

SUPPLEMENTAL INFORMATION

(p)ppGpp imposes graded transcriptional changes to impair motility and promote antibiotic tolerance in biofilms

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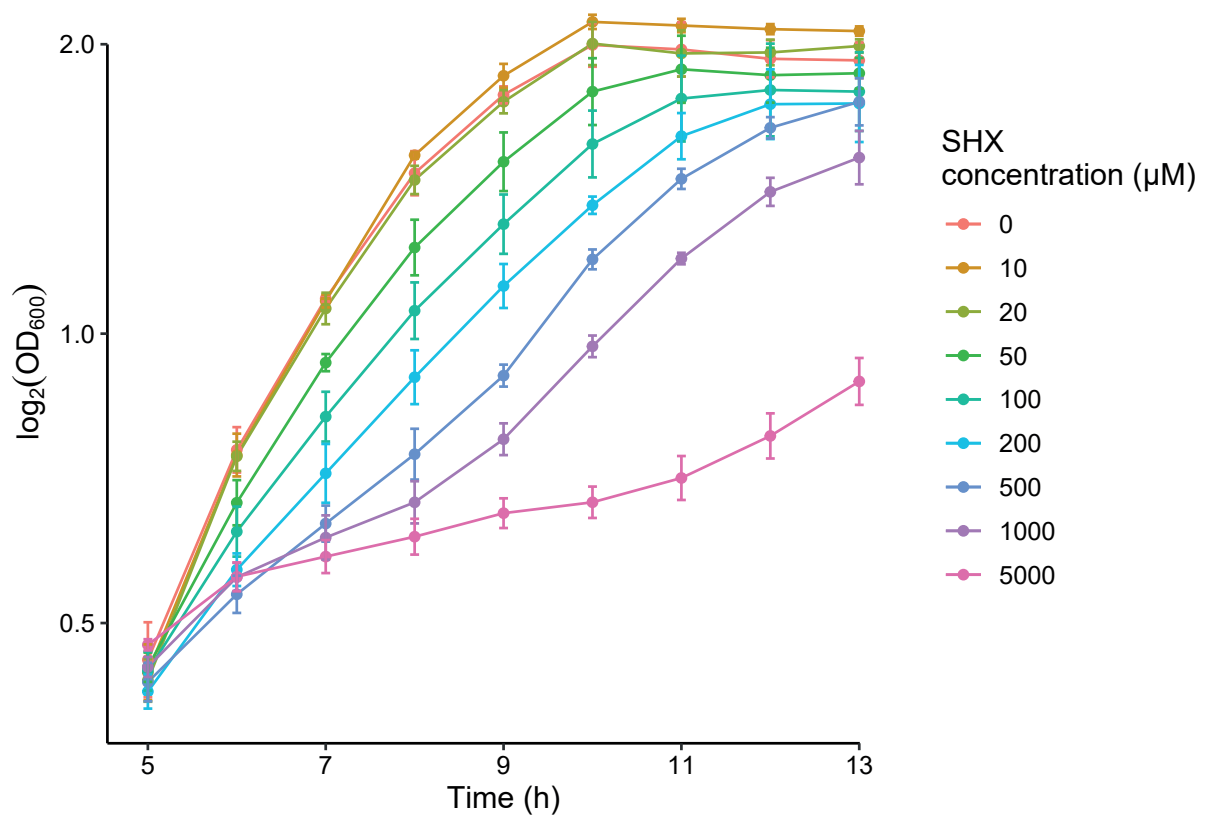
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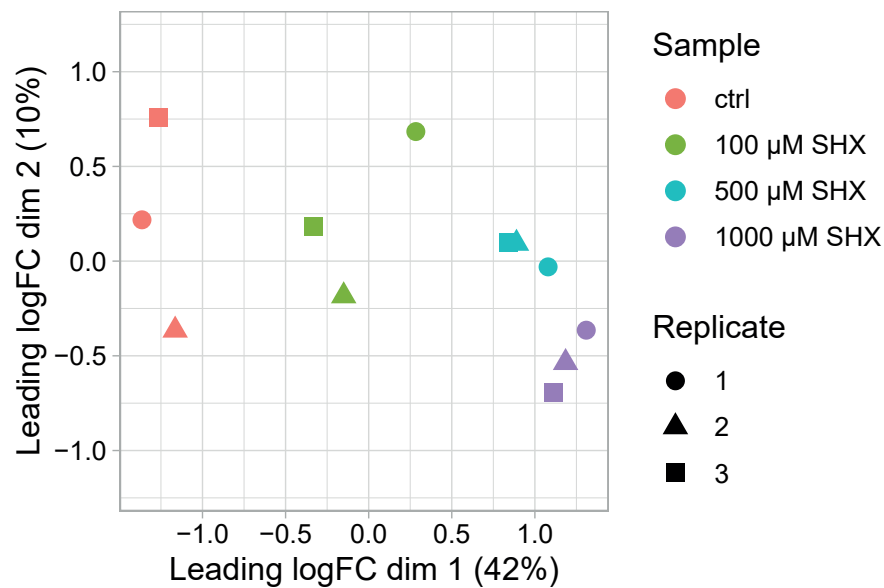
