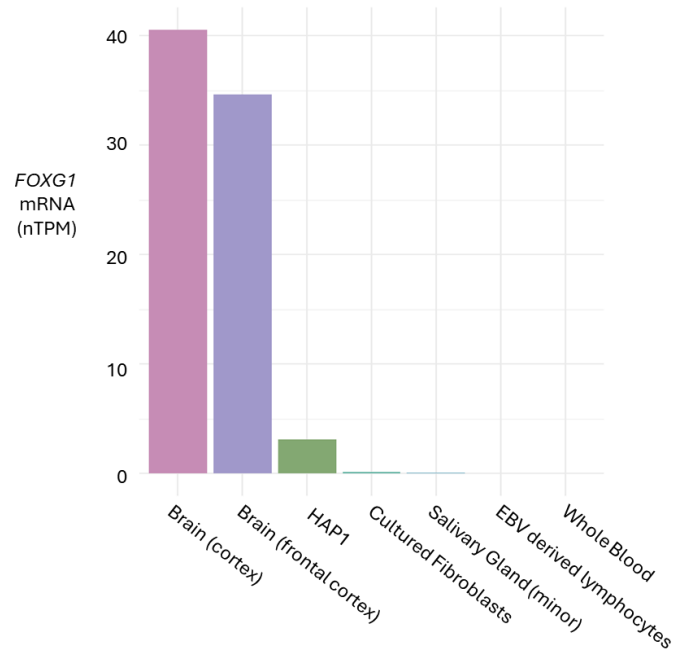


SUPPLEMENTARY INFORMATION

Analysis of 14q12 microdeletions reveals novel regulatory loci for the neurodevelopmental disorder-related gene, *FOXP1*

Ramamurthy et al.



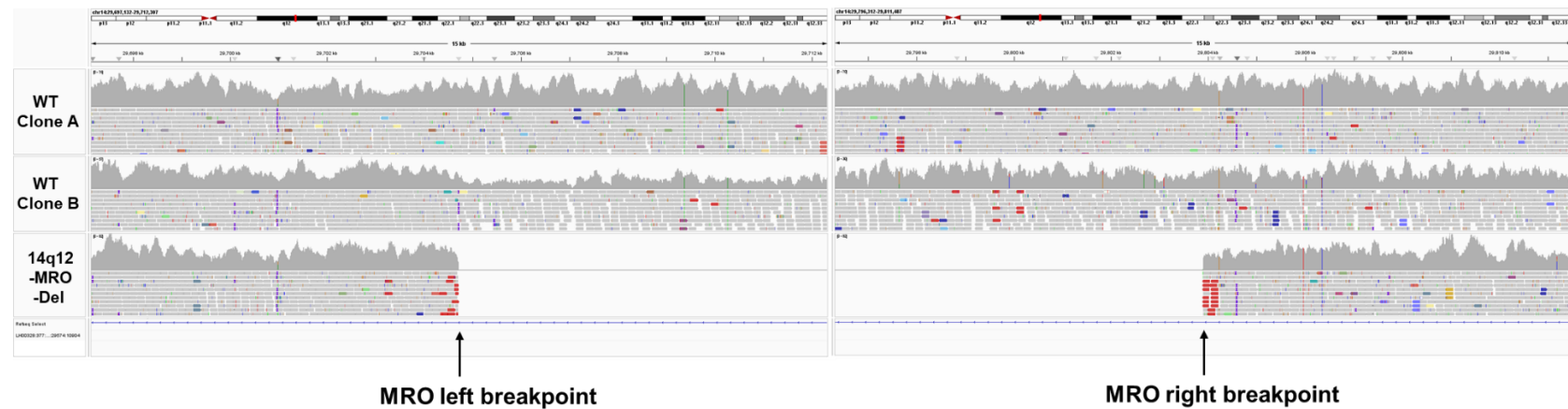
Supplementary Figure 1. *FOXG1* expression data. Normalized transcript per million (nTPM) values for *FOXG1* RNA were derived from GTEx for bulk tissues (GTEx 09.09.25, dbGaP accession number phs000424.vN.pN) and The Human Protein Atlas for HAP1 cell line (v24.proteinatlas.org). *FOXG1* nTPM values are provided in the Source Data file.



