

Youth-associated protein TIMP2 regulates microglial state and function in healthy and aged mice

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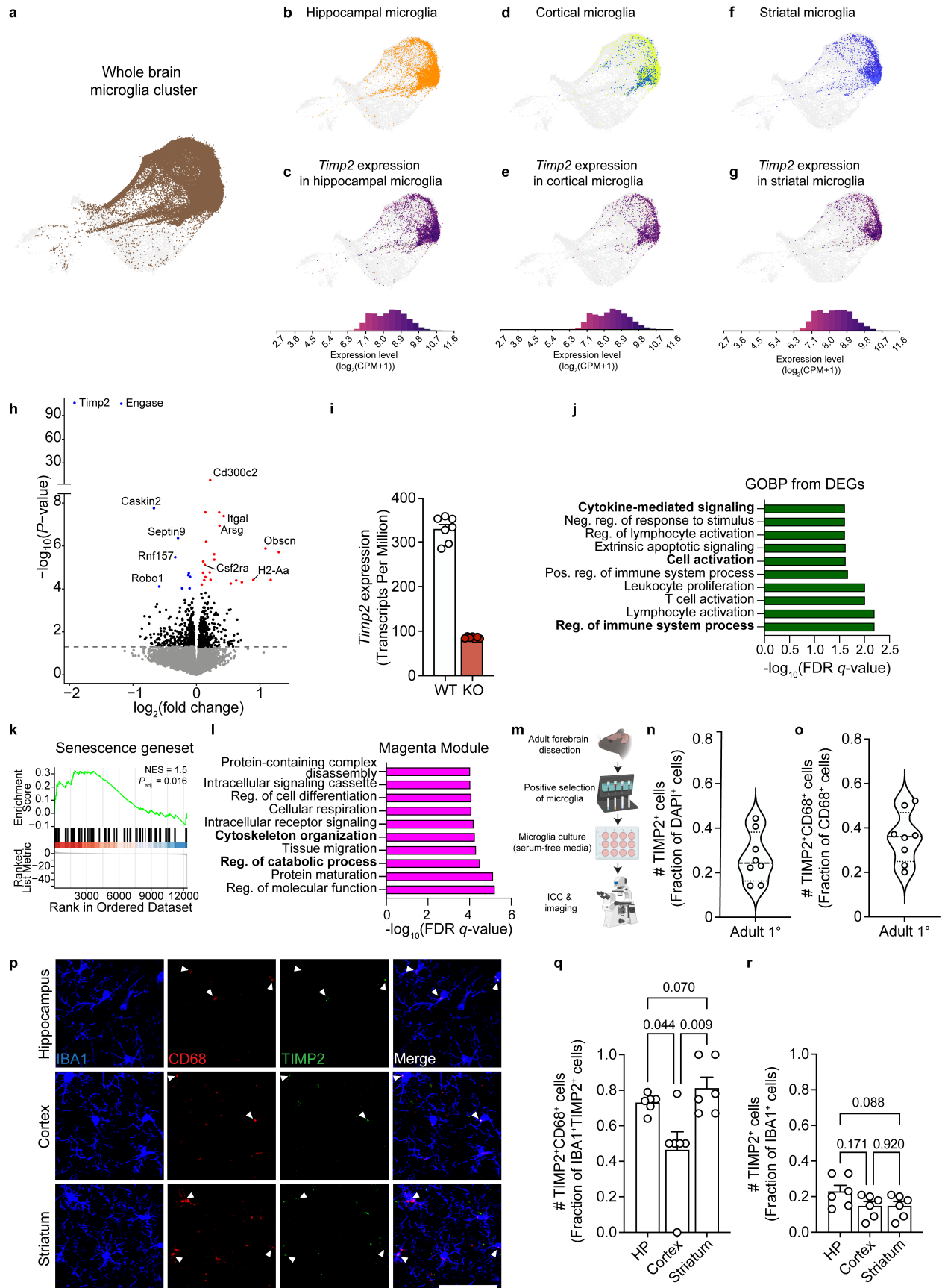
