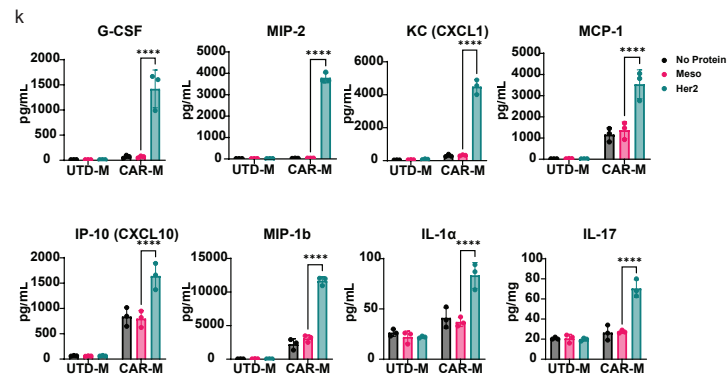
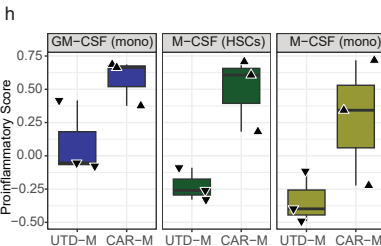
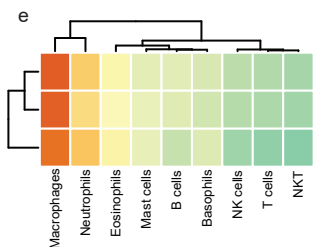


Extended Table 1. Antibody panel used to stain tumor and immune cells for IONpath multiplex ion beam imaging (MIBI)

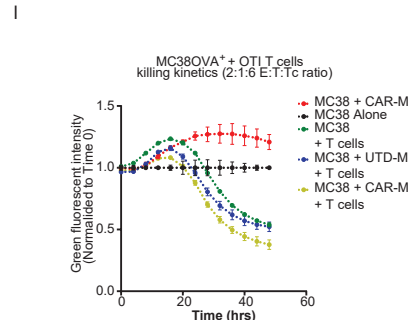
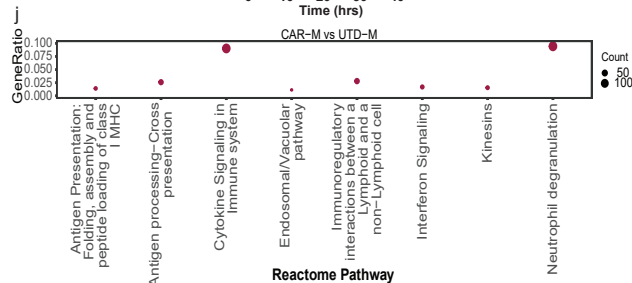
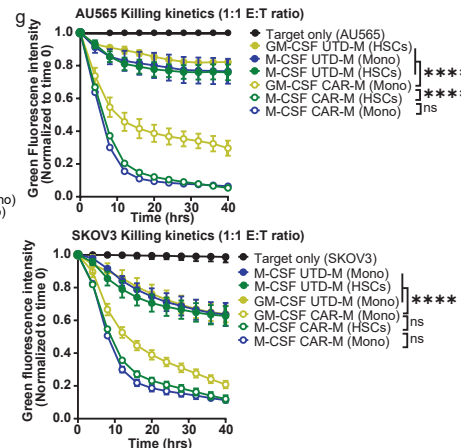
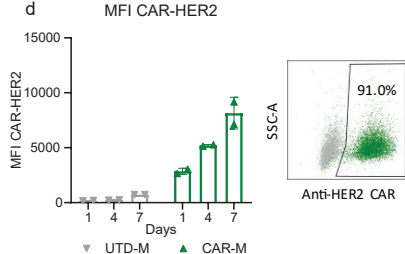
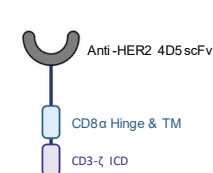
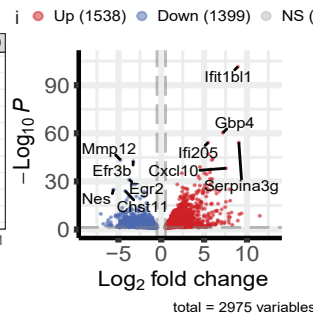
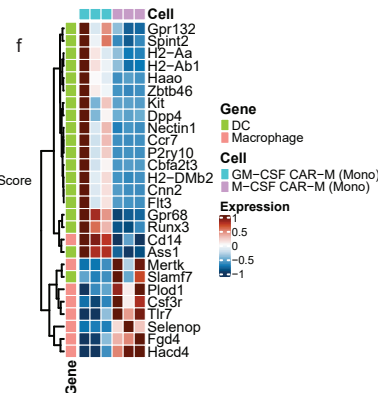
#	Antibody	Clone	Metal
1	dsDNA	35I9 DNA	89Y
2	beta-tubulin	D3U1W	113In
3	CD4	766	143Nd
4	CD11c	D1V9Y	144Nd
5	FOXP3	FJK-16s	147Sm
6	Granzyme B	EPR22645-206	150Nd
7	CD49b	EPR17338	151Eu
8	CD31	D8V9E	152Sm
9	Ki-67	SP6	153Eu
10	CD11b	EPR1344	155Gd
11	F4/80	D2S9R	156Gd
12	CD8	CAL38	158Gd
13	CD3e	EPR22667-12	159Tb
14	Vimentin	D21H3	163Dy
15	SMA	D4K9N	164Dy
16	PAX5	D7H5X	165Ho
17	Ly6G	1A8	173Yb
18	CD206	E2L9N	174Yb
19	CD45	D3F8Q	175 Lu
20	Na-K-ATPase	EP1845Y	176Yb

