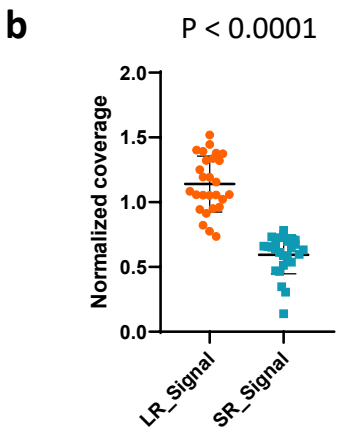
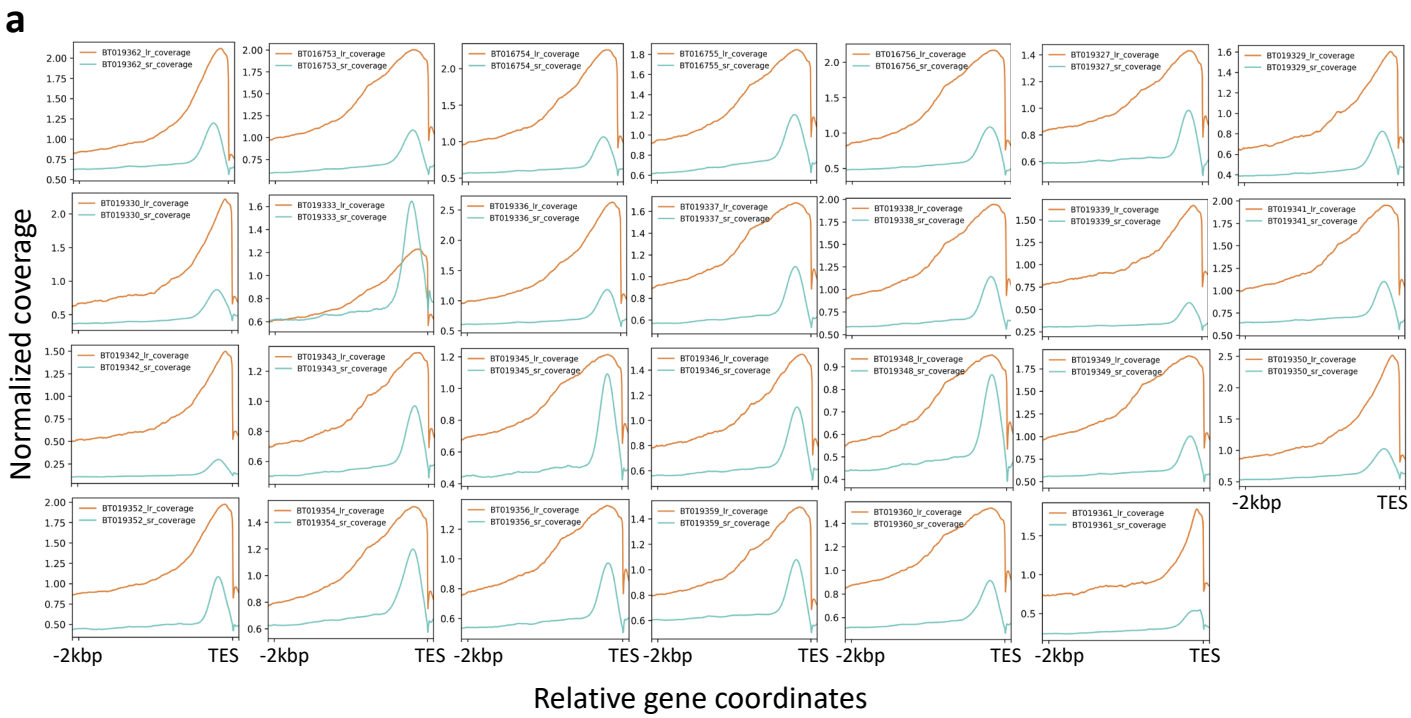
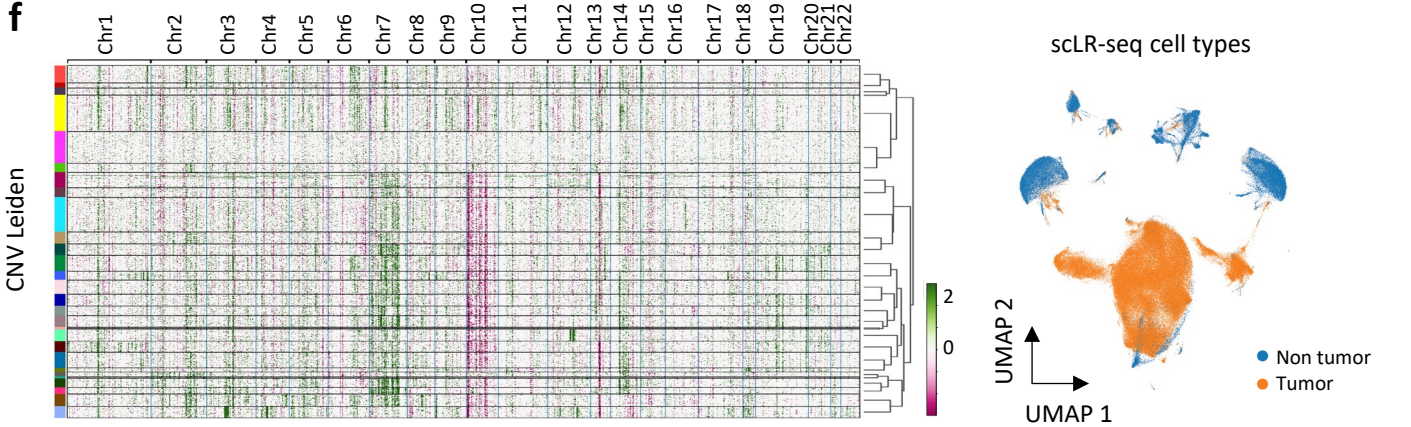