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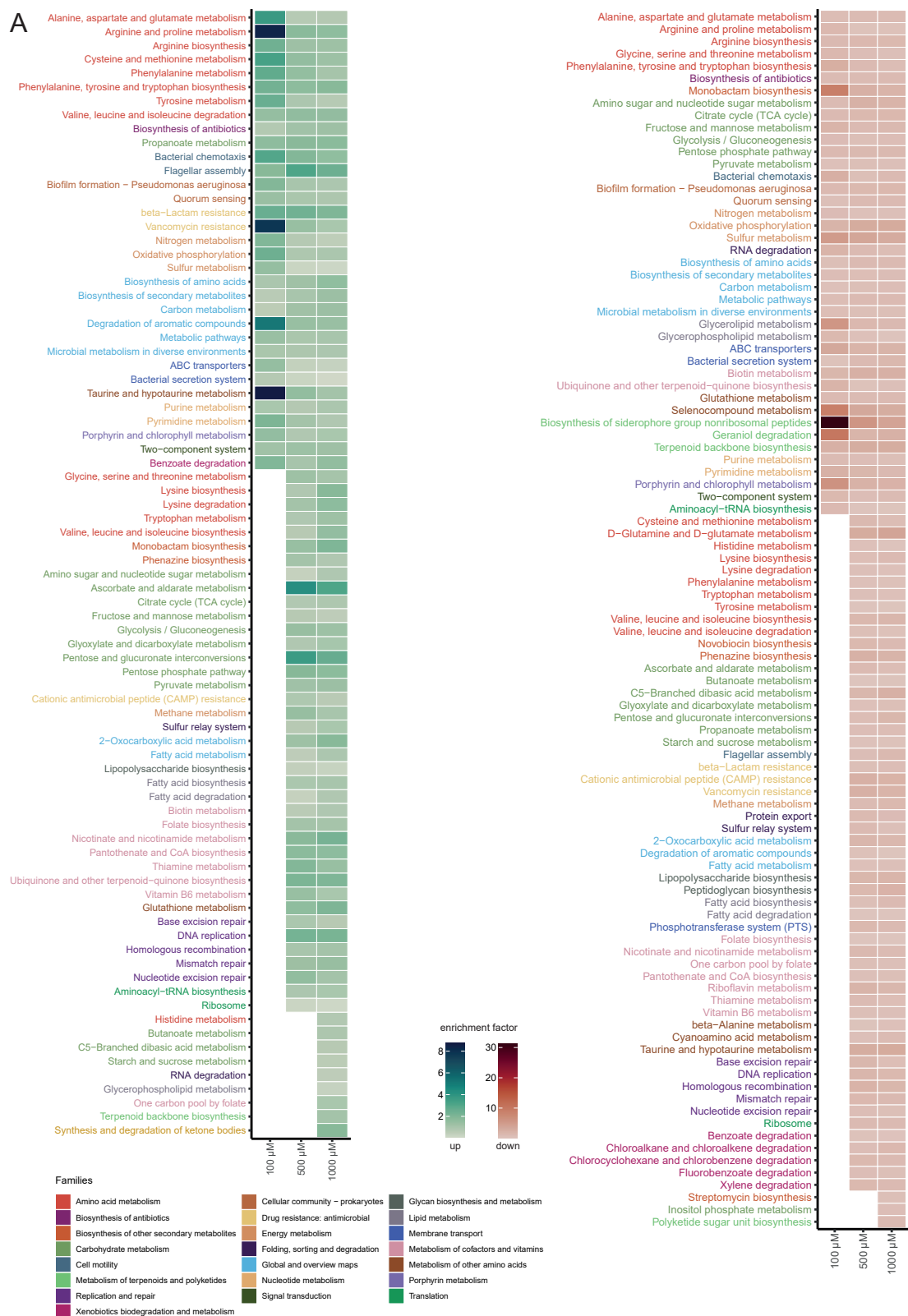
Supplementary Figure 1 (linked to Figure 1A). Effect of increasing concentrations of serine hydroxamate (SHX) on the growth of *P. aeruginosa* PA14.

