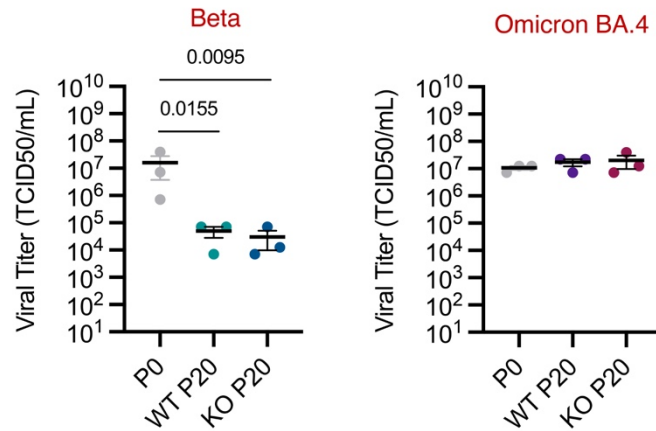
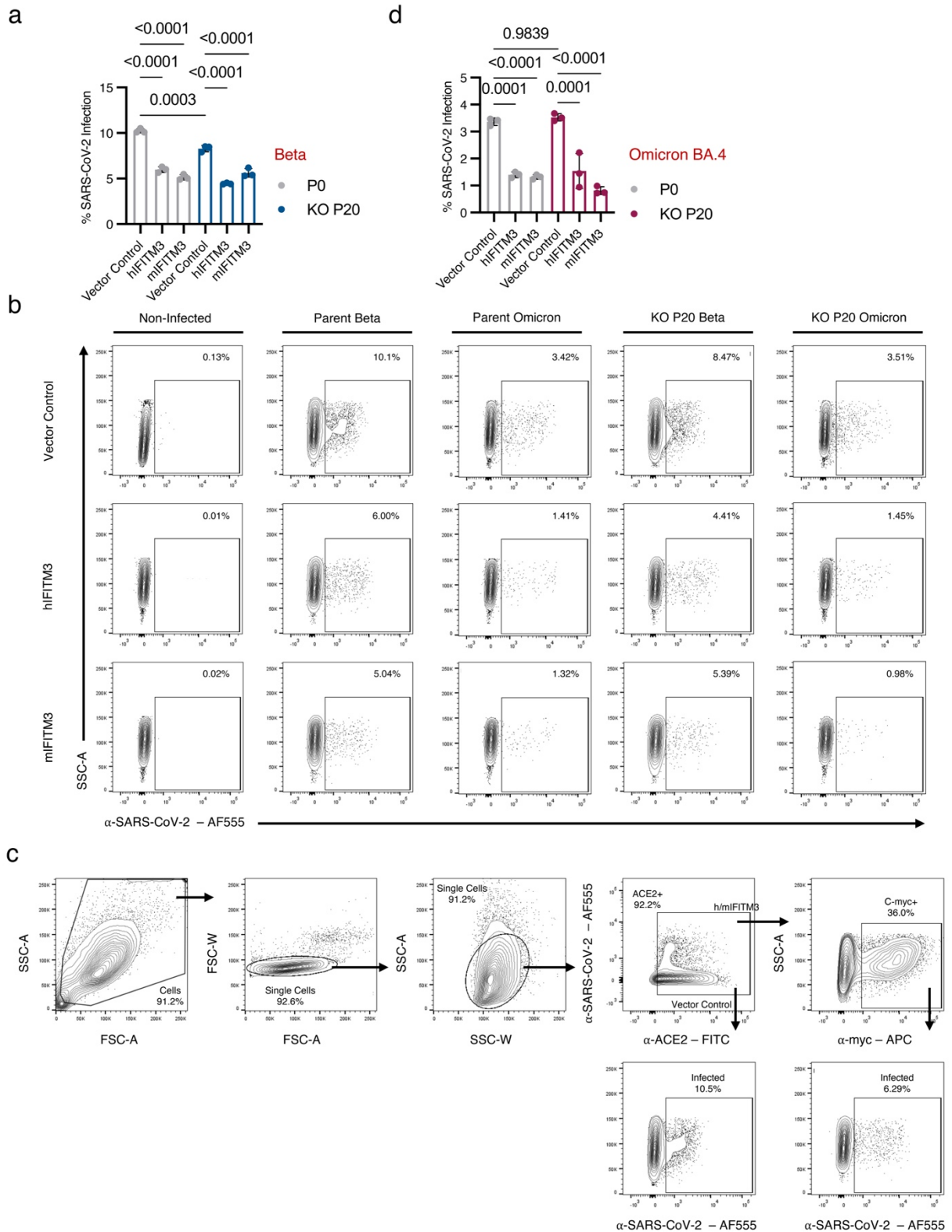


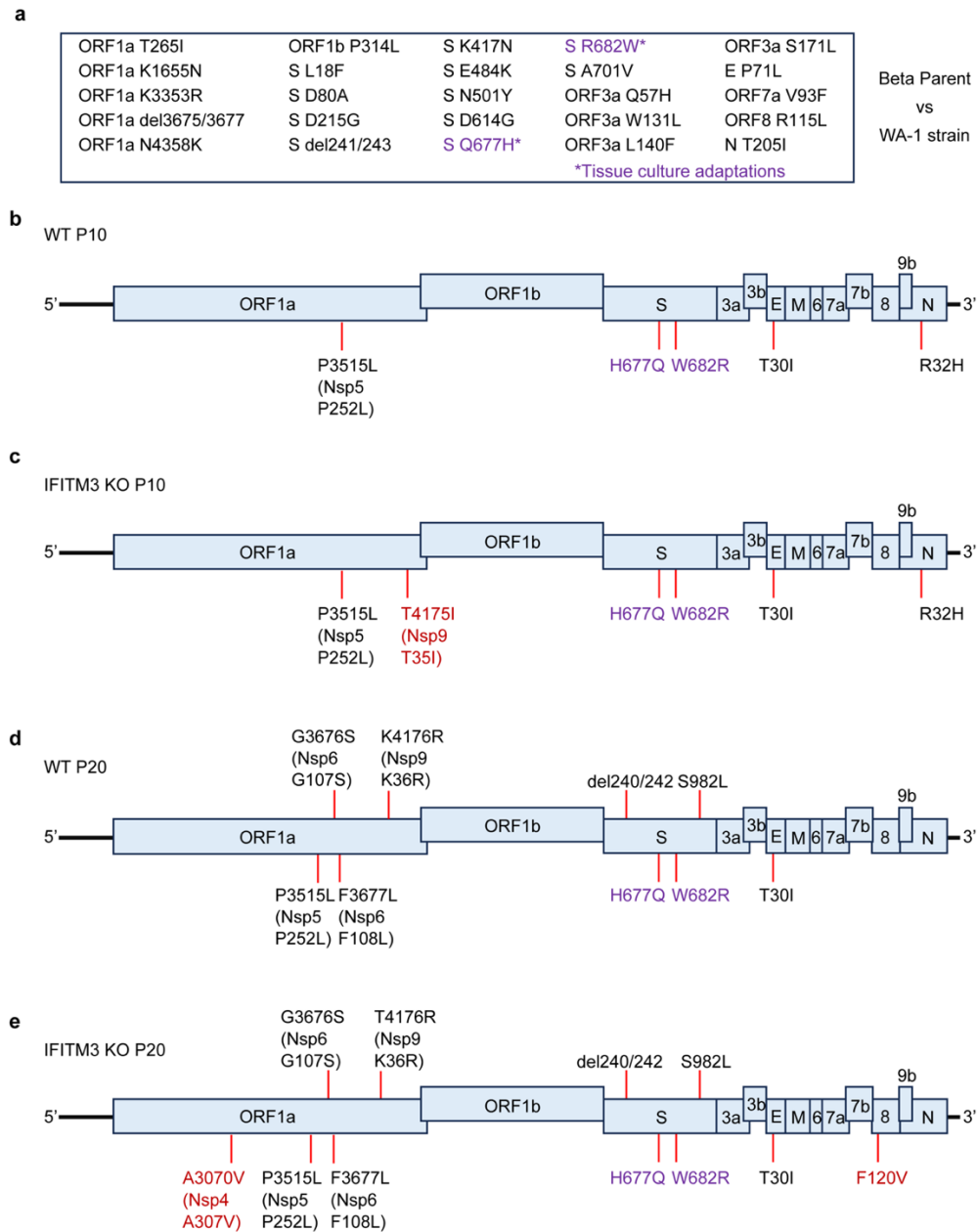


Supplementary Figure 1. Passaged virus in primary HBE cell cultures. Viral titers from SARS-CoV-2 infection of primary HBE cell cultures (error bars represent SEM, p-values by one-way ANOVA with multiple comparisons, n = 3 per group). Source data are provided as a Source Data file.



