

DNA transposon expansion drives genome plasticity in *Diutina catenulata*

Source Data File

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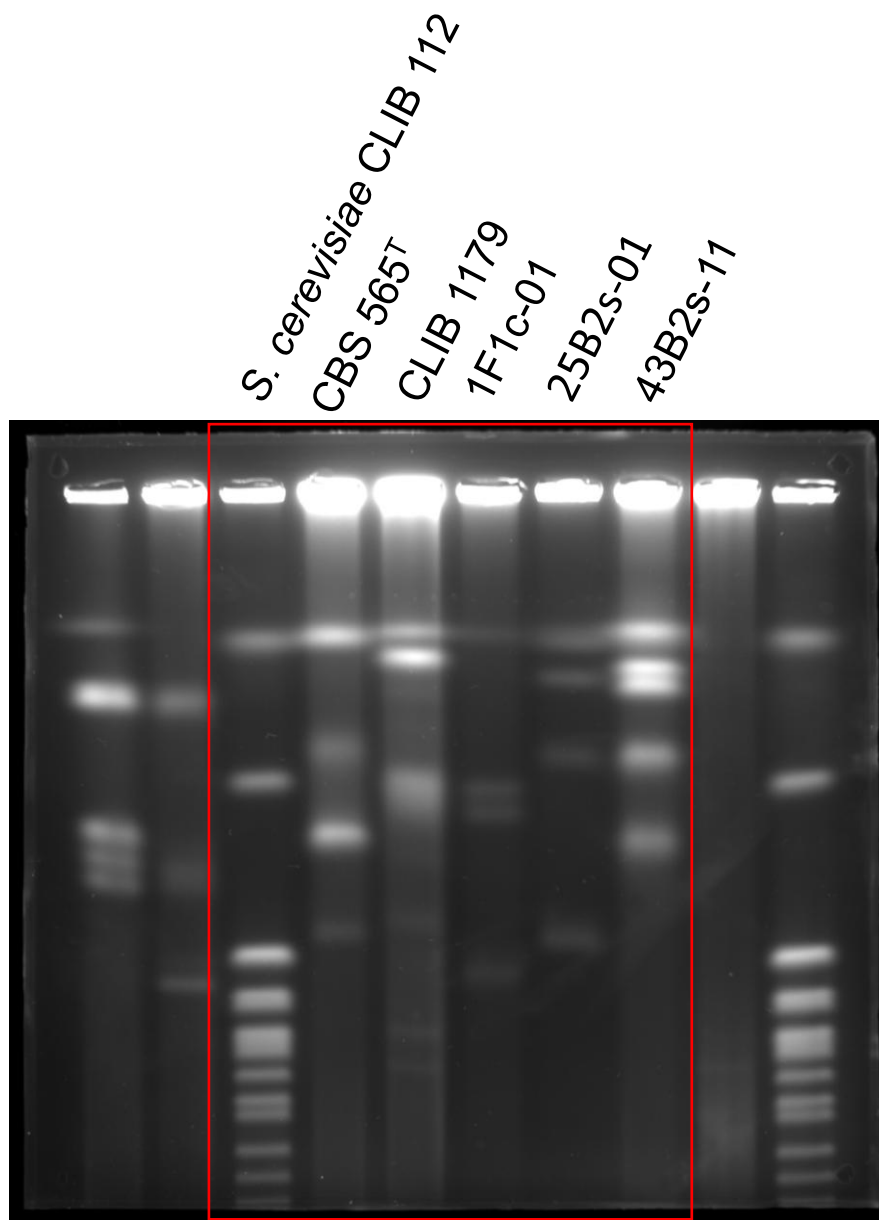
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Source data file 1A-E: uncropped pulsed-field gel electrophoresis (PFGE) images.

Source data file 2: Figure illustrating the gating strategy developed for ploidy determination using flow cytometry.

