

Supplementary Information

An essential role for TASL in mouse autoimmune pathogenesis and Toll-like receptor signaling

Laura Lau^{1,2}, Taryn A. Cariaga^{1,2}, Abraham B. Chang^{1,3}, Joan H. Lane¹, Whitney E. Purtha¹, Aaron S. Rapaport¹, Ruozhen Hu^{1,4}, Hiroyasu Konno¹, Daryl N. Bulloch¹, Matthew J. Rardin¹, Bradford W. Gibson¹, Jason Devoss¹, Wenjun Ouyang^{1,2}, Paolo S. Manzanillo^{1,2*}

¹Amgen Research, Amgen Inc., 720 Gateway Blvd, South San Francisco, California 94080, USA

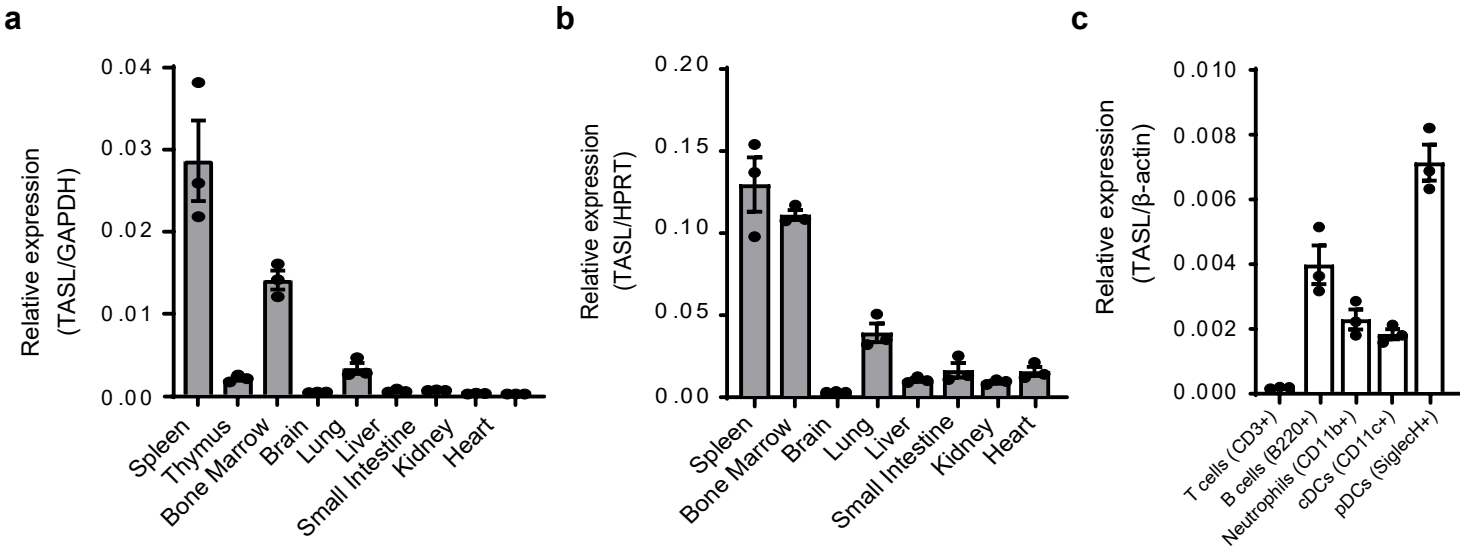
²Current address Gilead Inc, 333 Lakeside Dr, Foster City, California 94404, USA

³Current address Exelixis Inc, 1851 Harbor Bay Pkwy, Alameda, CA 94502, USA

⁴Current address, Genentech Inc, 1 DNA Way, South San Francisco, California 94080, USA

*Corresponding author: paolomanzanillo550@gmail.com (P.S.M.)

Supplementary Fig. 1



d Target Guide Design

Target sequence sgRNA 1 PAM

5' - AAAATTATAAGAAGACGCTGTGG - 3'

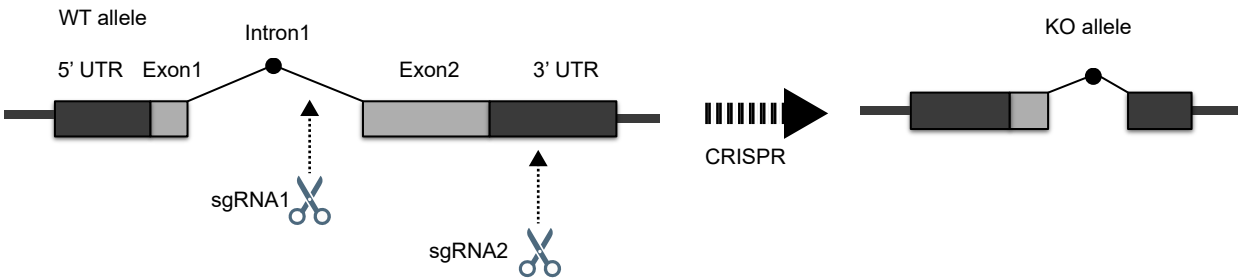
3' - TTTTAATATTCTTCTGCGAC - 5'