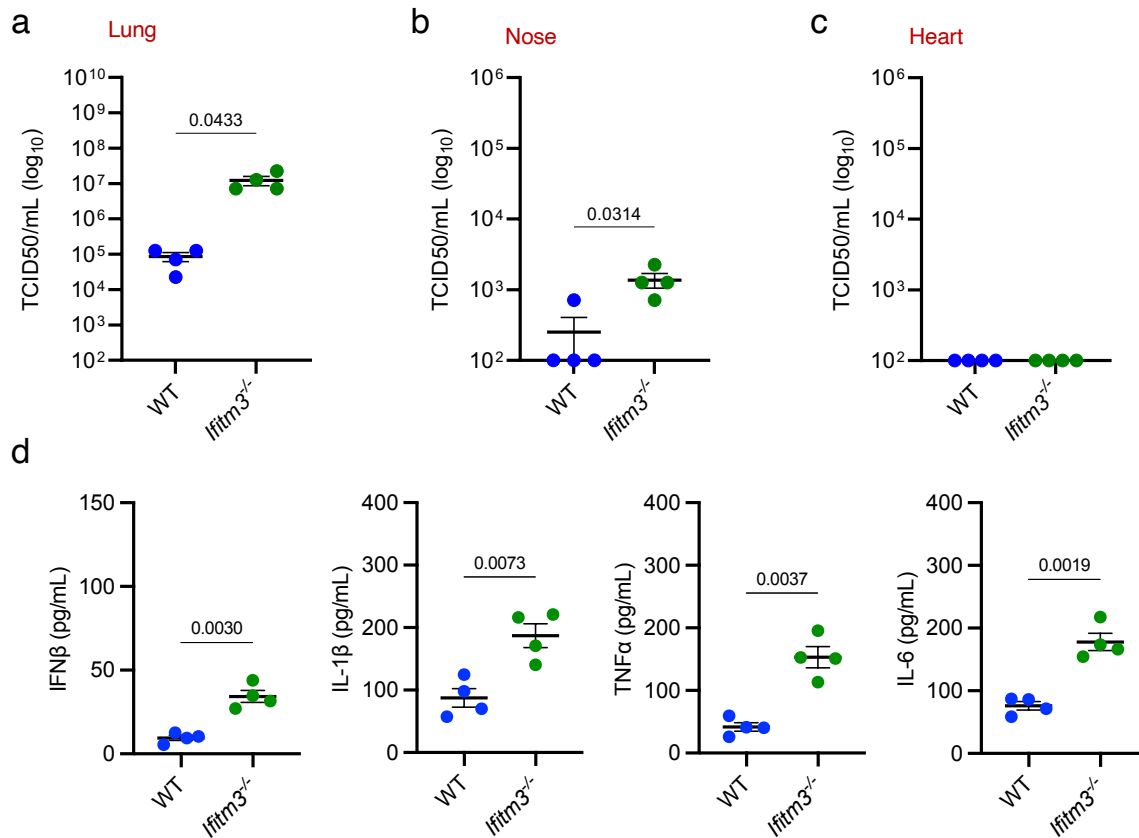
Supplementary Figure 2. Passaged virus in ACE2-HEK293T cell cultures. (a-b) Viral titers from SARS-CoV-2 infection of ACE2-HEK293T cell cultures (error bars represent SEM, p-values

represent one-way ANOVA with multiple comparisons, $n = 3$ per group). (c) Representative flow plots for a-b. (d) Representative gating strategy for (a-b). Source data are provided as a Source Data file.



Supplementary Figure 3. Mutations observed within the SARS-CoV-2 beta strain genome after passing through WT or *Ifitm3*^{-/-} mice. (a) List of amino acid differences between the SARS-CoV-2 Beta parent stock and the original WA1 strain. Tissue culture adaptations are indicated with an asterisk (*) and are colored purple. (b, c) Schematic of SARS-CoV-2 Beta strain genome annotated with amino acid substitutions found in the consensus sequence after

passaging 10 times (**b**) or 20 times (**c**) through WT mice. (**d, e**) Schematic of SARS-CoV-2 Beta strain genome annotated with amino acid substitutions found in the consensus sequence after passaging 10 times (**d**) or 20 times (**e**) through *Ifitm3*^{-/-} mice.

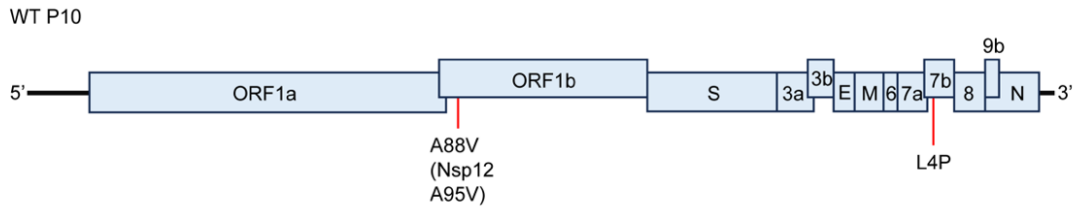


Supplementary Figure 4. IFITM3 restricts the BA.4 omicron virus in mice. Groups of WT or *Ifitm3*^{-/-} mice (n=4 per group) were challenged with 10⁵ TCID50 of the SARS-CoV-2 BA.4 strain. Viral titers from lung (**a**) or nose (**b**) homogenates collected at day 2 post infection. (**c**) ELISA quantification of IFNβ, IL-1β, TNFα, and IL-6 in lung homogenates collected at day 2 post infection. Each dot represents an individual mouse. Numbers above the graph represent p values. Source data are provided as a Source Data file.

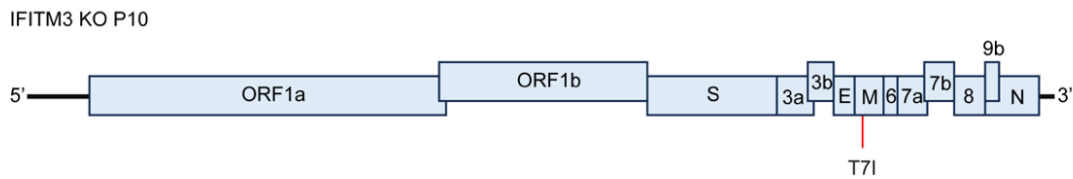
a

ORF1a S135R	ORF1b P314L	S G339D	S N440K	S Y505H	S Q954H	N P13L	Omicron Parent vs WA-1 strain
ORF1a del141/143	ORF1b R1315C	S S371F	S L452R	S D614G	S N969K	N R203K	
ORF1a T842I	ORF1b I1566V	S S373P	S S477N	S H655Y	ORF3a T223I	N G204R	
ORF1a G1307S	ORF1b T2163I	S S375F	S T478K	S N679K	E T9I		
ORF1a L3027F	S T19I	S T376A	S E484A	S P681H	M D3N		
ORF1a T3090I	S del25/27	S D405N	S F486V	S413R	M Q19E		
ORF1a T3255I	S G142D	S R408S	S Q498R	S N764K	M A63T		
ORF1a P3395H	S V213G	S K417N	S N501Y	S D796Y	ORF6 D61L		