Extended Table 2. Immune cell populations identified by IONpath multiplex ion beam imaging (MIBI)

#	Classified cell populations	Markers
1	Tumor cells & Fibroblast	Vimentin+, SMA- CD45-
2	Myofibroblasts	Vimentin+, SMA+
3	Immune cells	CD45+, CD3+, CD11b+, CD11c+, F4/80+, Ly6G+, PAX5+
4	Proliferating immune cells	CD45+, CD3+, CD11b+, CD11c+, F4/80+, Ly6G+, PAX5+, Ki-67+
5	T cells	CD45+, CD3+
6	Helper T cells	CD45+, CD3+, CD4+, FOXP3-
7	Activated Helper T cells	CD45+, CD3+, CD4+, FOXP3-, Granzyme B+
8	Cytotoxic T cells	CD45+, CD3+, CD8+
9	Activated Cytotoxic T cells	CD45+, CD3+, CD8+, Granzyme B+
10	Regulatory T cells	CD45+, CD3+, FOXP3+
11	CD4+ Regulatory T cells	CD45+, CD3+, CD4+, FOXP3+
12	Natural Killer cells	CD45+, CD3-, CD49b+
13	Activated Natural Killer cells	CD45+, CD3-, CD49b+, Granzyme B+
14	B cells	CD45+, PAX5+
15	Macrophages	CD45+, F4/80+
16	M1 Macrophages	CD45+, F4/80+, CD206-
17	M2 Macrophages	CD45+, F4/80+, CD206+
18	Dendritic cells	CD45+, CD11c+
19	CD8+ Dendritic cells	CD45+, CD11c+, CD8+, CD11b-
20	CD11b+ Dendritic cells	CD45+, CD11c+, CD8-, CD11b+
21	Other myeloid cells	CD45+, CD11b+, CD11c-, F4-80-
22	Granulocytes/PMN-MDSC	CD45+, CD11b+, CD11c-, F4-80-, Ly6G+
23	Monocytes/M-MDSC	CD45+, CD11b+, CD11c-, F4-80-, Ly6G-