Target sequence sgRNA 2 PAM

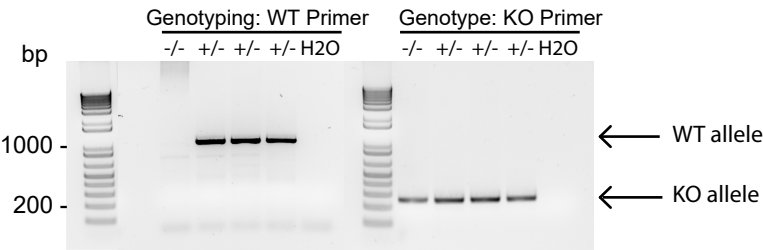
5' - AAGGCCTACCATTCCAGTGCAGG - 3'

3' - TTCCGGATGGTAAGGTCACG - 5'

e Targeting Schematic

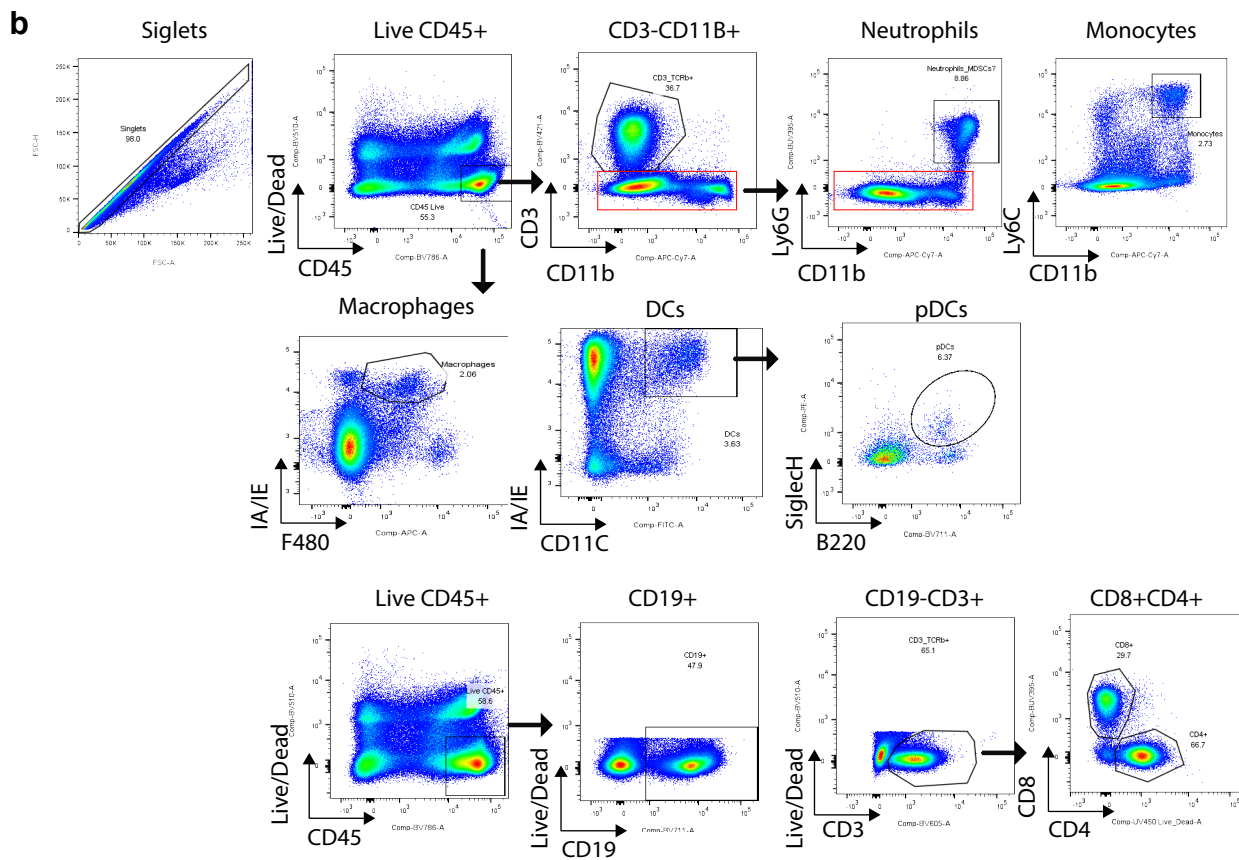
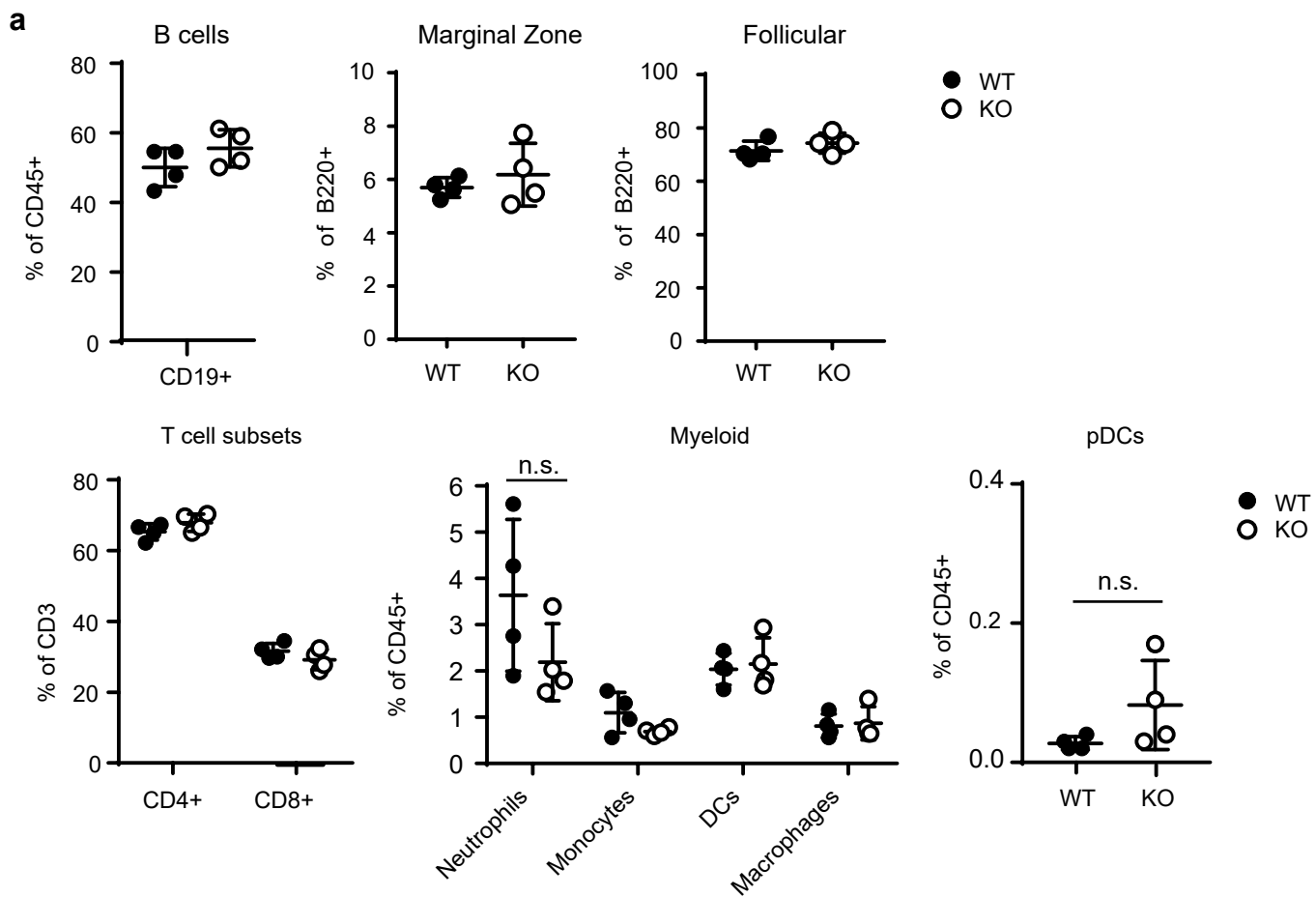


