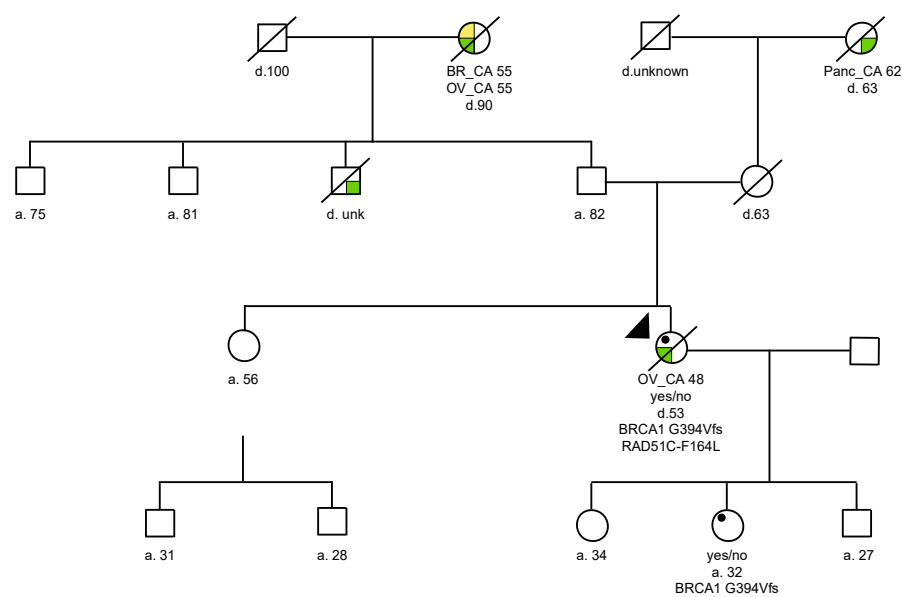
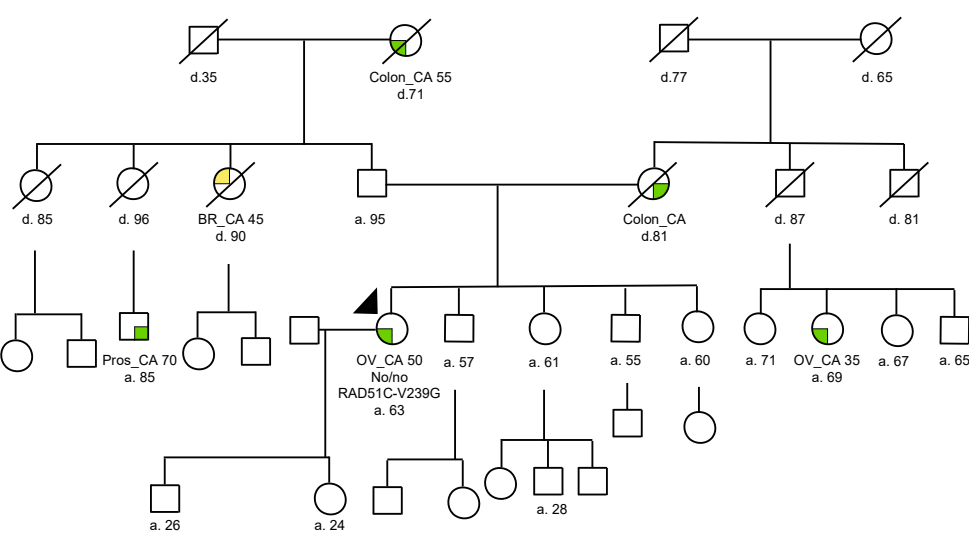