The growth of exponentially *P. aeruginosa* PA14 was measured following treatment with increasing concentration of SHX. The experiment was performed in triplicates. Data points are shown as mean $\pm$ standard deviation.



Supplementary Figure 2 (linked to Figure 2). Principal Component Analysis (PCA) of transcriptomic data from *P. aeruginosa* PA14 exponentially grown and treated with increasing concentrations of serine hydroxamate (SHX).

The analysis was performed on three biological replicates for each treatment condition. The first two principal components (dim 1 and dim2) are plotted, with each point representing a replicate sample. The plot shows the clustering of replicates from untreated and each SHX treatment group, highlighting the variation in gene expression profiles in response to different SHX concentrations. The distance between the clusters indicates the degree of transcriptional variation across treatments.



B

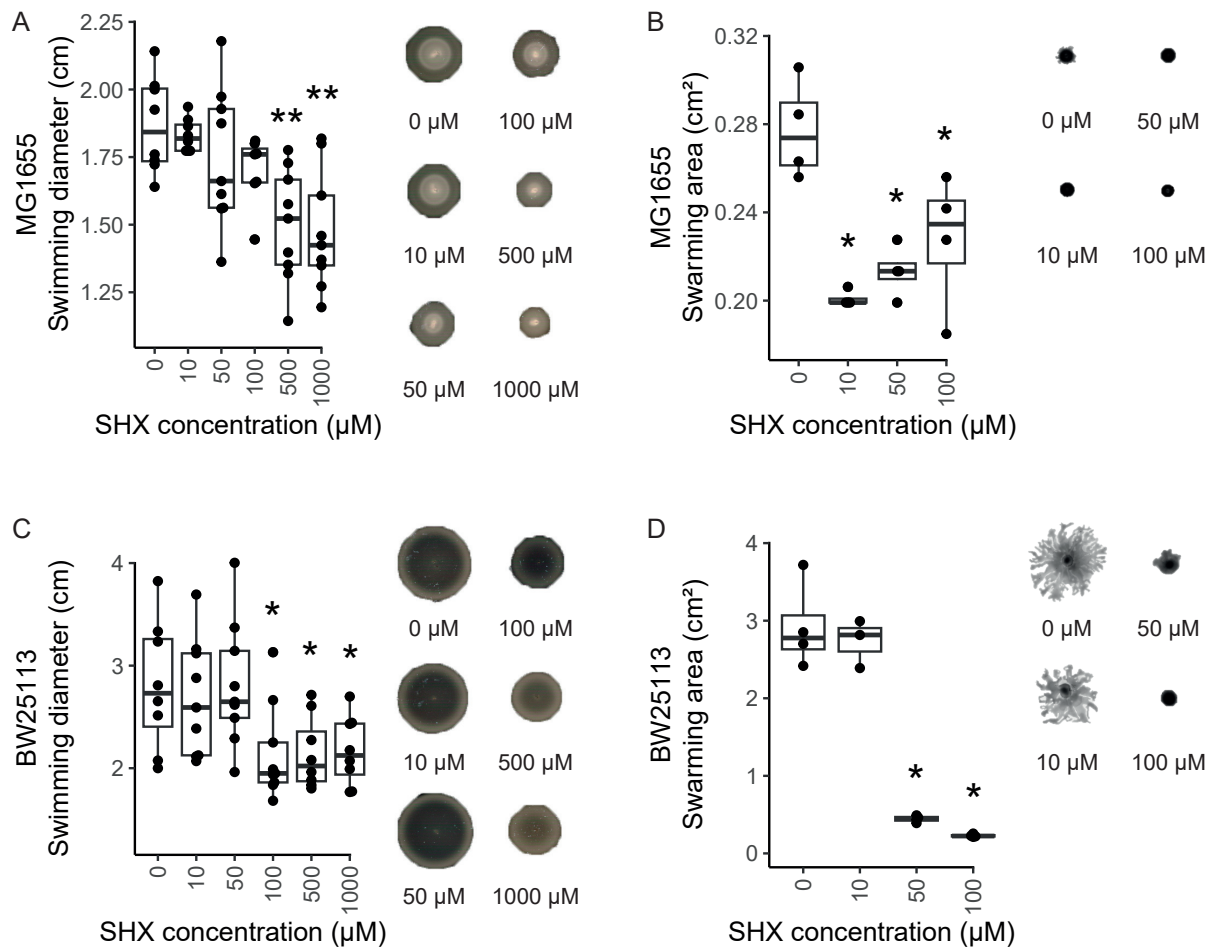


Supplementary Figure 3 (linked to Figure 2). Pathway analysis of gene expression changes in *P. aeruginosa* PA14 treated with increasing concentrations of serine hydroxamate (SHX).

(A) KEGG and (B) Gene Ontology (GO) pathway enrichment analysis showing cellular pathways whose expression is dependent on increased (p)ppGpp levels. The pathways that were upregulated are shaded in green, and those that were downregulated are shaded in red, with the intensity of the shading representing the enrichment factor (i.e., the significance of the pathway's differential regulation). The analysis was performed with SHX treatments at 100 μ M, 500 μ M, and 1000 μ M, with changes relative to the exponentially grown, untreated control sample.

