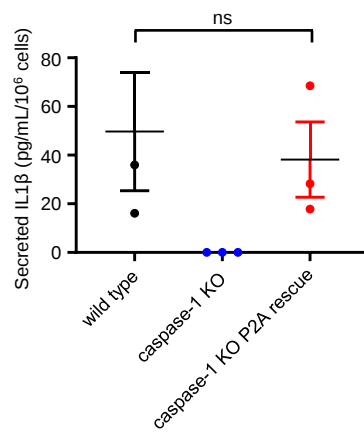
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Fig. S7. IL1 β secretion, growth and macrophage recruitment of KBP *IL1 β -P2A* variant tumors phenocopy wild type. Growth curve (**a**), tumor weights at end point (n = 3) (**b**), and flow cytometry quantification of macrophages (F4/80 and CD11b double positive cells as a proportion of CD45 positive immune cells) (n = 3) (**c**) of wild type and P2A rescue KBP orthotopic tumors (n = 3). **d** ELISA measuring secreted IL1 β from wild type and P2A rescue KBP orthotopic tumors (n = 3). **e** ELISA measuring secreted IL1 β from wild type, caspase-1 KO, and caspase-1 KO P2A rescue KBP orthotopic tumors (n = 3). Mean with SEM, two-way ANOVA with Bonferroni's multiple comparisons test for growth curve, two-tailed, unpaired t-test with equal variances for tumor weight, macrophage quantification and ELISA, ns = not significant. Source data are provided as a Source Data file.

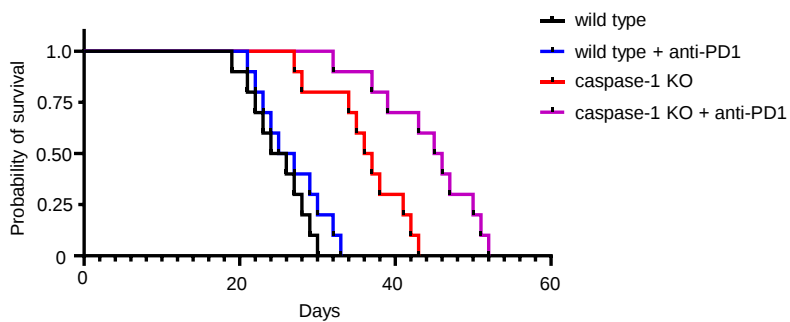
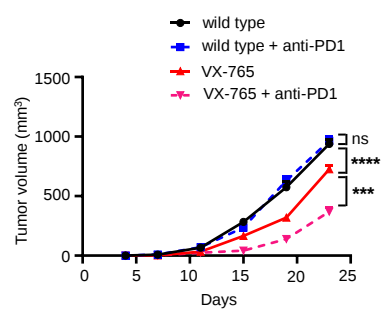
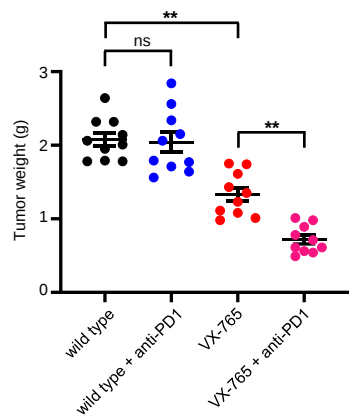
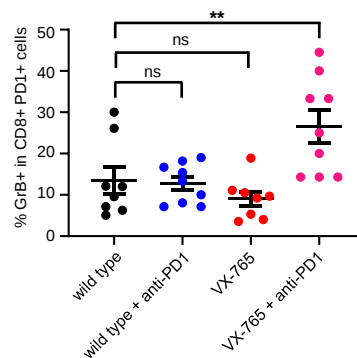
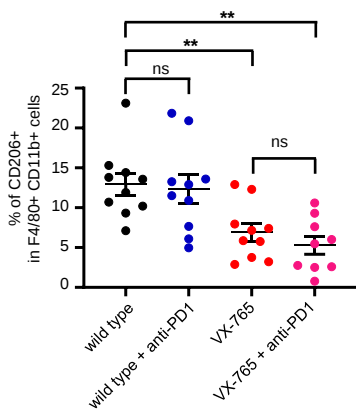
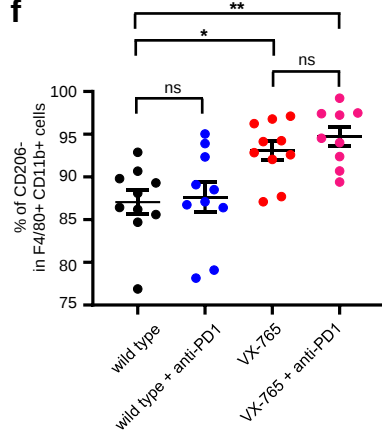
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Fig. S8. Caspase-1 neutralization improves the effect of anti-PD1 treatment in 4T1 and KBP TNBC mouse models. **a** Kaplan-Meier survival curves of wild type and caspase-1 KO KBP orthotopic tumors treated with IgG control or anti-PD1 antibody. A log-rank test was used to determine the differences between wild type and wild type + anti-PD1; wild type and caspase-1 KO; wild type and caspase-1 KO + anti-PD1; caspase-1 KO and caspase-1 KO + anti-PD1 groups ($p = 0.24$, $p < 0.0001$, $p < 0.0001$, and $p = 0.0023$, respectively, $n = 10$). Growth curve (** : $p = 0.0002$; **** : $p < 0.0001$) (**b**), tumor weight (wild type vs VX-765: ** : $p = 0.002$; VX-765 vs VX-765 + anti-PD1: ** : $p = 0.002$, $n = 10$) (**c**), flow cytometry quantification of GrB+ CD8+ PD1+ T cells (** : $p = 0.0071$) (**d**), F4/80+ CD11b+ CD206+ macrophages (wild type vs VX-765: ** : $p = 0.0031$; wild type vs VX-765 + anti-PD1: ** : $p = 0.0086$, $n = 10$) (**e**), and F4/80+ CD11b+ CD206- macrophages (* : $p = 0.0187$; ** : $p = 0.0027$, $n = 10$) (**f**) in wild type and VX-765-treated 4T1 orthotopic tumors treated with IgG control or anti-PD1 antibody. Mean with SEM, two-way ANOVA with Tukey's multiple comparisons test for growth curves, one-way ANOVA with Tukey's multiple comparisons test for tumor weights and flow cytometry. ns = not significant. Source data are provided as a Source Data file.