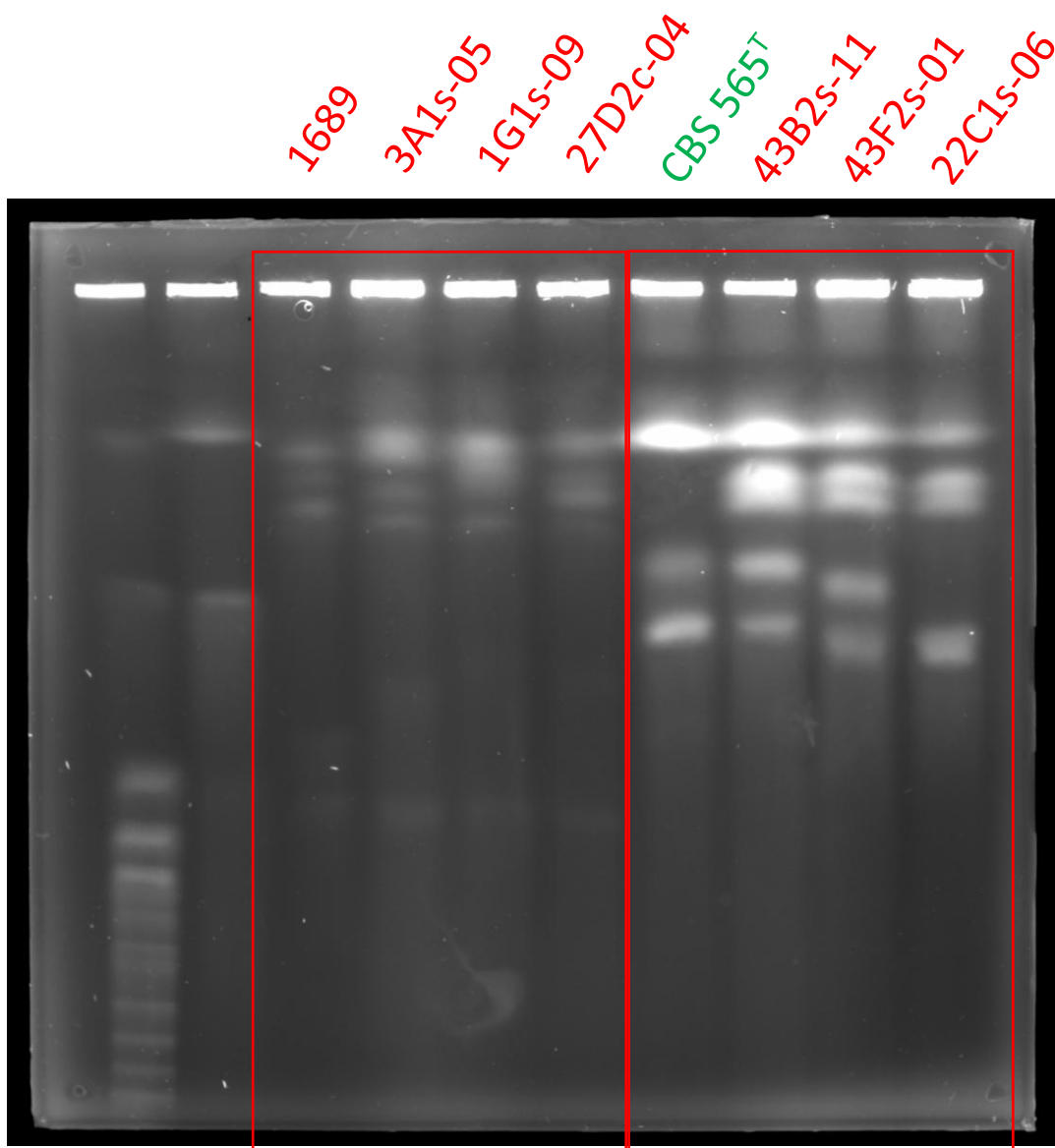
A



Source data file 1A: uncropped pulsed-field gel electrophoresis (PFGE) images.

This caryotype corresponds to Figure 4a

B



Source data file 1B: uncropped pulsed-field gel electrophoresis (PFGE) images.