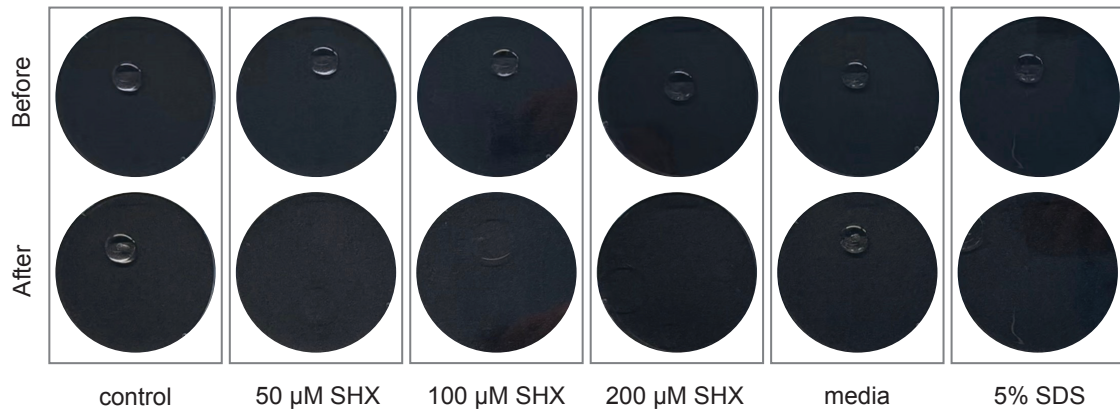
Supplementary Figure 4: Visualisation of a selection of cellular pathways affected upon (p)ppGpp induction.

This figure visualises the transcriptional effects of (p)ppGpp on genes from selected cellular pathways derived from KEGG enrichment analysis. Each gene is color-coded to reflect the observed effects at increasing SHX treatments (100 μ M, 500 μ M, and 1000 μ M) relative to the exponentially grown untreated control sample. The colours red and green indicate $\log_2$ decreases or increases in gene expression, respectively, while genes in yellow are unaffected by (p)ppGpp. These pathways are selected to illustrate the graded effects of (p)ppGpp induction on cellular processes, but do not represent all regulated pathways or genes.



Supplementary Figure 5 (linked to Figure 3). (p)ppGpp induction negatively affects motility.

A-D) The ability of *E. coli* MG1655 and BW25113 to swim (A and C, respectively) or swarm (B and D, respectively) was determined in the presence of increasing serine hydroxamate (SHX) concentrations. Increase in the SHX concentration abolished both motility phenotypes. The experiments were repeated at least four times and representative images of swimming and swarming motility agar plates are shown on the right side of each panel. Significance was tested by using the Wilcoxon test (* $P \leq 0.05$, ** $P \leq 0.005$).



Supplementary Figure 6. (p)ppGpp induction promotes biosurfactant production.