Variant ID	Allele Count	Allele Frequency
DEL_14_140953	1	0.00006
DEL_14_140954	2109	0.126424
DEL_14_140956	1	0.00006
DEL_14_140957	1	0.00006

Supplementary Figure 2. gnomAD variants, in particular deletions (grey bars), are mapped in the 14q12 MRO region (purple bar). The table reports allele count and minor allele frequency for each of these gnomAD deletions.

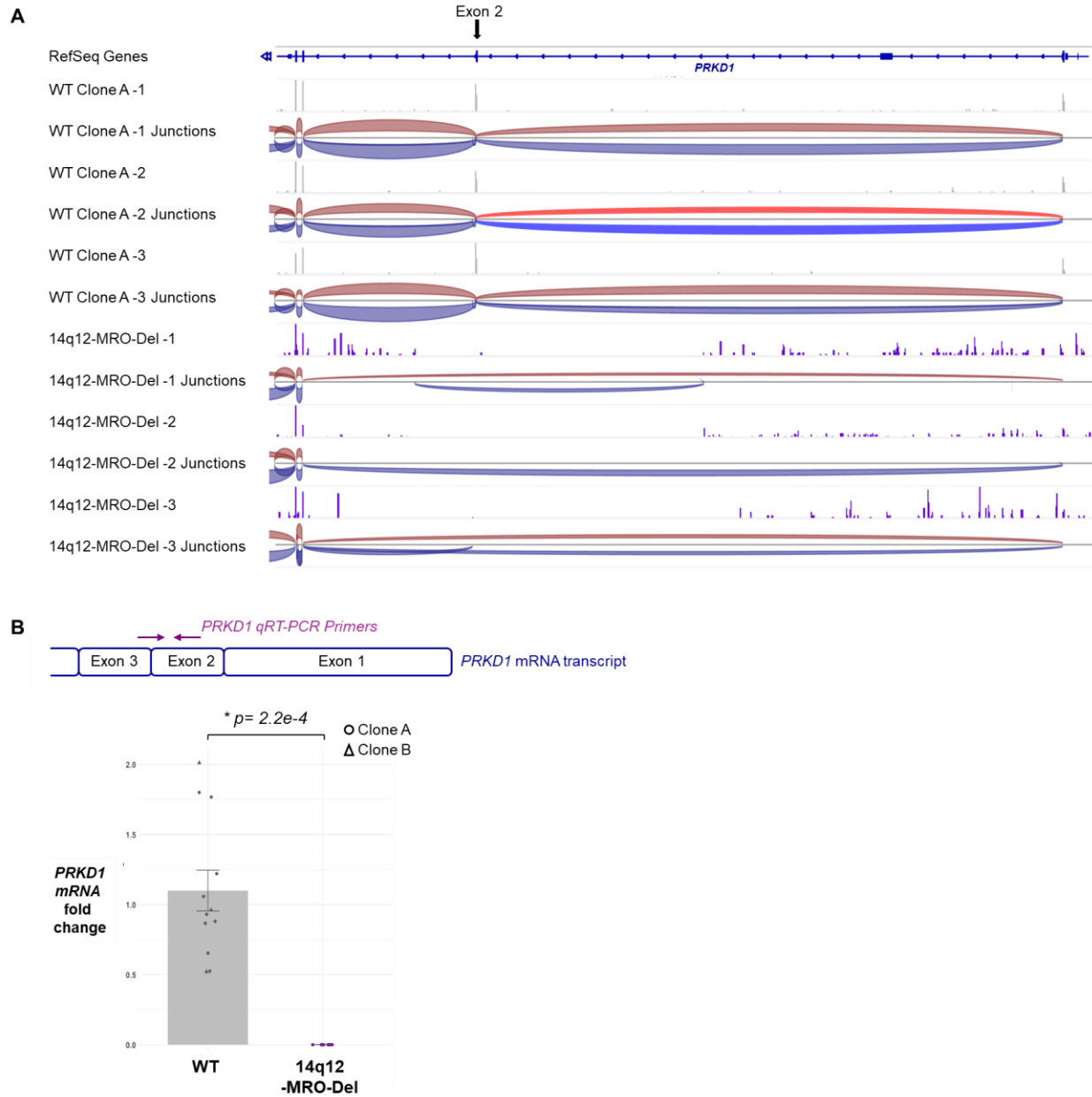
A



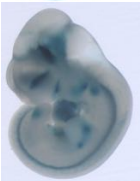
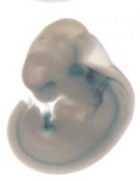
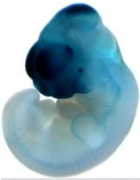
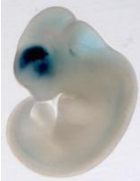
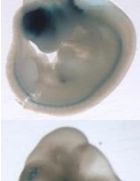
B



