

Supplementary information for

**Cryo-EM Structure and Oligomerization of the Human Planar Cell Polarity Core Protein
Vangl1**

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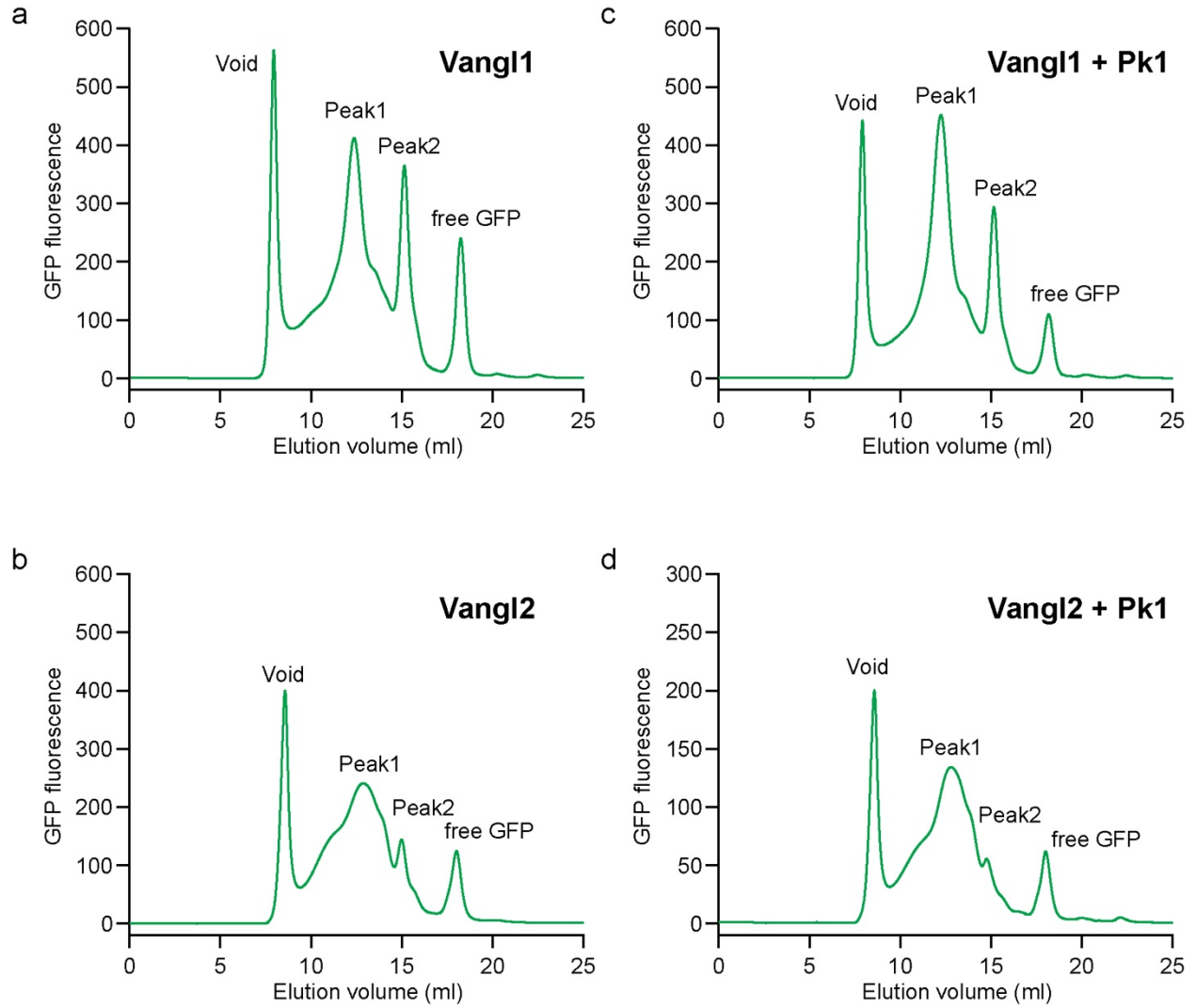
Supplementary Figure 9 | Electrophysiological recordings consistent with the notion that Vangl1 is unlikely to be an ion channel.

Supplementary Figure 10 | Comparison of the structures of Vangl1 determined by cryo-EM (cyan) and predicted by AlphaFold (light pink).

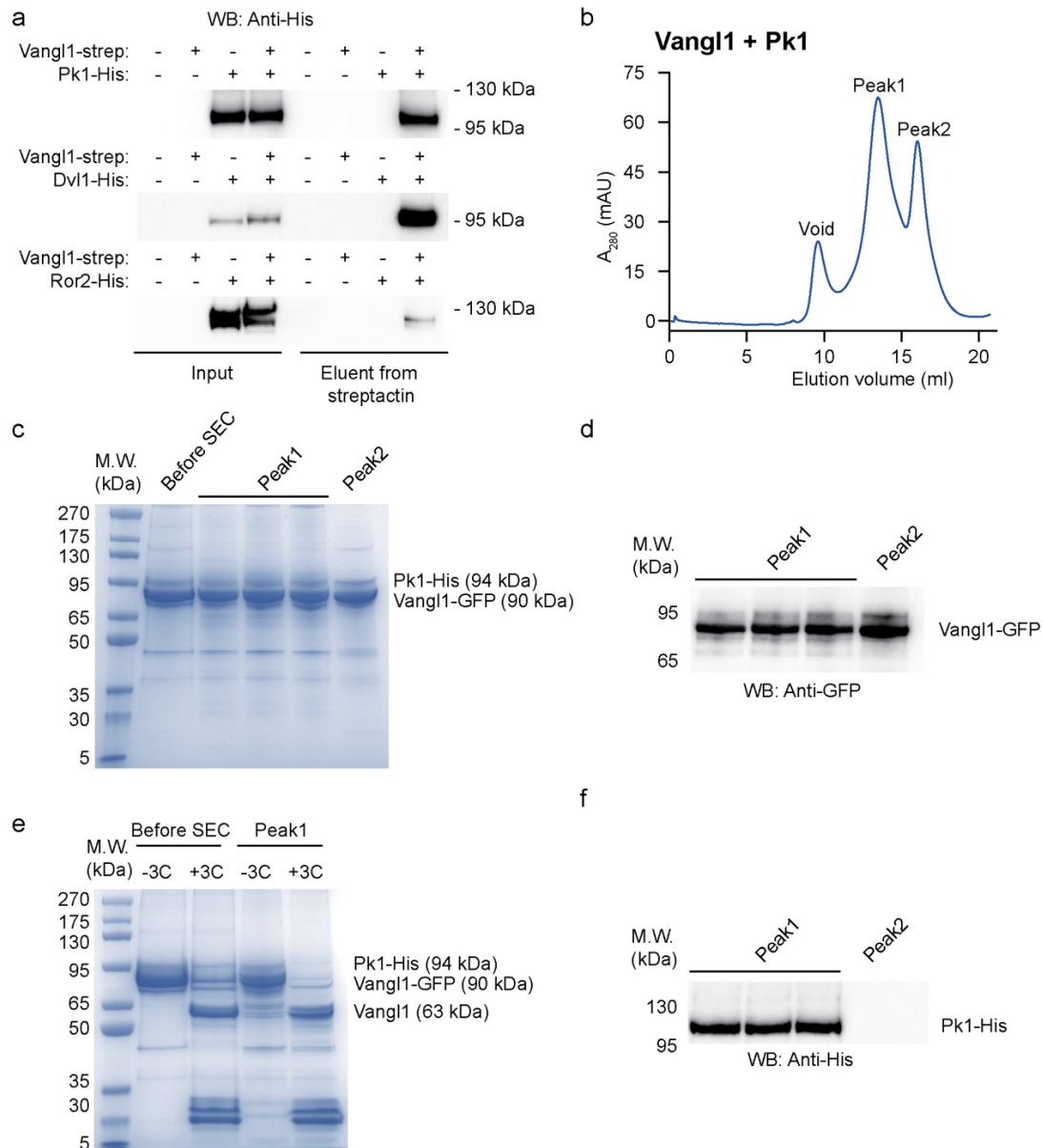
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Supplementary Table 2 | Human disease-associated point mutations of Vangl1 and Vangl2 and their structural locations.