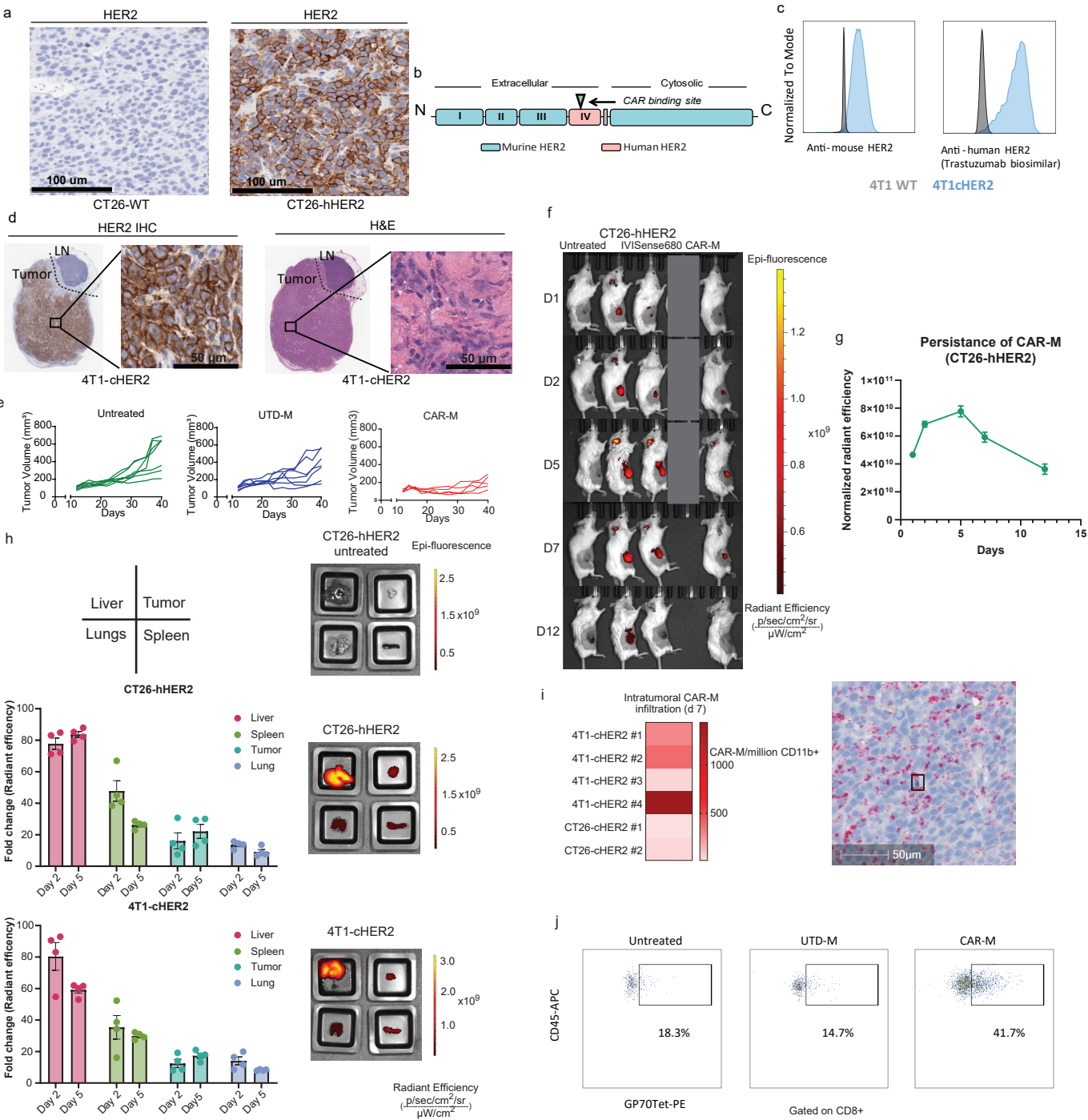


% of CD45+	M-CSF UTD-M (Mono)	GM-CSF UTD-M (Mono)	M-CSF UTD-M (HSCs)
CD11b+ CD14+	99.3 $\pm$ 0.1	91.2 $\pm$ 0.9	94.1 $\pm$ 0.1
Ly6G+	0.1 $\pm$ 0	2.7 $\pm$ 0.5	2.1 $\pm$ 0
CD3+	0.0 $\pm$ 0	0.0 $\pm$ 0	0.0 $\pm$ 0
CD19+	0.0 $\pm$ 0	0.0 $\pm$ 0	0.0 $\pm$ 0
CD49b+	0.0 $\pm$ 0	0.0 $\pm$ 0	0.0 $\pm$ 0



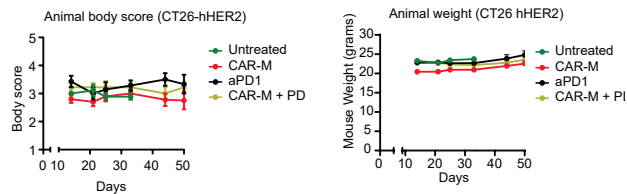
Supplementary Fig. 1 Generation of murine CAR-Ms and assessment of CAR-mediated tumor cytotoxicity

a, BMDM yield. Data is represented with a truncated violin plot showing all points ($n=5-9$ biological replicate). Statistical analysis was calculated using the ordinary one-way ANOVA. **b**, Purity evaluated by flow cytometry. Data are represented as the mean $\pm$ SD of $n=2$ biological replicate. **c**, Diagram of the anti-HER2 CAR construct engineered into the Ad5f35 vector under the control of a CMV promoter. Created in BioRender. Minutolo, N. (2025) <https://BioRender.com/i18y106>. **d**, Anti-HER2 CAR MFI represented as mean $\pm$ SD of $n=2$ technical replicate and representative FACS plot of anti-HER2 CAR expression in M-CSF UTD-M (HSCs). **e**, SingleR prediction analysis assign reference populations to HSC, monocytes, and the three CAR-M cultures ($n=3$ biological replicate). **f**, DESeq2⁷⁰ was used to identify significant differentially expressed DC and macrophage genes between GM-CSF CAR-M (mono) and M-CSF CAR-M (mono) ($n=3$ biological replicate). **g**, Incucyte-based killing assays of UTD-M or CAR-M cocultured with SKOV3 NLG or AU565 NLG at E:T, 1:1 for 40 hr. Green fluorescence intensity (GFI) values were normalized to the 0 hr. timepoint and then, further normalized to the target cells. Data is represented as the mean $\pm$ SD of $n=4$ technical replicate, and graph is representative of at least three experiments. Ordinary one-way ANOVA was performed at 40 hr. **h**, The pro-inflammatory phenotype was assessed using the proinflammatory score. Data is represented as the mean with Min to Max whiskers of $n=3$ biological replicate. **i**, Volcano plot of M-CSF UTD-M (HSC) vs M-CSF CAR-M (HSC) response gene. Red denotes significantly upregulated genes, and blue indicates significantly downregulated genes ($n=3$ biological replicate). Statistical significance was calculated using DESeq2. **j**, Pathway analysis and gene expression in M-CSF UTD-M (HSC) vs M-CSF CAR-M (HSC) ($n=3$ biological replicate). **k**, CAR-M specificity in response to no protein, mesothelin (Meso; irrelevant antigen control) and HER2. Data are represented as the mean $\pm$ SD of $n=3$ biological replicate. Statistical analysis was performed using the two-way ANOVA Tukey's multiple comparisons test. **l**, Killing kinetics of UTD-M and CAR-M cocultured with MC38OVA⁺ (HER2 negative) and transgenic CD8⁺ T cells (OT-1), at E:T:T (Effector:Target:T cells), 2:1:6. GFI values were normalized to the 0 hr. timepoint and then, further normalized to the target cells. Data represents the mean $\pm$ SEM of $n=3$ technical replicates and is representative of two experiments. For all images: * $p<0.05$, ** $p<0.01$, *** $p<0.001$, **** $p<0.0001$. Source data are provided as a Source Data file.