f Genotyping



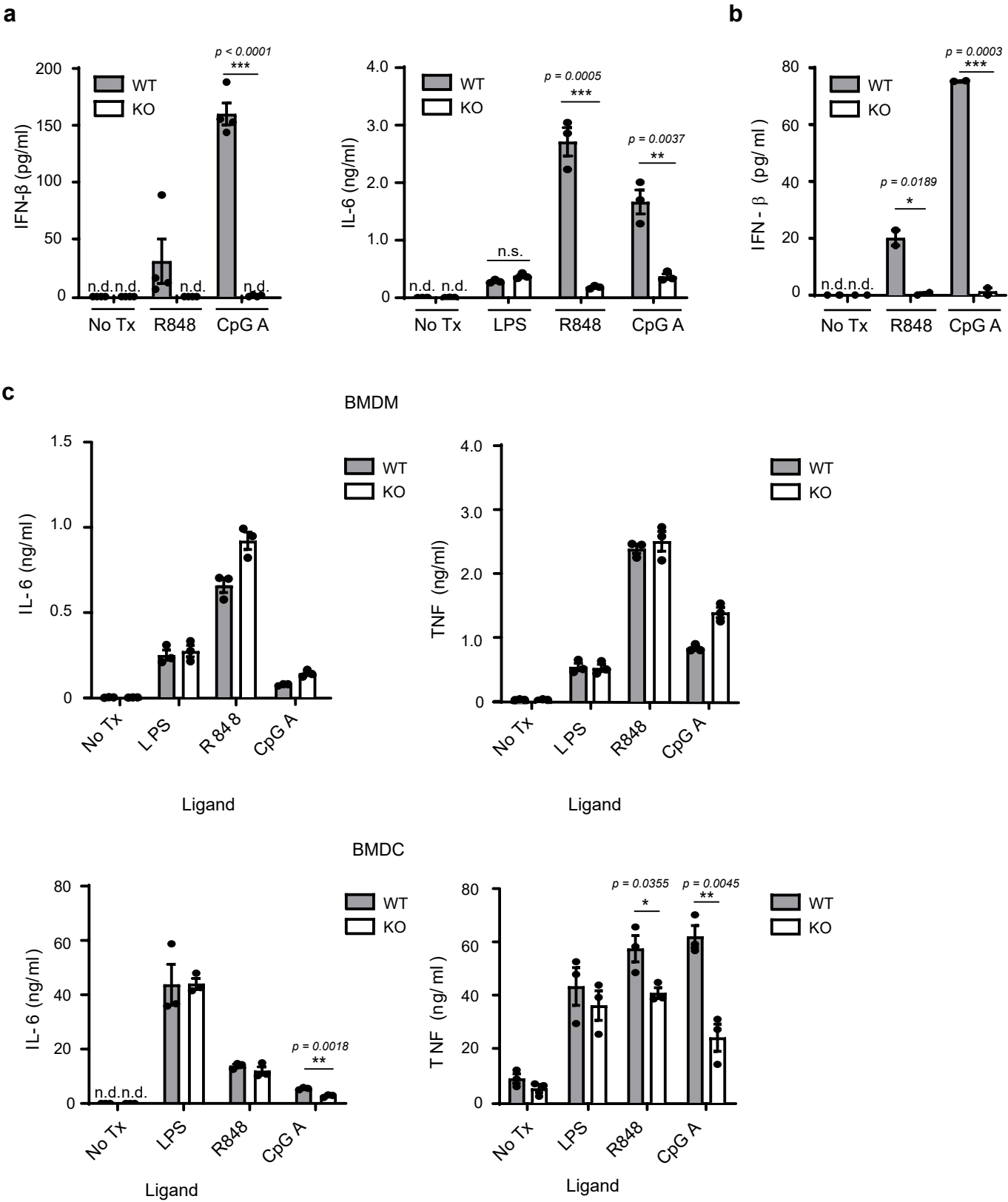
Supplementary Figure 1. *TASL* expression and knockout mouse strategy. RT-qPCR for *TASL* expression in (a) indicated human tissues. (b) indicated mouse tissues. (c) RT-qPCR analysis of *Tasl* expression from various immune cells sorted from mouse (n=3) spleens. (d). Target sequence (red) and PAM site (blue) used to generate sgRNA oligos against *Tasl* loci. (e). Schematic representation of the generation of *Tasl*^{-/-} mice illustrating sgRNA1 targeting the intronic region and sgRNA2 targeting 3'UTR region for removal of exon 2 of *Tasl*. (f) Agarose gel depicting genotyping results from DNA samples obtained from knockout mice (-/-) or heterozygote mice (+/-). Arrows indicate presence of wild type (WT) or knockout (KO) specific alleles. Samples loaded with H₂O was used as a negative control. In (a-c), data are means ± SEM. In (a-c) and (f) data are representative of 2 independent experiments.

Supplementary Figure 2



Supplementary Figure 2. Immune cell populations in *Tas^{fl/-}* mice. (a) FACS analysis of spleens from wildtype (WT) and *Tas^{fl/-}* (KO) mice was performed and gated on CD45+(immune cells) or CD3+ (T cells) positive cells. Shown are percentage of B cells, marginal zone B cells, follicular B cells, T cells, myeloid cell populations, and pDCs. Data are means $\pm$ SEM and are representative of 2 independent experiments with 4 mice per genotype individually represented. (b) Representative FACS gating strategy for defined populations represented in (a).