This caryotype corresponds to Supplementary Figure 2, clades 2 and 5

C

CBS 2014

CBS 2015

CBS 2821

CBS 6174

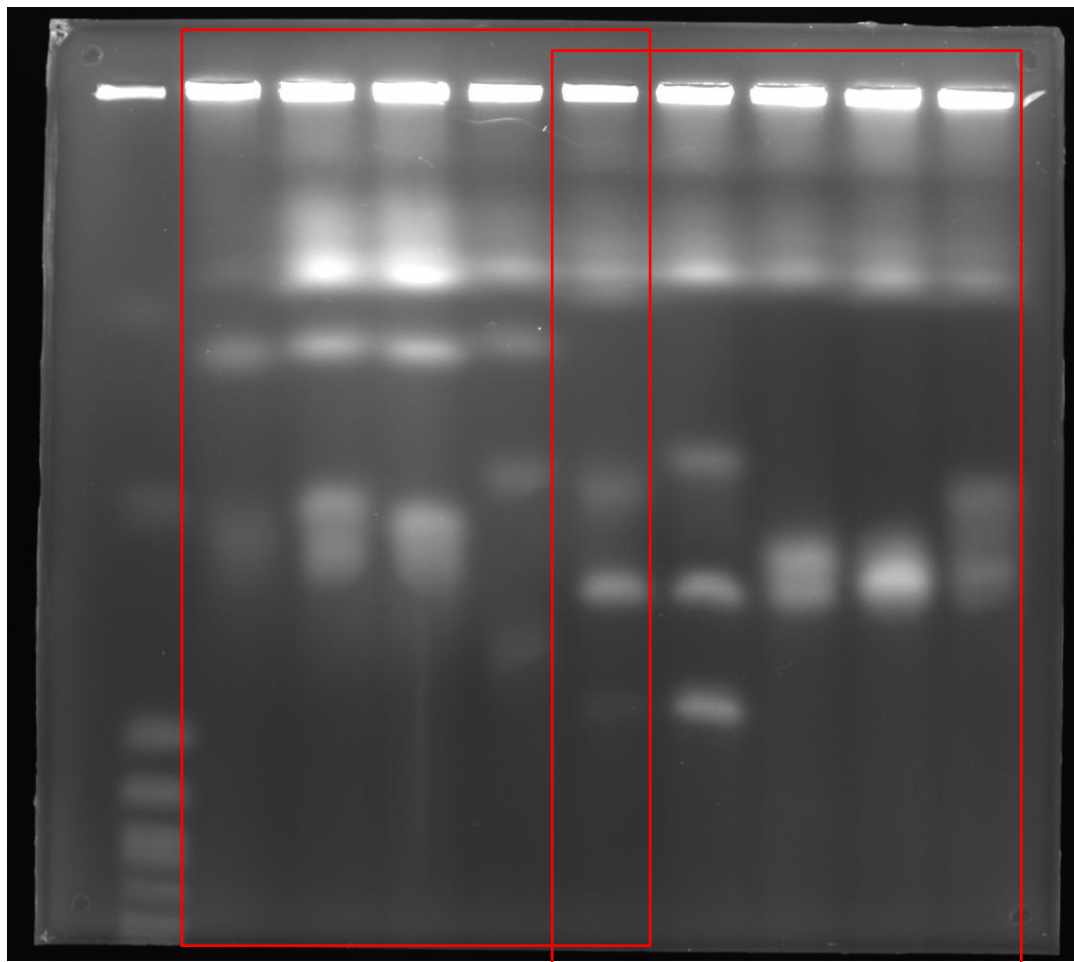
CBS 565T

line #1, clone #A