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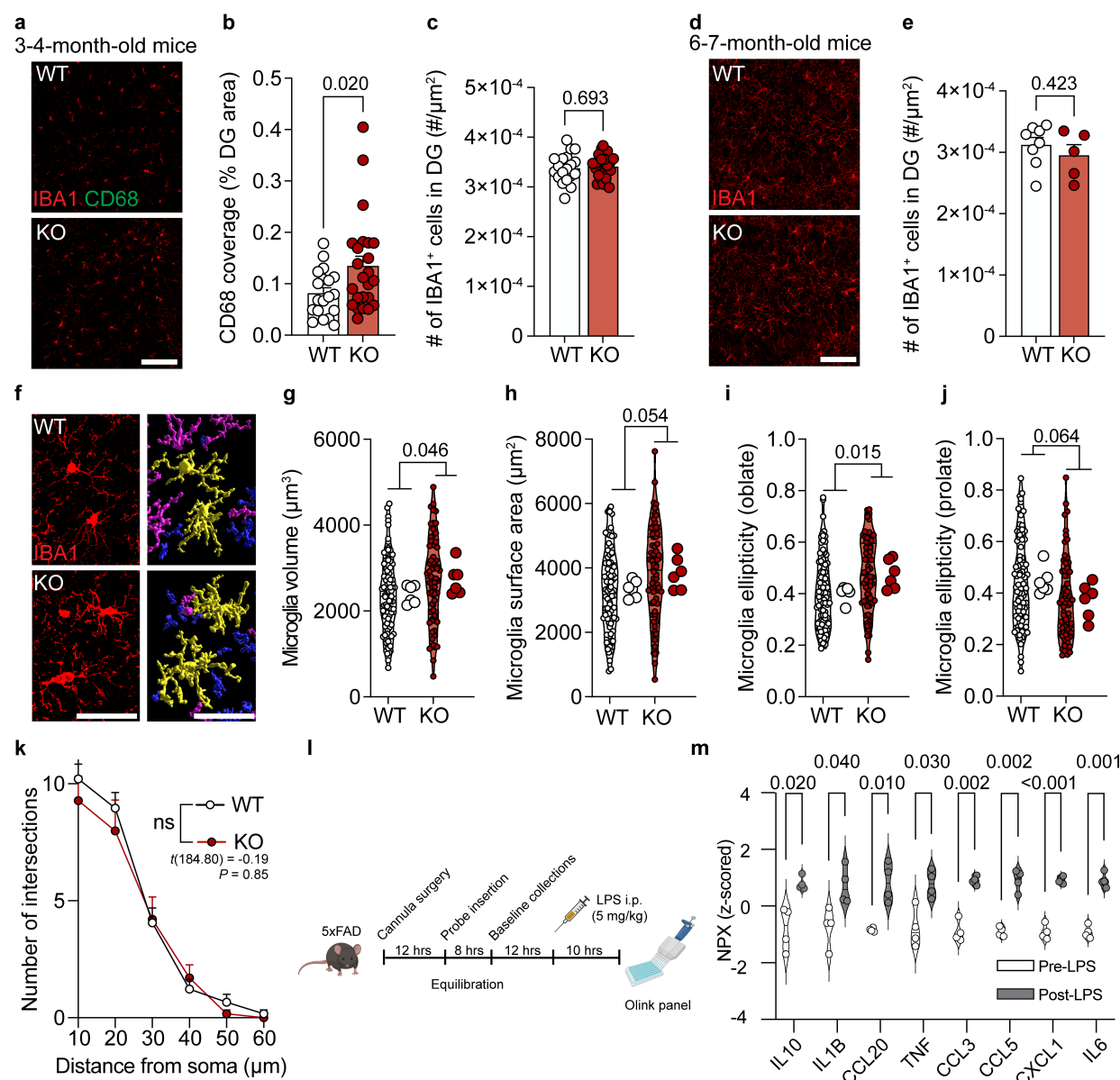
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Supplementary Figure 1. TIMP2 is expressed in microglia across regions and in cells with CD68 expression.