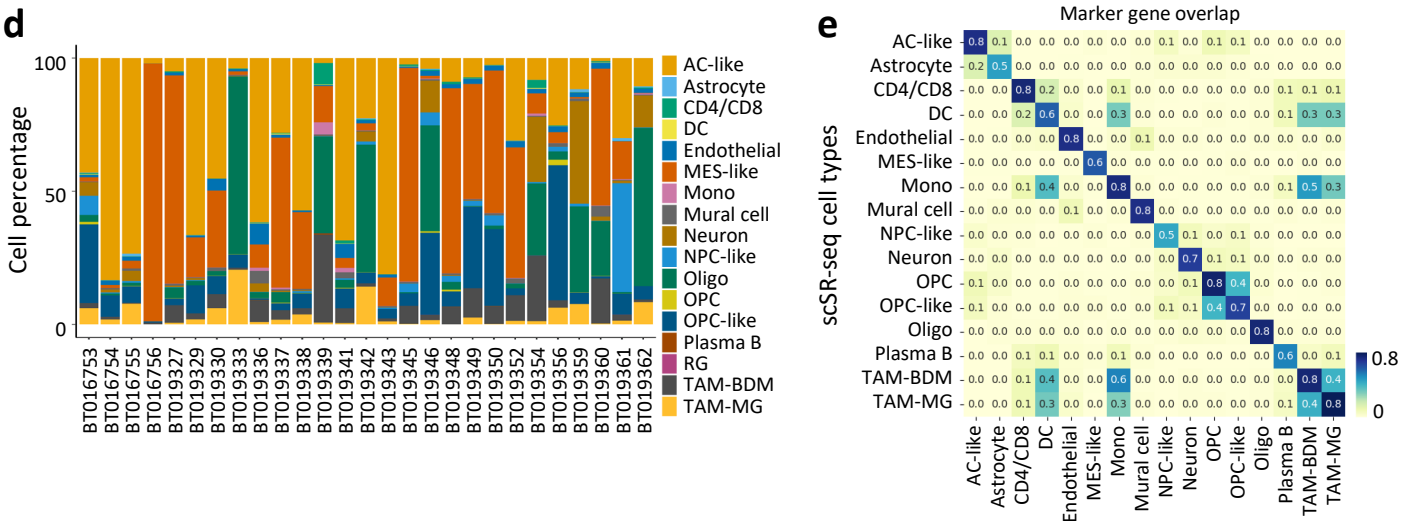
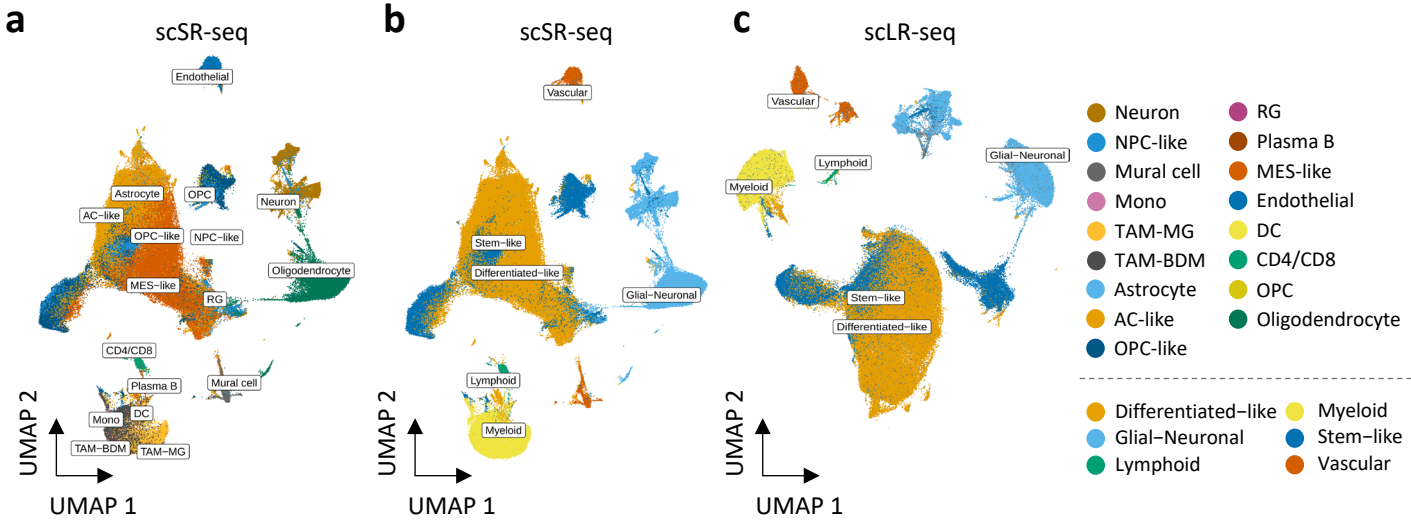