Supplementary Figure 3



Supplementary Figure 3. Cytokine responses from *Tas1^{-/-}* mice. IFN- β and IL-6 levels from supernatants of (a) total mouse splenocytes (b) total mouse marrow cells stimulated with either LPS (5 μ g/ml), R848 (10 μ g/ml) or CpG A (3 μ M) overnight. (c) M-CSF bone marrow-derived macrophages (BMDM) and GM-CSF conventional dendritic cells (BMDC) isolated from wildtype (WT) and *Tas1^{-/-}* (KO) mice were stimulated with LPS (500ng/ml), R848 (1 μ g/ml) and CpG A (10 μ g/ml) and IL-6 and TNF were measured 24 hours later by CBA. Representative of 2 independent experiments. Data are means $\pm$ SEM and are representative of at least 2 independent experiments, 5 mice per genotype were pooled per experiment. Unpaired t- test (2-paired) *** $P < 0.0005$; ** $P < 0.005$; * $P < 0.05$, n.d. not detected.

a

Gene	WT	KO
IFIT1	~0.03	~0.03
ISG15	~0.16	~0.16
Mx1	~0.07	~0.06

b

c

Time (minutes)	WT	KO
0	~25000	~25000
60	~45000	~45000
120	~42000	~40000
180	~42000	~40000
240	~42000	~40000