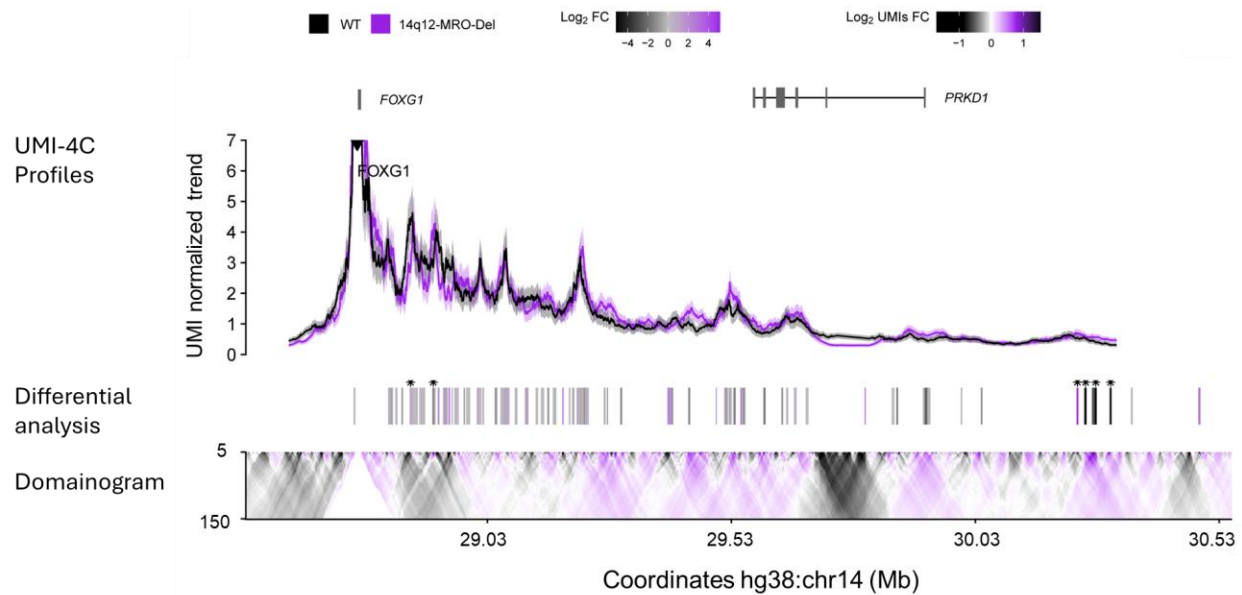
Supplementary Figure 3. Genotyping assays for validating CRISPR-Cas9 edited lines in this study. (A) Genome Sequencing data visualized using the Integrative Genomics Viewer confirm the deletion of MRO locus in 14q12-MRO-Del. (B) Deletion of the MRO in 14q12-MRO-Del and *FOXG1* coding sequence in *FOXG1*^{KO} verified by Sanger sequencing, visualized using SnapGene Viewer. The guide sequence at each breakpoint is highlighted in blue within the reference sequence.



Supplementary Figure 4. mRNA analysis of *PRKD1*. (A) IGV plot from RNA-seq of WT and 14q12-MRO-Del HAP1s demonstrating the deletion of exon 2 (arrow) of *PRKD1*, which renders the transcript out of frame, likely subjecting it to nonsense mediated decay. The junction reads (blue and red) confirm skipping of this exon (n=3) (B) *PRKD1* mRNA expression assayed by qRT-PCR in WT and 14q12-MRO-Del HAP1s, with primers spanning *PRKD1* exon 2 as shown. *PRKD1* mRNA fold change data are provided in the Source Data file. (Replicate count: WT Clone A = 9, WT Clone B = 3, 14q12-MRO-Del = 9). *Two sample T-test (two-tailed) was performed for statistical significance, WT vs 14q12-MRO-Del, $p = 2.2e-4$. Bar height corresponds to mean; error bars represent one standard error of the mean.