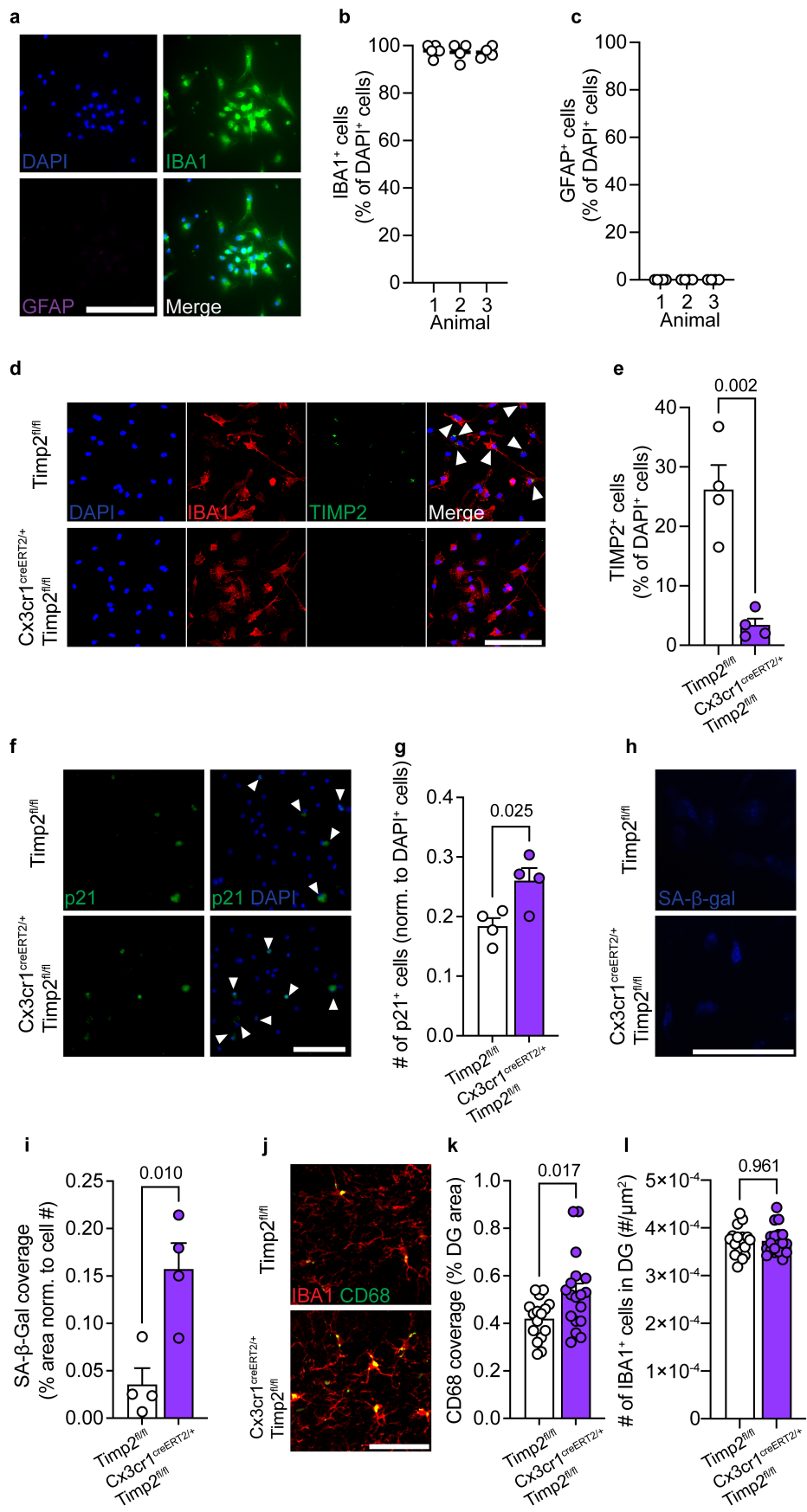
(a) Data from Allen Brain Cell scRNAseq and MERFISH Atlas from whole brain microglia (brown), with subsets corresponding to **(b)** hippocampal microglia (orange), and **(c)** *Timp2* expression within hippocampal microglia. Additional microglia subsets including **(d)** cortical microglia (green = retrosplenial cortex; blue = visual cortex), with **(e)** corresponding *Timp2* expression, and **(f)** striatal microglia (dark blue = ventral striatum; light blue = dorsal striatum), with **(g)** *Timp2* expression within striatum. **(h)** Volcano plot of bulk RNAseq DEGs from early postnatal TIMP2 KO vs. WT primary microglia (N = 7 samples per genotype, with each sample representing a pool of 5 pups (P6-P7); sex-matched; mean $\pm$ SEM), showing $\log_2$ fold-change versus $-\log_{10} P_{\text{adj}}$ -value for each gene, highlighting significant DEGs ($P_{\text{adj}} < 0.05$) in red (upregulated) or blue (downregulated). **(i)** Gene expression (transcripts per million) values for *Timp2* from bulk RNAseq experiments in “h”. **(j)** Significant GOBP pathways ($P_{\text{adj}} < 0.05$, FDR) and NES from over-representation analysis of significant ($P_{\text{adj}} < 0.05$) DEGs from “h” (calculated from DESeq2, two-sided Wald test, Benjamini-Hochberg-adjusted). **(k)** GSEA enrichment plot for senescence gene set from ranked genes (TIMP2 KO vs WT, using two-tailed Wald statistic-ranked DEGs) with green lines representing running enrichment score, black vertical bars indicating position of genes in ranked list (upregulated