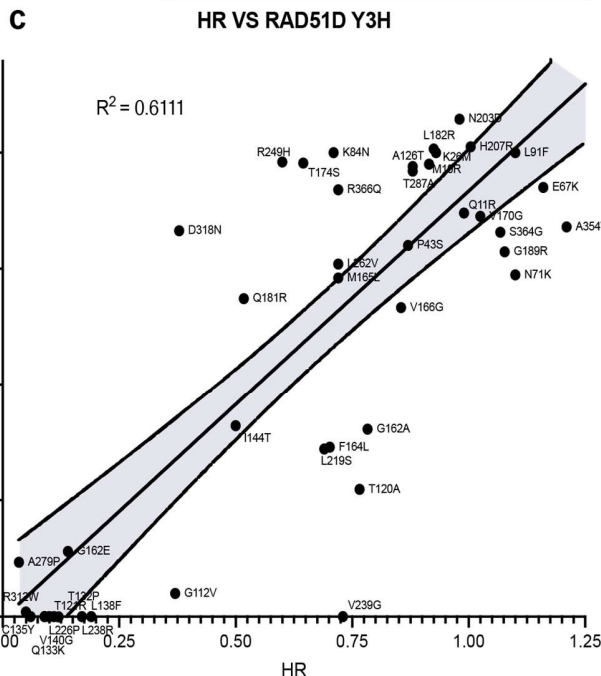
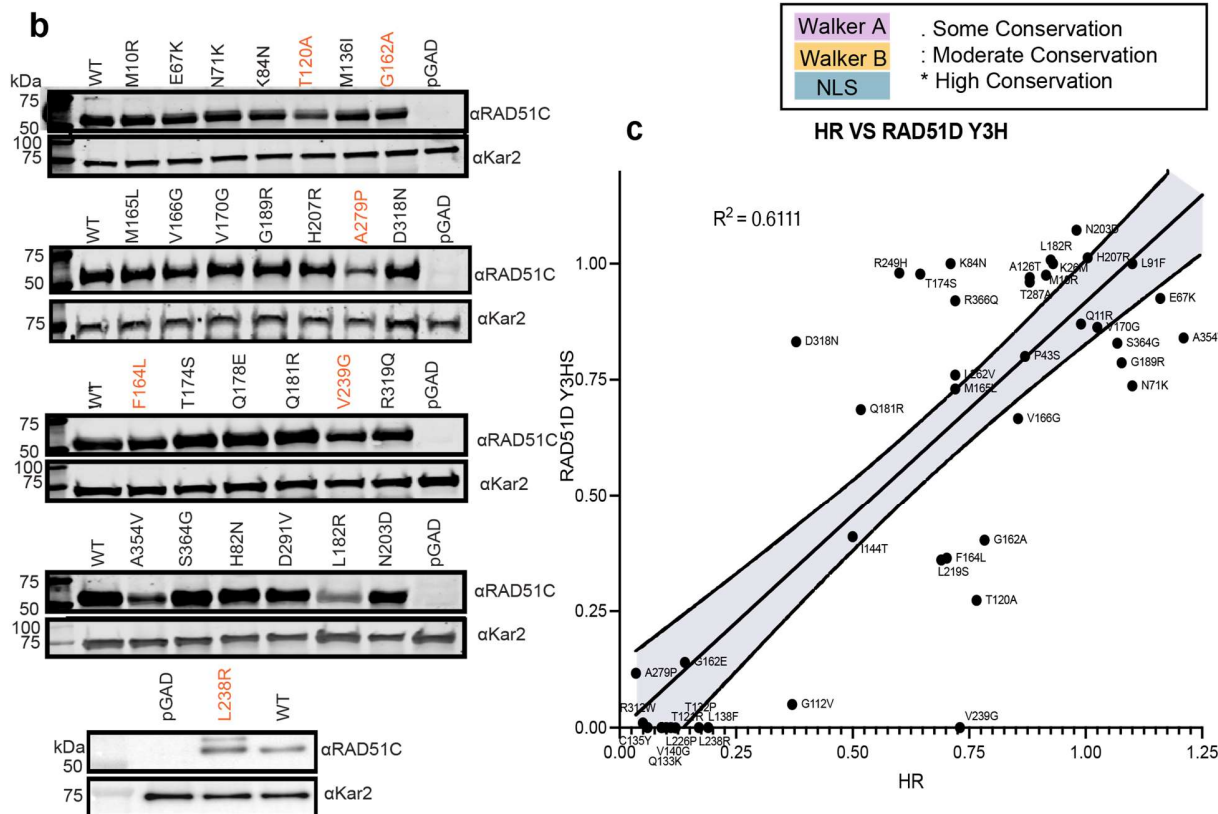
a RAD51C-F164L Family Pedigree