VISTA Enhancer	Activity	Example embryo (E11.5)
Hs1064	Forebrain, Hindbrain	
Hs566	Forebrain	
Hs1539	Hindbrain	
Hs1168	Facial mesenchyme, Hindbrain, Cranial nerve	
Hs1523	Branchial arch, Facial mesenchyme, Forebrain, Midbrain, Hindbrain, Eye	
Hs342	Forebrain	
Hs598	Neural tube	
Hs433	Forebrain, Hindbrain	
Hs344	Forebrain	

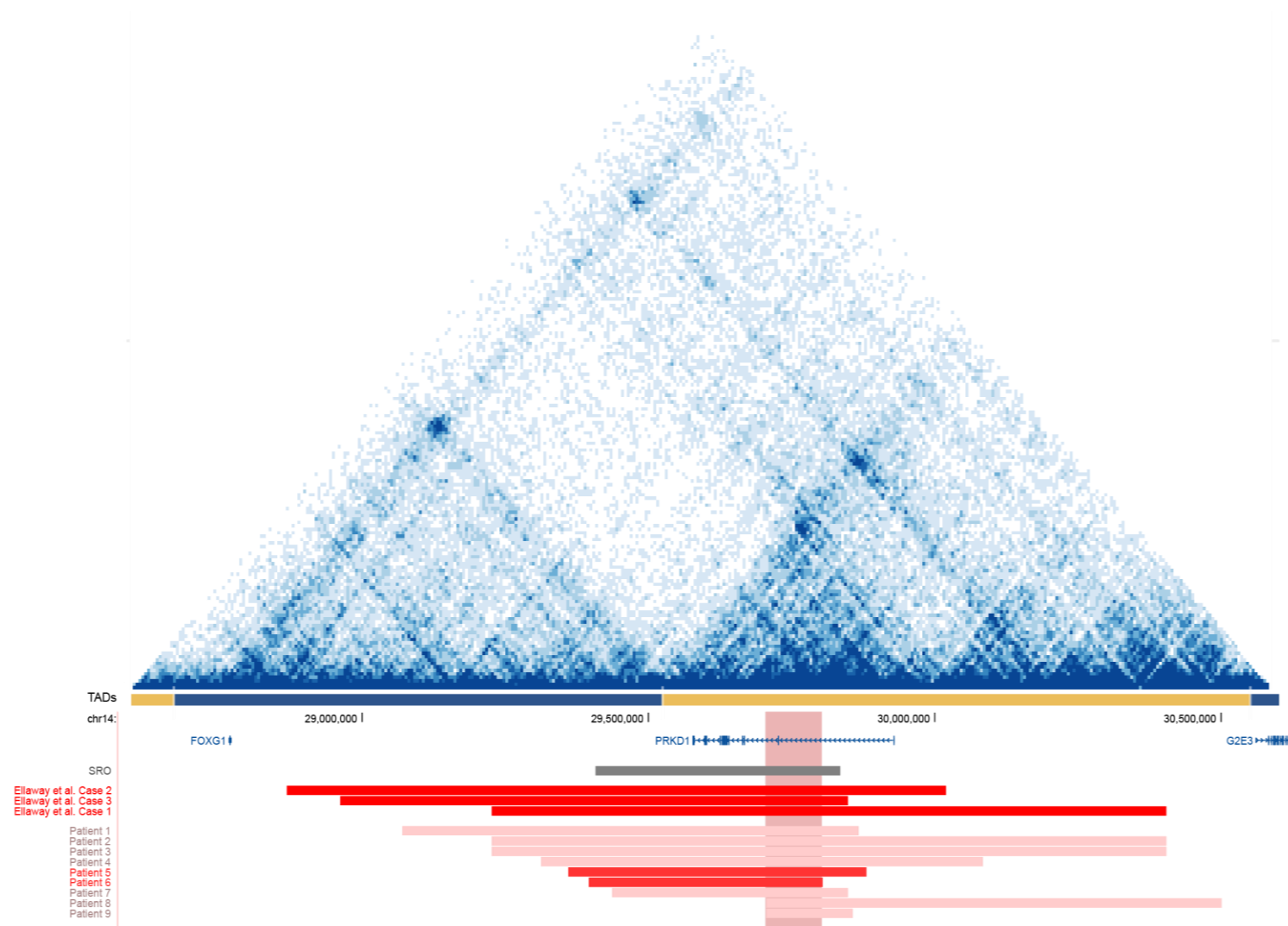
Supplementary Figure 5. Active VISTA enhancers in the 14q12 regulatory region downstream of *FOXG1* ¹.