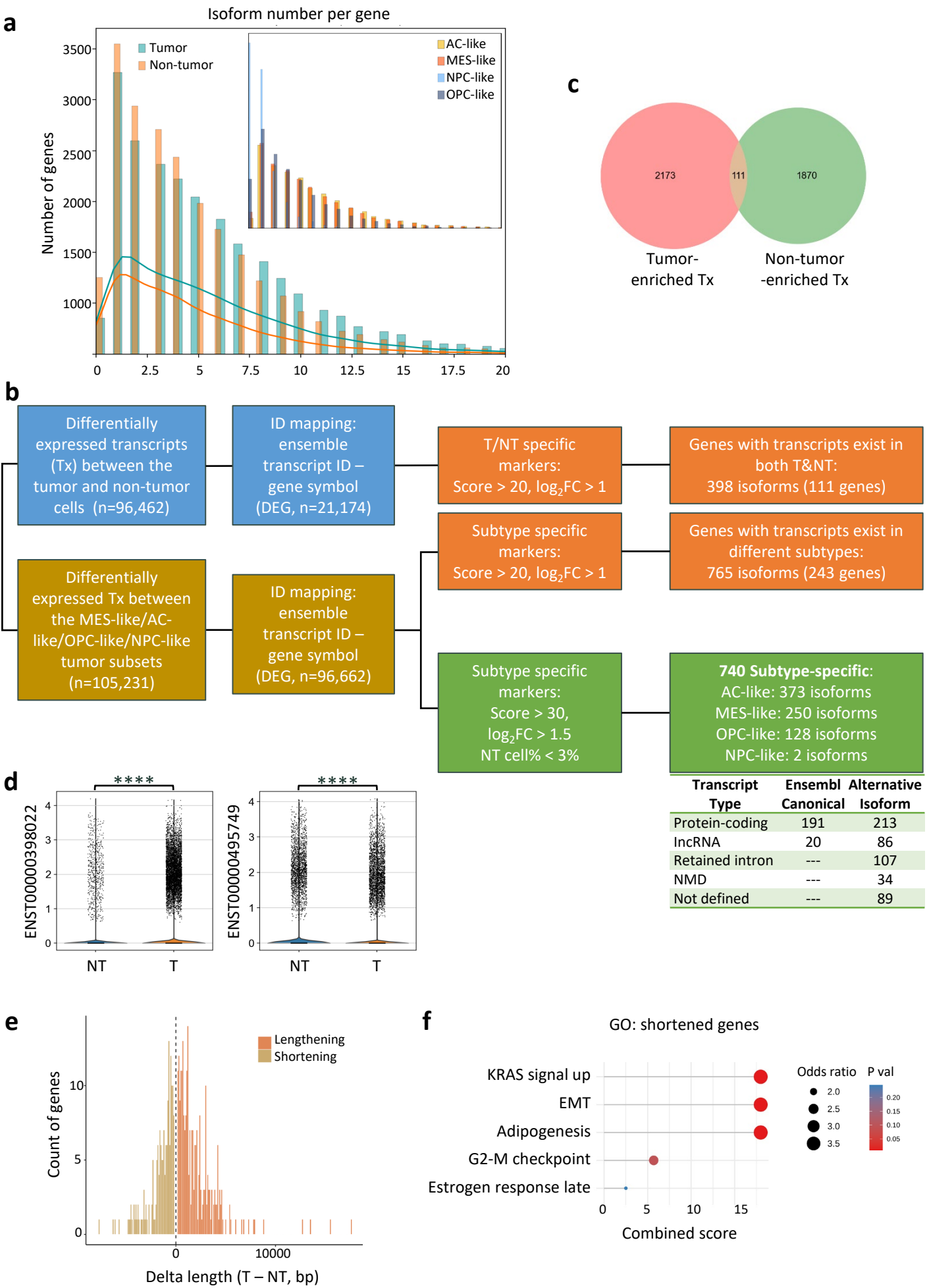
Supplementary Fig. 1. Normalized coverage comparison between scSR-seq and scLR-seq
 (a) Read-depth-normalized coverage profiles from short-read and long-read BAM files (27 samples) spanning -2,000 bp upstream to the transcription end site (TES). (b) Dot plot showing normalized coverage distributions from TSS to TES (per 10 bp bin) for scSR-seq and scLR-seq (27 samples). Statistical significance between methods was assessed using a two-sided Mann-Whitney U test (exact P value). Data are presented as mean values $\pm$ SD.