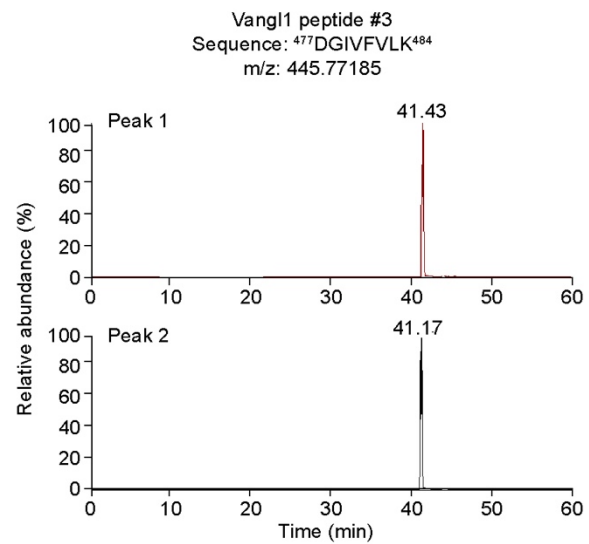
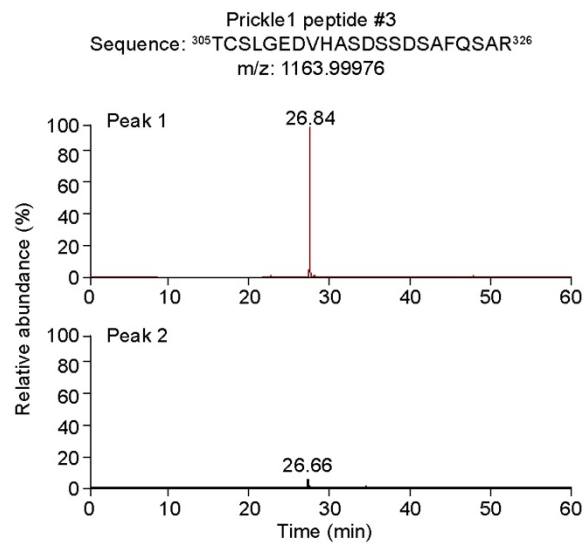
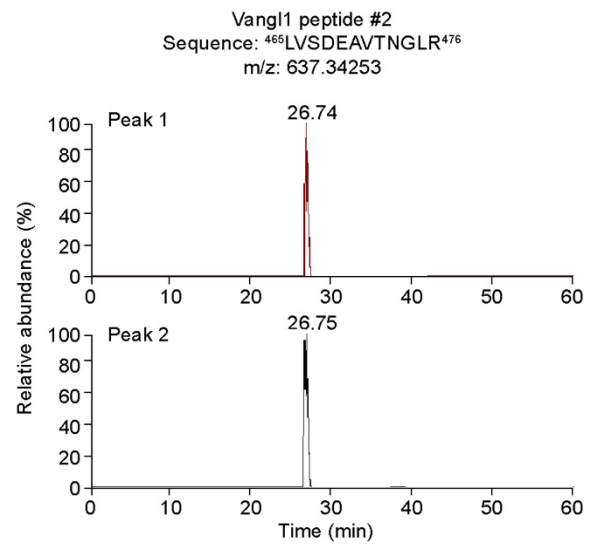
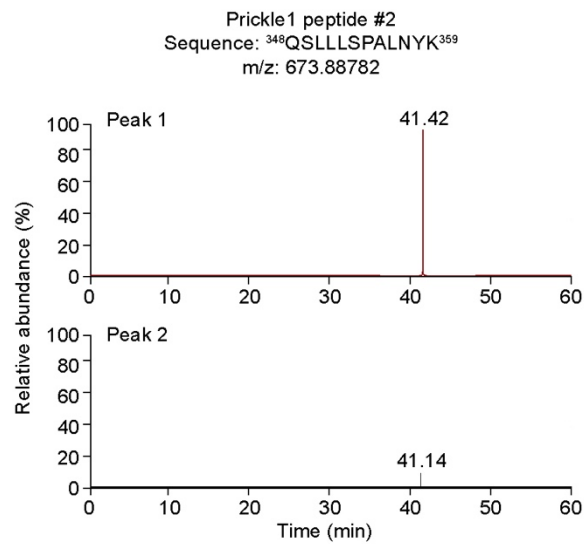
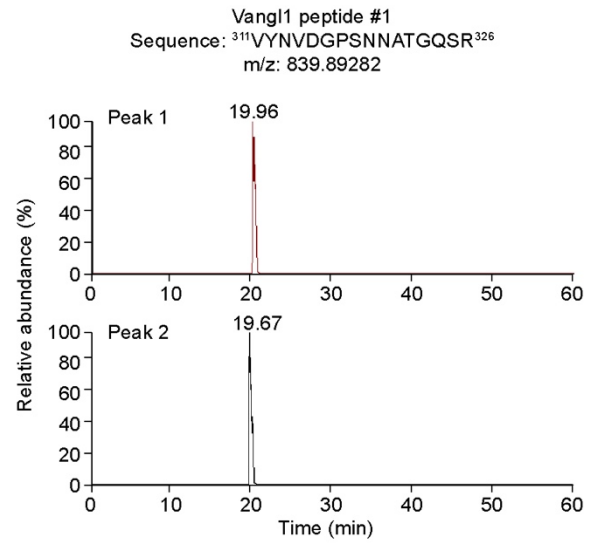
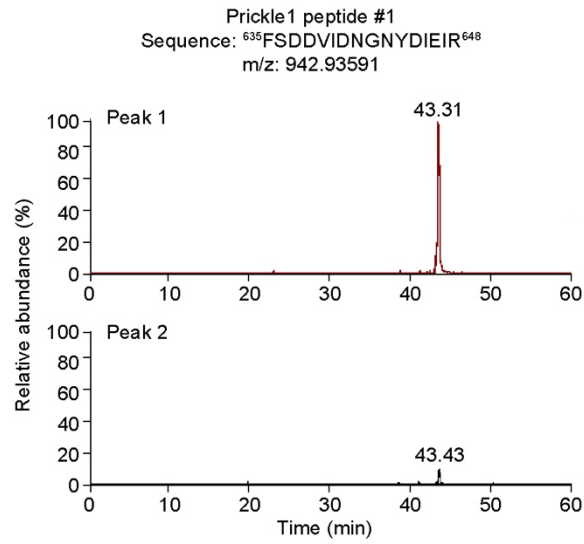
Uncropped images of Western-blot and gels