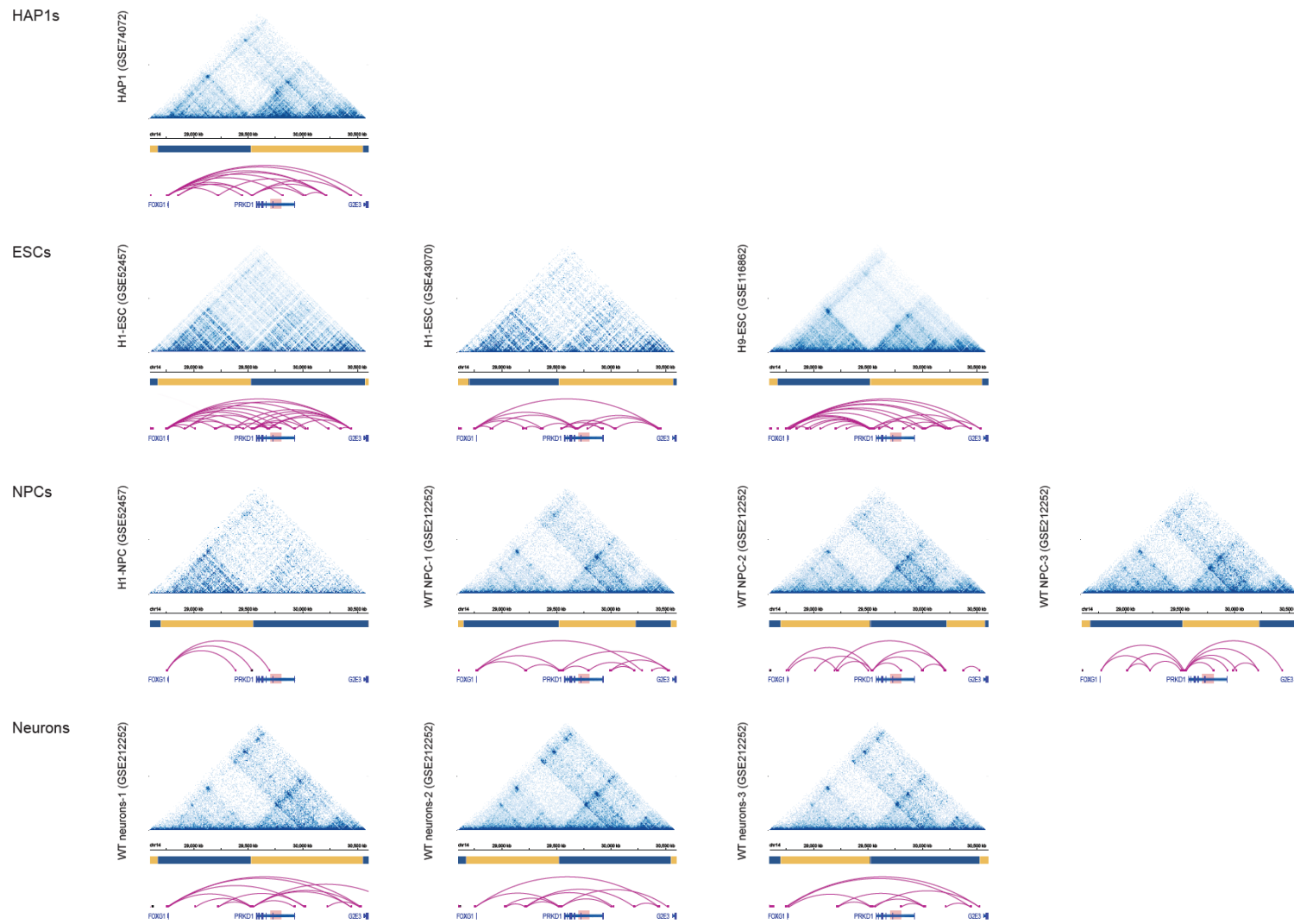
***Genomic intervals of differential UMI-4C contacts between WT and 14q12-MRO-Del**