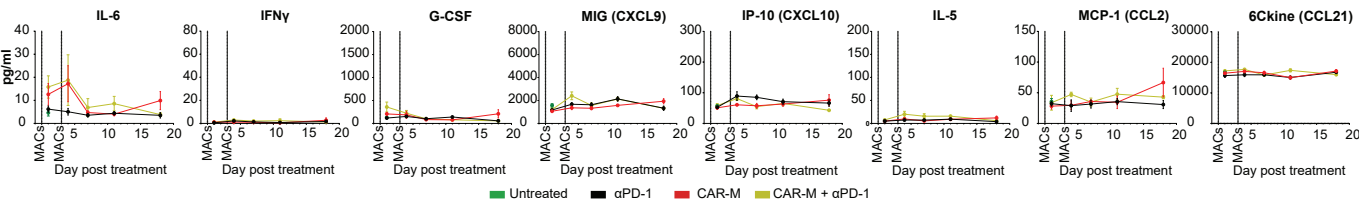


Supplementary Fig. 2 Generation of the CT26-hHER2 and the 4T1-cHER2 models and assessment of CAR-M persistence and biodistribution

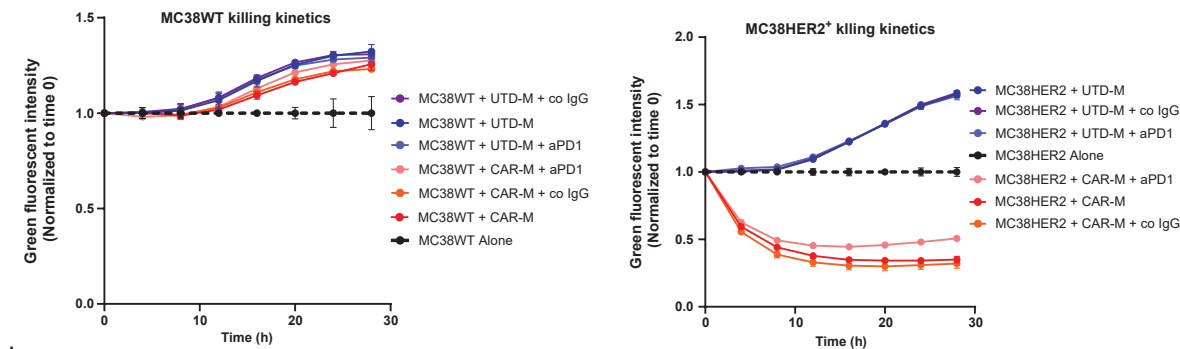
a, IHC analysis on FFPE tumor specimens confirm HER2 expression on CT26-hHER2 implanted subcutaneously in syngeneic BALB/c mice (CT26WT included as a control) evaluated at x20 high power fields per slide. **b**, Diagram of cHER2 construct engineered into a lentiviral vector under the control of CMV promoter. **c**, Representative FACS plots show cHER2 surface expression in 4T1-cHER2 cells upon staining with anti-mouse and anti-human (Trastuzumab biosimilar) antibodies. **d**, IHC analysis on ex vivo FFPE tumor specimen to confirm cHER2 expression on cells orthotopically implanted in the MFP of BALB/c. Lymph nodes (LN) were included as a negative control for HER2 expression and were assessed at x40 high power fields per slide. **e**, 4T1-cHER2 tumor was established orthotopically in the MFP of BALB/c mice for 13 days and treated with IV CAR-M. CAR-M ($n=5$) controlled tumor growth in 4T1-cHER2 compared to untreated ($n=7$) and UTD-M ($n=7$). Tumor growth curves from the individual mouse are shown. **f, g**, 5×10^6 of fluorescently labeled (IVISense680) murine CAR-M were IV injected into tumor-bearing BALB/c mice. Fluorescence was tracked over a period of 12 days. **f**, IVIS images. **g**, Quantification represented as the mean $\pm$ SEM of $n=3$. Values are normalized by subtracting the baseline signal from untreated mouse. **h**, organs and tumors were explanted for ex vivo imaging 2 and 5 days after a single injection of 5×10^6 CAR-M. Fold change values were calculated as radiant efficiency of treated organ divided by its respective untreated organ. $n=4$ mice per treatment group per tumor type; the experiment was performed once. **i**, Mice with established tumors were treated with three doses of 5×10^6 CAR-M/mouse every 3-4 days. RNAscope was used to detect CD11b gene (red) and CAR sequence (green) on FFPE sections from tumors collected seven days after the last injection. Heatmap shows the number of CAR-M per 1×10^6 CD11b (4T1-cHER2 $n=4$ and CT26-hHER2 $n=2$). Representative images at 40x magnification. The square designates a CAR-M+ event. **j**, Representative FACS plots show % of gp70+ tetramer within intratumoral CD8⁺ T cells from CT26-hHER2 tumor treated with reginal CAR-M. Source data are provided as a Source Data file.