Method	Bio conservation					Batch correction					Aggregate score		
	Isolated labels	KMeans NMI	KMeans ARI	Silhouette label	cLISI	Silhouette batch	iLISI	KBET	Graph connectivity comparison	PCR	Batch correction	Bio conservation	Total
x_harmony scSR	0.52	0.54	0.30	0.53	0.99	0.83	0.07	0.26	0.65	0.68	0.50	0.58	0.55
x_harmony scLR	0.60	0.53	0.30	0.53	0.99	0.85	0.08	0.36	0.64	0.71	0.53	0.59	0.56

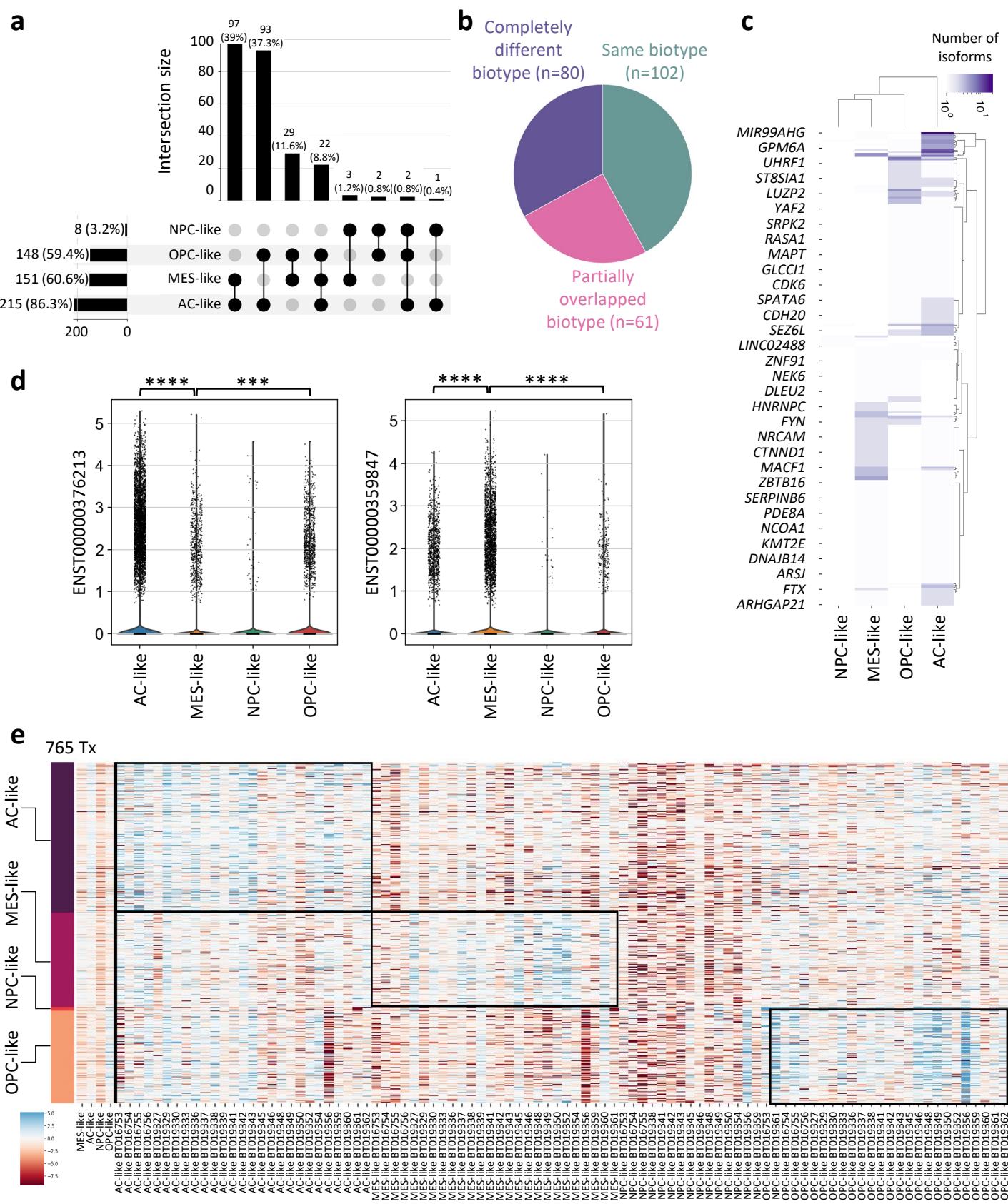
Supplementary Fig. 2. Cell clustering, annotation, and concordance between scSR-seq and scLR-seq

(a,b) UMAP projections of 182,222 single cells clustered using scSR-seq data, colored by fine-grained (a) and broad (b) cell type annotations (n=27 samples). (c) UMAP visualization of 182,222 single cells generated using scLR-seq, colored by broad cell type categories (n=27 samples). (d) Bar plot comparing cell type proportions across individual patients. (e) Heatmap showing Jaccard Similarity Indices for marker genes identified in scSR-seq versus scLR-seq. Scores were calculated based on the top 100 of significantly enriched markers for each cell state (AC-like: 56,795 cells; Astrocyte: 340 cells; CD4/CD8: 940 cells; DC: 142 cells; Endothelial: 3,661 cells; MES-like: 50,287 cells; Mono: 671 cells; Mural: 1,875 cells; NPC-like: 4,565 cells; Neuron: 8,485 cells; OPC: 414 cells; OPC-like: 18,912 cells; Oligodendrocyte: 19,373 cells; Plasma B: 59 cells; TAM-BDM: 9,753 cells; TAM-MG: 5,943 cells). (f) Copy-number variation profiling across individual chromosomes and corresponding UMAP projection of scLR-seq data, with cells (182,222 from 27 samples) classified as normal or malignant based on inferCNV. (g) Dot plot showing Biological Conservation Scores and Batch Correction Scores between scSR-seq and scLR-seq modalities.



