

Supporting Information for

Harnessing a Germ-Free Mouse Gut Bioreactor for Directed Evolution of Probiotics to Combat Non-Alcoholic Fatty Liver Disease

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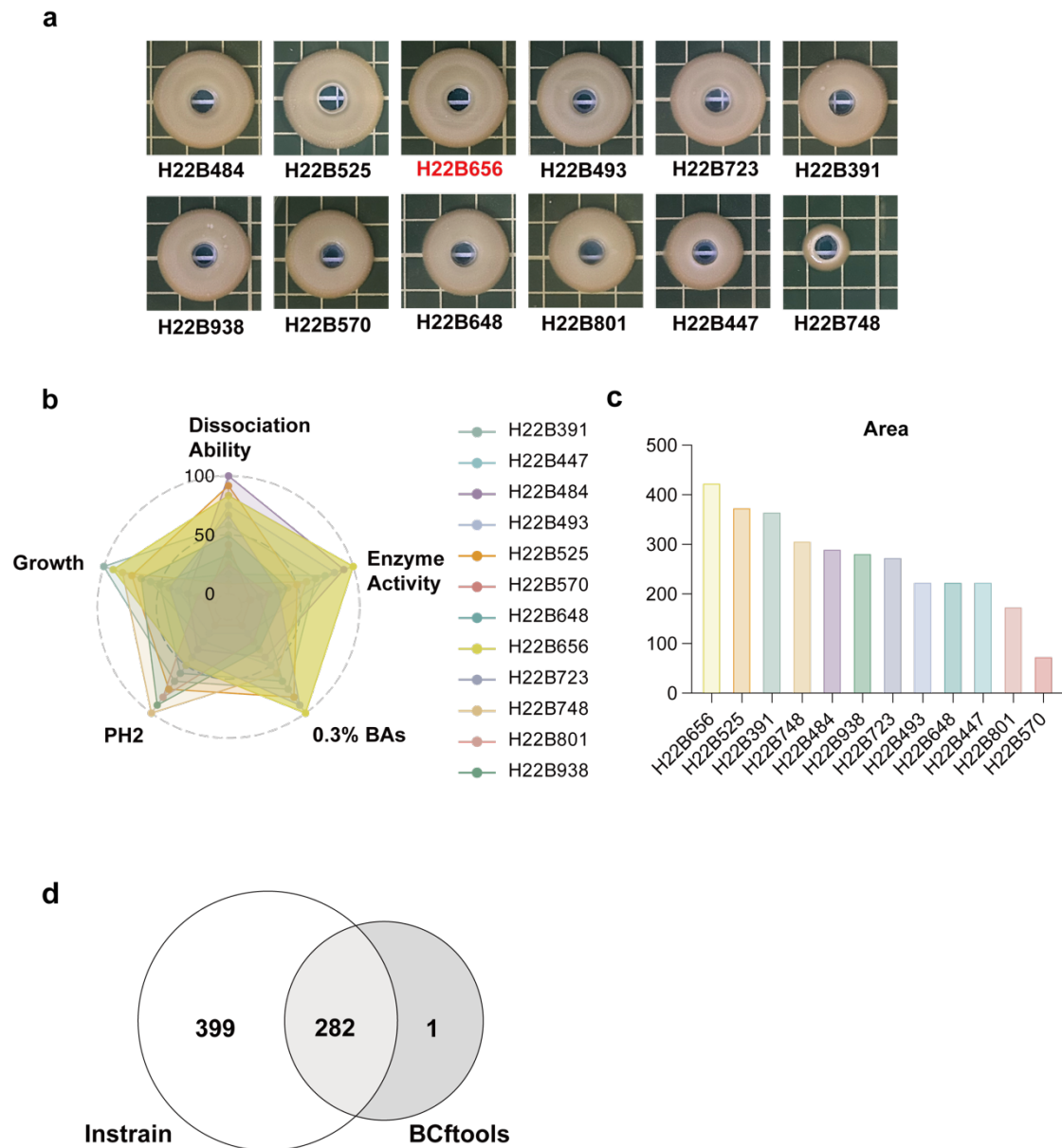
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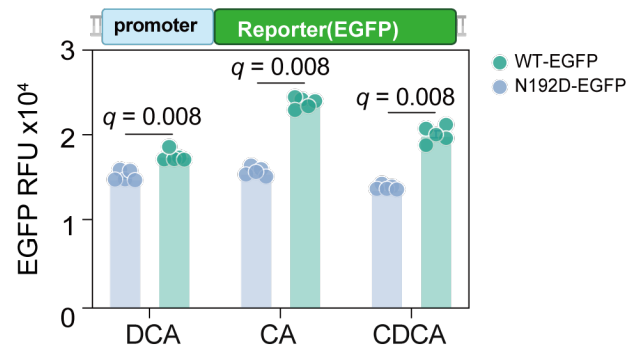
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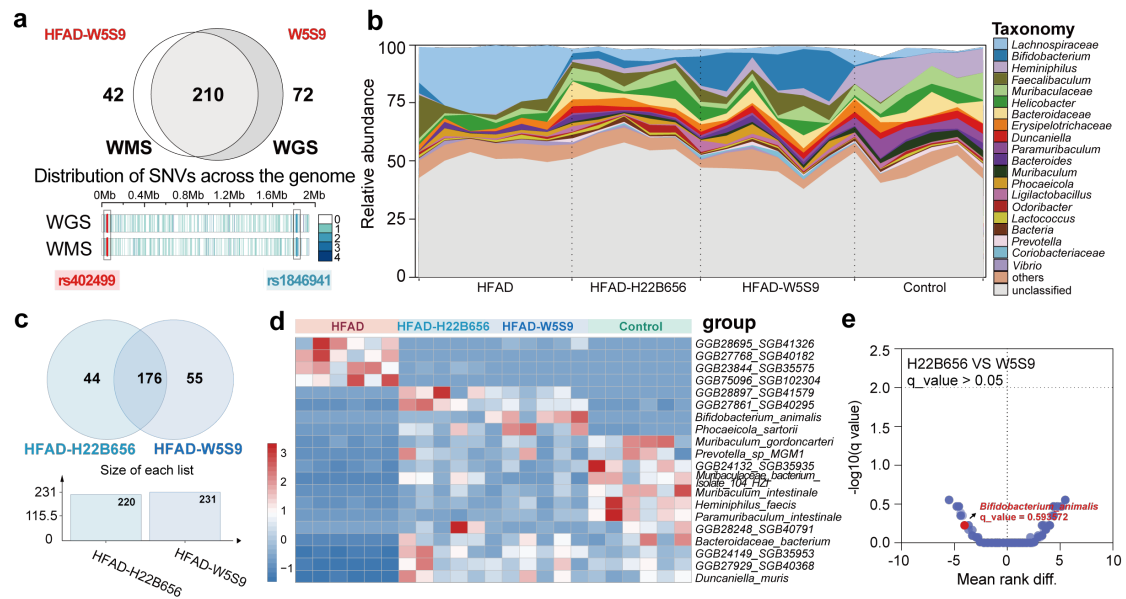
Supplementary Figures 1 to 8