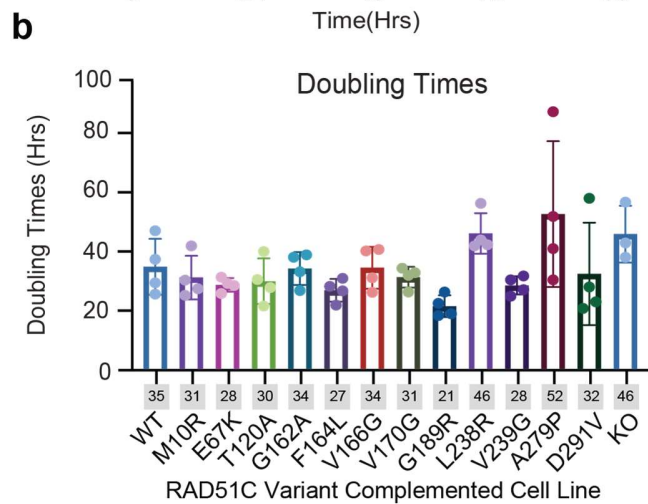
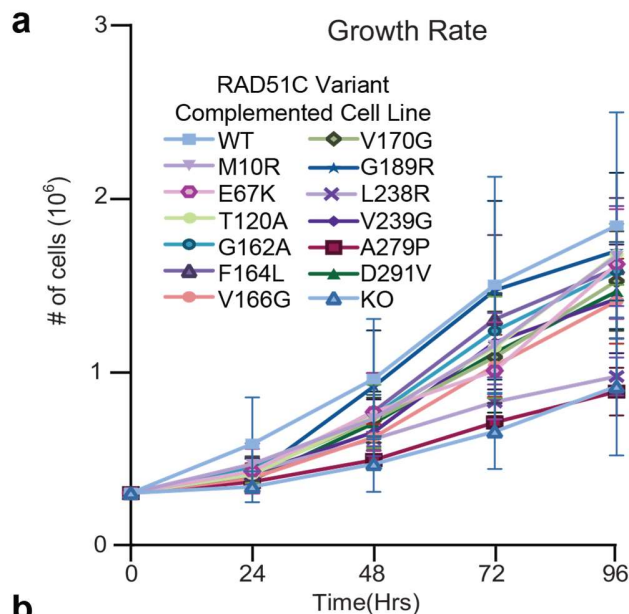
b RAD51C-V239G Family Pedigree



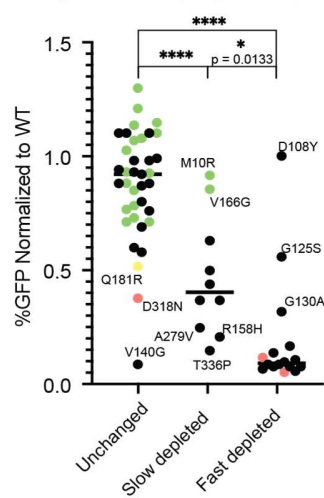
Supplementary Figure 1. RAD51C F164L and V239G variant inheritance and cancer occurrence. Pedigrees showing family inheritance of RAD51C-F164L **(a)** or RAD51C-V239G **(b)** and cancer occurrence. The patient of interest (detailed in **Supplementary Table 1**) is identified by the black arrow. Females are indicated with a circle and males are indicated with a square. Slashes indicate deceased individuals, and age of death is shown below by (d.). Cancer status is detailed below each individual as well as age of diagnosis and current age (a.). If available, BRCA allele status and *RAD51C* variant status is also indicated. Cancer type is indicated by the colored quadrant (top left yellow, unilateral breast cancer; bottom left green, ovarian cancer; bottom right green, cancer type other than breast or ovarian cancer). BRCA variant carriers are also indicated by a black dot.