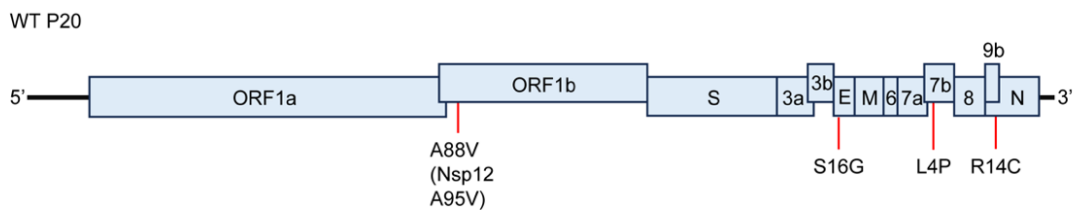
b



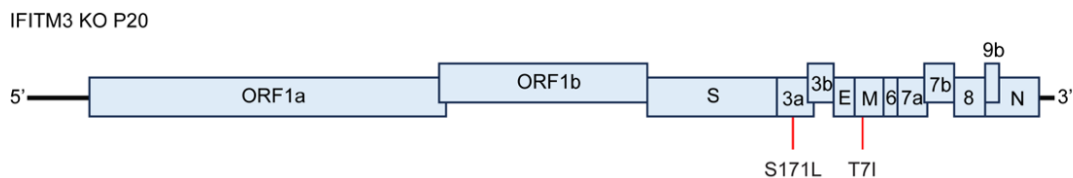
c



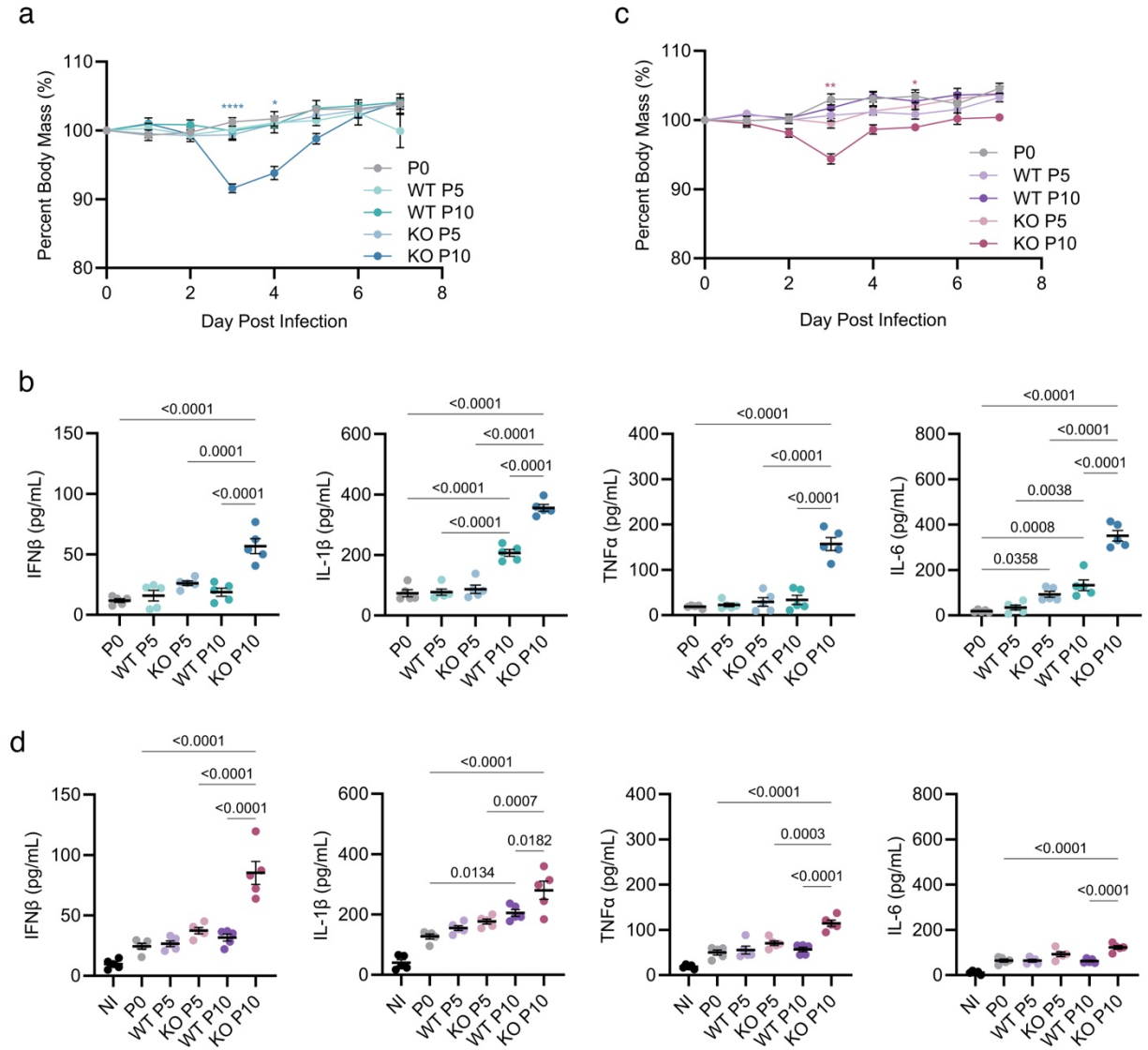
d



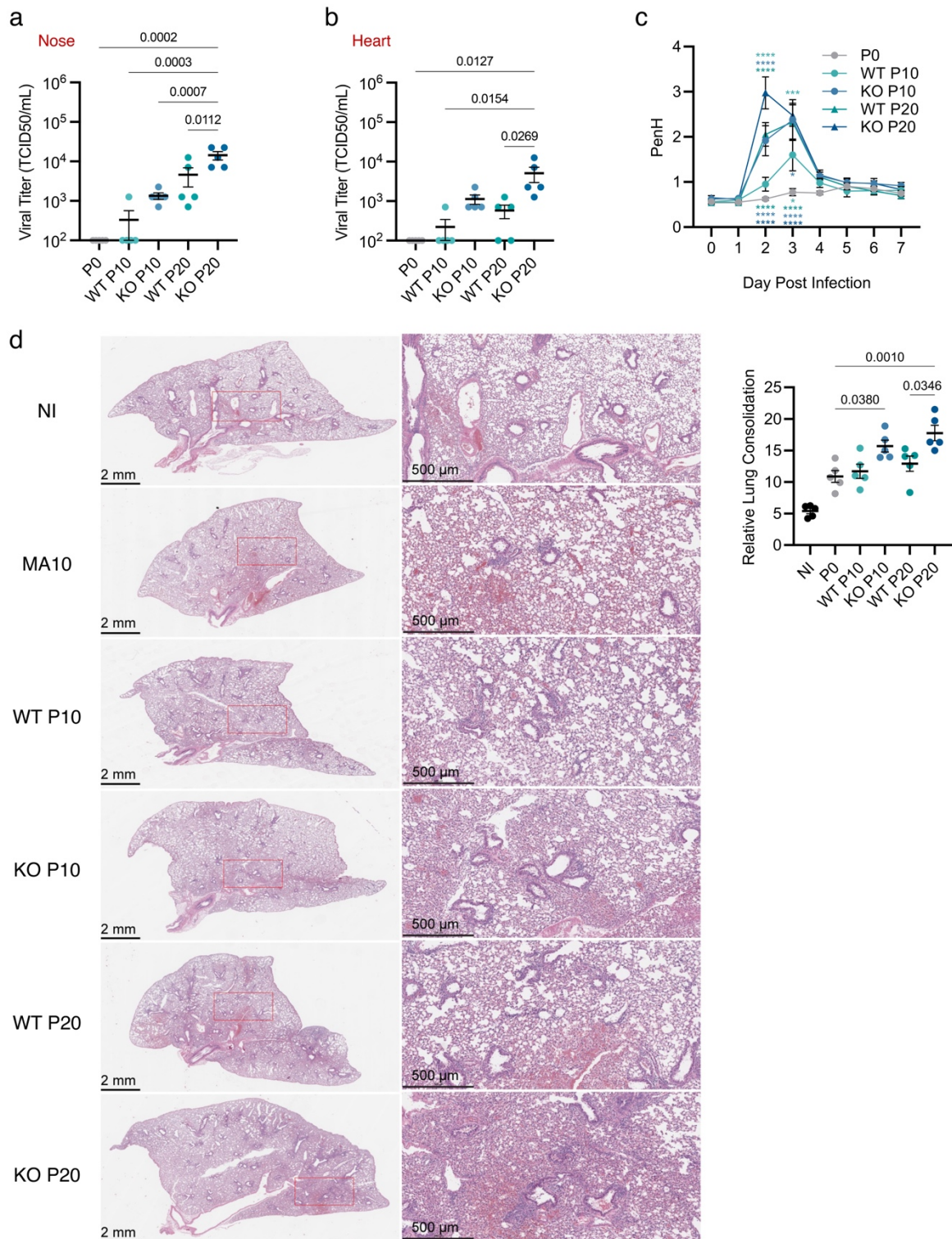
e



Supplementary Figure 5. Mutations observed within the SARS-CoV-2 BA.4 strain genome after passaging through WT or *Ifitm3*^{-/-} mice. (a) List of amino acid differences between the SARS-CoV-2 Beta parent stock and the original WA1 strain. (b, c) Schematic of SARS-CoV-2 BA.4 strain genome annotated with amino acid substitutions found in the consensus sequence after passaging 10 times (b) or 20 times (c) through WT mice. (d, e) Schematic of SARS-CoV-2 BA.4 strain genome annotated with amino acid substitutions found in the consensus sequence after passaging 10 times (d) or 20 times (e) through *Ifitm3*^{-/-} mice.

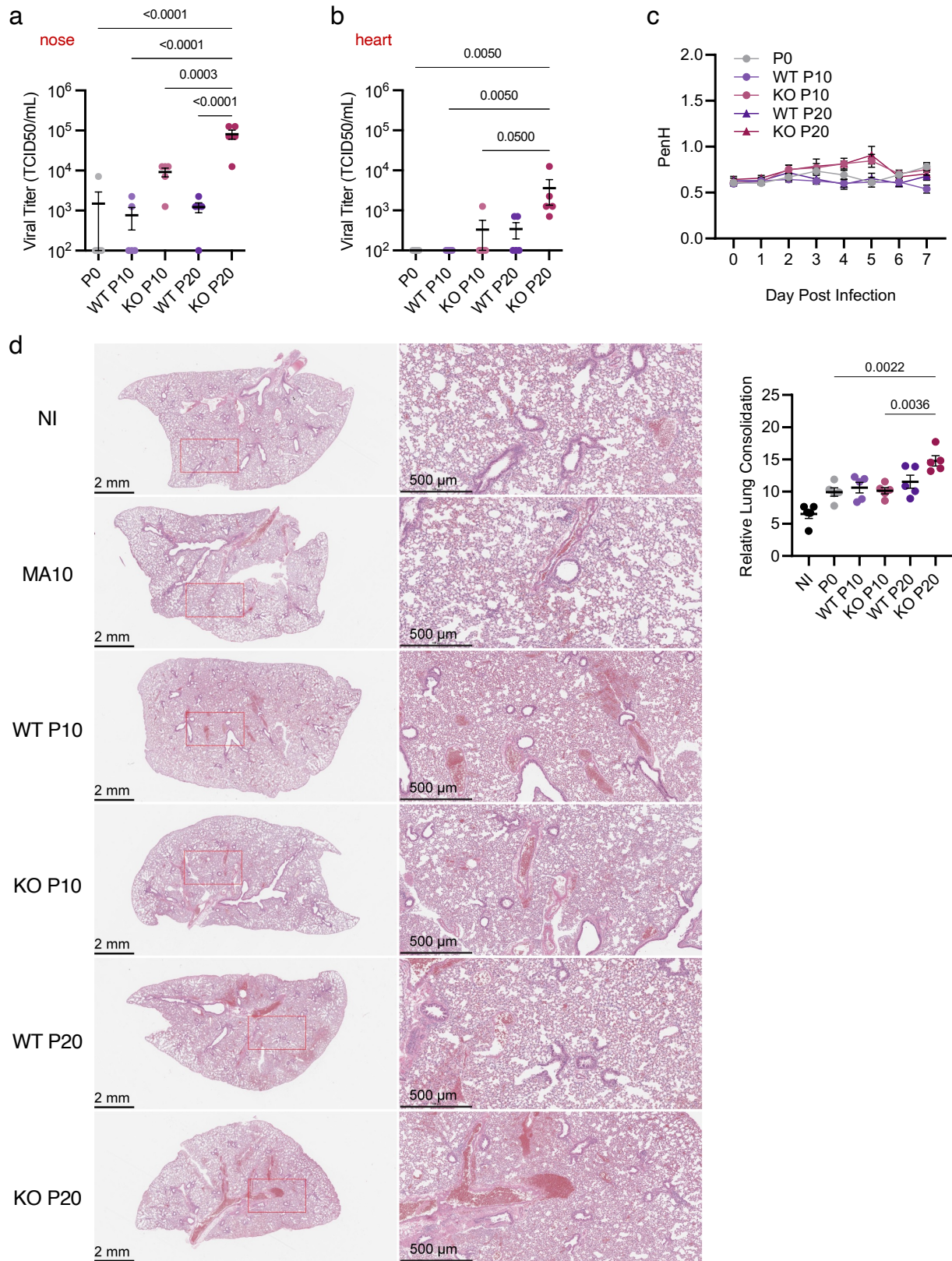


Supplementary Figure 6. Passaging SARS-CoV-2 variants 10 times through *Ifitm3*^{-/-} mice is sufficient to induce greater weight loss and inflammatory cytokines. Groups of WT mice (n=5 per group) were challenged with 10⁵ TCID₅₀ of the SARS-CoV-2 Beta (**a**, **b**) or