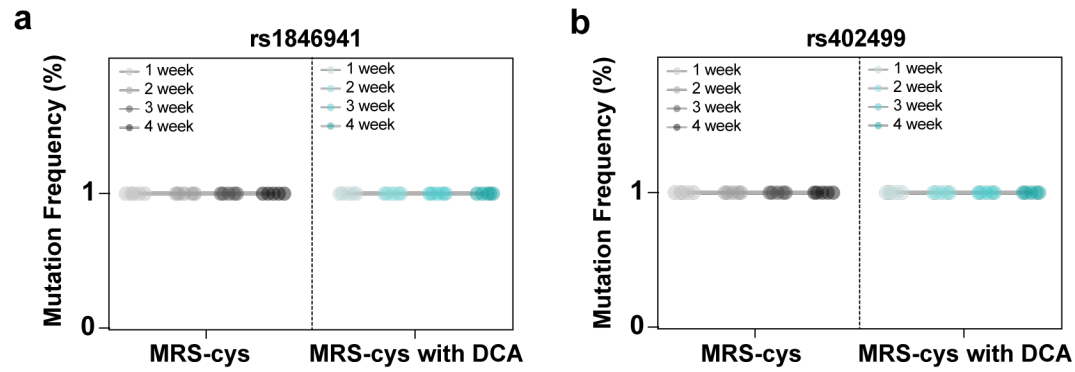
Supplementary Fig. 1 Phenotypic Profiling and High-Confidence SNV Detection. (a) Calcium solubilization by each strain after 48 h of anaerobic incubation in bile-salt medium supplemented with sodium thioglycolate and CaCl_2 ($n = 3$). (b, c) Composite scores for each strain, calculated by converting individual phenotypic metrics to percentile ranks and summing them. (d) Application of InStrain and BCftools pipelines to the W5S9 sample, jointly identifying 282 high-confidence SNVs. Source data are provided as a Source Data file.