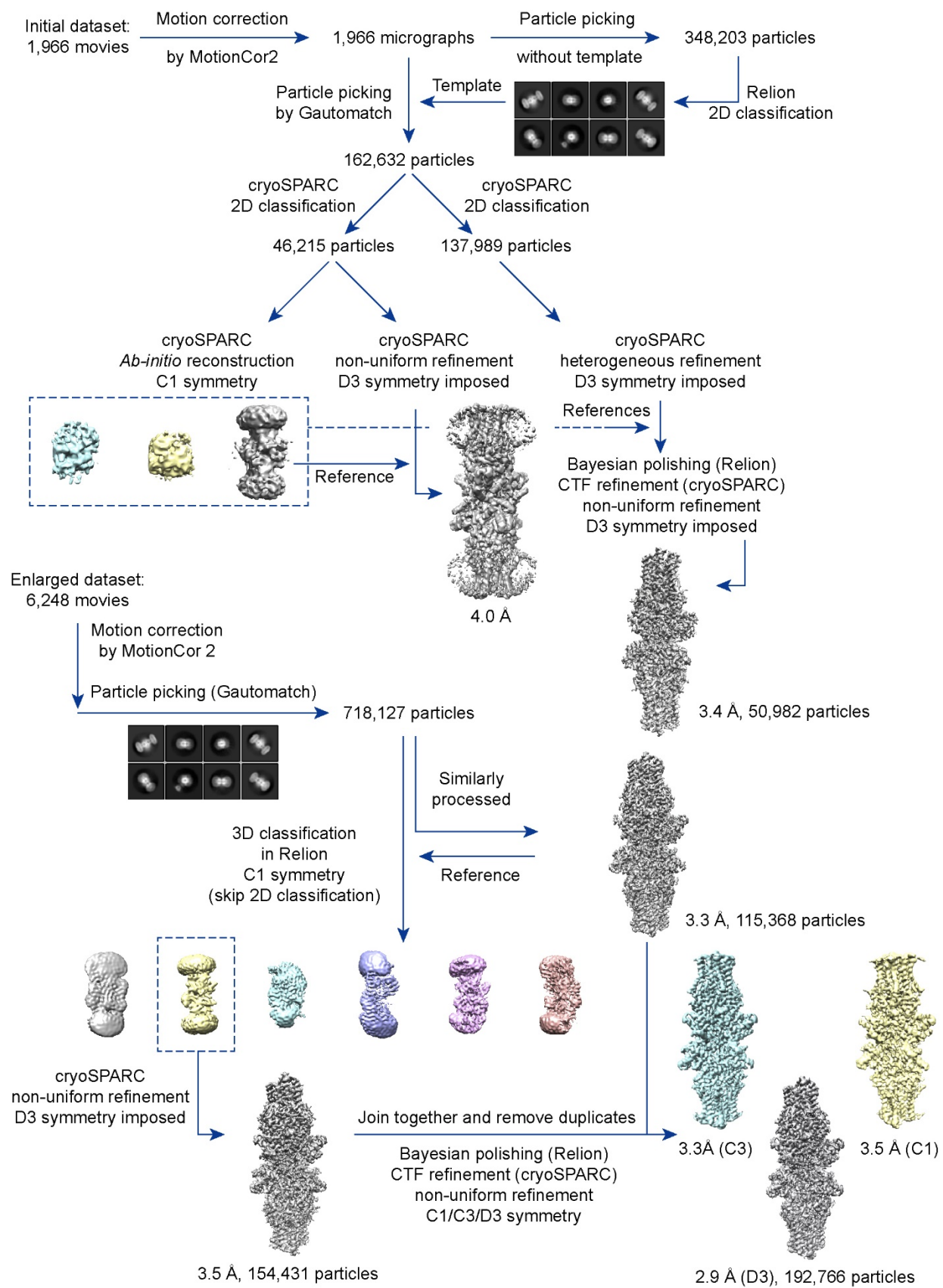
Supplementary Figure 1 | Homogeneity and oligomerization equilibrium of Vangl revealed by FSEC. a-d, FSEC profiles of Vangl1 alone (a), Vangl2 alone (b), Vangl1 co-expressed with Pk1 (c), and Vangl2 co-expressed with Pk1 (d).