line #2, clone #A

line #3, clone #A

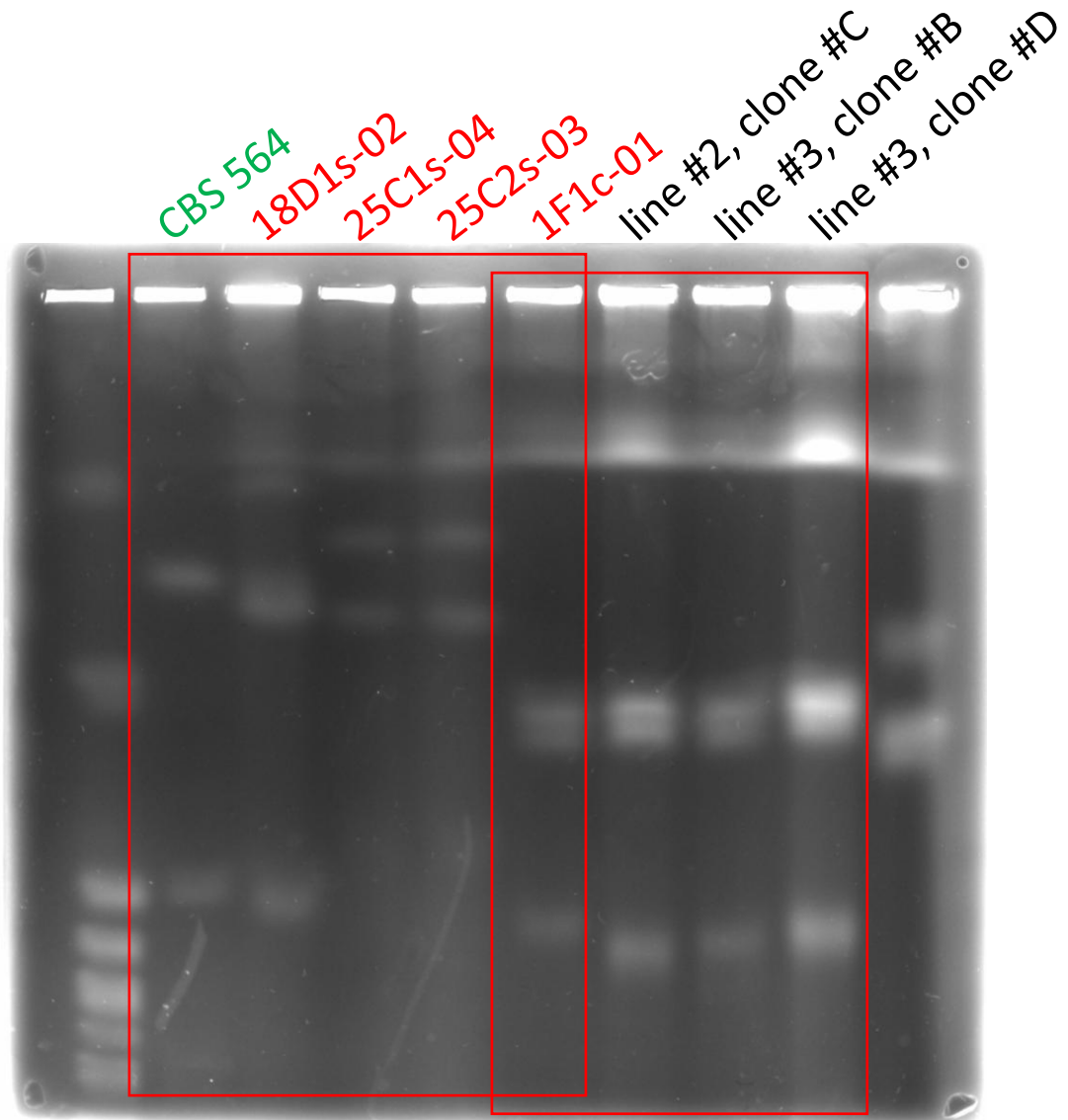
line #3, clone #B



Source data file 1C: uncropped pulsed-field gel electrophoresis (PFGE) images.