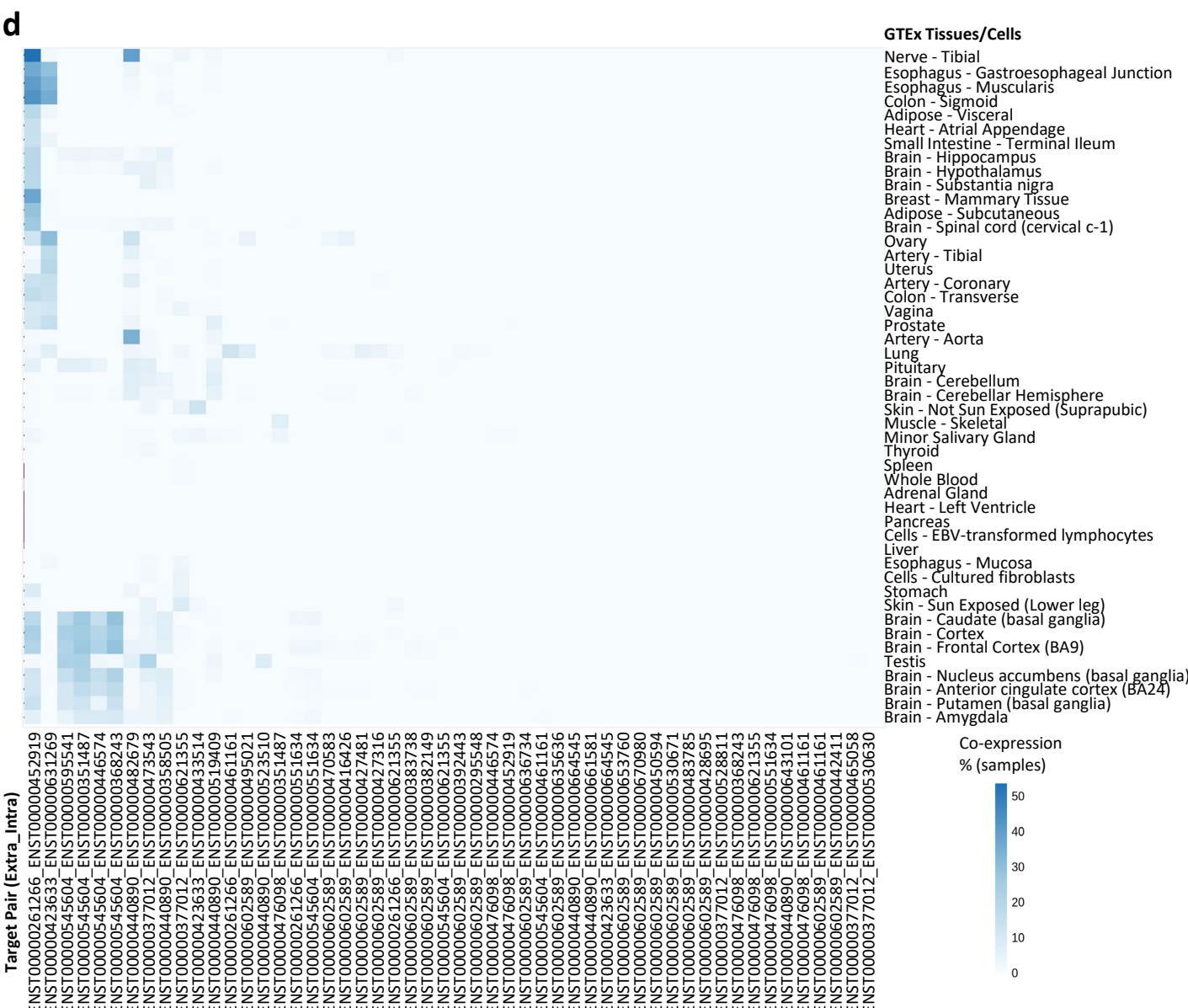
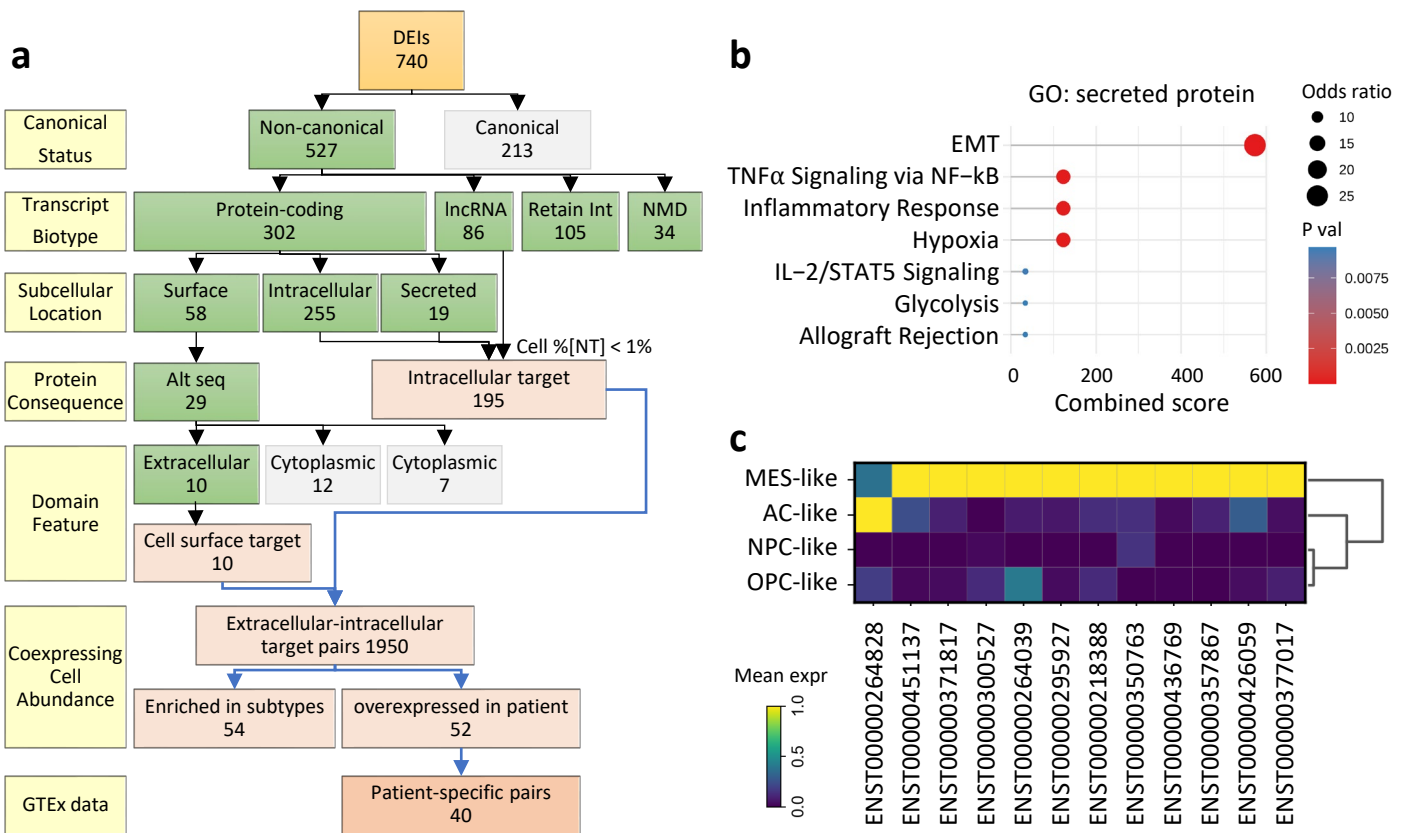
Supplementary Fig. 3. Transcript diversity and differential transcript usage analysis in GBM