Supplementary Figure 2 | Purification and characterization of Vangl. **a**, Vangl1 directly interacts with Pk1, Dvl1 and Ror2, as shown by pull down experiments. **b**, Size-exclusion chromatography (SEC) profile of tandem affinity purified Vangl1/Pk1. **c**, SDS-PAGE analysis of the SEC peak fractions from (b). Protein bands were visualized by Coomassie blue staining. **d**, Western blot confirming the presence of Vangl1 in both SEC peaks. **e**, the GFP can be cleaved from Vangl1 by 3C protease revealed by SDS-PAGE analysis. **f**, Western blot showing the presence of Pk1 in Peak 1 but not in Peak 2.



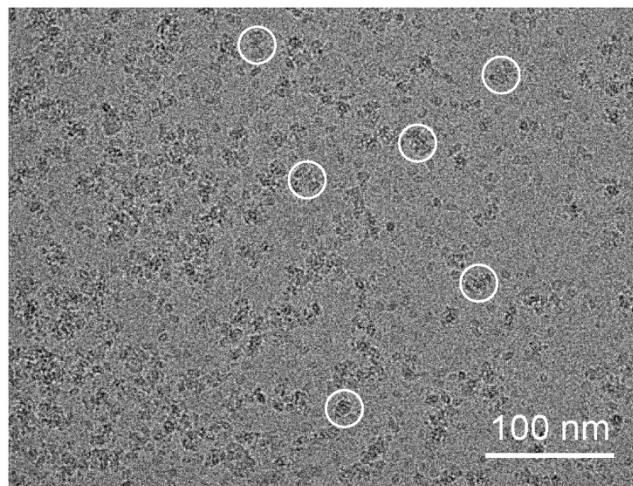
Supplementary Figure 3 | Mass spectrometry analysis of SEC peaks. The relative abundance of Pk1 in Peak 1 and Peak 2 is estimated to be ~10 times different using three independent Pk1 peptides (three panels on the left). In contrast, that of Vangl1 in both peaks is similar (three panels on the right).



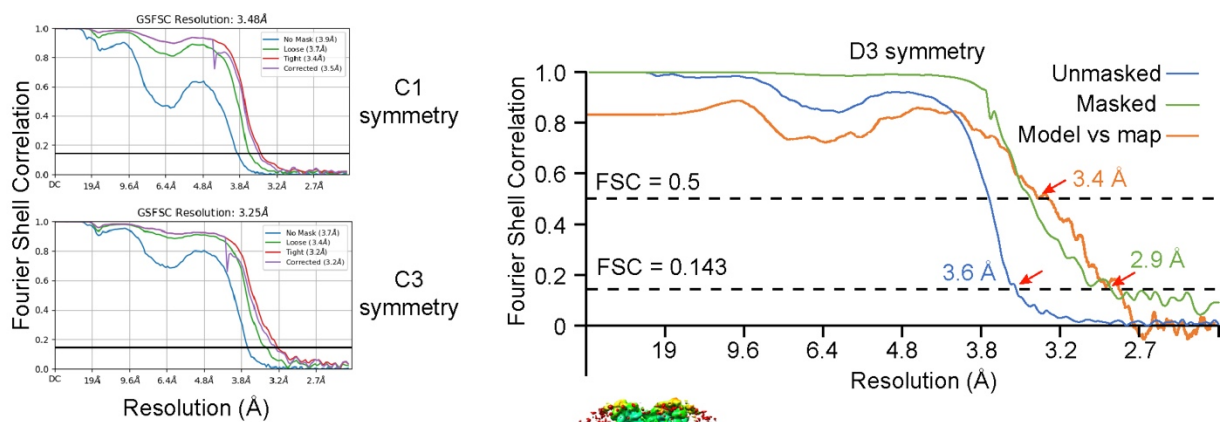
Supplementary Figure 4 | Workflow of cryo-EM data processing.

Please refer to Methods for details.