This caryotype corresponds to Supplementary Figure 2, clade 1 and Figure 4b

D

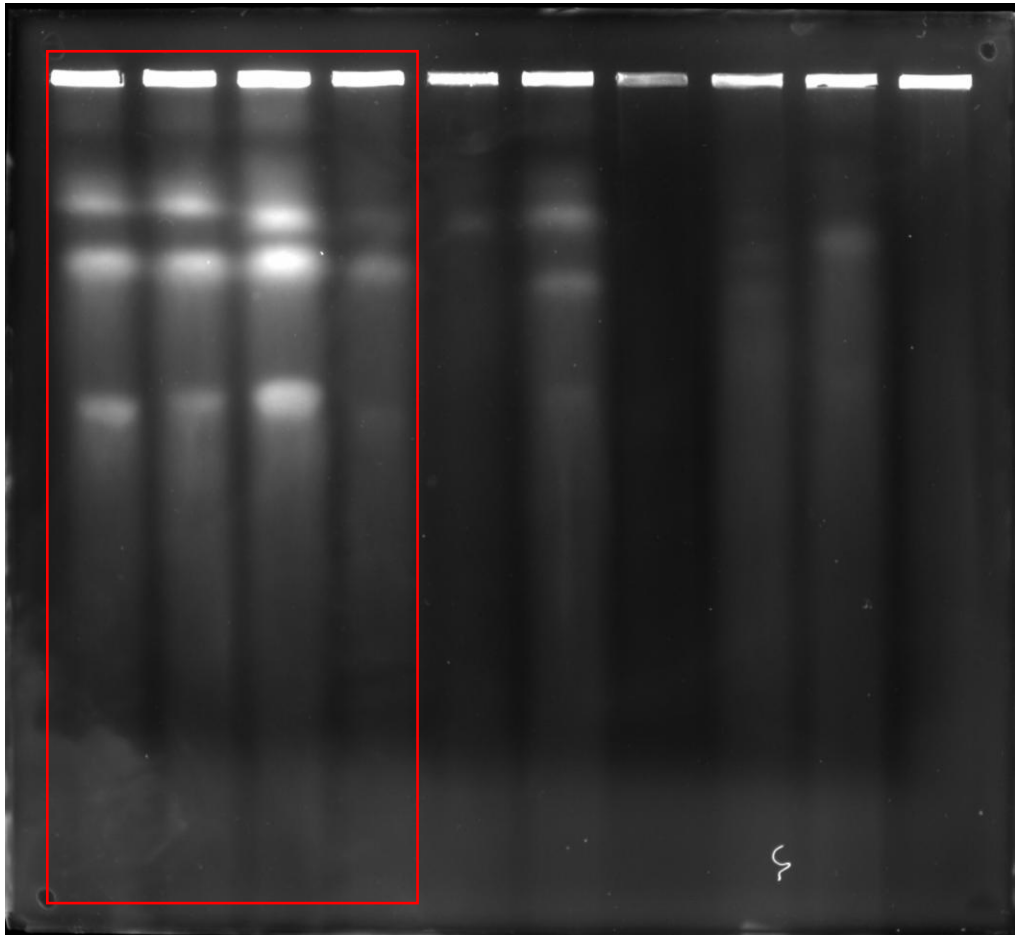


Source data file 1D: uncropped pulsed-field gel electrophoresis (PFGE) images.