Coordinates (hg19)	Coordinates (hg38)	p-value	log2FoldChange
chr14:29,340,992-29,344,417	chr14:28,871,786-28,875,211	0.0021	1.13
chr14:29,387,328-29,391,265	chr14:28,918,122-28,922,059	0.0016	-1.42
chr14:30,707,603-30,710,998	chr14:30,238,397-30,241,792	0.0013	5.14
chr14:30,724,104-30,728,011	chr14:30,254,898-30,258,805	0.0015	-4.98
chr14:30,745,035-30,748,796	chr14:30,275,829-30,279,590	0.0019	-4.89
chr14:30,775,521-30,779,248	chr14:30,306,315-30,310,042	0.0012	-5.14

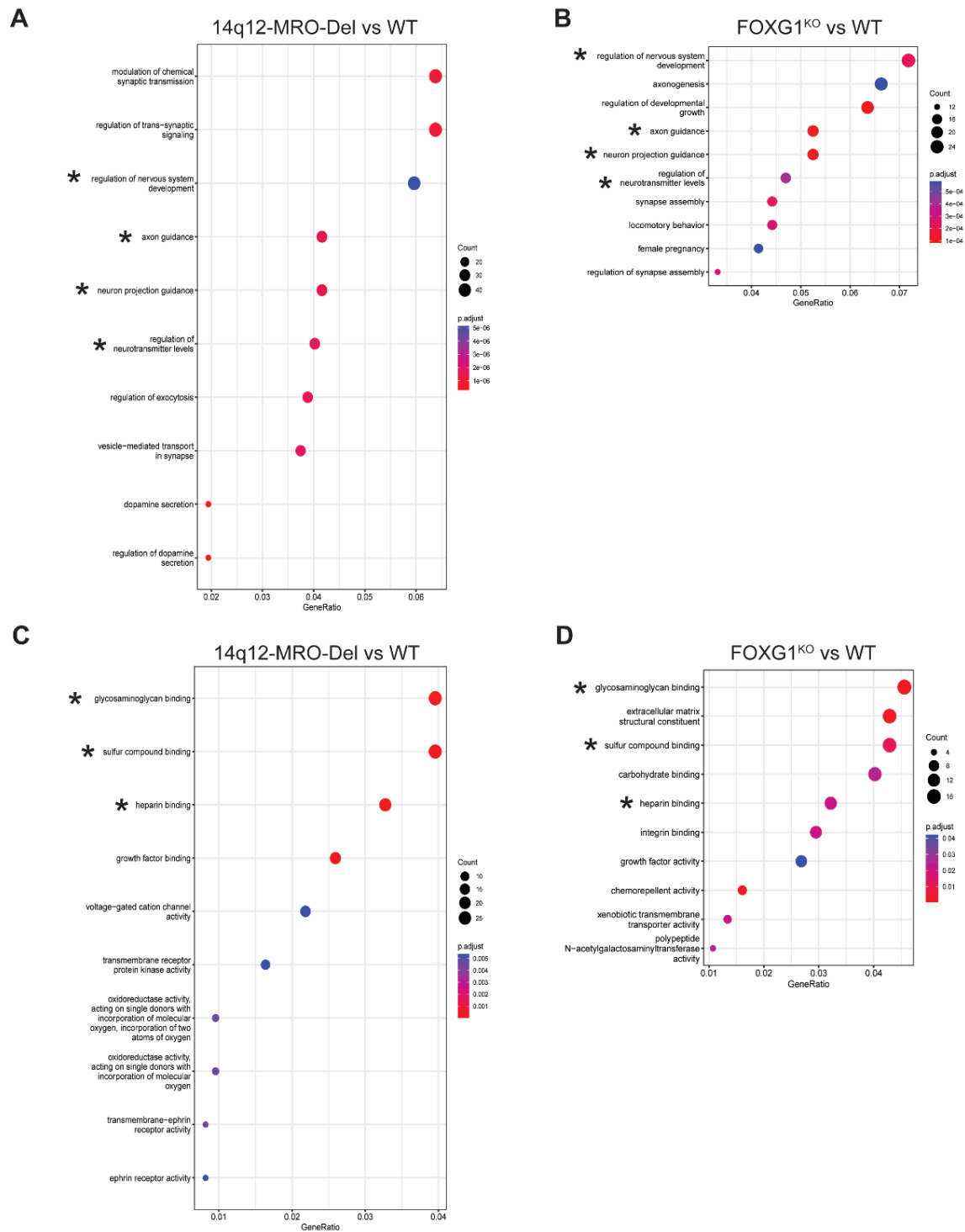
Supplementary Figure 6. Differential UMI-4C contact analysis. The top panel of UMI-4C profiles represents a smoothed trend of normalized counts of *FOXG1* genomic interactions from WT (black) and 14q12-MRO-Del (purple) samples. The black inverted triangle is directed towards the *FOXG1* viewpoint. The differential analysis panel represents log₂ fold change (FC) of differential UMI-4C contacts in the 14q12 region from WT and 14q12-MRO-Del samples. Asterisks in black indicate significant differences in contacts between samples (FDR adjusted *p*-value < 0.05). The bottom panel is a domainogram that illustrates the mean contact intensity log₂ fold changes between WT and 14q12-MRO-Del. * refer to genomic coordinates of the differential UMI contacts where negative log₂FC is indicative of increased contacts in the 14q12-MRO-Del line, positive values are loci that have decreased contacts in the 14q12-MRO-Del line (higher in WT). Processed data for differential genomic interactions between samples are provided in the Source Data file.