Omicron BA.4 (**c**, **d**) strains passaged 5 or 10 times through WT or *Ifitm3*^{-/-} mice and compared to the parent virus (passage 0). (**a**, **c**) Weight loss during challenge by passaged SARS-CoV-2 viruses. Error bars represent SEM, comparisons were made using the Mann-Whitney test (only comparisons between P0, WT P10, and KO P10 are shown (p values: *** <0.0005, ** <0.005, * <0.05). Dots represent averages of individual mice (n=5 per group). (**b**, **d**) ELISA quantification of IFN β , IL-1 β , TNF α , and IL-6 in lung homogenates collected at day 2 post infection. Each dot represents an individual mouse. Numbers above the graph represent p values. Source data are provided as a Source Data file.

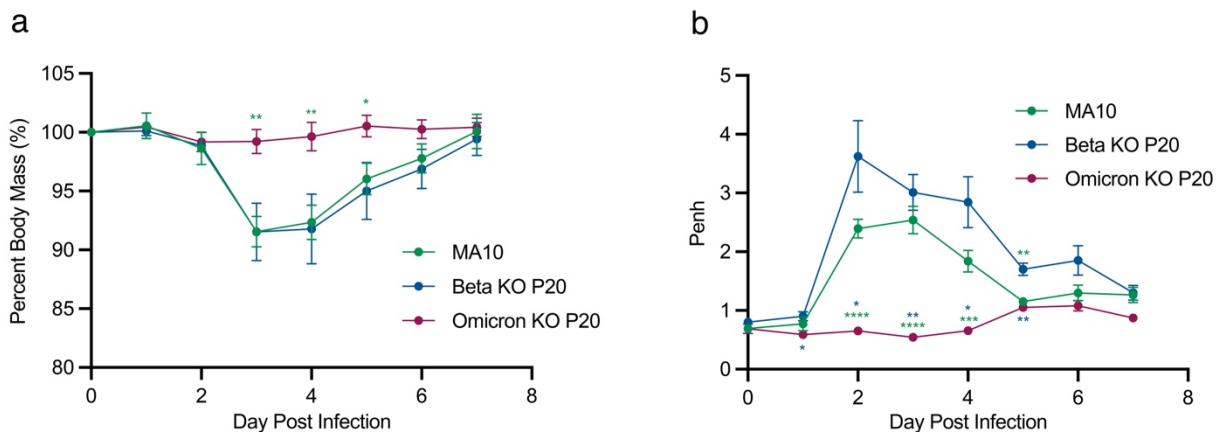


Supplementary Figure 7. Comparing lung pathogenesis and inflammation induced by the SARS-CoV-2 Beta strain passaged 10 or 20 times through WT or *Ifitm3*^{-/-} mice. Groups of WT mice (n=5 per group) were challenged with 10⁵ TCID₅₀ of the SARS-CoV-2 Beta strain passaged 10 or 20 times through WT or *Ifitm3*^{-/-} mice and compared to the parent virus (passage