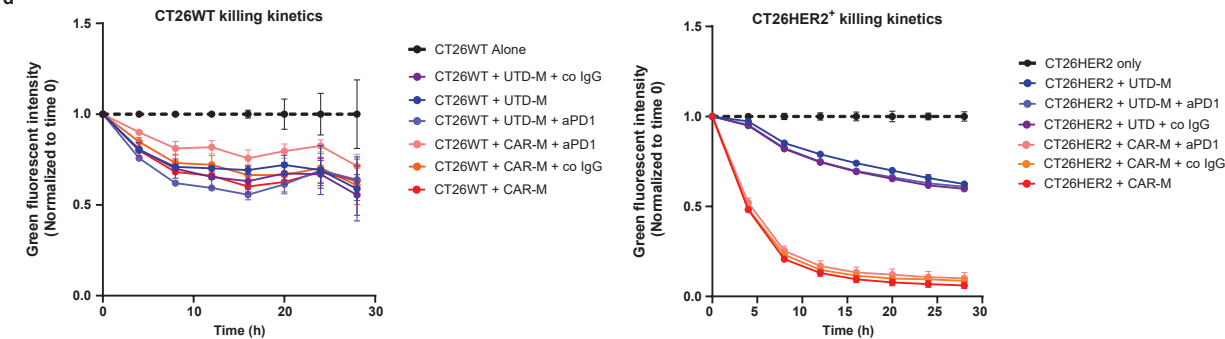
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c



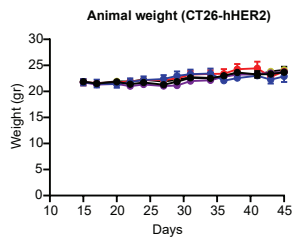
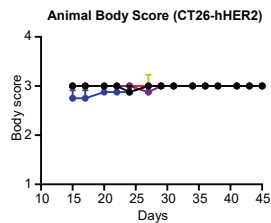
d



Supplementary Fig. 3. Regional CAR-M + aPD1 combination therapy preserves body condition score, body weight, and serum cytokines/chemokines levels

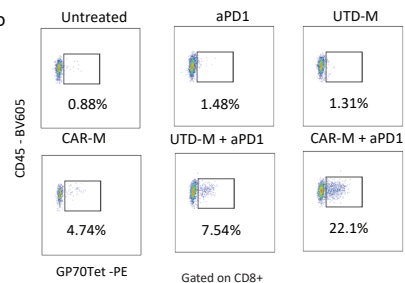
CT26-hHER2 tumor was established SQ in BALB/c and treated with IT CAR-M starting at day 12 and IP PD1 from day 12 until day 36 (approximately every 3 days). **a**, Body condition score and body weight were evaluated in CAR-M ($n=10$), anti-PD1 ($n=7$), CAR-M + aPD1 ($n=9$), or untreated ($n=9$) groups. Data represents the mean $\pm$ SEM. **b**, Serum IL-6, IFN γ , G-CSF, MIG (CXCL9), IP-10 (CXCL10), IL-5, MCP-1 (CCL2) and 6Ckine (CCL21) levels were quantified by multiplexed Luminex assay up to 18 days post treatment. Dotted lines represent CAR-M doses. Data represents the mean $\pm$ SEM of $n=6-7$ /group and is representative of two experiments. **c,d**, Incucyte-based killing assays of UTD-M or CAR-M co-cultured with either MC38 or CT26, at E:T ratio 2:1 in the presence 10 μ g/mL IgG control or 10 μ g/mL aPD1. Green fluorescence intensity (GFI) values were normalized to the 0 hr. timepoint and then, further normalized to the target cells. Data represents the mean $\pm$ SEM of $n=3$ technical replicate and is representative of two experiments. Source data are provided as a Source Data file.

