

Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our [Editorial Policies](#) and the [Editorial Policy Checklist](#).

Statistics

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

n/a	Confirmed
☐	☒ The exact sample size (<i>n</i>) for each experimental group/condition, given as a discrete number and unit of measurement
☐	☒ A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
☐	☒ The statistical test(s) used AND whether they are one- or two-sided <i>Only common tests should be described solely by name; describe more complex techniques in the Methods section.</i>
☒	☐ A description of all covariates tested
☒	☐ A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
☐	☒ A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
☐	☒ For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>
☒	☐ For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
☒	☐ For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
☒	☐ Estimates of effect sizes (e.g. Cohen's <i>d</i> , Pearson's <i>r</i>), indicating how they were calculated

Our web collection on [statistics for biologists](#) contains articles on many of the points above.

Software and code

Policy information about [availability of computer code](#)

Data collection	Scanning microscope images were collected by Scanning electron microscopy (SEM; Philips FEG SEM XL30). Zeta size and zeta potential data were collected by Particle Size Analyzer software (Zeta sizer, Malvern Panalytical). Fluorescence microscopy images were collected by Leica SP8LIA++ software (Leica Confocal microscope). Fluorescence/UV-vis absorption intensity data were collected by SoftMax® Pro Software (The SpectraMax® M5e Microplate Reader). Infrared data was collected by FLIR SC600 Infrared Camera (FLIR™).
Data analysis	Scanning microscope data were analyzed by ImageJ (Fiji), OriginPro 2019 b (64-bit) and Microsoft Excel 365. Zeta size and zeta potential data were analyzed by OriginPro 2019 b (64-bit) and Microsoft Excel 365. Immunofluorescence data were analyzed by using ImageJ (Fiji), OriginPro 2019 b (64-bit) and Microsoft Excel 365. Fluorescence/UV-vis absorption intensity data were analyzed by OriginPro 2019 b (64-bit) and Microsoft Excel 365. Infrared data were analyzed by Windows Media Player, OriginPro 2019 b (64-bit) and Microsoft Excel 365.

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio [guidelines for submitting code & software](#) for further information.

Data

Policy information about [availability of data](#)

All manuscripts must include a [data availability statement](#). This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our [policy](#)

The authors declare that all data are available in the main text and supporting information.

Research involving human participants, their data, or biological material

Policy information about studies with [human participants or human data](#). See also policy information about [sex, gender \(identity/presentation\), and sexual orientation](#) and [race, ethnicity and racism](#).

Reporting on sex and gender

Reporting on race, ethnicity, or other socially relevant groupings

Population characteristics

Recruitment

Ethics oversight

Note that full information on the approval of the study protocol must also be provided in the manuscript.

Field-specific reporting

Please select the one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.

☒ Life sciences ☐ Behavioural & social sciences ☐ Ecological, evolutionary & environmental sciences

For a reference copy of the document with all sections, see [nature.com/documents/nr-reporting-summary-flat.pdf](https://www.nature.com/documents/nr-reporting-summary-flat.pdf)

Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

Sample size

Data exclusions

Replication

Randomization

Blinding

Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems

Methods

n/a	Involved in the study
☐	☒ Antibodies
☐	☒ Eukaryotic cell lines
☒	☐ Palaeontology and archaeology
☒	☐ Animals and other organisms
☒	☐ Clinical data
☒	☐ Dual use research of concern
☒	☐ Plants

n/a	Involved in the study
☒	☐ ChIP-seq
☒	☐ Flow cytometry
☒	☐ MRI-based neuroimaging

Antibodies

Antibodies used

Anti- α -Tubulin anti-body in Tubulin-Tracker Green kit, (Beyotime, C1051S).
 Anti-mouse IgG H&L with Alexa Fluor 488 in Tubulin-Tracker Green kit, (Beyotime, C1051S).
 Phalloidin with Alexa Fluor 594 in Actin-Tracker Red-594 kit, (Beyotime, C2205S).
 Anti-GAPDH (Beyotime, AG019).
 Anti-actin (Beyotime, AA128).

Validation

All antibodies are commercially available and have been validated by the manufacturers.

Eukaryotic cell lines

Policy information about [cell lines and Sex and Gender in Research](#)

Cell line source(s)

Human lung cancer cells A549 cell (CRM-CCL-185 TM) line was purchased from ATCC, USA.

Authentication

Each cell line used was morphologically confirmed according to the information provided by ATCC website.

Mycoplasma contamination

The cell line tested negative for mycoplasma.

Commonly misidentified lines
(See [ICLAC](#) register)

N.A.

Plants

Seed stocks

N.A.

Novel plant genotypes

N.A.

Authentication

N.A.