d

e

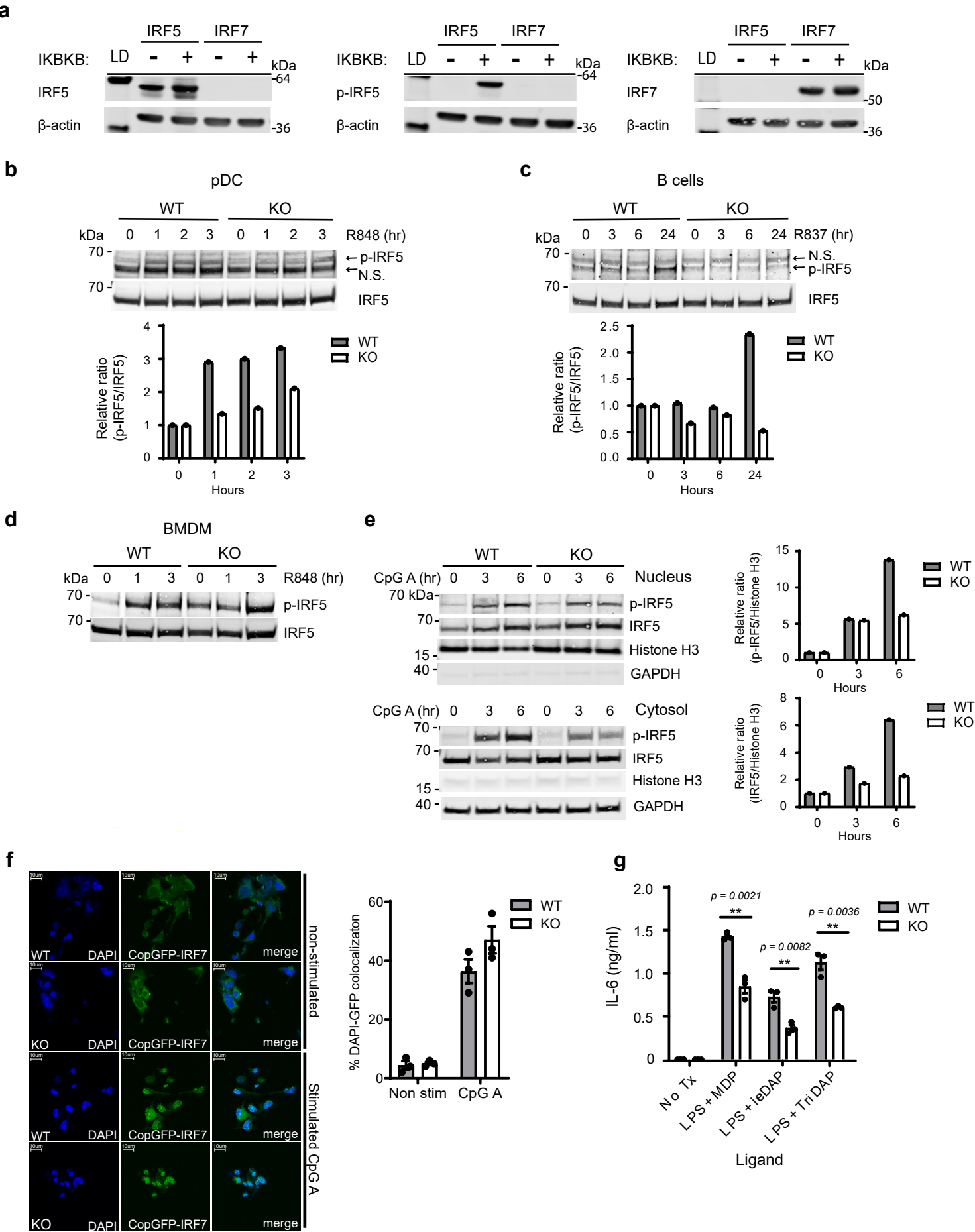
f

g

h

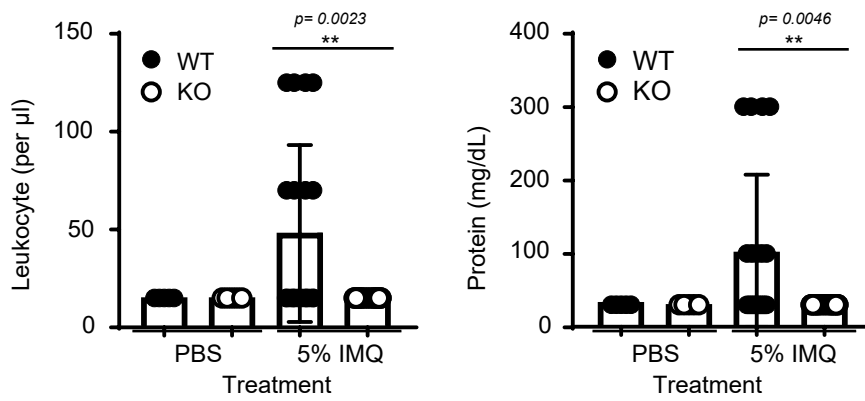
Supplementary Figure 4. TLR7/9 expression and signaling pathways in *Tas1^{-/-}* mice. (a) RT-qPCR analysis of ISG expression of pDCs isolated from wildtype (WT) or *Tas1^{-/-}* (KO) spleens and treated with Type I IFN (500U) for 5 hours. (b) Western blot analysis of pSTAT1 and STAT1 in splenic pDCs with Type I IFN for 30 mins. (c) WT or KO Flt3L pDCs treated with LysoTracker were stimulated with CpG A (3μM) for the indicated time periods and fluorescence was analyzed by flow cytometry. (d) RT-qPCR analysis for TLR7 and TLR9 expression in spleens from wild type (WT) or *Tas1^{-/-}* (KO) mice stimulated with R848 (10μg/ml) or CpG A (μM) (n=3, biological replicates). (e) Endogenous TLR9 from Flt3L derived pDCs from wild type (WT) or *Tas1^{-/-}* (KO) mice was immunoprecipitated and lysates were subjected to western blot analysis with TLR9 antibody. β-actin in whole cell lysate (WCL) was detected as internal control. (f) Immunoblots of p-JNK and JNK (g) p-p38 and p38 and (wild type and knockout Flt3L pDCs stimulated with R848 (10μg/ml) for the indicated time periods (0-3hr). Representative of at least 2 independent experiments, 5 mice per genotype were pooled per experiment. (h) Immunoblots of plkBα, IκBα, pSTAT1, and STAT1 in CpG A (3μM) stimulated Flt3L pDCs. In (a) and (c-d), data are means ± SEM and in (a-c), and (e-h), 5 mice per genotype were pooled per experiment. All data are representative of at least 2 independent experiments.

Supplementary Figure 5