Table S1. Sequences of primers, siRNAs, shRNAs, and sgRNAs.

RTqPCR	Assay ID
human caspase-1	Hs.PT.56a.22997425.g
human ER α	Hs.PT.58.14846478
human ETS1	Hs.PT.58.39917763
human β -actin	Hs.PT.39a.22214847
mouse caspase-1	Mm.PT.58.12299102.g
mouse ER α	Mm.PT.58.8025728
mouse IL1 β	Mm.PT.58.41616450
human β -actin	Mm.PT.58.28904620.g
ChIP assay	
human caspase-1 promoter	Forward primer: CGACATTCTCATTCCAGAGCCTATG Reverse primer: GACAGGGTCTCCTTGTGTTTCCTA
Cloning	
human ER α overexpressing plasmid	Forward primer: CGCGGATCCATGACCATGACCCTCCACAC Reverse primer: CCGGAATTCTCAGACCGTGGCAGGGAAAC
human ETS1 overexpressing plasmid	Forward primer: CGCGGATCCATGCCAACTTTGTACAAAAA Reverse primer: CCGGAATTCTCACTCGTCGGCATCTGGCT
mouse ER α overexpressing plasmid	Forward primer: AATCCTGAATTCATGACCATGACCCTTCACAC Reverse primer: AATAGGATCCTCAGATCGTGTTGGGGAAGC
caspase-1 CRISPR plasmid	Forward primer: CACCGAGGGCAAGACGTGTACGAG Reverse primer: AAACCTCGTACACGTCTTGCCCTC

Table S1 cont'd.

siRNA	Sequence
siscr	UGGUUUACAUGUCGACUAA UGGUUUACAUGUUGUGUGA UGGUUUACAUGUUUUCUGA UGGUUUACAUGUUUCCUA
siER α	GAUCAAACGCUCUAAGAAG GAAUGUGCCUGGCUAGAGA GAUGAAAGGUGGGAUACGA GCCAGCAGGUGCCCUACUA
siETS1	AUAGAGAGCUACGAUAGUU GAAUGAUGUCUCAAGCAU GUGAAACCAUAUCAAGUUA CAGAAUGACUACUUUGCUA
shRNA	
caspase-1	ACACGTCTTGCTCTCATTA
ETS1	AGGTGTAACAGGATTCTGG
CRISPR sgRNA	
caspase-1 (cloning method)	CTCGTACACGTCTTGCCCTC
caspase-1 (Synthego method)	UUAAACAGACAAGAUCUGA
IL1 β	CCAUUAGACAACUGCCAC
CRISPR ssODN	
P2A rescue (red = P2A sequence)	TGATAACCTGCTGGTGTGTGACGTTCCCATTAGACAACTGCAC GGCAGCGGAGCTACTAACTTCAGCCTGCTGAAGCAGGCTGGAGA CGTGGAGGAGAACCCTGGACCTGTTCCCATTAGACAACTGCACTA CAGACTCCGAGATGAACAACAAAAAGCCTCGTGCTGTCGGA

Table S2. Antibodies.