(a) Bar plot showing the number of expressed transcripts per gene in tumor versus non-tumor cells (isoforms expressed in at least 10 cells), as well as across malignant subtypes. (b) Schematic workflow for detecting DTU between tumor and non-tumor cells, as well as across malignant subtypes including AC-like, MES-like, NPC-like, and OPC-like cells. The pipeline also identifies subtype-specific marker isoforms. (c) Venn diagram showing genes exhibiting differential transcript usage between tumor and non-tumor cells. The overlapping region indicates genes for which distinct isoforms are detected as differentially expressed in both tumor and non-tumor populations. (d) Dot plot showing the expression of *TM9SF4* isoforms in tumor (130,559) and non-tumor (51,663) cells. (e) Histogram showing the number of genes and corresponding changes in 3' UTR length between tumor (130,559) and non-tumor (51,663) cells. (f) Top enriched gene ontology terms (MSigDB: Hallmark gene sets) for 171 genes with shortened 3' UTRs in 27 samples. Statistical significance was assessed using a one-sided Fisher's exact test.



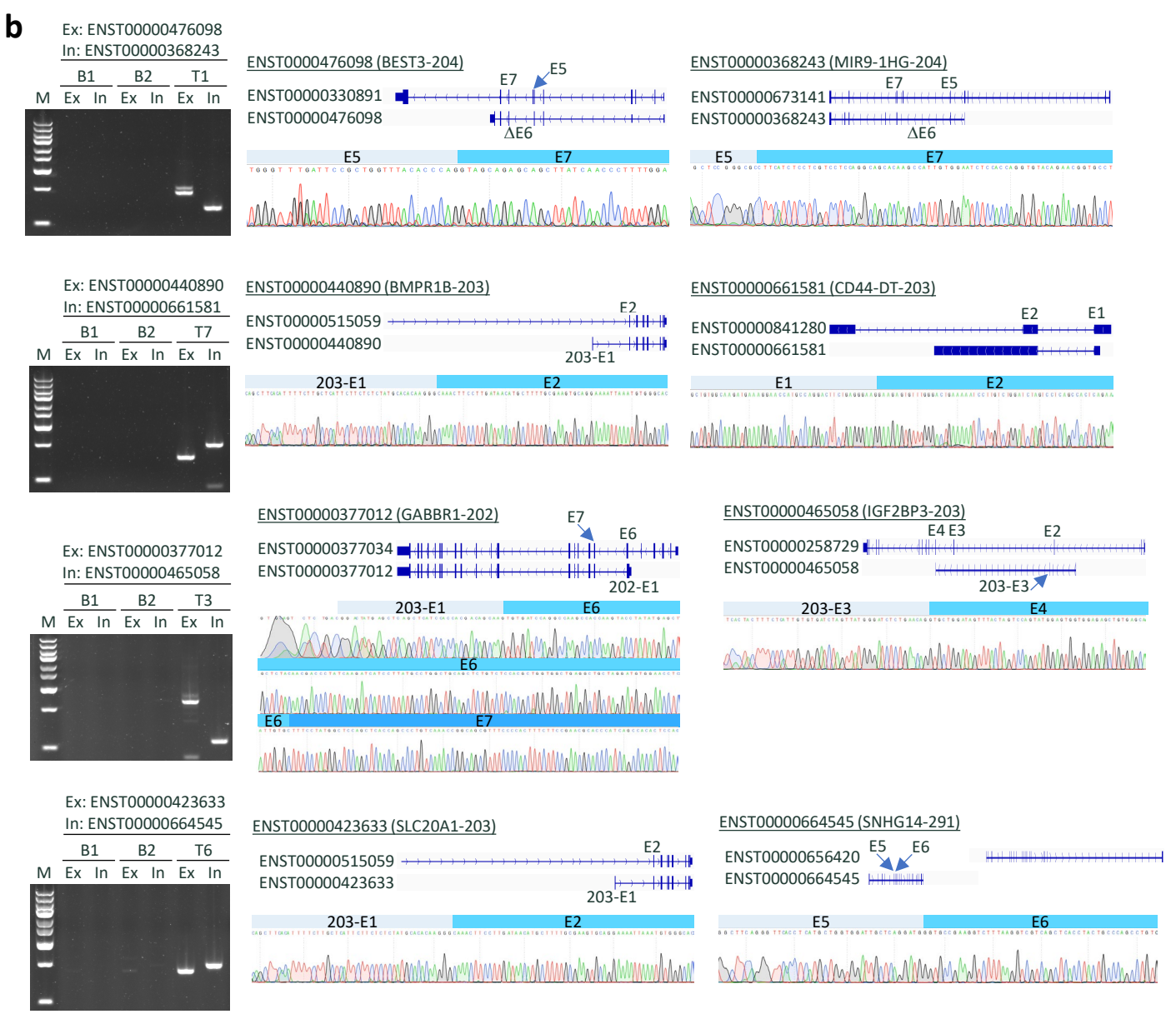
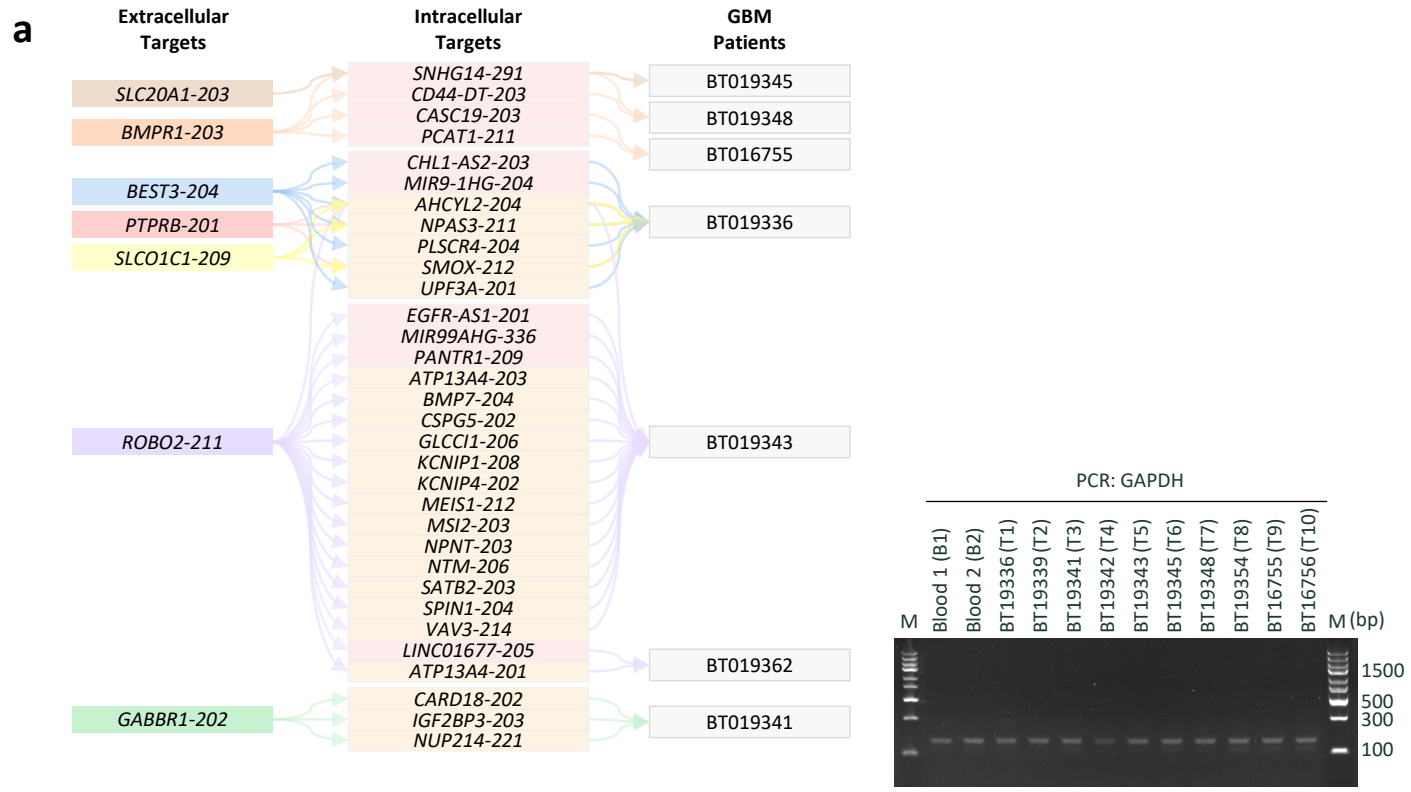
Supplementary Fig. 4. Isoform diversity and subtype-specific transcript usage in malignant GBM cells