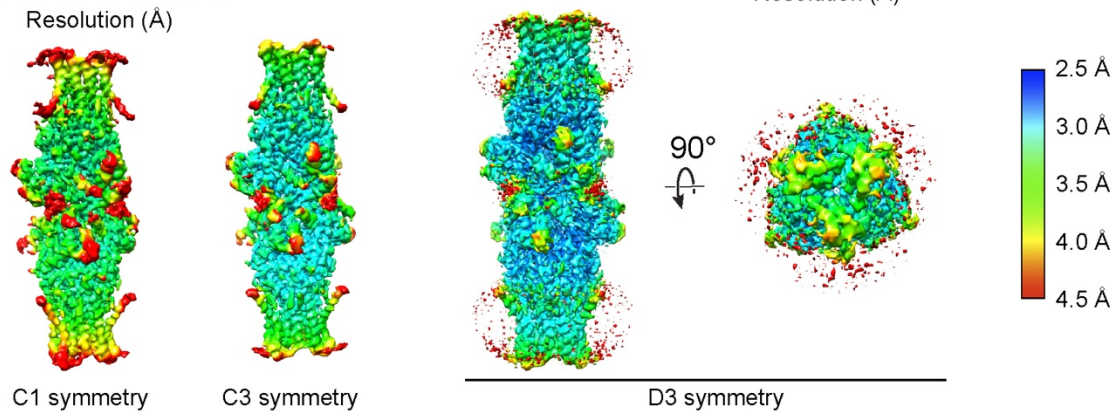
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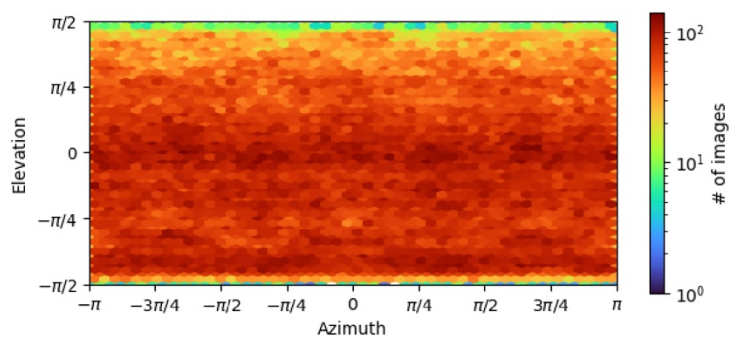
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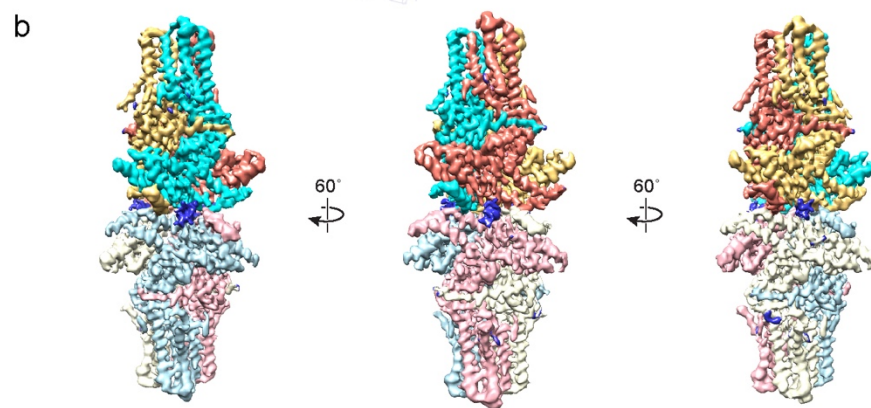
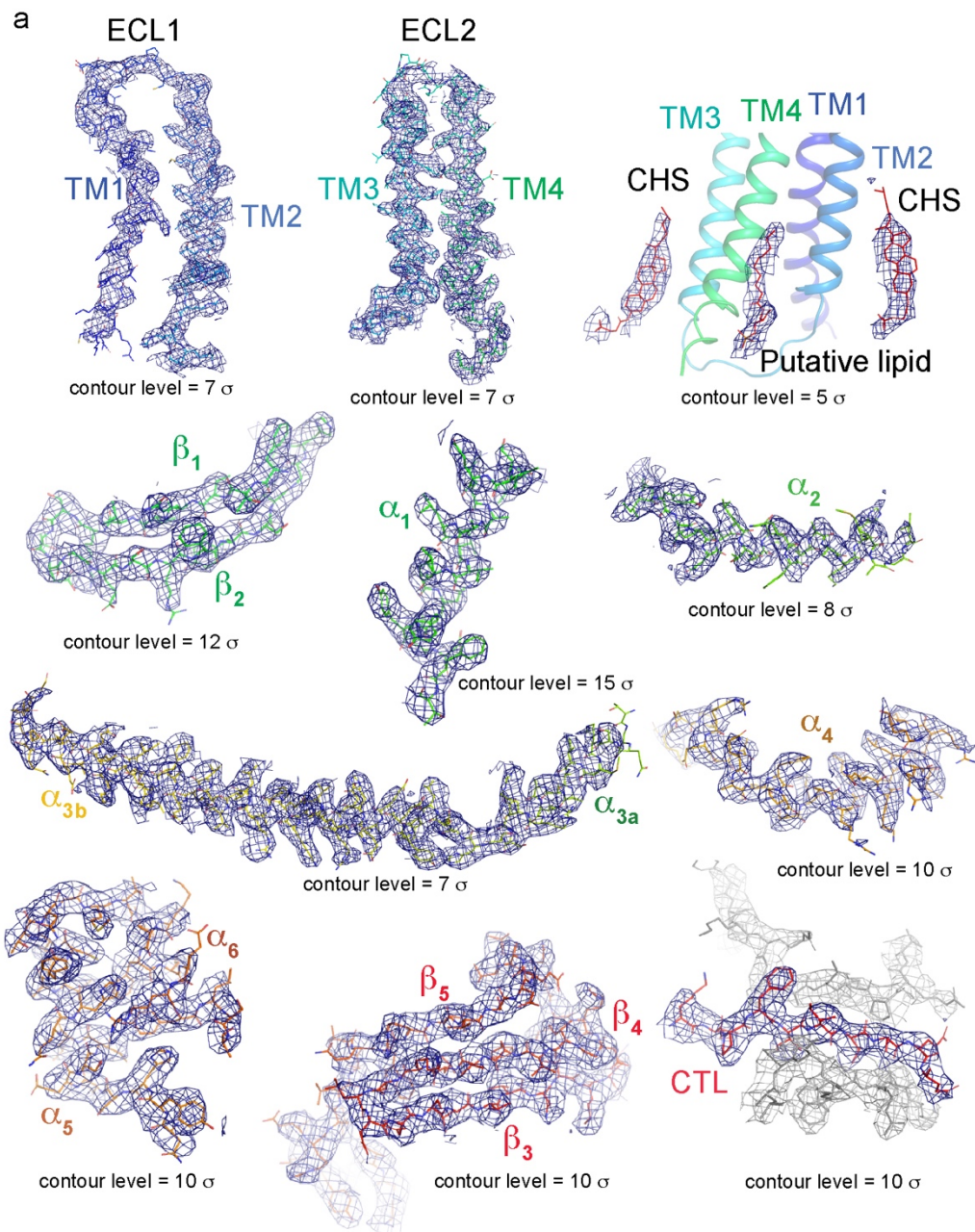
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d



Supplementary Figure 5 | Validation of cryo-EM data processing. **a**, A representative cryo-EM micrograph after motion-correction, with several single particles circled. A scale bar for 100 nm is shown in the corner. **b**, Fourier shell correlation between cryo-EM half maps with or without mask with C1, C3 and D3 symmetry imposed. That between the structural model and the D3-symmetrical cryo-EM reconstruction of D3 symmetry was also plotted. **c**, Local resolution of reconstructions of C1, C3 and D3 symmetry estimated using cryoSPARC. **d**, Angular distribution of the particles used in the final reconstruction.