Supplementary Figure 5. IRF5 expression in *Tas1^{-/-}* mice. (a) Western blot analysis of HEK293T cells transfected in combination with vectors encoding mouse *Irf5*, *Irf7*, and *Ikbbk*. L.D. indicates protein ladder. Beta actin was used as a loading control. Western blot analysis of p-IRF5 and total IRF5 in (b) Western blot analysis of pIRF5 and IRF5 in Flt3L pDCs stimulated with R848 (10µg/ml) for the indicated time periods (0-3hr). N.S. indicates a non-specific band. Bottom panel indicates quantification of p-IRF5 band normalized to total IRF5 protein band and representative of two independent experiments. (c) Western blot analysis of p-IRF5 and IRF5 in splenic B cells stimulated with R837 (10µg/ml) for the indicated time periods (0-24hr). N.S. indicates a non-specific band. Bottom panel indicates quantification of p-IRF5 band from two independent experiments normalized to total IRF5 protein band. (d) Western blot analysis of pIRF5 and IRF5 in M-CSF derived BMDMs stimulated with R848 (1µg/ml) for the indicated time periods (0-3hr). (e) Western blot analysis of cytosolic and nuclear fractions obtained from Flt3L pDCs stimulated with CpG A (3µM) for the indicated time periods (0-6hrs). Graphs indicate quantification of p-IRF5 or IRF5 bands normalized to Histone H3 protein band. Quantification is normalized to zero time point and is representative of two independent experiments. (f) Microscopy image of Flt3L pDCs transfected with mRNA encoding CopGFP-IRF7(Green) and stimulated with CpG A (3µM) for 6 hrs followed by DAPI staining for nuclei (Blue). Data plotted is the average percent colocalization of CopGFP-IRF7 and nuclei from three cover slips and is representative of two independent experiments. (g) Flt3L derived pDCs were stimulated with LPS (5ng/ml) and the indicated NOD1/2 ligands (10µg/ml). IL-6 secretion was measured 24hrs post stimulation (n=5 pooled, 3 replicates per condition) Data are means ± SEM. p-values; unpaired *t*-test (2-tailed), ***P*<0.005.