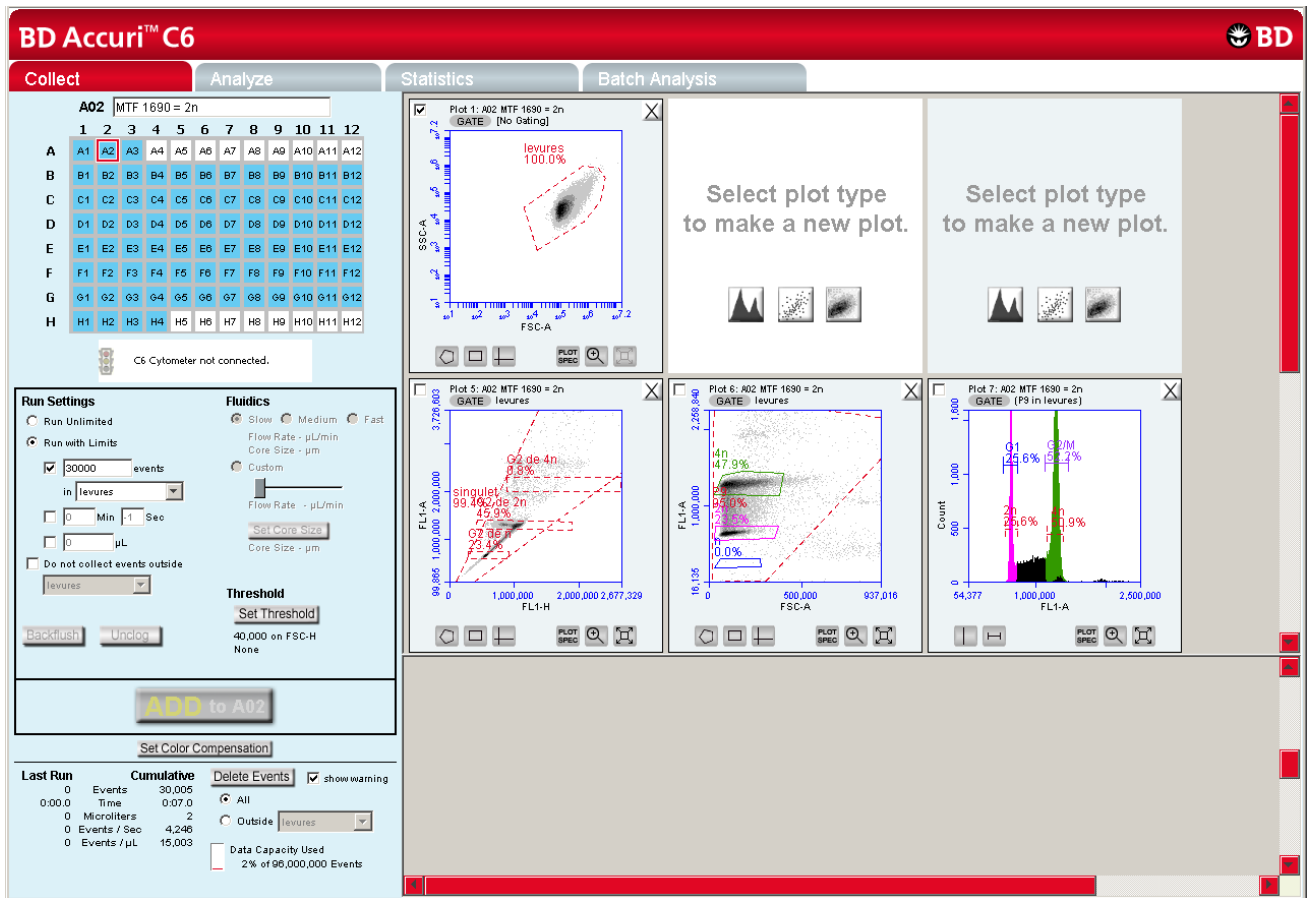
This caryotype corresponds to Supplementary Figure 2, clade 6 and Figure 4c

E

CLIB 1179
PYCC 7045
CBS 7135
CNRMA12-149



Source data file 1E: uncropped pulsed-field gel electrophoresis (PFGE) images.
This caryotype corresponds to Supplementary Figure 2, clade 3



Source data file 2: Figure illustrating the gating strategy developed for ploidy determination using flow cytometry. Cells were analyzed by gating on forward scatter (FSC-A) and side scatter (SSC-A) signals, and an initial gate was defined to identify yeast cells. Gates were then established on forward scatter versus fluorescence intensity plots according to the ploidy levels of the reference strains; the blue, pink, and green gates represent haploid, diploid, and tetraploid cells, respectively. Finally, the ploidy of the studied strains was determined directly in the software by comparison with the reference strains.