(a) UpSet plot showing genes with expressed transcripts shared across different malignant subtypes. (b) Pie chart categorizing these genes based on whether they exhibit completely distinct biotypes, partially overlapping biotypes, or the same biotype across malignant subtypes. (c) Heatmap showing the number of expressed isoforms per gene across tumor subtypes. Each row represents a gene, and the color scale indicates the number of isoforms detected within each subtype. (d) Dot plot showing the expression of *NTRK2* isoforms across tumor subtypes. (e) Heatmap showing mean expression scores of 765 differentially used transcripts across malignant subtypes for each individual patient.



Supplementary Fig. 5. Workflow and validation for prioritizing extracellular–intracellular target pairs in GBM

(a) Schematic illustrating the multi-step workflow used to identify and prioritize DEIs for patient-specific and subtype-specific target pairs. (b) Top enriched gene ontology terms (MSigDB Hallmark gene sets) for the 61 identified secreted proteins. Statistical significance was assessed using a one-sided Fisher’s exact test. (c) Heatmap showing the expression of EMT-associated secreted proteins across four cell groups (AC-like: 56,795 cells; MES-like: 50,287 cells; OPC-like: 18,912 cells; NPC-like: 4,565 cells). (d) The heatmap presents a descriptive analysis of gene co-occurrence frequencies across human tissues. It shows the percentage of GTEx samples co-expressing each of the 52 prioritized target pairs across normal human tissues, based on bulk RNA-sequencing data. The heatmap was generated using hierarchical clustering of the proportion of samples per tissue exhibiting concurrent expression of both genes, defined as TPM > 1. Tissues with fewer than 100 samples in the GTEx v8 dataset were excluded to ensure robust percentage estimates.