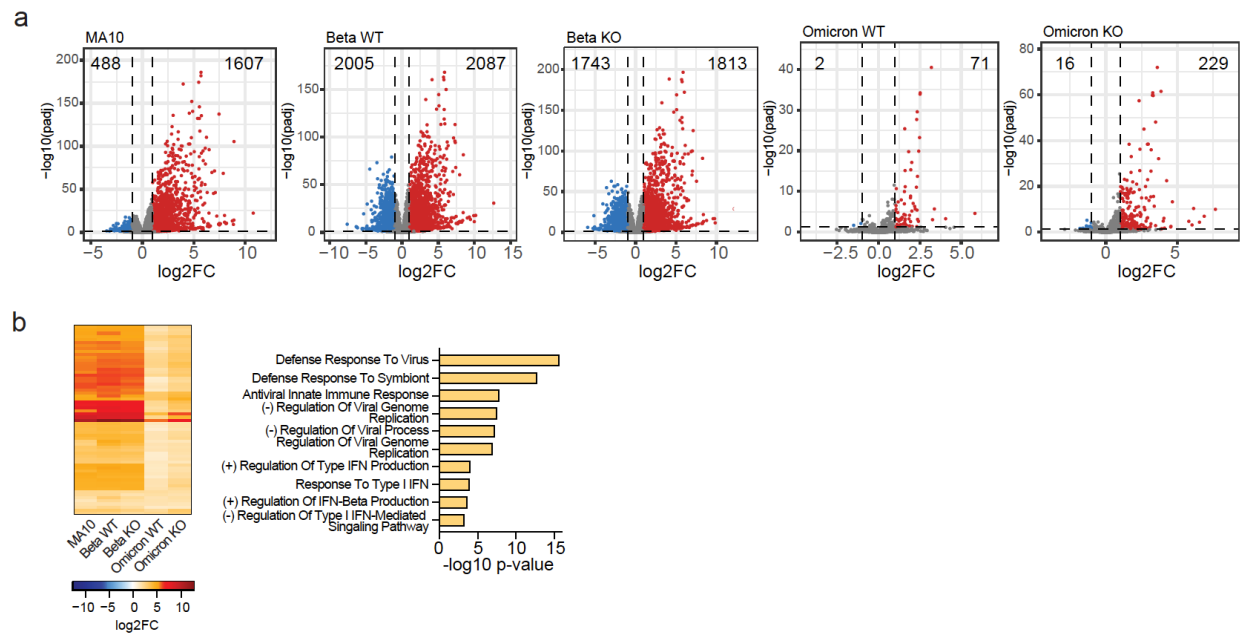
0). Viral titers from nose (**a**), or heart (**b**) homogenates collected at day 2 post infection. (**c**) Daily whole-body plethysmography enhanced pause (PenH) measurements. Dots represent averages of individual mice (n=5 per group) (p values: **** <0.0001, *** <0.0005, ** <0.005, * <0.05). (**d**) Representative histology images of infected lungs at day 7 post infection. Whole lung images are shown on the left (scale bars represent 2 mm) and 4x zoomed images are shown on the right (scale bars represent 500 μ m). The red box present in the whole lung images corresponds to its matched zoomed in image. (**e**) Quantification of consolidated tissue versus open airspace in H&E staining for entire lung sections of multiple animals infected at day 7 post infection. (**a**, **b**, **e**) Each dot represents an individual mouse. (**c**) Dots represent averages of individual mice (n=5 per group). Source data are provided as a Source Data file.



Supplementary Figure 8. Comparing lung pathogenesis and inflammation induced by the SARS-CoV-2 Omicron BA.4 strain passaged 10 or 20 times through WT or *Ifitm3*^{-/-} mice. Groups of WT mice (n=5 per group) were challenged with 10⁵ TCID50 of the SARS-CoV-2 BA.4 strain passaged 10 or 20 times through WT or *Ifitm3*^{-/-} mice and compared to the parent virus (passage 0). Viral titers from nose (a), or heart (b) homogenates collected at day 2 post infection. (c) Daily whole-body plethysmography enhanced pause (PenH) measurements. Dots represent averages of individual mice (n=5 per group). (d) Representative histology images of infected lungs at day 7 post infection. Whole lung images are shown on the left (scale bars represent 2 mm) and 4x zoomed images are shown on the right (scale bars represent 500 μm). The red box present in the whole lung images corresponds to its matched zoomed in image. (e) Quantification of consolidated tissue versus open airspace in H&E staining for entire lung sections of multiple animals infected at day 7 post infection. (a, b, e) Each dot represents an individual mouse. (c) Dots represent averages of individual mice (n=5 per group). Source data are provided as a Source Data file.



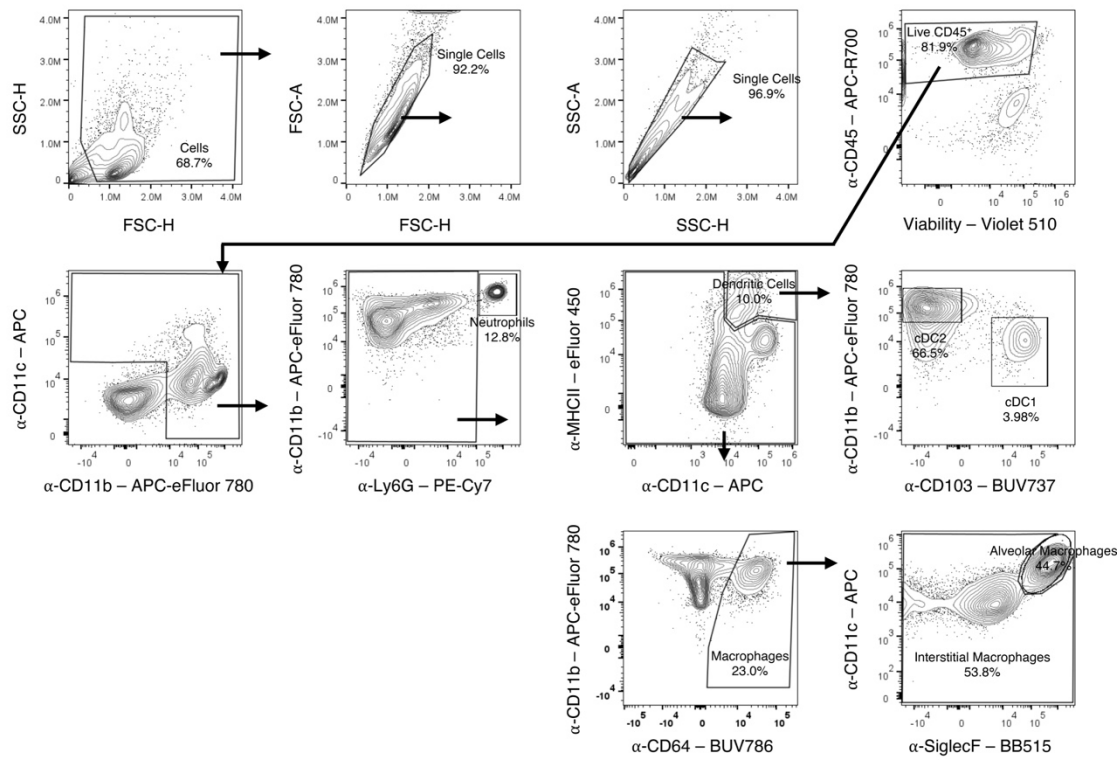
Supplementary Figure 9. Comparative analysis of mouse-adapted Beta, Omicron, and MA10 strains in WT Balb/c mice. Groups of WT mice (WT n=5, Beta KO P20 n=5, Omicron KO P20 n=5) were challenged with 10⁵ TCID50 of the SARS-CoV-2 BA.4 strain or Beta strain passaged 20 times through *Ifitm3*^{-/-} mice and compared to infection by an equal dose of MA10. (a) Weight loss during challenge by passaged SARS-CoV-2 viruses. (b) Daily whole-body plethysmography enhanced pause (PenH) measurements. (a, b) Dots represent averages of individual mice. Error bars represent SEM, comparisons were made using a two-way ANOVA with Tukey's multiple comparisons. (p values: **** <0.0001, *** <0.0005, ** <0.005, * <0.05, exact p values are provided in Source Data). Source data are provided as a Source Data file.