[illegible]

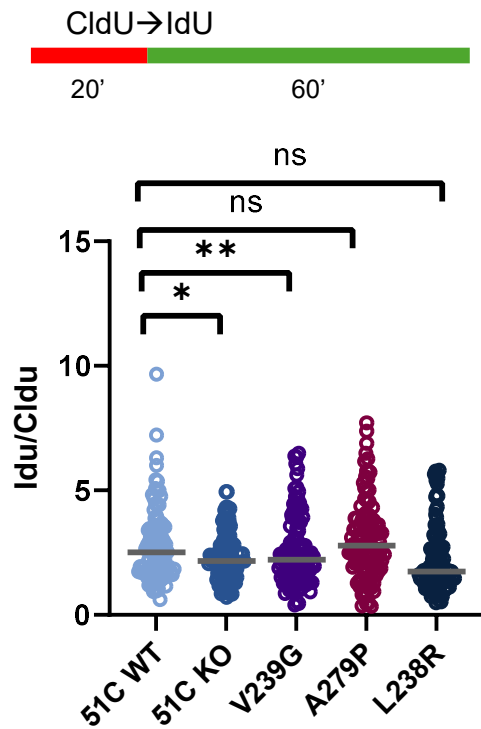
Supplementary Figure 2. Comparison of RAD51 conservation, protein expression, and yeast-3-hybrid among RAD51C ovarian cancer variants with deficient interaction with RAD51D. **(a)** Four out of five RAD51C-RAD51D interaction deficient variants moderately or highly conserved with RAD51 by multiple sequence alignment. Variants are indicated above the alignment, with ATPase motifs labeled in purple (Walker A) and gold (Walker B) and the nuclear localization sequence labeled in light blue. Conservation is indicated with an asterisk (*) for high conservation and a colon (:) for moderate conservation (conserved within amino acid groups of strongly similar properties) and a period (.) for some conservation (conserved within amino acid groups of weakly similar properties). RAD51D interaction capabilities are indicated in red (deficient) or green (proficient). **(b)** RAD51C ovarian cancer variants examined in **Fig. 1c** are expressed in yeast. Western blot analysis of wild-type RAD51C, the variant of interest, or an empty plasmid performed. RAD51C protein levels were analyzed using an α RAD51C antibody and equal protein loading was determined using the loading control Kar2 (α Kar2). The experiments were performed in duplicate. **(c)** RAD51C variant HR proficiency results from this study and Prakash et al. compared against the yeast-three-hybrid (Y3H) RAD51C-RAD51D interaction results. The average relationship is represented by the black line, with 95% confidence intervals in green. R^2 and Pearson correlation values are shown. Note that results for L27P and Q178E have been excluded due to lack of protein expression.