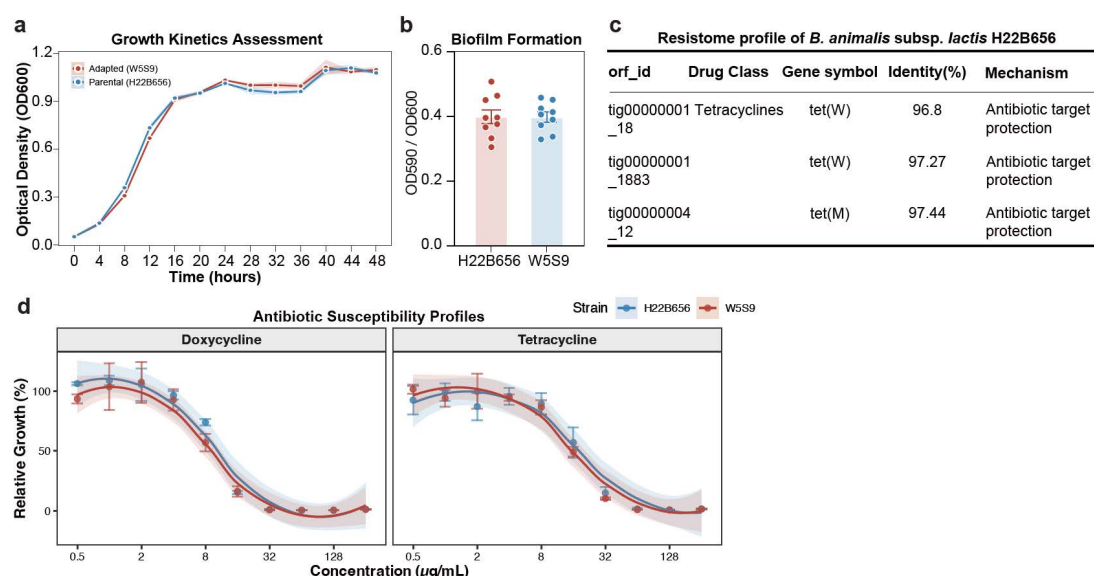
Supplementary Fig. 2 N192D substitution enhances *mdr* translation under bile acid stress in *E. coli*. EGFP fluorescence intensities in *E. coli* BL21(DE3) cells harboring reporter plasmids encoding WT-EGFP or N192D-EGFP following treatment with DCA, CA, and CDCA. Data are presented as mean $\pm$ SEM ($n = 5$). Differences between groups were analyzed using multiple Mann-Whitney tests. To correct for multiple comparisons, the False Discovery Rate (FDR) was controlled using the two-stage step-up method (Benjamini, Krieger, and Yekutieli), with a desired FDR (Q) set at 1.00%. Dots represent individual data. Source data are provided as a Source Data file.