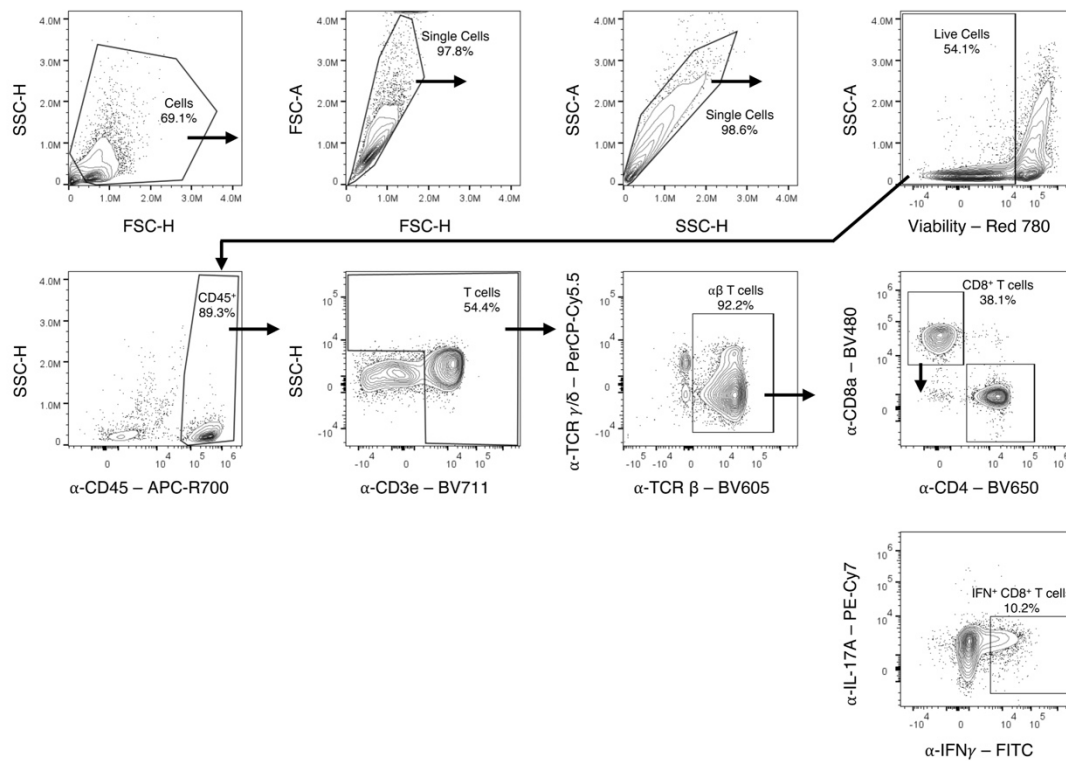
Supplementary Figure 10. SARS-CoV-2 variants elicit innate immune activation in the lungs of infected mice. (a) Volcano plots depict the magnitude (x axis) and significance (y axis) of change in gene expression at day two post-infection. Each data point represents a unique transcript, where color denotes upregulation (red) or downregulation (blue) in expression relative to mock-inoculated mice. Dashed lines depict significance cutoffs ($|lfc| \geq 1$, $padj < 0.05$) and grey data points depict non-significant genes. The number of DEG in either is highlighted in the top corners of the graphs. (b) Euclidean distance-based hierarchical clustering of the 63 DEG that overlap across all infectious conditions. Color indicates upregulation (red) in gene expression (left). Bar graphs represent the enrichment scores of GO Biological Pathways terms identified by EnrichR. Each node represents a unique term, edges represent connectivity across terms. Color indicates significant enrichment across infectious groups as indicated by the color scale.

x-axis denoted the $-\log_{10}$ padj value of enrichment of the top 10 GO Biological Pathways for each cluster as indicated by color. (e) Negatively enriched ImmGen cell types as inferred by DCQ (same as **Figure 7f**). (f-g) Heatmap representing expression, relative to PBS control mice, of the top genes contributing to the (f) GN.BM and (g) GN.THIO.PC ImmGen cell types in the DCQ algorithm. Red indicates greater expression, and blue indicates lower expression, compared to PBS-inoculated control mice.