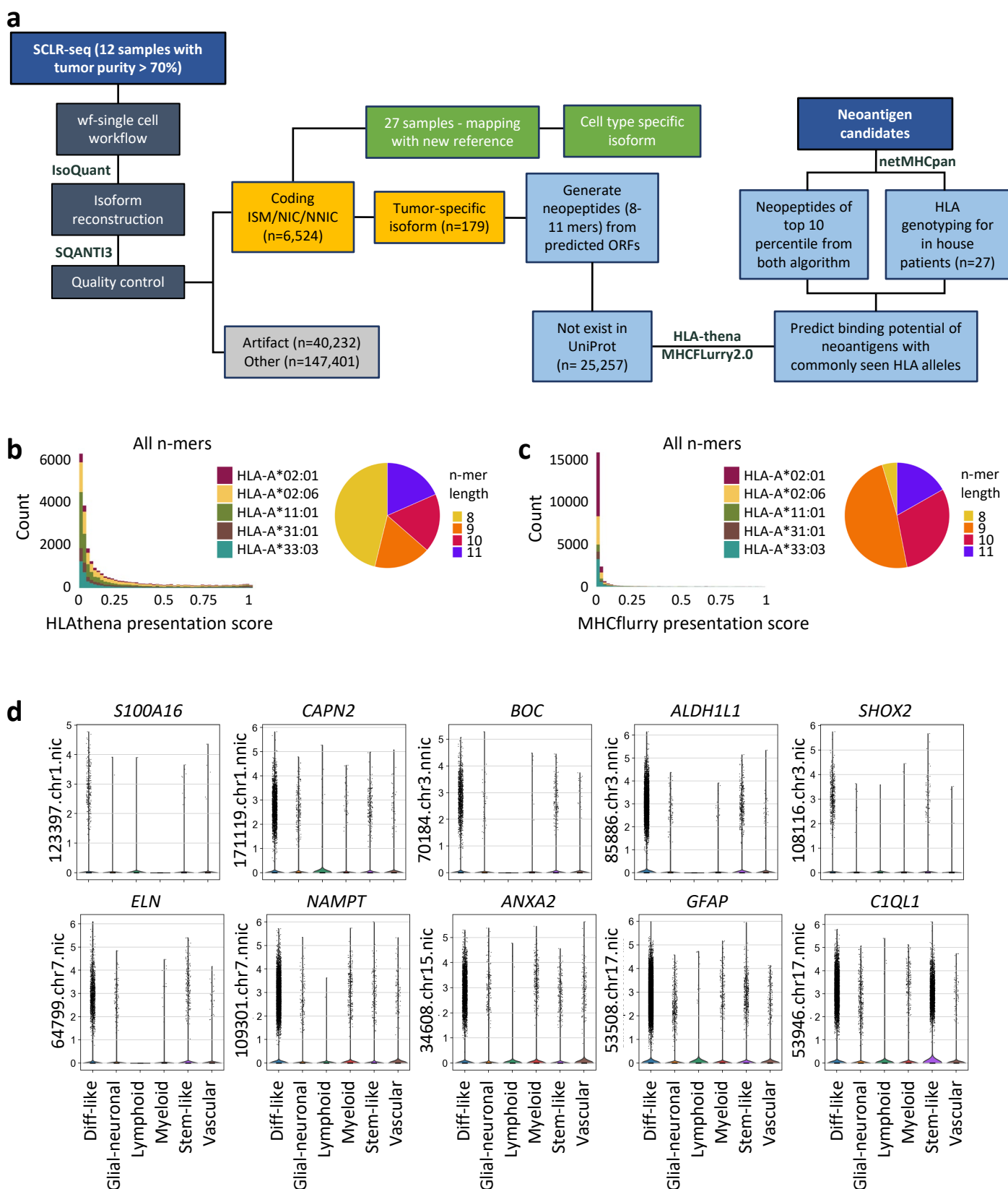


Supplementary Fig. 6. PCR and Sanger sequencing validation of selected extracellular–intracellular target pairs in tumor samples and control blood

(a) Schematic representation of extracellular-intracellular target pair combinations identified across GBM patients. The diagram illustrates patient-specific pairing patterns between extracellular targets (left) and intracellular targets (middle), with connecting lines indicating the pairing relationships. (b) Sanger sequencing was performed as an orthogonal validation of selected junctions of extracellular–intracellular target pairs in tumor samples and control blood. The experiment was performed once, and the chromatograms and Sanger images are shown. E, exon; Ex, extracellular; In, intracellular; B, blood control; T: tumor sample.

Supplementary Fig. 7. Characterization of isoforms not present in existing annotations and their cell-type-specific expression

(a) Histograms showing the distribution of transcript abundance (TPM, left) and transcript length (right) across all isoforms in 27 samples. (b) Sankey plot illustrating changes in transcript structural annotation for 6,524 transcripts between GENCODE v32 and v49. (c) Sankey plot summarizing classification of 6,524 isoforms reconstructed in barcode-aware single-cell mode relative to isoforms reconstructed from pseudobulk (sample-level aggregated) reads. Class codes indicate the structural relationship between single-cell-derived isoforms and pseudobulk models: “=” denotes exact intron-chain matches; “j” indicates isoforms sharing at least one splice junction match; “k” denotes containment of reference; “m” indicates retained matched intron(s); “c” represents containment within a larger transcript; “n” denotes retained novel intron structure; “o” indicates generic exon overlap on the same strand; “u” represents unknown/intergenic transcripts; “x” denotes overlap with exons on the opposite strand; and “y” indicates transcripts containing a reference within its intron. (d) Bar plots showing the distribution of splicing event types stratified by major cell types (Differentiated-like: 108,300; Glial-Neuronal: 28,049; Stem-like: 22,831; Myeloid: 16,535; Vascular: 5,511; Lymphoid: 996 cells) in 27 samples. (e,f) Dot plots highlighting the top differentially expressed isoforms across fine-resolution cell states (e) and broader non-malignant cell populations, including myeloid, vascular, and glial-neuronal cells in 27 samples (f). Differential expression was evaluated using a two-sided t-test with Benjamini–Hochberg correction, and isoforms with adjusted $P < 0.05$ are shown.



Supplementary Fig. 9. Prediction and characterization of peptides derived from isoforms not present in existing annotations
 (a) Schematic workflow outlining generation of peptides from isoforms and prediction of their MHC-binding affinities. (b,c) Histograms and pie charts showing the distribution of peptide presentation scores by HLA allele (left) and peptide length (right) as predicted by HLathena (b) and MHCflurry 2.0 (c). (d) Dot plot displaying expression patterns of select isoforms across major GBM cell types.