a

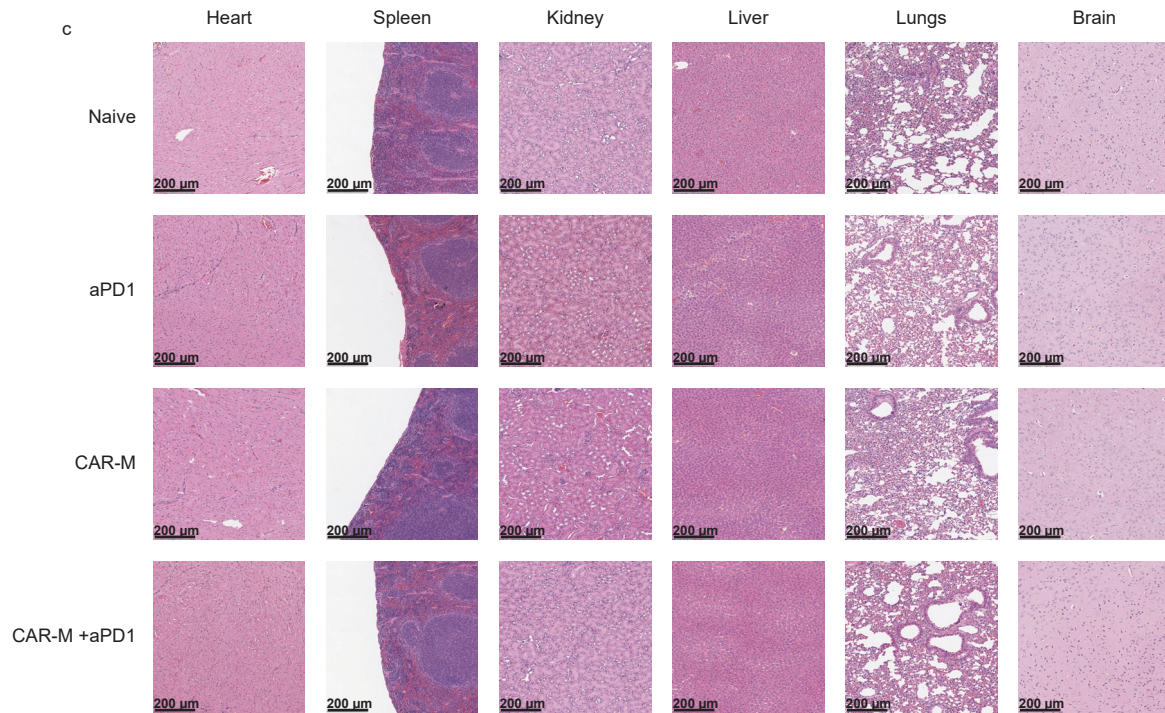


● aPD1
 ◆ UTD-M
 ◆ UTD-M + aPD1
 ◆ CAR-M
 ◆ CAR-M + aPD1

b

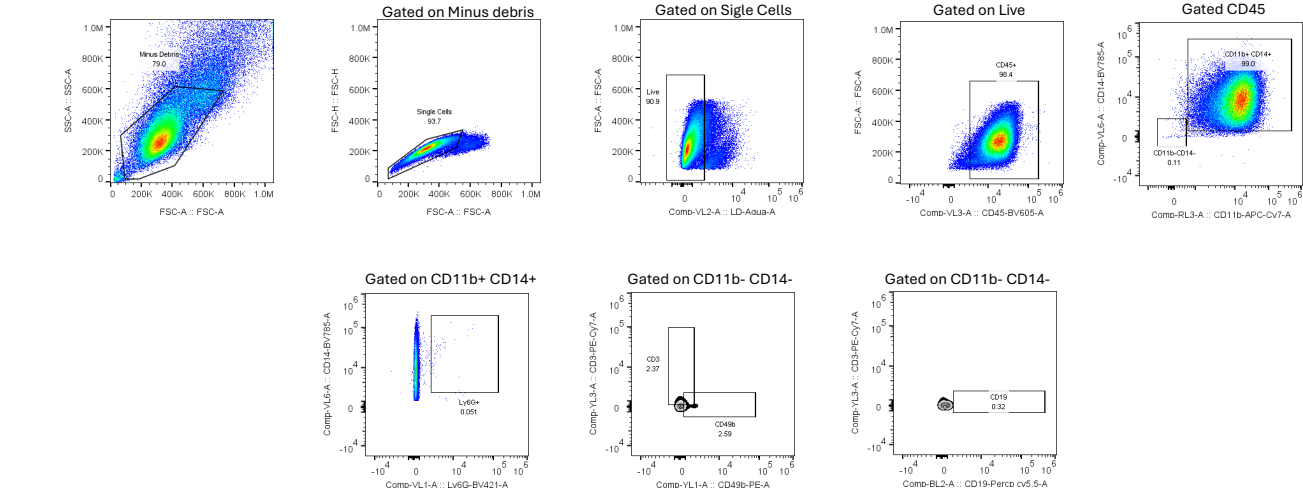


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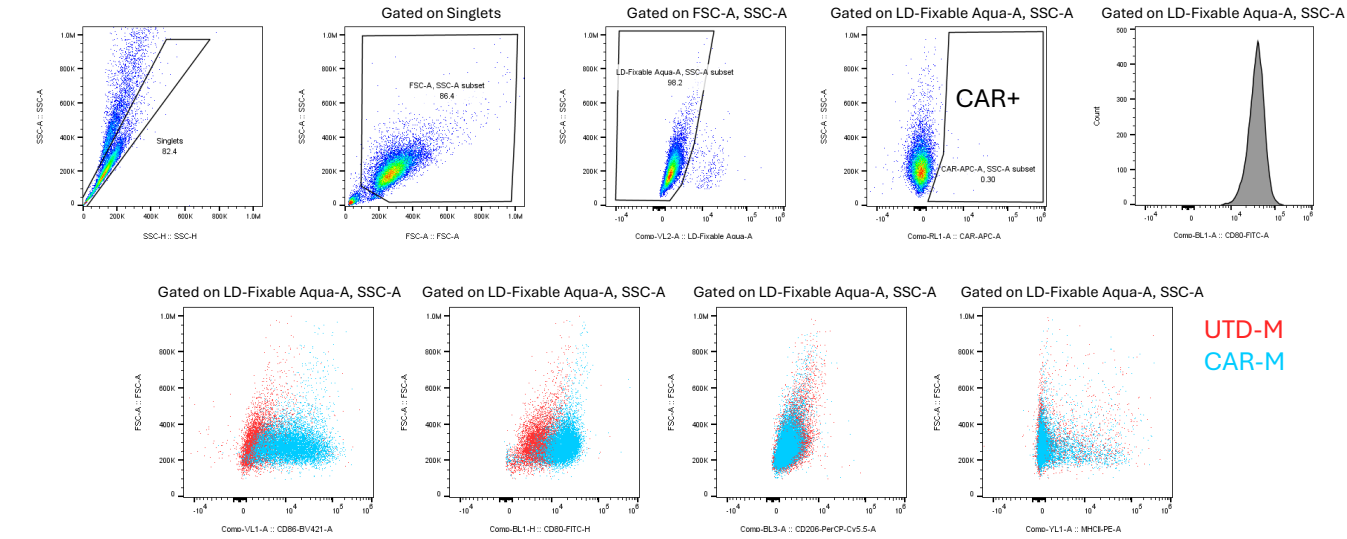


Supplementary Fig. 4. Systemic CAR-M and aPD-1 combination therapy is well tolerated