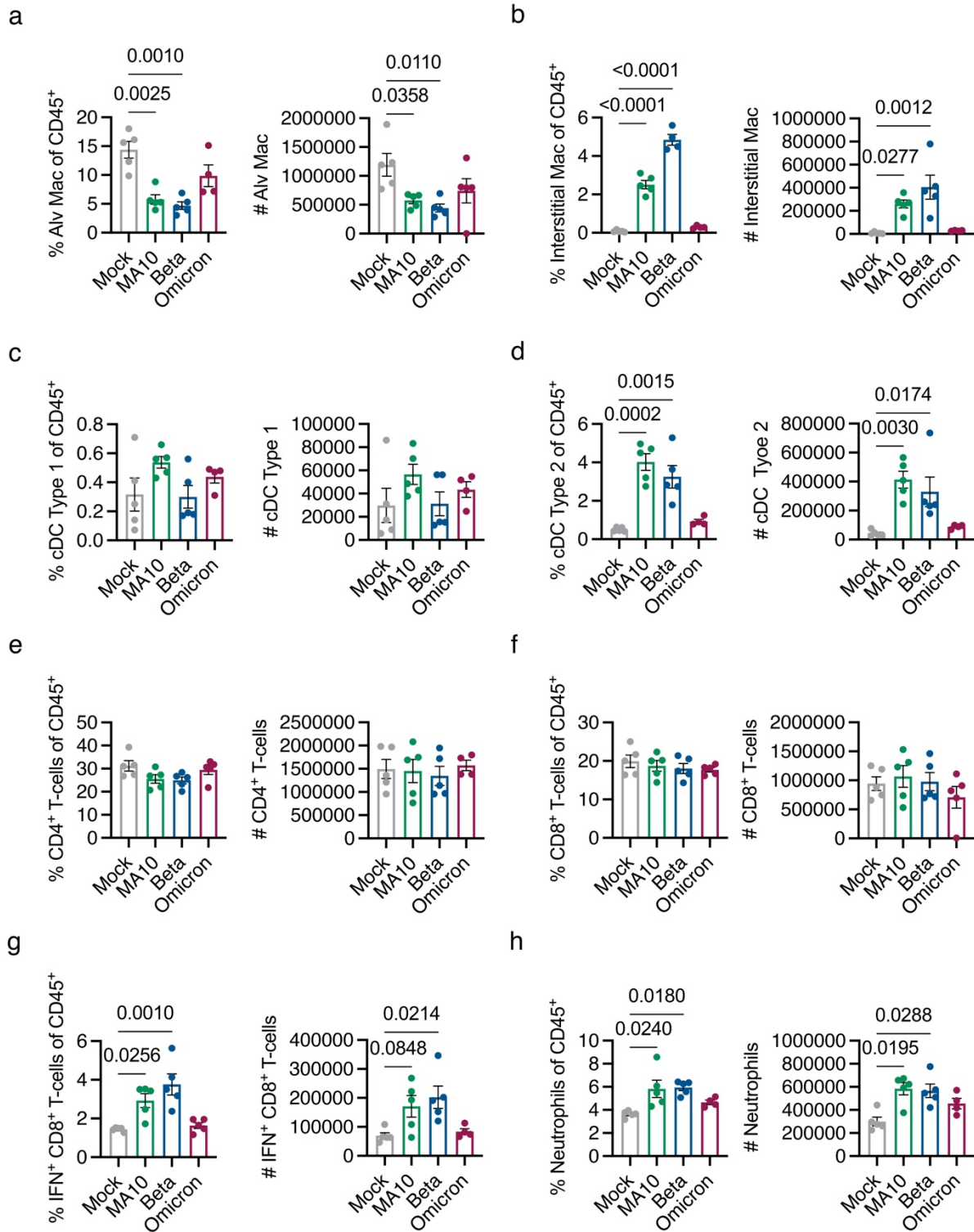
a



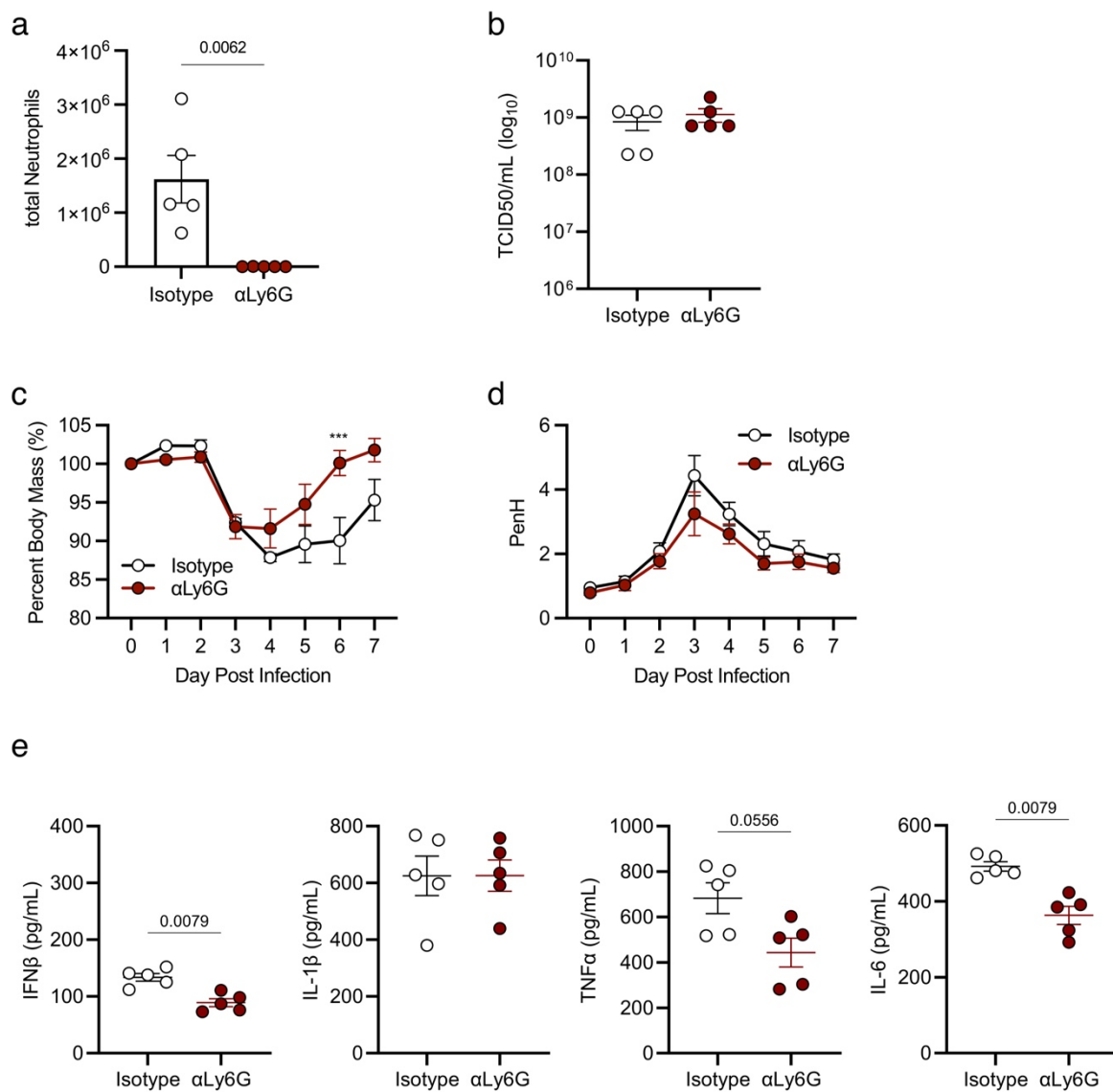
b



Supplementary Figure 13. Flow cytometry lung immunophenotyping gating strategy. (a) Gating strategy for data generated in Supplementary Fig 14a, b, c, d, and h. (b) Gating strategy for data generated in Supplementary Fig 14e, f, and g.



Supplementary Figure 14. Lung immunophenotyping. (a-h) Groups of WT mice (n=5 Mock, MA10; n=4 Beta, Omicron) were challenged with 10^5 TCID₅₀ of the indicated mouse adapted SARS-CoV-2 strain. On day 4 post infection, lungs were harvested for immunophenotyping via flow cytometry. Data is presented as % CD45+ (left) or total number (right) for alveolar macrophages (a), interstitial macrophages (b), conventional dendritic cells type 1 (c), conventional dendritic cells type 2 (d), CD4+ T cells (e), CD8+ T cells (f), IFN+ CD8+ T cells (g), and neutrophils (h) (error bars represent SEM, p-values by one-way ANOVA with multiple comparisons test). Source data are provided as a Source Data file.



Supplementary Figure 15. Neutrophil depletion reduces disease severity in mice infected with SARS-CoV-2. Groups of WT mice (n=5 per group) were challenged with 10^5 TCID₅₀ of the SARS-CoV-2 Beta strain passaged 20 times through *Ifitm3*^{-/-} mice. To deplete neutrophils, mice were treated with antibodies targeting neutrophils (αLy6G) or an isotype control antibody, starting at day 1 post infection through day 6 post infection. **(a)** Neutrophil number in the lungs of infected mice on day 4 post infection by flow cytometry; error bars indicate SEM, p value above the bar determined by two-tailed unpaired t-test. **(b)** Viral titers from lung homogenates collected at day 2 post infection. **(c)** Weight loss during challenge by passaged SARS-CoV-2 viruses. Error bars represent SEM. Comparisons made by two-way ANOVA followed by Sidak multiple comparisons, exact p-value provided in source data. **(d)** Daily whole-body plethysmography enhanced pause (PenH) measurements. **(e)** ELISA quantification of IFNβ, IL-1β, TNFα, and IL-6 in lung homogenates collected at day 2 post infection. **(a, b, e)** Dots represent individual mice (n=5). **(c, d)** Dots represent averages of individual mice (n=5 per group). Source data are provided as a Source Data file.

Supplementary Table 1. Mouse-adaptive SARS-CoV-2 mutations from the current study and published works.

Protein		
S	Citations	Mutation in current study
N30T	Willet et al. Commun. Biol. 2024	Identical mutation in multiple studies
ins 215KLRS	Rathnasinghe et al. Nat. Commun. 2022	Residue mutated in multiple studies, with different resulting amino acid changes
del 240/242	Current work	*Reversion of known tissue-culture adaptations
T250A	Beer et al. J. Exp. Med. 2022	
S371F	Willet et al. Commun. Biol. 2024	
K417N	Sun et al. Nat. Commun. 2021, Beer et al. J. Exp. Med. 2022, & Muruato et al. PLoS Biol. 2021	
K417T	Gawish et al. Elife 2022	
K417M	Wong et al. Nature 2022	
E484K	Wong et al. Nature 2022	
Q493H	Sun et al. Nat. Commun. 2021	
Q493K	Huang et al. EBioMedicine 2021, Leist et al. Cell 2020, Bader et al. PNAS 2023, & Ueki et al. Viruses 2024	
Q493R	Gawish et al. eLife 2022 & Wong et al. Nature 2022	
Q498H	Huang et al. EBioMedicine 2021, Beer et al. J. Exp. Med. 2022, Zhang et al. Cell Discov. 2021 & Gawish et al. Elife 2022	
Q498R	Wong et al. Nature 2022	
N501Y	Gu et al. Science 2020 & Rathnasinghe et al. Nat. Commun. 2022	
H655Y	Muruato et al. PLoS Biol. 2021, Zhang et al. Cell Discov. 2021, & Rathnasinghe et al. Nat. Commun. 2022	
H677Q*	Current work	
Q677H	Willet et al. Commun. Biol. 2024	
R682W	Willet et al. Commun. Biol. 2024	
W682R*	Current work	
A684V	Willet et al. Commun. Biol. 2024	
A892V	Willet et al. Commun. Biol. 2024	
S982L	Current work	
E1182G	Willet et al. Commun. Biol. 2024	