Immunoblot	Source	Catalog no.	Working dilutions
Primary			
ER α	Cell Signaling	8644S	1: 500
ETS1	Cell Signaling	14096S	1: 1000
caspase-1	Cell Signaling	3866S	1: 500
IL1 β	Cell Signaling	83186	1: 200
IL18	Abcam	Ab68435	1: 500
β -actin	Sigma	A5441	1: 2000
mouse caspase-1	Invitrogen	14-9832-82	1: 500
mouse IL1 β	R&D Systems	AF-401-NA	1: 200
Secondary			
Donkey anti-rabbit-800CW	Mandel Scientific	926-32213	1: 2000
Donkey anti-goat-800CW	Mandel Scientific	926-32214	1: 2000
Goat anti-rat-800CW	Mandel Scientific	926-32219	1: 2000
Donkey anti-mouse-800CW	Mandel Scientific	923-33212	1: 2000
Goat anti-mouse IgG2a-680LT	Mandel Scientific	926-68051	1: 2000
Donkey anti-rabbit-680LT	Mandel Scientific	926-68023	1: 2000
Donkey anti-goat-680LT	Mandel Scientific	926-68024	1: 2000
Donkey anti-mouse-680LT	Mandel Scientific	926-68022	1: 2000
ChIP assay			
Isotype control rabbit monoclonal	Cell Signaling	3900S	1: 200
IgG			
ETS1	Cell Signaling	14069S	1: 200
IL1 β immunoprecipitation			
IL1 β	R&D Systems	AF-401-NA	1: 200
IHC			
Primary			
F4/80	ThermoFisher	MF48000	1: 400
IL1 β	Abcam	ab9722	1: 400
Pan-CK	Abcam	ab86734	1: 1000
Secondary			
Rabbit IgG Biotinylated	Vector Laboratories	AK-5001	1: 1000
Mouse IgG Peroxidase	Vector Laboratories	PK-4002	1: 1000
Rat IgG Peroxidase	Vector Laboratories	PK-4004	1: 1000
ELISA			
IL1 β	R&D Systems	DY401-05	
Flow cytometry			
viability	eBioscience	65-0865-18	1: 1000
mCD45	eBioscience	63-0451-82	1: 100
mCD19	eBioscience	67-0193-82	1: 100

Table S2 cont'd

Flow cytometry			
mF4/80	BioLegend	123128	1: 100
mCD11b	BioLegend	101212	1: 100
mCD3	BioLegend	100234	1: 100
mCD8	eBioscience	78-0081-82	1: 100
mPD1	eBioscience	78-9985-82	1: 100
mGranzyme B	eBioscience	48-8898-82	1: 100
mCD206	ebioscience	25-2601-80	1: 100
hCD45	Biolegend	304048	1: 100
hCD14	Invitrogen	12-0149-42	1: 100
hHLA-DR	Biolegend	307644	1: 100
hCD163	Biolegend	333618	1: 100
hCD8	Invitrogen	25-0087-42	1: 100
hGranzyme B	Invitrogen	MHGB04	1: 100
Multiplex IHC	Source	Catalog no.	Working dilutions
panCK	Agilent	M351529-2	1: 200
caspase-1	EMD Millipore	06-503	1: 200
CD3	Roche	790-4341	1: 200
CD8	Roche	790-4460	1: 200
CD68	Agilent	M0814	1: 200
CD163	Biocare Medical	CM353M	1: 200

Table S3. Clinical details of patient samples obtained from the UHN Biobank

IDC = invasive ductal carcinoma, All patient samples were obtained from surgical specimens.

Sample IDs	Age at diagnosis	Breast cancer type	ER status
63015	50-55	IDC	Negative
63082	60-65	IDC	Negative
64231	50-55	IDC	Negative
65044	80-85	IDC	Negative
66719	40-45	IDC	Negative
68316	75-80	IDC	Negative
69342	60-65	IDC	Negative
69489	40-45	IDC	Negative
71307	70-75	IDC	Negative
1036564	40-45	IDC	Negative
1054120	55-60	IDC	Negative
1095672	60-65	IDC	Negative
60195	70-75	IDC	Positive
62343	65-70	IDC	Positive
63080	40-45	IDC	Positive
64313	80-85	IDC	Positive
65811	70-75	IDC	Positive
66475	60-65	IDC	Positive
66501	60-65	IDC	Positive
67901	70-75	IDC	Positive
69739	50-55	IDC	Positive
919853	45-50	IDC	Positive
1368383	55-60	IDC	Positive
1441649	55-60	IDC	Positive

Supplementary References

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