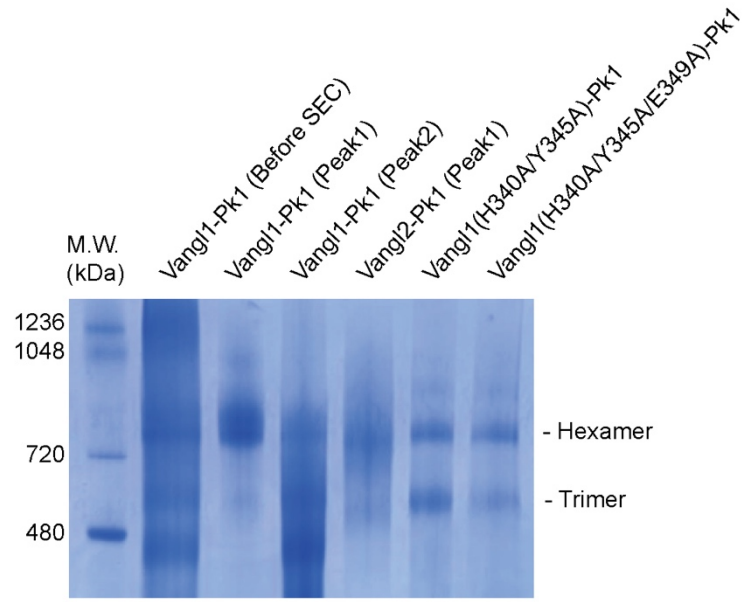


Supplementary Figure 6 | The EM map density of Vangl1. **a**, Consistent with the 2.9 Å nominal resolution, EM map density for most side chains is clearly visible, thus allowing accurate registry and unambiguous model building. **b**, Three views of the Vangl1 reconstruction without imposed symmetry, 60° apart from each other, where density for Vangl1 were colored in red, cyan and yellow of two different shades, and that putatively for Pk were shown in blue.

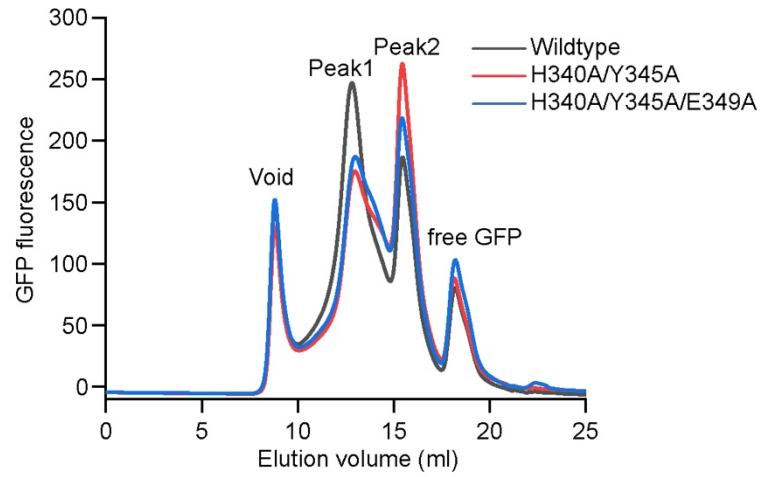


Supplementary Figure 7 | Sequence alignment of human Vangl1 and Vangl2. Identical residues are boxed in red, similar residues are represented in red font with white background, variable residues are colored in black. α -helices, β -strands, loops and disordered regions are indicated by cylinders, arrows, solid and dashed lines, respectively, according to the structure.