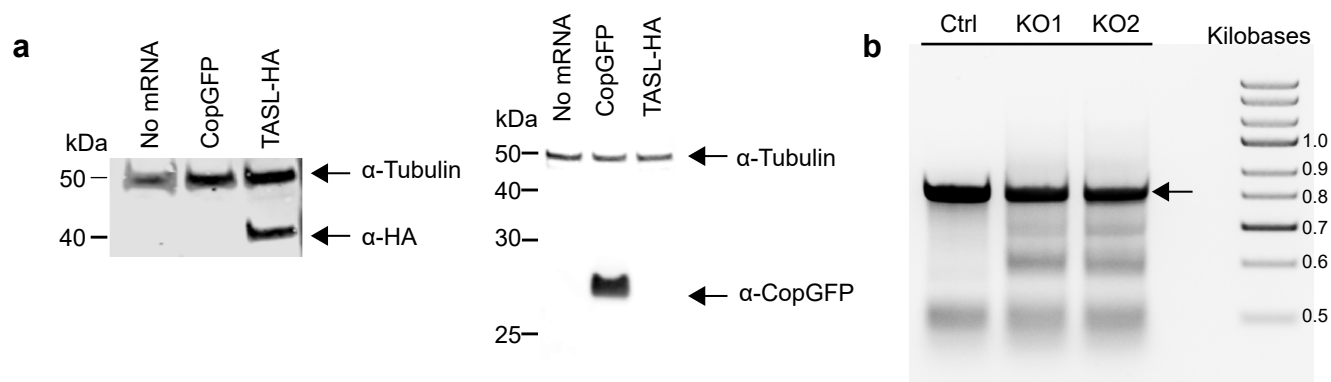
Supplementary Figure 6



Supplementary Figure 6. *Tas1^{-/-}* mice are resistant to Aldara model of SLE.

Urinalysis of leukocytes and protein levels from wild type (WT) and *Tas1^{-/-}* (KO) mice that were treated with either PBS (n= 5) or 5% imiquimod (IMQ; Aldara) (n = 20) for 8 weeks. Data are means $\pm$ SEM and representative of 2 independent experiments. Unpaired t- test (2 tailed) $**P < 0.005$.

Supplementary Fig. 7



Supplementary Figure 7. *TASL* mRNA and CRISPR Analysis. (a) HEK293T cells were transfected with in vitro synthesized mRNA encoding either CopGFP or TASL-HA. Western blot analysis was performed using anti-tubulin, anti-HA, and anti-CopGFP antibodies (b) Surveyor mutation analysis of *TASL* loci in BLCL line transfected with Cas9 complexed with either control gRNAs (ctrl) or gRNAs targeting *TASL* (KO1, KO2). Arrow indicates full length *TASL* product.

Supplementary Table 1 Source and catalog numbers of BLCL lines used in this study