in red and downregulated in blue). **(l)** Significant GOBP terms (FDR q -value < 0.05) from over-representation analysis on genes from significant “Magenta” module from Fig. 1e. **(m)** Schematic diagram of adult microglia isolation, culturing, and imaging, with **(n)** quantification of the number of TIMP2⁺ cells as a fraction of DAPI⁺ cells in WT primary microglia cultures stained by ICC, or **(o)** the number of TIMP2⁺CD68⁺ cells as a fraction of CD68⁺ cells (N = 8 mice, 5-6 month-old mice; sex-matched; 2 independent experiments combined; scale bar = 100 μ m; violin plots of median with quartiles). **(p)** Representative confocal images of hippocampus, cortex, and striatum from 6-7-month-old mice stained with anti-IBA1, anti-CD68, and anti-TIMP2 antibodies (scale bar = 50 μ m; arrowheads indicate IBA1⁺CD68⁺TIMP2⁺ cells), with **(q)**

48 quantification of number of TIMP2⁺CD68⁺ cells as fraction of IBA1⁺TIMP2⁺ cells for each region
49 (N = 6 mice per group) and (r) number of TIMP2⁺ cells as fraction of IBA1⁺ cells across brain
50 regions; 1-way ANOVA, Tukey's post test. Source data are provided as Source Data file. Created
51 in BioRender: Philippi, S. (2026) <https://BioRender.com/2bm4j2d>.



Supplementary Figure 2. TIMP2 KO mice exhibit altered state and morphology associated with activation.

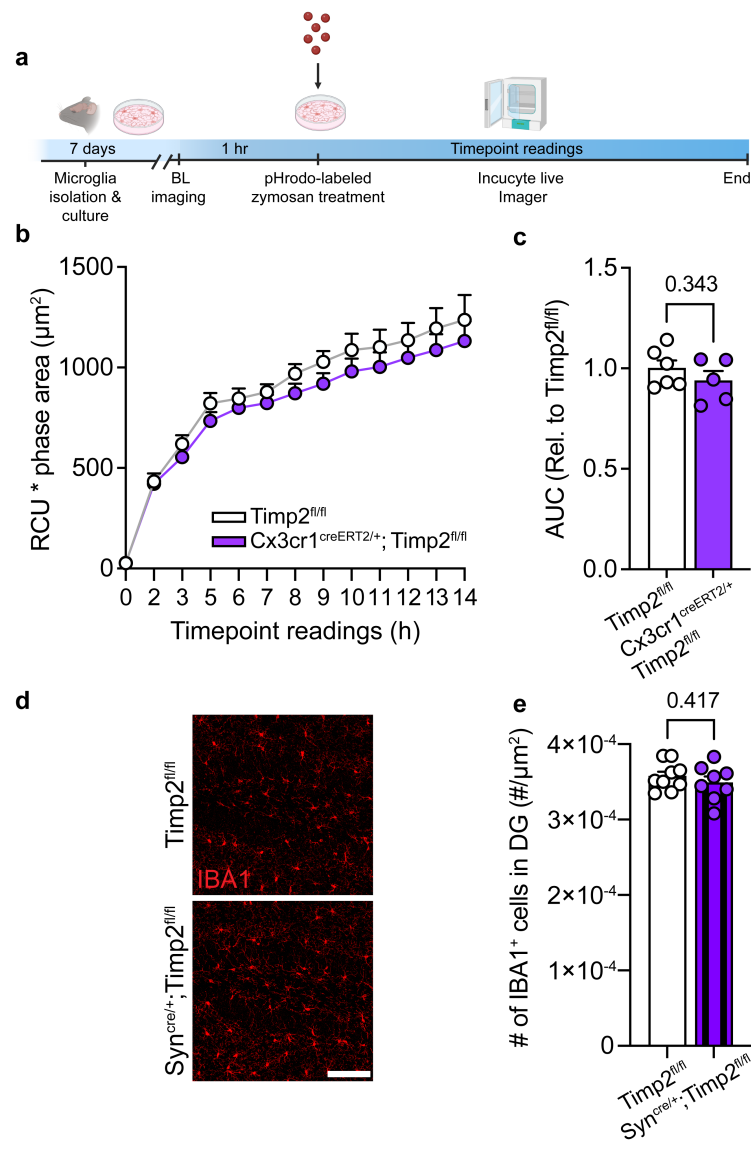