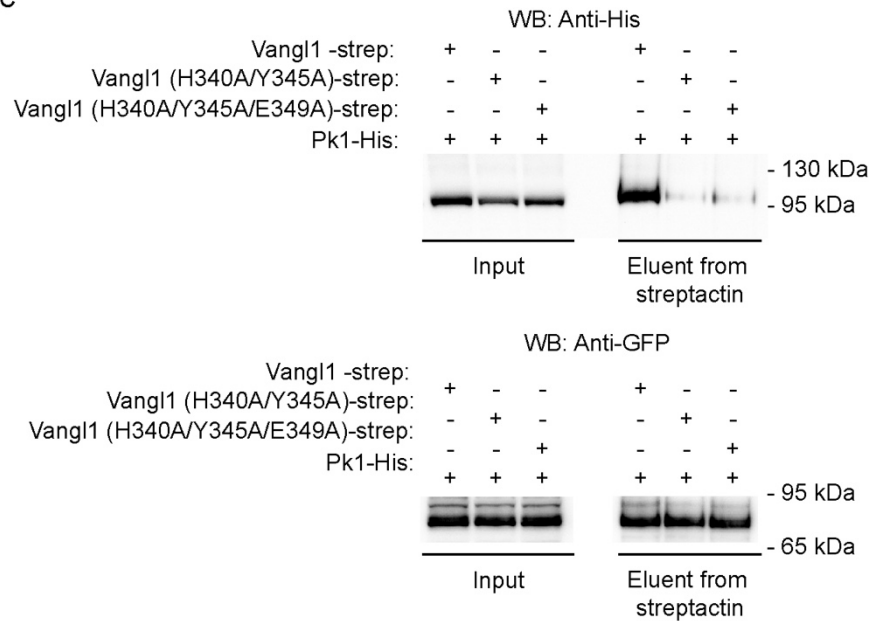
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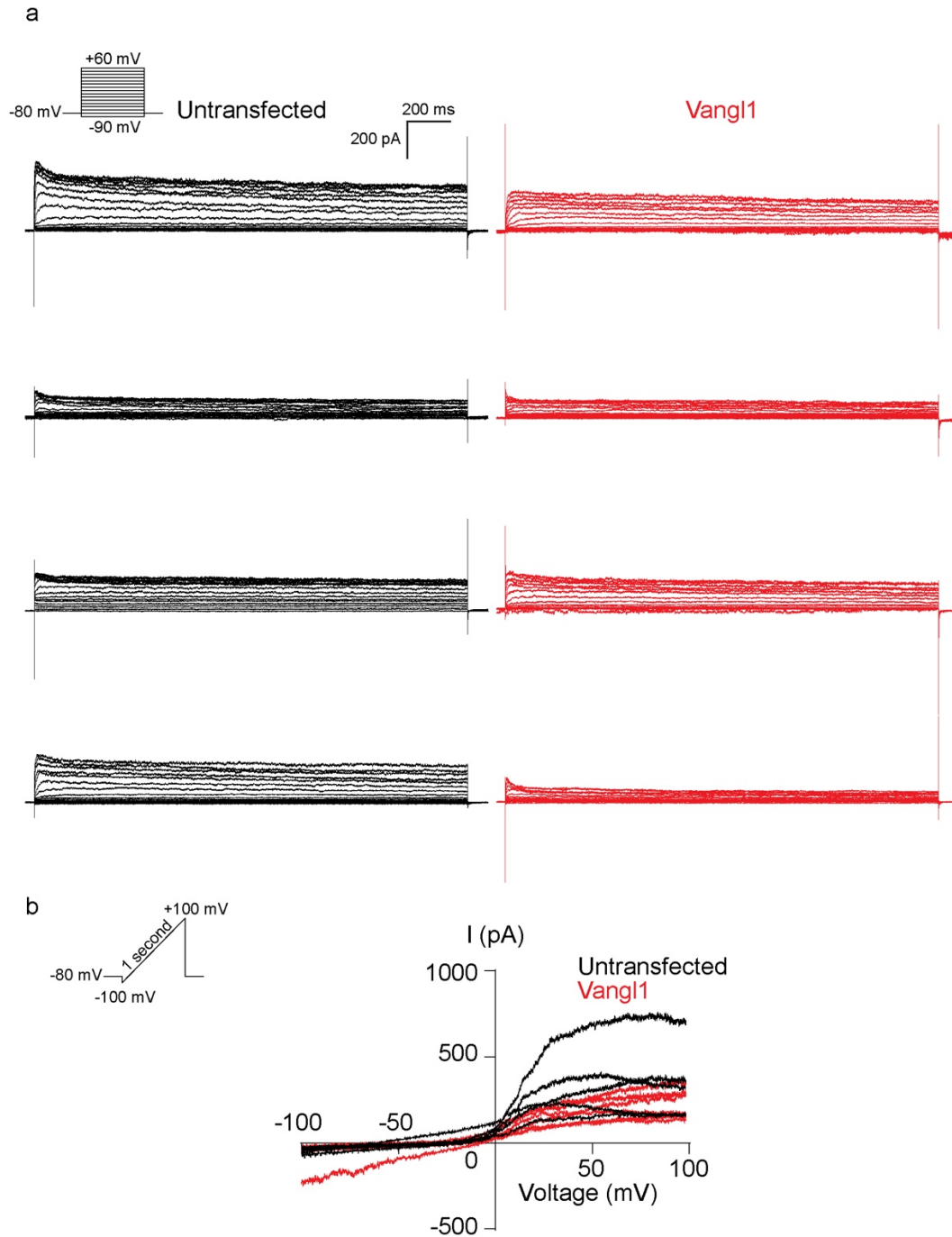
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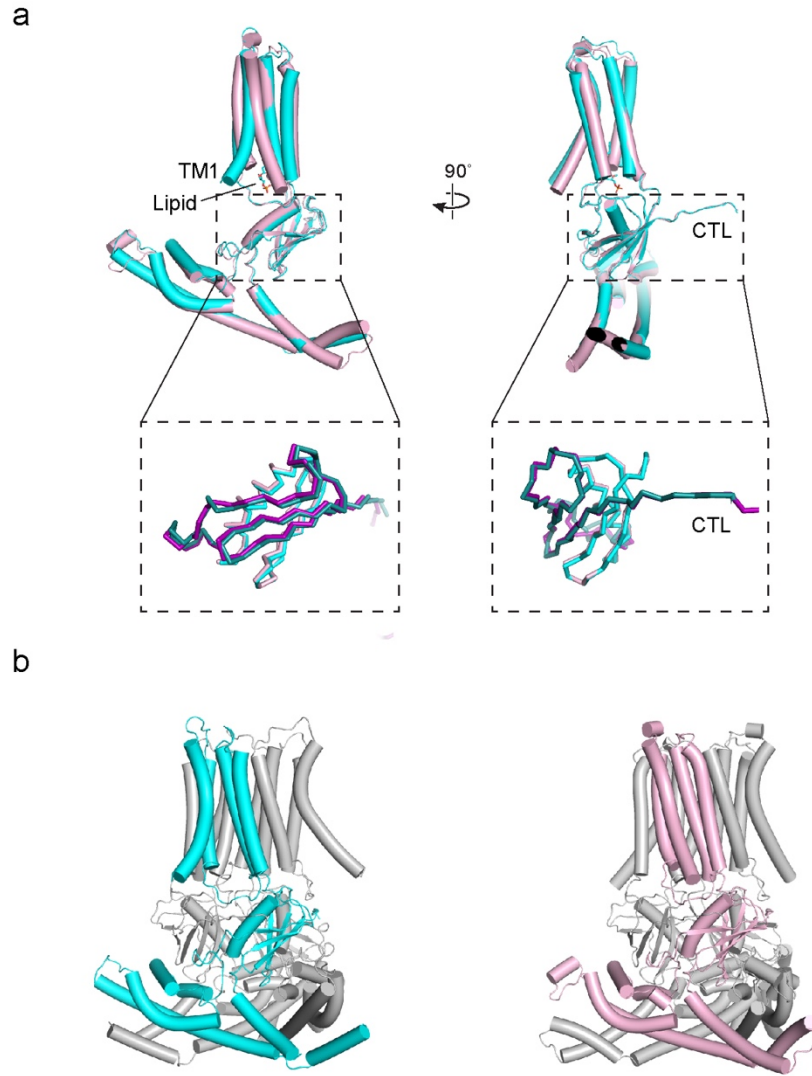
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Supplementary Figure 8 | Oligomerization equilibrium of Vangl1. **a**, Blue native PAGE of wild-type Vangl1, Vangl2 and Vangl1 mutants showing the presence of hexmers and trimers. Purification and native PAGE analysis were reproducibly replicated three times. **b**, FSEC profiles of wildtype (WT) Vangl1 in black, Vangl1 with two point mutations (H340A/Y345A) in red and Vangl1 with three point mutations (H340A/Y345A/E349A) in blue, showing mutations introduced at the trimer-trimer interface promoted the presence of trimers. Overexpression and FSEC analysis were reproducibly replicated six times. **c**, The same Vangl1 mutations render compromised ability of pulling down Pk1.



Supplementary Figure 9 | Electrophysiological recordings consistent with the notion that Vangl1 is unlikely to be an ion channel. a-b, Current-voltage relationship was recorded using untransfected HEK293T cells (black) or Vangl1-expressing HEK293T cells (red), which were held at -80 mV before stimulated by a series of voltage steps (a) or a voltage ramp (b).



Supplementary Figure 10 | Comparison of the structures of Vangl1 determined by cryo-EM (cyan) and predicted by AlphaFold (light pink). a, Comparison of a protomer revealed consistent overall architecture including the TM domain, the split hand domain, the CTL (inset) and the stick domain, but the kinked TM1 adopts different configuration. **b,** The trimerization of Vangl can be accurately predicted.

Supplementary Table 1 | Statistics of the cryo-EM data collection, refinement and validation.