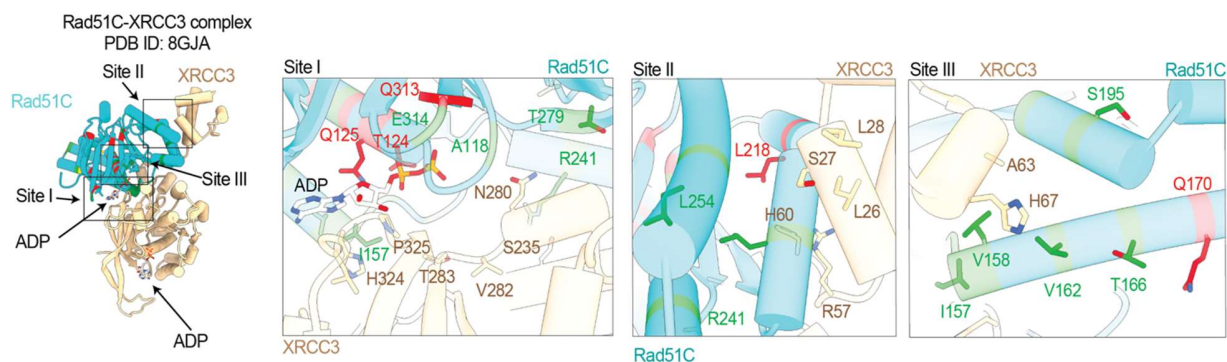
c HR proficiency by category



Supplementary Figure 3. Growth and comparison of HR analysis of RAD51C ovarian cancer variants of interest expressed in RAD51C knockout U2OS cells. (a) The growth rates of *RAD51C* variant expressing cells. The growth rate of *RAD51C* KO cell line and *RAD51C* wild-type complemented or variant expressing cells were analyzed for cell number every 24 hours for four days. Growth of each cell line was measured using a coulter counter and logged over 96 hours and average growth of four experiments was graphed with standard deviations plotted. The experiment was repeated four times per cell line. (b) The doubling times of *RAD51C* variant expressing cells. Doubling times were calculated from the 24-hour log-phase period determined in (a). Experiments were repeated four times for every variant cell line; in the *RAD51C*-knockout cell line, the experiment was performed three times. Average doubling time was plotted with standard deviations. (c) Comparison of growth to HR proficiency from ²⁶. Unchanged indicates a variant that has WT growth whereas a slow depleted variant has intermediate growth and a fast depleted variant is inviable. Variants analyzed in this study for HR proficiency are color coded in green (proficient), red (deficient), and yellow (intermediate). The average HR proficiency within the variants categorized as unchanged is significantly higher than the slow-depleted and fast-depleted variants. Statistical significance determined by t-test is indicated (**** indicates $p < 0.0001$). Outliers are indicated.



Supplementary Figure 4. The replication fork speed of RAD51C KO and V239G complemented cells is reduced. *RAD51C* CRISPR/Cas9 U2OS cells were complemented with the WT *RAD51C* or the indicated variant and DNA replication fork progression was determined. The cells were first pulsed with CldU for 20 min followed by IdU for 60 min. Each data point represents IdU/Cldu ratio for one fiber measurement. From left to right, n = 204, 205, 205, 218, or 218 fibers analyzed from two experiments. Bars represent median of each data set. Statistical significance was determined by Kruskal-Wallis with multiple Dunn's comparison and is indicated (ns is not significant, * p = 0.0375, ** p = 0.0079).



Supplementary Figure 5. Location of conserved RAD51C variants on *Alvinella pompejana* CX3 structure. The Rad51C-XRCC3 structure (³⁶:PDB ID:8GJA) with the indicated human RAD51C ovarian cancer identified variants are shown. Site I indicates residues that were identified in the ATPase region, Sites II and III are at the interface with XRCC3. Red indicates an HR deficient variant whereas green indicates an HR proficient variant. Note that *Alvinella pompejana* residues T124, Q125, Q170, L218, and Q313 correspond to human T132, Q133, Q178, L226, and D318, respectively.

Supplementary Table 1. Clinical information for ovarian cancer patients with either RAD51C-F164L, L238R, or V239G variants. The variant is indicated as well as the nucleotide change (Nuc. Change) and the prior classification status. The allele origin including variant allele frequency (VAF) is shown. The age of diagnosis, diagnosis, BRCA status, as well as treatment history and clinical outcomes are shown.

Variant	Nuc. Change	Classification	Allele Origin	Age at diagnosis	Diagnosis	BRCA Status	Treatment	Outcome
F164L	492T>G	Likely Benign (Myriad)	Germline	48	IIIC - Papillary serous cystadenocarcinoma	+	• carbo/taxol • bevacizumab • olaparib • carbo/gem	4 years Died - 2017
L238R	713T>G	VUS	Likely Germline, 99% VAF	54	Platinum resistant HGSO C, stage unknown	-	• cisplatin/taxotere • carbo/taxotere • carbo/gem/taxol • niraparib • CHK1 inhibitor (prexasertib) phase 2 trial	Recurrent HGSO C, improved with prexasertib before trial withdrawal, current status unknown
V239G	716T>A	VUS	Germline	50	IIIC - Papillary serous cystadenocarcinoma	-	• IV carbo/taxol • IP cisplatin	13 years Currently NED

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