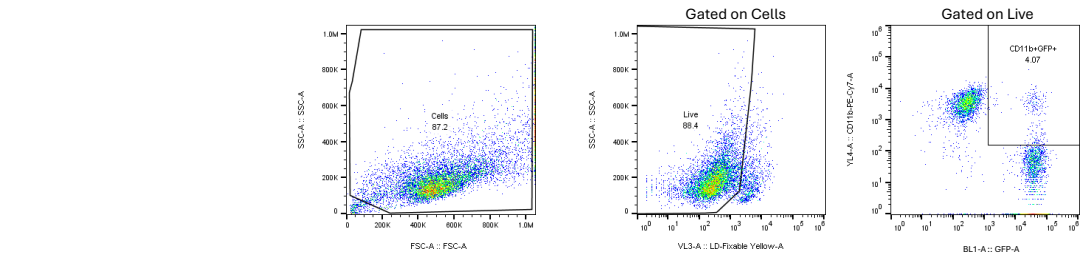
CT26-hHER2 tumor-bearing mice were treated with systemic IV CAR-M from day 10 and IP aPD1 from day 14. **a**, Body condition score and body weight are maintained in response to CAR-M + aPD1 combination therapy. Data represents the mean $\pm$ SEM of $n=8$ /group except UTD-M + aPD1 $n=7$ and is representative of three experiments. **b**, Representative FACS plots show % of gp70+ tetramer within intratumoral CD8⁺ T cells from CT26-hHER2 tumor treated with systemic CAR-M. **c**, Histological assessment of heart, spleen, kidney, liver, lung, and brain harvested from CT26-hHER2 tumor-bearing BALB/c mice treated with systemic CAR-M ($n=4$ /group). Organs from tumor-free (naïve) mice were included as controls $n=4$. No signs of treatment-related toxicity were observed. Source data are provided as a Source Data file.