Catalog Number	Description	Product	SEX
HG01500	1000 GENOMES PROJECT - IBERIAN POPULATIONS IN SPAIN	LCL from B-Lymphocyte	M
HG01606	1000 GENOMES PROJECT - IBERIAN POPULATIONS IN SPAIN	LCL from B-Lymphocyte	M
HG01625	1000 GENOMES PROJECT - IBERIAN POPULATIONS IN SPAIN	LCL from B-Lymphocyte	M
HG01669	1000 GENOMES PROJECT - IBERIAN POPULATIONS IN SPAIN	LCL from B-Lymphocyte	M
HG01682	1000 GENOMES PROJECT - IBERIAN POPULATIONS IN SPAIN	LCL from B-Lymphocyte	M
HG01756	1000 GENOMES PROJECT - IBERIAN POPULATIONS IN SPAIN	LCL from B-Lymphocyte	M
HG01761	1000 GENOMES PROJECT - IBERIAN POPULATIONS IN SPAIN	LCL from B-Lymphocyte	M
HG02224	1000 GENOMES PROJECT - IBERIAN POPULATIONS IN SPAIN	LCL from B-Lymphocyte	M
HG02231	1000 GENOMES PROJECT - IBERIAN POPULATIONS IN SPAIN	LCL from B-Lymphocyte	M
HG02238	1000 GENOMES PROJECT - IBERIAN POPULATIONS IN SPAIN	LCL from B-Lymphocyte	M
HG01503	1000 GENOMES PROJECT - IBERIAN POPULATIONS IN SPAIN	LCL from B-Lymphocyte	M
HG01608	1000 GENOMES PROJECT - IBERIAN POPULATIONS IN SPAIN	LCL from B-Lymphocyte	M
HG01624	1000 GENOMES PROJECT - IBERIAN POPULATIONS IN SPAIN	LCL from B-Lymphocyte	M
HG01672	1000 GENOMES PROJECT - IBERIAN POPULATIONS IN SPAIN	LCL from B-Lymphocyte	M
HG01680	1000 GENOMES PROJECT - IBERIAN POPULATIONS IN SPAIN	LCL from B-Lymphocyte	M
HG01765	1000 GENOMES PROJECT - IBERIAN POPULATIONS IN SPAIN	LCL from B-Lymphocyte	M
HG01767	1000 GENOMES PROJECT - IBERIAN POPULATIONS IN SPAIN	LCL from B-Lymphocyte	M
HG02221	1000 GENOMES PROJECT - IBERIAN POPULATIONS IN SPAIN	LCL from B-Lymphocyte	M
HG02233	1000 GENOMES PROJECT - IBERIAN POPULATIONS IN SPAIN	LCL from B-Lymphocyte	M
HG02236	1000 GENOMES PROJECT - IBERIAN POPULATIONS IN SPAIN	LCL from B-Lymphocyte	M
HG01786	1000 GENOMES PROJECT - IBERIAN POPULATIONS IN SPAIN	LCL from B-Lymphocyte	F
HG01746	1000 GENOMES PROJECT - IBERIAN POPULATIONS IN SPAIN	LCL from B-Lymphocyte	F
HG01702	1000 GENOMES PROJECT - IBERIAN POPULATIONS IN SPAIN	LCL from B-Lymphocyte	F
HG01628	1000 GENOMES PROJECT - IBERIAN POPULATIONS IN SPAIN	LCL from B-Lymphocyte	F
HG01784	1000 GENOMES PROJECT - IBERIAN POPULATIONS IN SPAIN	LCL from B-Lymphocyte	F
HG01757	1000 GENOMES PROJECT - IBERIAN POPULATIONS IN SPAIN	LCL from B-Lymphocyte	F
HG01704	1000 GENOMES PROJECT - IBERIAN POPULATIONS IN SPAIN	LCL from B-Lymphocyte	F
HG01632	1000 GENOMES PROJECT - IBERIAN POPULATIONS IN SPAIN	LCL from B-Lymphocyte	F
HG00111	1000 GENOMES PROJECT - BRITISH FROM ENGLAND AND SCOTLAND, UK	LCL from B-Lymphocyte	F
HG00102	1000 GENOMES PROJECT - BRITISH FROM ENGLAND AND SCOTLAND, UK	LCL from B-Lymphocyte	F
GM20795	1000 GENOMES PROJECT - PANEL OF 114 TOSCANI IN ITALIA	LCL from B-Lymphocyte	F
GM20799	1000 GENOMES PROJECT - PANEL OF 114 TOSCANI IN ITALIA	LCL from B-Lymphocyte	F
GM20832	1000 GENOMES PROJECT - PANEL OF 114 TOSCANI IN ITALIA	LCL from B-Lymphocyte	F
GM20804	1000 GENOMES PROJECT - PANEL OF 114 TOSCANI IN ITALIA	LCL from B-Lymphocyte	F
GM20797	1000 GENOMES PROJECT - PANEL OF 114 TOSCANI IN ITALIA	LCL from B-Lymphocyte	F
GM20822	1000 GENOMES PROJECT - PANEL OF 114 TOSCANI IN ITALIA	LCL from B-Lymphocyte	F
GM12006	1000 GENOMES PROJECT-CEPH	LCL from B-Lymphocyte	F
GM12761	1000 GENOMES PROJECT-CEPH	LCL from B-Lymphocyte	F
GM11994	1000 GENOMES PROJECT-CEPH	LCL from B-Lymphocyte	F
GM12004	1000 GENOMES PROJECT-CEPH	LCL from B-Lymphocyte	F
GM12763	1000 GENOMES PROJECT-CEPH	LCL from B-Lymphocyte	F
GM11993	1000 GENOMES PROJECT-CEPH	LCL from B-Lymphocyte	F