(a) Representative confocal images of DG from 3-4-month-old WT and TIMP2 KO mice stained with anti-CD68 and anti-IBA1 antibodies (scale bar = 100 μ m) with **(b)** quantification of thresholded DG area covered by CD68⁺ staining and **(c)** the number of IBA1⁺ cells normalized to DG area (N = 19 WT (9 female, 10 male), 24 KO (13 female, 11 male)). **(d)** Representative confocal images of DG from 6-7-month-old WT and TIMP2 KO mice stained with anti-IBA1 antibody (scale bar = 100 μ m) with **(e)** quantification of the number of IBA1⁺ cells normalized to DG area (N = 8 WT, 5 KO female mice; mean $\pm$ SEM). **(f)** Representative confocal staining (left; scale bar = 15 μ m) of DG from 6-7-month-old TIMP2 KO and WT mice stained with anti-IBA1 antibody, with Imaris-based reconstruction (right; scale bar = 15 μ m) of microglial surfaces, with corresponding quantification from TIMP2 KO and WT microglia of **(g)** volume, **(h)** surface area, **(i)** oblate and **(j)** prolate ellipticity, and **(k)** number of Sholl intersections over distance from soma (N = 6 mice per group; sex-matched; two-tailed nested *t*-test using cell (left replicates) and mouse (right replicates) as levels; linear mixed-effects model for Sholl analysis in (k)). **(l)** Timeline of LPS treatment paradigm in mice undergoing high molecular-weight cut-off (HMWCO, 1-MDa) *in vivo* microdialysis (N = 4; 6-7-month-old male 5xFAD mice) with **(m)** brain ISF protein measurements, represented as z-scored NPX values for significant proteins (paired two-tailed *t*-test; median with quartiles). Two-tailed Student's *t*-test used in (b-c), (e), with Welch's correction applied in (b). Source data are provided as Source Data file. Created in BioRender: Philippi, S. (2026) <https://BioRender.com/ruyfrku>.