Supplementary Figure 7. Deletions downstream of *FOXG1* in the context of native 14q12 chromatin architecture. HAP1 Hi-C data matrix (previously generated by ²) with TAD annotations reveal the SRO ³ as well as a majority of the 14q12 deletions (this study and ⁴), span both the TAD containing *FOXG1* and a neighboring TAD. The MRO (vertical highlight in red) resides in the TAD neighboring *FOXG1*. Hi-C data was visualized using the 3D Genome Browser ⁵.



Supplementary Figure 8. Native 14q12 chromatin architecture across various cell types. Hi-C data from HAP1s, H1 and H9 (ESCs), neural progenitor cells (NPCs) and NPC-derived neurons reveal a highly conserved *FOXG1* TAD boundary across cell types, with chromatin interactions spanning TAD boundaries. TADs are annotated by alternating horizontal blue and yellow bars. The MRO is highlighted in red. Hi-C data was visualized using the 3D Genome Browser ⁵.



Supplementary Figure 9. Functional enrichment (FE) analysis of DEGs. (A-B) Biological pathways and (C-D) molecular functions that are shared between 14q12-MRO-Del vs WT and FOXG1^{KO} vs WT datasets are designated as *.

Supplementary Table 1. Catalog of gRNAs and oligos

Protocol	Specific Usage	Oligo Sequence (5' to 3')
CRISPR/ Cas9 gene editing	MRO left breakpoint gRNA	ACATATAGAATGGAAGTTTG
	MRO right breakpoint gRNA	CTTCAGCTCACTGCTTACTG
	FOXG1 ^{KO} left breakpoint gRNA	AGCATCACCCAGTCGTCGGG
	FOXG1 ^{KO} right breakpoint gRNA	TCACTTACAGTCTGGTCCCA
Genotyping PCR primers	MRO breakpoint PCR F	TCATTCTTGCCAAAGTAAAACA
	MRO breakpoint PCR R	TGCCCCGGCTAATTTTTGTAT
	FOXG1 ^{KO} breakpoint PCR F	GGCCAGACCAGTTACTTTTTCC
	FOXG1 ^{KO} breakpoint PCR R	TTTAGGTTGTTCTCAAGGTCTGC
qRT-PCR primers	FOXG1 Primer F	CCTGCCCTGTGAGTCTTTAAG
	FOXG1 Primer R	GTTCACTTACAGTCTGGTCCC
	PRKD1 Primer F	CTTTTTCGCCATGACCCTACC
	PRKD1 Primer R	GGAAGCTGACAAGACCACTTCA
	EAR Primer F	GAGGCTGAGGCAGGAGAATCG
	EAR Primer R	GTCGCCCAGGCTGGAGTG
UMI-4C primers	FOXG1 viewpoint US primer	GGGTGGGGAGGATGGAATAA
	FOXG1 viewpoint DS primer	AATGATACGGCGACCACCGAGATCTAC ACTCTTTCCCTACACGACGCTCTTCCG ATCTTGAGTATTCATCAACCGCATTCT
	Illumina Universal Primer 2	CAAGCAGAAGACGGCATACGA