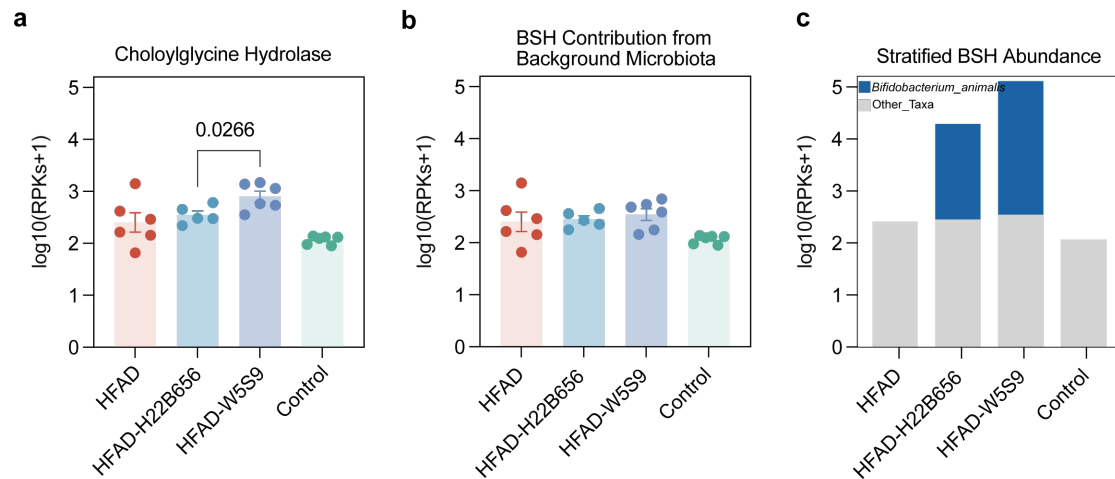
Supplementary Fig. 3 Effects of *B. animalis* subsp. *lactis* H22B656 and *B. animalis* subsp. *lactis* W5S9 interventions on intestinal microbiota composition. (a) The Venn diagram depicts the overlap of SNVs called by whole-metagenome shotgun (WMS) sequencing in the HFAD-W5S9 group and by whole-genome sequencing (WGS) of the adapted W5S9 strain. Two key SNVs were annotated in the HFAD-W5S9 samples but were not detected in the HFAD-H22B656 group, indicating that no cross-contamination occurred between the gavaged strains. (b) Stacked area chart showing the relative abundance of bacterial taxa at the family and genus levels across the HFAD, HFAD-H22B656, HFAD-W5S9, and Control group. (c) Venn diagram illustrating the number of shared and unique species between the HFAD-H22B656 and HFAD-W5S9 groups. (d) Heatmap showing the relative abundance of specific bacterial species across all groups. The color scale represents the normalized abundance (Z-score), with red indicating higher abundance and blue indicating lower abundance. (e) Volcano plot comparing the differential abundance of gut microbiota between the HFAD-H22B656 and HFAD-W5S9 groups. Differences between groups (e.g., H22B656 vs. W5S9) were analyzed using multiple Mann-Whitney tests. To correct for multiple comparisons, the False Discovery Rate (FDR) was controlled using the two-stage step-up method (Benjamini, Krieger, and Yekutieli), with a desired FDR (Q) set at 1.00%. The red dot highlights *Bifidobacterium animalis*, indicating no significant difference (q -value > 0.05) between the two probiotic treatments. Data are presented as $n = 6$ mice per group. Source data are provided as a Source Data file.



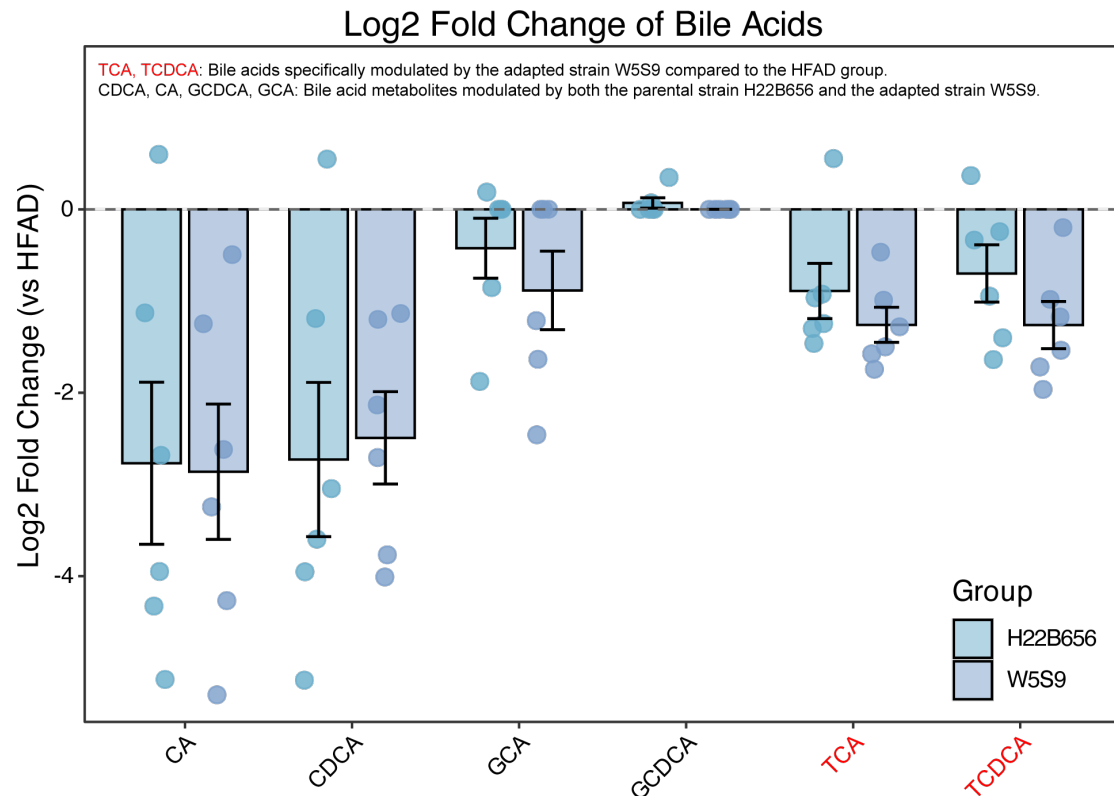
Supplementary Fig. 4 Genetic Stability of rs1846941 and rs402499. Mutation frequency of the key SNVs (a) rs1846941 and (b) rs402499 in the adapted strain over a 4-week period under continuous passage conditions. The mutation frequency was assessed in MRS-cys medium (control) and in MRS-cys medium supplemented with 0.3% deoxycholic acid (DCA) as the bile salt stress ($n = 5$ per week). Source data are provided as a Source Data file.