Supplementary Figure 3. Purity of primary microglia cultures and efficiency of microglia

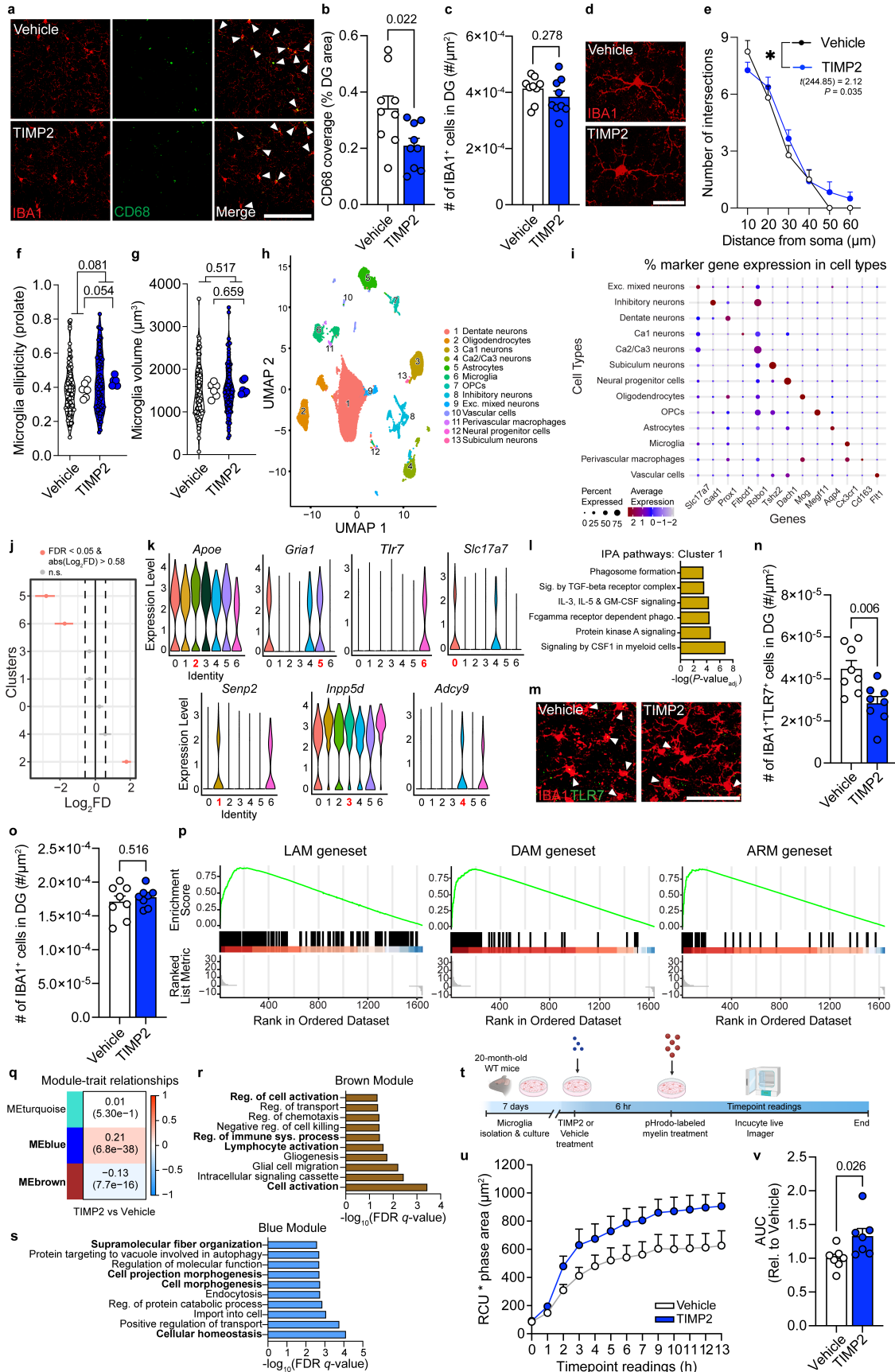
***Timp2* deletion**

(a) Representative microscopy images of primary microglia cultures from WT mice stained with DAPI, anti-IBA1, and anti-GFAP antibodies (scale bar = 100 μ m), with **(b)** quantification of IBA1⁺ cells as percentage of DAPI⁺ cells or **(c)** quantification of GFAP⁺ cells as percentage of DAPI⁺ cells (N = 5 replicates for mouse 1, 4 replicates each for mice 2 and 3; 9-month-old). **(d)** Representative images from primary microglia cultures from 5-month-old *Timp2*^{fl/fl} control and *Cx3cr1*^{CreERT2/+};*Timp2*^{fl/fl} littermates stained by ICC with DAPI, anti-IBA1 antibody, and anti-TIMP2 antibody (arrowheads indicate TIMP2⁺ microglia; N = 4 mice per group; sex-matched; scale bar = 100 μ m), with **(e)** quantification of TIMP2⁺ cells as percentage of DAPI⁺ cell number (mean $\pm$ SEM). **(f)** Representative images from primary microglia cultures isolated from 10-month-old *TIMP2*^{fl/fl} control and *Cx3cr1*^{CreERT2/+};*Timp2*^{fl/fl} littermates stained by ICC with DAPI and anti-p21 antibody (arrowheads indicate p21⁺ cells; scale bar = 100 μ m;), with **(g)** quantification of number of p21⁺ cells normalized to total number of cells (DAPI⁺) (N = 4 mice per group). **(h)** Representative images from primary microglia cultures isolated from 10-month-old *TIMP2*^{fl/fl} control and *Cx3cr1*^{CreERT2/+};*Timp2*^{fl/fl} littermates stained with SA- β -gal (scale bar = 100 μ m), with **(i)** SA- β -gal staining calculated as percentage of thresholded area normalized to cell number (N = 4 mice per group). **(j)** Representative confocal images of DG from 6-7-month-old *TIMP2*^{fl/fl} control and *Cx3cr1*^{CreERT2/+};*Timp2*^{fl/fl} littermates stained with anti-IBA1 and anti-CD68 antibodies with **(k)** quantification of DG area covered by CD68⁺ staining (N= 16 *TIMP2*^{fl/fl} control, N=18 *Cx3cr1*^{CreERT2/+};*Timp2*^{fl/fl}) and **(l)** quantification of the number of IBA1⁺ cells per DG area (N= 17 *TIMP2*^{fl/fl} control, N=19 *Cx3cr1*^{CreERT2/+};*Timp2*^{fl/fl}) sex-matched mice; mean $\pm$ SEM; scale bar = 50 μ m. Two-tailed Student's *t*-test. Source data are provided as Source Data file.



Supplementary Figure 4. Microglial TIMP2 deletion does not affect zymosan uptake, and neuronal deletion of TIMP2 does not alter microglia number in the dentate gyrus

(a) Schematic diagram of treatment of adult primary microglia with pHrodo-labeled zymosan and subsequent Incucyte imaging to assess uptake across timepoint readings. **(b)** Timepoint readings with total integrated density (red object mean intensity [RCU, red, x average phase object area (cell, μm^2)] of pHrodo-labeled zymosan signal in primary microglia cultured from tamoxifen-treated 4-5-month-old $\text{Cx3cr1}^{\text{CreERT2/+}};\text{Timp2}^{\text{fl/fl}}$ and $\text{Timp2}^{\text{fl/fl}}$ control mice, with **(c)** corresponding uptake quantified using area under the curve (AUC; $N = 6$ $\text{Timp2}^{\text{fl/fl}}$, $N = 5$ $\text{Cx3cr1}^{\text{CreERT2/+}};\text{Timp2}^{\text{fl/fl}}$ sex-matched mice; 2 independent experiments combined; mean $\pm$ SEM). **(d)** Representative confocal images of DG from 2-3-month-old $\text{Timp2}^{\text{fl/fl}}$ and $\text{Syn}^{\text{Cre/+}};\text{Timp2}^{\text{fl/fl}}$ littermates stained with anti-IBA1 antibody with corresponding **(e)** quantification of the number of IBA1⁺ cells normalized to DG area ($N = 9$ $\text{Timp2}^{\text{fl/fl}}$, $N = 8$ $\text{Syn}^{\text{Cre/+}};\text{Timp2}^{\text{fl/fl}}$ female mice; scale bar = 100 μm ; mean $\pm$ SEM). Two-tailed Student's *t*-test. Source data are provided as Source Data file. Created in BioRender: Philippi, S. (2026) <https://BioRender.com/fblpd8f>.



Supplementary Figure 5. Systemic TIMP2 treatment in aged mice alters microglial state and shifts microglial subclusters.