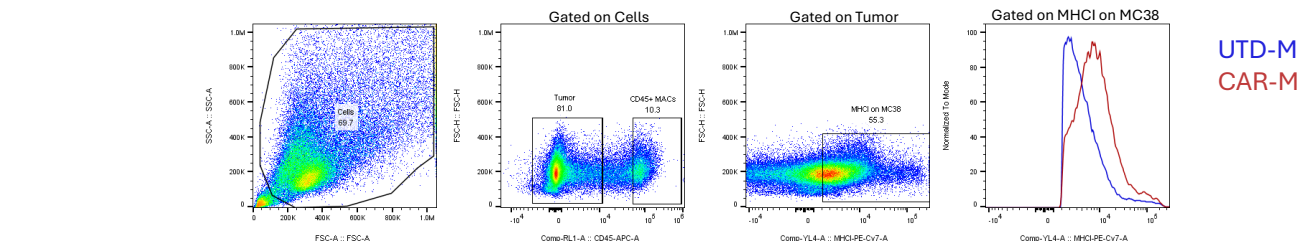
Supplementary Figure 5a



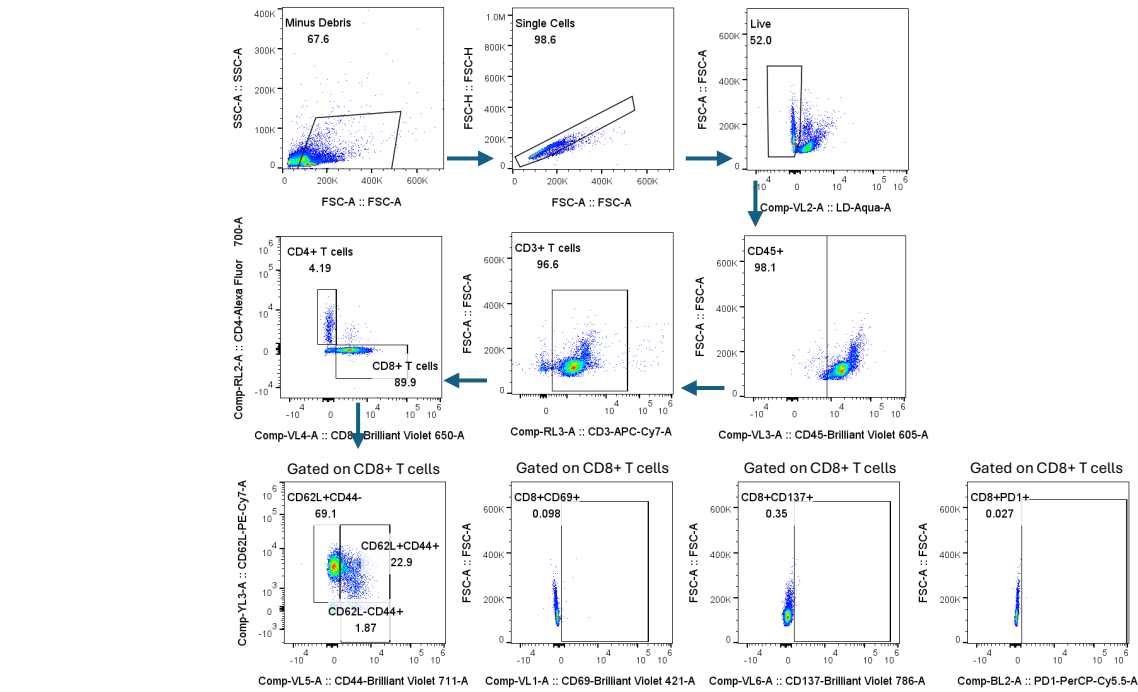
Supplementary Figure 5b



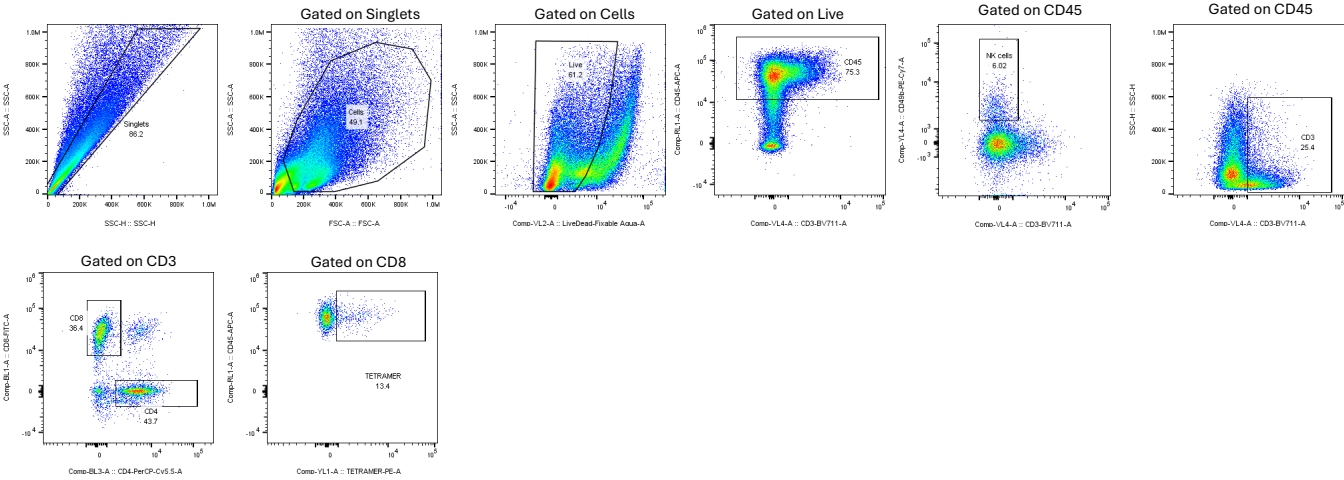
Supplementary Figure 5c



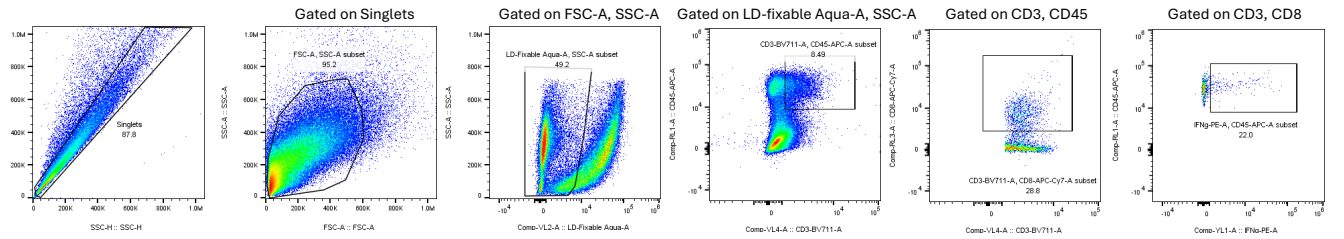
Supplementary Figure 5d



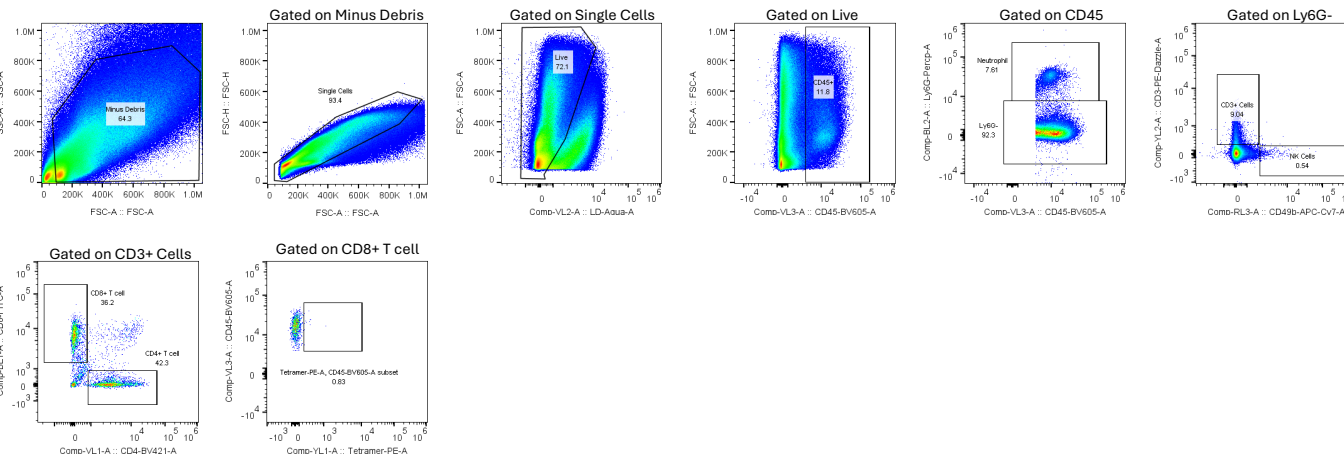
Supplementary Figure 6a



Supplementary Figure 6b



Supplementary Figure 6c



Supplementary Figure 6d