Supplementary Table 2. Catalog of cell lines

Cell Line	Genotype
WT-Clone A (unedited)	FOXG1 ⁺ .MRO ⁺
WT-Clone B	FOXG1 ⁺ .MRO ⁺
14q12-MRO-Del	FOXG1 ⁺ .MRO ⁻
FOXG1 ^{KO}	FOXG1 ⁻ .MRO ⁺

Abbreviations: WT, wildtype; KO, knock-out, Del, deletion

Supplementary Table 3. Catalog of antibodies

Antibody	Experiment	Binding Condition	Product Information
FOXG1 (epitope within the last 100 amino acids)	Western (primary)	1:1000 overnight at 4°C	Abcam 196868 Clone# EPR18987
TAF5	Western (primary)	1:1000 overnight at 4°C	Bethyl Labs A303-686A
Goat anti-Rabbit IgG H&L (HRP)	Western (secondary)	1:20,000 1 hr at RT	Abcam 205718

Supplementary Table 4. Genomic intervals of interest in UMI-4C analysis

Interval ID	Coordinates (hg19)	Coordinates (hg38)	Size
FOXG1 promoter bait	chr14: 29,234,251-29,234,254	chr14: 28,765,045-28,765,048	4 bp
14q12-MRO	chr14: 30,173,942-30,273,103	chr14: 29,704,736-29,803,897	99 kb
14q12-MRO-Del	chr14: 30,173,921-30,273,106	chr14: 29,704,715-29,803,900	99 kb
SRO	chr14: 29,875,672-30,303,083	chr14:29406466-29833877	427kb

Supplementary Table 5. Genomic intervals of cCREs tested in luciferase assay

CRE ID	Coordinates (hg38)
cCRE1	chr14:29,736,248-29,737,089
cCRE2	chr14:29,799,340-29,799,959

Supplementary Data 1. Differentially expressed genes (DEGs) and direct FOXG1 targets from RNA-seq

List of items in Source Data file:

- Fig. 2A. *FOXG1* mRNA Fold Change Values, normalized to WT.
- Fig. 2B. *FOXG1* Protein Fold Change Values, normalized to WT. Densitometry data and Western Blots
- Fig. 3. Genomic coordinates of interest including GWAS and MPRA hits.
- Fig. 4B. Luciferase Reporter Assay Fold Change Values, normalized to Empty.
- Supplementary Fig. 1. Normalized transcript per million (nTPM) values for *FOXG1* across various cell/tissue types.
- Supplementary Fig. 4B. *PRKD1* mRNA Fold Change Values, normalized to WT.
- Supplementary Fig. 6. Differential genomic interactions between samples from UMICats.

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1. Kosicki, M. *et al.* VISTA Enhancer browser: an updated database of tissue-specific developmental enhancers. *Nucleic Acids Res* **53**, (2025).
2. Sanborn, A. L. *et al.* Chromatin extrusion explains key features of loop and domain formation in wild-type and engineered genomes. *Proc Natl Acad Sci U S A* **112**, E6456–E6465 (2015).
3. Allou, L. *et al.* 14q12 and severe Rett-like phenotypes: new clinical insights and physical mapping of FOXP1-regulatory elements. *Eur J Hum Genet* **20**, 1216–1223 (2012).
4. Ellaway, C. J. *et al.* 14q12 microdeletions excluding FOXP1 give rise to a congenital variant Rett syndrome-like phenotype. *European Journal of Human Genetics* **21**, 522–527 (2012).
5. Wang, Y. *et al.* The 3D Genome Browser: A web-based browser for visualizing 3D genome organization and long-range chromatin interactions. *Genome Biol* **19**, 1–12 (2018).