Supplementary Fig. 5 Safety evaluation of the parental *B. animalis* subsp. *lactis* H22B656 and the adapted *B. animalis* subsp. *lactis* W5S9. (a) Growth curves over 48 h showing the growth kinetics of the two strains. Data are presented as mean $\pm$ SD. Differences between strains at each time point were analyzed using multiple unpaired t-tests. To correct for multiple comparisons, the False Discovery Rate (FDR) was controlled using the two-stage step-up method (Benjamini, Krieger, and Yekutieli), with a desired FDR (Q) set at 1.00%. (b) Crystal-violet biofilm formation assay after 12 h of anaerobic incubation; biofilm biomass is expressed as OD590/OD600. Data are presented as mean $\pm$ SEM. Statistical significance was determined using an unpaired two-tailed Student's t-test. (c) Resistome profile based on annotation with the Resistance Gene Identifier against the CARD database; only hits with >95% amino-acid identity to reference sequences are shown. orf_id, open reading frame identifier; Identity (%), percentage amino-acid identity to the reference gene. (d) Dose-response growth-inhibition curves of the two strains in the presence of doxycycline and tetracycline. Data are presented as mean $\pm$ SD. Statistical analysis was performed using multiple unpaired t-tests with FDR correction (Q = 1.00%), as described in (a). Data represent $n = 3$ independent biological replicates per group. Source data are provided as a Source Data file.



Supplementary Fig. 6. Metagenomic quantification and source attribution of bile salt hydrolase genes. (a) Total abundance of *cbh*-encoding genes (EC 3.5.1.24) in the gut microbiome across groups. Data are expressed as log₁₀-transformed Reads Per Kilobase (RPKs). Data are presented as mean $\pm$ SEM. Statistical significance was determined using an unpaired two-tailed Student's t-test. (b) Abundance of *cbh* genes attributed exclusively to the resident background microbiota (excluding the administered *Bifidobacterium animalis*). Data are presented as mean $\pm$ SEM. Statistical significance was determined using an unpaired two-tailed Student's t-test. (c) Stratified analysis of BSH sources. The stacked bar chart illustrates that the elevated BSH capacity in the HFAD-W5S9 group is primarily driven by the direct contribution of *Bifidobacterium animalis*, while the contribution from other taxa remains comparable to the control and HFAD groups. Data represent $n = 6$ biological replicates per group. Source data are provided as a Source Data file.



Supplementary Fig.7 Targeted bile acid metabolomics showing log₂ fold change in fecal bile acid concentrations relative to the HFAD group. Bar plots represent the mean $\pm$ SEM for each bile acid metabolite, with individual dots indicating biological replicates. Differences in log₂ fold change of bile acid concentrations between groups were analyzed using multiple unpaired t-tests. To correct for multiple comparisons, the False Discovery Rate (FDR) was controlled using the two-stage step-up method (Benjamini, Krieger, and Yekutieli), with a desired FDR (Q) set at 1.00%. Data represent $n = 6$ biological replicates per group. Source data are provided as a Source Data file.



Supplementary Fig.8 Co-occurrence network analysis revealing the interaction patterns of *Bifidobacterium animalis* subsp. *lactis* with the gut ecosystem and host physiological parameters. The networks illustrate the specific correlations between *B. animalis* subsp. *lactis* and gut microbiota species, serum biochemical indices, and metabolites in the **(a)** HFAD-H22B656 and **(b)** HFAD-W5S9 groups. Nodes represent microbial species, serum biochemical parameters, bile acid metabolites, and microbial functional genes. Edges indicate significant associations determined by Spearman's rank correlation analysis ($|r| > 0.4$, $p < 0.05$). Red edges denote positive correlations, and blue edges denote negative correlations, with edge transparency corresponding to the magnitude of the correlation coefficient. Data are derived from $n = 6$ biological replicates per group. Source data are provided as a Source Data file.