N	
R14C	Current work
T16M	Willet et al. Commun. Biol. 2024
R32H	Current work & Willet et al. Commun. Biol. 2024
R32C	Sun et al. Nat. Commun. 2021
D128Y	Gu et al. Science 2020 & Muruato et al. PLoS Biol. 2021
S194T	Rathnasinghe et al. Nat. Commun. 2022
E	
E8V	Muruato et al. PLoS Biol. 2021
S16G	Current work
T30I	Current work
M	
T7I	Current work, Ueki et al. Viruses 2024, Beer et al. J. Exp. Med. 2022, & Rathnasinghe, et al. Nat. Commun. 2022
H125Y	Bader et al. PNAS 2023
Nsp2	
I42V	Willet et al. Commun. Biol. 2024
Nsp3	
P108L	Willet et al. Commun. Biol. 2024
N262T	Zhang et al. Cell Discovery 2021
H682Y	Willet et al. Commun. Biol. 2024
A1197V	Beer et al. J. Exp. Med. 2022
I1258V	Sun et al. Nat. Commun. 2021
Nsp4	
T285I	Leist et al. Cell 2020
T295I	Ueki et al. Viruses 2024, Beer et al. J. Exp. Med 2022, & Wong et al. Nature 2022
A307V	Current work
H313Y	Bader et al. PNAS 2023
H470Y	Sun et al. Nat. Commun. 2021
Nsp5	
P96L	Willet et al. Commun. Biol. 2024
C160F	Willet et al. Commun. Biol. 2024
P252L	Current work & Willet et al. Commun. Biol. 2024
F294L	Ueki et al. 2024 & Huang et al. EBioMedicine 2021
S301L	Sun et al. Nat. Commun. 2021

Nsp6	
I7T	Huang et al. EBioMedicine 2021
L37F	Gu et al. Science 2020
G107S	Current work
F108L	Current work
A128V	Sun et al. Nat. Commun. 2021
Nsp7	
K2R	Leist et al. Cell 2020
S8F	Sun et al. Nat. Commun. 2021
Nsp8	
Q19R	Bader et al. PNAS 2023 & Zhang et al. Cell Discov. 2021
E23G	Leist et al. Cell 2020
S76F	Wong et al. Nature 2022
E155A	Willet et al. Commun. Biol. 2024
Nsp9	
D25Y	Beer et al. J. Exp. Med 2022
T34A	Beer et al. J. Exp. Med 2022
T35I	Current work
K36R	Current work
R39K	Ueki et al. Viruses 2024
T67A	Wong et al. Nature 2022, Willet et al. Commun. Biol. 2024, & Zhang et al. Cell Discov. 2021
T77I	Huang et al. EBioMedicine 2021
Nsp10	
P84S	Gu et al. Science 2020
P87S	Muruato et al. PLoS Biol. 2021
Nsp12	
A95V	Current work
Nsp13	
A362V	Zhang et al. Cell Discov. 2021
P504S	Bader et al. PNAS 2023
Nsp14	
V269I	Zhang et al. Cell Discov. 2021
Orf3a	
S171L	Current work
ORF6	
F7S	Leist et al. Cell 2020
Orf7a	
V82A	Willet et al. Commun. Biol. 2024

T120I	Willet et al. Commun. Biol. 2024
Orf7b	
L4P	Current work
Orf8	
T11I	Willet et al. Commun. Biol. 2024
L84S	Rathnasinghe et al. Nat. Commun. 2022
R115L	Willet et al. Commun. Biol. 2024
F120V	Current work

Supplementary Table 2. Gene Expression Modules.

Module	Gene Counts	Top Hubs	Gene Symbol	kWithin
brown	324	ENSMUSG0000007360	Prob1	143.28586
		0		2
		ENSMUSG0000003234	Gsta4	139.92693
		8		9
		ENSMUSG0000002223	Cmbl	138.53185
		5		9
		ENSMUSG0000006342	Ddo	138.31869
		8		7
		ENSMUSG0000002123	Entpd5	135.55148
		6		5
		ENSMUSG0000005617	Col8a2	135.26774
		4		7
		ENSMUSG0000006132	Dnaic1	134.17373
		2		3
		ENSMUSG0000003403	Ccdc17	133.82238
		5		8
		ENSMUSG0000005297	Cyp2f2	133.79285
		4		8
		ENSMUSG0000006420	4430402I18Rik	133.67169
		2		6
turquoise	2342	ENSMUSG0000004002	Saa3	1263.1094
		6		9
		ENSMUSG0000001917	Rab5c	1246.6254
		3		1
		ENSMUSG0000002798	Hadh	1242.2370
		4		1
		ENSMUSG0000003342	Mcee	1239.3562
		9		2
		ENSMUSG0000000265	Gtf2f1	1230.8075
		8		7
		ENSMUSG0000006298	Cped1	1227.7504
		0		2
		ENSMUSG0000005517	C1ra	1223.9543
		2		9
		ENSMUSG0000003782	Tgm2	1223.0032
		0		4
		ENSMUSG0000004020	Zfp704	1221.3390
		9		2
		ENSMUSG0000003154	Sfrp1	1219.9022
		8		4

blue	2049	ENSMUSG0000006259	Lilrb4a	1189.1564
		3		5
		ENSMUSG0000000113	Timp1	1186.0969
		1		5
		ENSMUSG0000001251	Mkl	1182.8187
		9		6
		ENSMUSG0000002439	Aif1	1180.4139
		7		4
		ENSMUSG0000004582	Serpinb9	1178.7616
		7		6
		ENSMUSG0000003232	Pstpip1	1178.6992
		2		6
		ENSMUSG0000007042	Il18bp	1178.6153
		7		1
		ENSMUSG0000003494	Tmem106a	1178.5832
green	44	7		4
		ENSMUSG0000003846	Chmp4b	1175.9998
		7		9
		ENSMUSG0000002742	Rrbp1	1175.9221
		2		5
		ENSMUSG0000006434	mt-Tn	8.6627665
		8		
		ENSMUSG0000006434	mt-Tc	8.5553748
		9		4
		ENSMUSG0000009069	Apold1	7.6831621
		8		6
		ENSMUSG0000006434	mt-Ta	7.6726623
		7		4
		ENSMUSG0000006434	mt-Tl1	7.3073819
		0		
yellow	128	ENSMUSG0000004478	Zfp36	7.1654447
		6		3
		ENSMUSG0000006434	mt-Tw	6.9002537
		6		8
		ENSMUSG0000006542	Mir142b	6.4974383
		0		3
		ENSMUSG0000004848	8430408G22Ri	6.3759558
		9	k	3
		ENSMUSG0000009734	Gm17275	6.3713121
		7		6
		ENSMUSG0000005602	Clca3a1	55.669152
		5		2
		ENSMUSG0000003150	Col4a2	53.498565
		3		2

		ENSMUSG0000002664	Plxna2	51.199328
		0		
		ENSMUSG0000006412	Mocs1	49.710265
		0		9
		ENSMUSG0000001770	Serinc3	49.534956
		7		1
		ENSMUSG0000002422	Fkbp5	49.293232
		2		1
		ENSMUSG0000006187	Sphk1	48.288020
		8		7
		ENSMUSG0000003368	Qsox1	47.236073
		4		5
		ENSMUSG0000002068	Rasl10b	47.222874
		4		2
		ENSMUSG0000002293	Fam3b	45.651529
		8		
Unassigned genes (grey)	7	ENSMUSG00000010498	1600023N17Ri	0.1549473
		6	k	8
		ENSMUSG0000008805	Mir1968	0.0962048
		4		3
		ENSMUSG0000008540	Gm13068	0.0850594
		3		6
		ENSMUSG0000008046	Snord98	0.0429642
		9		7
		ENSMUSG0000008923	Gm22200	0.0378685
		9		
		ENSMUSG00000010376	Gm37856	0.0281152
		8		2
		ENSMUSG0000008779	Gm25965	0.0246193
		5		4