Statistics for data collection and structural refinement	
	Human Vangl1 PDB 8ZXD EMDB 60540
Data collection and processing	
Microscope	FEI Arctica
Magnification	81000
Voltage (kV)	200
Detector	Falcon 4
Electron exposure (e ⁻ /Å ²)	50
Defocus range (μm)	1.5 to 2.5
Pixel size (Å)	1.2
Symmetry imposed	D3
Initial particle images (no.)	718,127
Final particle images (no.)	192,766
Map resolution (Å)	2.9
FSC threshold	0.143
Map resolution range (Å)	2.7-3.9
Refinement	
Initial model used	cryoSPARC <i>Ab-initio</i>
Model resolution (Å)	3.4
FSC threshold	0.5
Map sharpening <i>B</i> factors (Å ²)	-80
Model composition	
Non-hydrogen atoms	19518
Protein residues	2328
<i>B</i> factors (Å ²)	
protein	175.03
Ligand	194.48
R.m.s. deviations	
Bond lengths (Å)	0.001
Bond angles (°)	0.403
Validation	
MolProbity score	1.29
Clash score	5.16
Poor rotamers (%)	0
Ramachandran plot	
Favored (%)	97.92
Allowed (%)	2.08
Disallowed (%)	0.00

Supplementary Table 2 | Structural mapping of human disease-associated Vangl mutations.

Diseases/phenotype	Vangl subtype	Mutation Sites (ref.)	Structural location
Neural tube defects	hVangl1	G25K ⁵⁶	N-terminal disordered region
	hVangl1	S83L ⁵⁶	N-terminal disordered region
	hVangl1	A116T ⁵⁴	Transmembrane domain TM1
	hVangl1	F153S ⁵⁶	Transmembrane domain TM2
	hVangl1	R173H ⁵⁷	Intracellular loop
	hVangl1	R175Q ⁵⁶	Intracellular loop
	hVangl1	R181Q ⁵⁶	Intracellular loop
	hVangl1	R186H ⁵⁷	Transmembrane domain TM3
	hVangl1	A187V ⁵⁸	Transmembrane domain TM3
	hVangl1	L202F ⁵⁶	Transmembrane domain TM3
	hVangl1	G205R ⁵⁷	Transmembrane domain TM3
	hVangl1	R207H ⁵⁸	Extracellular Loop 2
	hVangl1	V239I/F ³²	Transmembrane domain TM4
	hVangl1	T251M ⁵⁶	Hand domain β_1
	hVangl1	R274Q ³²	Hand domain α_1
	hVangl1	Y290H ⁵⁶	Loop region between α_1 and α_2
	hVangl1	M328T ³²	Stick domain α_{3a}
	hVangl1	H350H ⁵⁸	Stick domain α_{3b}
	hVangl1	K356K ⁵⁸	Stick domain α_{3b}
	hVangl1	R374H ⁶²	Stick domain α_{3b}
	hVangl1	P384R ⁸⁰	Stick domain $\alpha_{3b} - \alpha_4$
	hVangl1	D389H ⁵⁸	Stick domain $\alpha_{3b} - \alpha_4$
	hVangl1	A404S ⁵⁶	Stick domain α_4
	hVangl1	D468E ⁵⁶	Hand domain $\beta_3 - \beta_4$
	hVangl1	R517H ⁵⁸	C-terminal disordered region
	hVangl2	S84F ³³	N-terminal disordered region
	hVangl2	R105C ⁵⁹	Transmembrane domain TM1
	hVangl2	R135W ⁵⁹	Extracellular Loop 1
	hVangl2	R177H ⁵⁹	Intracellular loop
	hVangl2	V178I ⁵⁹	Intracellular loop
	hVangl2	L242V ⁵⁹	Transmembrane domain TM4
	hVangl2	T247M ⁵⁹	Hand domain β_1
	hVangl2	R270H ⁵⁹	Hand domain α_1
	hVangl2	R353C ³³	Stick domain α_{3b}
	hVangl2	V363E ⁶²	Stick domain α_{3b}
	hVangl2	F437S ³³	Stick domain α_6
	hVangl2	S464R ⁶²	Hand domain $\beta_3 - \beta_4$
	hVangl2	R482H ⁵⁹	Hand domain β_4
Adolescent idiopathic scoliosis	hVangl1	I136N ⁵⁵	Transmembrane domain TM1
Sacral agenesis Klippel-Feil syndrome Congenital anomalies of the kidney and urinary tract	hVangl1	F440V ⁵⁵	Stick domain α_6
	hVangl1	S338Term ⁸¹	Stick domain $\alpha_{3a} - \alpha_{3b}$
	hVangl1	P384R ⁸⁰	Stick domain $\alpha_{3b} - \alpha_4$
	hVangl2	E70K ⁸²	N-terminal disordered region
Congenital scoliosis	hVangl2	T433M ⁸²	Stick domain $\alpha_5 - \alpha_6$
	hVangl1	R256H ⁶⁰	Hand domain β_2
	hVangl1	E281G ⁶⁰	Hand domain α_2
	hVangl1	R443W ⁶⁰	Stick domain α_6
Heterotaxy and congenital heart disease	hVangl2	L226F ⁶⁰	Transmembrane domain TM4
	hVangl2	R169H ³⁵	Intracellular loop
Non-syndromic hearing loss	hVangl2	E465A ³⁵	Hand domain $\beta_3 - \beta_4$
Congenital heart and brain defects Looptail	hVangl2	R135W ³⁵	Extracellular Loop 1
	mVangl2	D255E ³⁶	Hand domain $\beta_1 - \beta_2$
	mVangl2	R259L ⁸³	Hand domain β_2
	mVangl2	S464N ³⁶	Hand domain $\beta_3 - \beta_4$
Curly Bob	mVangl2	I268N ⁶¹	Hand domain α_1
NTD-associated mammalian mutations found in <i>Drosophila</i> Vang	dVang	D317E ³⁴	Hand domain $\beta_1 - \beta_2$
	dVang	R321L ³⁴	Hand domain β_2
	dVang	R332H ³⁴	Hand domain α_1
	dVang	V391T ³⁴	Stick domain α_{3a}
	dVang	K418C ³⁴	Stick domain α_{3b}
	dVang	K577H ³⁴	C-terminal disordered region

Uncropped images of Western-blots and gels