(a) Representative confocal images of DG from 20-month-old WT mice treated systemically with TIMP2 or vehicle, stained with anti-IBA1 and anti-CD68 antibodies (scale bar = 100 μ m; arrowheads indicate IBA1⁺CD68⁺ cells), with corresponding **(b)** quantification of the percentage of DG area covered by CD68⁺ staining and **(c)** quantification of the number of IBA1⁺ cells normalized to DG area (N = 9 male mice per group; mean $\pm$ SEM). **(d)** Representative high-magnification confocal images anti-IBA1 antibody-stained microglia from DG of 20-month-old mice treated with vehicle or TIMP2 (scale bar = 20 μ m) for Imaris-based morphology analyses, with **(e)** quantification of number of Sholl intersections over distance (μ m) from soma, **(f)** microglia ellipticity (prolate) and **(g)** volume (N = 6 male mice per group; *P*-values reflect two-tailed nested *t*-tests, using cell and mouse replicates as levels, as well as standard mouse-level replicate comparison by two-tailed Student's *t*-test; linear mixed-effects model for Sholl analysis in (e)). **(h)** Uniform Manifold Approximation and Projection (UMAP) plot of hippocampal nuclei isolated from 20-month-old WT mice treated systemically with vehicle or TIMP2 (N = 4 male mice per group combined as 2 samples per group). **(i)** Dot plot of scaled expression of selected marker genes across annotated cell types. Dot size represents percentage of cells expressing the gene within each cell type, while color indicates the scaled average expression level of the gene across cells in the respective cluster. **(j)** Differences in cell proportions for each cluster between treatment conditions. Significant clusters are denoted in salmon (FDR < 0.05, Log₂ fold-enrichment > 0.58, relative to vehicle-treated mice). **(k)** Violin plots of marker genes across microglial subclusters from Fig. 5h. **(l)** Significant canonical IPA pathways (Benjamini-Hochberg-adjusted) for marker genes significantly upregulated in subcluster 1 (from one-tailed Wilcoxon Rank Sum *P*_{adj}-values (Bonferroni-adjusted)). **(m)** Representative confocal images of DG from 20-month-old WT mice treated systemically with TIMP2 or vehicle, stained with anti-IBA1 and anti-TLR7 antibodies (scale bar = 50 μ m; arrowheads indicate IBA1⁺TLR7⁺ cells), with corresponding **(n)** quantification of

141 IBA1⁺TLR7⁺ cells normalized to DG area and **(o)** number of IBA1⁺ cells normalized to DG area
142 (N = 8 mice per group; males; mean $\pm$ SEM). **(p)** GSEA enrichment plots for LAM, DAM, and ARM
143 gene sets based on the ranked list of microglial genes (TIMP2 vs. vehicle). Green lines represent
144 running enrichment score, with vertical black bars indicating the position of genes from indicated
145 gene sets in the ranked list and gene ranking direction indicated by color bar (red = upregulated,
146 blue = downregulated). **(q)** Module-trait relationships plot based on high-dimensional (hd)WGCNA
147 from snRNAseq dataset (TIMP2 vs. vehicle), with colored module assignments and
148 corresponding membership values and module significance ($P < 0.05$, two-tailed Student's
149 asymptotic t -test) in parentheses, with **(r)** significant GOBP terms (FDR < 0.05) from over-
150 representation analysis on significant "Brown" and (s) "Blue" modules on extracted genes. **(t)**
151 Schematic diagram of treatment of adult primary microglia with pHrodo-labeled myelin and
152 subsequent Incucyte imaging to assess uptake. **(u)** Timepoint readings, with total integrated
153 density (red object mean intensity [RCU, red, $\times$ average phase object area (cell, μm^2)] of pHrodo-
154 labeled myelin signal in primary microglia cultured and treated with TIMP2 or vehicle from 20-
155 month-old WT mice, with corresponding **(v)** uptake quantification using area under the curve
156 (AUC; N = 7 mice per group; 2 independent experiments combined; sex-matched; mean $\pm$ SEM);
157 Two-tailed Students t -test in (b-c), (n-p), (v). Source data are provided as Source Data file.
158 Created in BioRender: Philippi, S. (2026) <https://BioRender.com/13xrppj>.