P. aeruginosa PA14 cultures treated with increasing concentrations of serine hydroxamate (SHX) (50 μ M, 100 μ M, and 200 μ M) were added to mineral oil on the surface of distilled water (A). A 5% SDS solution was also added as a positive control. This resulted in oil displacement by SHX-treated cultures and SDS (B). Untreated cultures and media showed no effect on oil displacement, confirming the biosurfactant production induced by (p)ppGpp. These are representative images from at least three independent experiments.

Supplementary Table 1: List of genes differentially regulated upon activation of the stringent response. Genes with unknown function are italicised and in bold.

Supplementary Table 2: Minimal inhibitory concentrations (MIC) of ciprofloxacin or tobramycin for *Pseudomonas aeruginosa* PA14 or *Escherichia coli* MG1655 in the presence of increasing concentrations of serine hydroxamate (SHX).

SHX (μM)	Ciprofloxacin (μg/mL)		Tobramycin (μg/mL)	
	PA14	MG1655	PA14	MG1655
0	0.125	0.016	2	0.5
50	0.125	0.016	2	0.5
100	0.063	0.016	1	0.5
200	0.063	0.016	1	0.5
500	0.063	0.016	1	0.25
1000	0.031	0.008	0.5	0.25

Supplementary Table 3: List of bacterial strains, chemicals, reagents, kits, devices, and informatics tools used in this study.

Reagent or Resource	Source	Identifier
Bacterial Strains		
<i>Pseudomonas aeruginosa</i> UCBPP-PA14	Laboratory stock	N/A
<i>Pseudomonas aeruginosa</i> PA01	Laboratory stock	N/A
<i>Pseudomonas aeruginosa</i> CH7480	Laboratory stock	N/A
<i>Escherichia coli</i> MG1655	Laboratory stock	N/A
<i>Escherichia coli</i> BW25113	Laboratory stock	N/A
Chemicals		
DL-serine hydroxamate	Merck	Cat#S4503
RNAprotect Bacteria Reagent	Qiagen	Cat#76506
FastAP buffer	ThermoFisher Scientific	Cat#EF0651
TurboDNase	Invitrogen	Cat#AM1907
T4 RNA Ligase 1	New England Biolabs	Cat#M0437
SMARTScribe Reverse Transcriptase	Takara	Cat#639536
Exonuclease I	New England Biolabs	Cat#M0293
AccuPrime Taq DNA Polymerase	Invitrogen	Cat#12346-086
CAS Amino Acids	ThermoFisher Scientific	Cat#228820
Tobramycin	Merck	Cat#T1783

Ciprofloxacin	Merck	Cat#17850
Congo red	Merck	Cat#C6767
Coomassie Brilliant blue	Merck	Cat#112553
Phosphorus-32 Radionuclide	Perkin Elmer	Cat#NEX053H001MC
Yeast extract	Carl Roth	Cat#2363.2
Sodium chloride (NaCl)	Carl Roth	Cat#3957.1
Tryptone	Carl Roth	Cat#8952.2
Magnesium sulphate (MgSO ₄)	Carl Roth	Cat#1A99.2
Calcium chloride (CaCl ₂)	Carl Roth	Cat#A119.1
Ammonium chloride (NH ₄ Cl)	Carl Roth	Cat#5470.1
Potassium dihydrogen phosphate (KH ₂ PO ₄)	Carl Roth	Cat#P018.1
di-Sodium hydrogen phosphate heptahydrate (Na ₂ HPO ₄ · 7H ₂ O)	Carl Roth	Cat#X987.2
Iron(II) sulphate heptahydrate (FeSO ₄ · 7H ₂ O)	Carl Roth	Cat#3722.1
D(+)-Glucose monohydrate	Carl Roth	Cat#6887.1
TE Buffer	ThermoFisher Scientific	Cat#12090015
Lysozym	Carl Roth	Cat#8259.2
2-Mercaptoethanol	Carl Roth	Cat#4227.3
BD Bacto Agar	ThermoFisher Scientific	Cat#10455513
Casamino acids	MP Biomedicals	Cat#113060012-CF
Mineral oil	Merck	Cat#330760
Other		
Fragment Analyzer	Agilent	https://www.agilent.com
Qubit 2.0	ThermoFisher Scientific	https://www.thermofisher.com
QIAshredder column	Qiagen	Cat#79656
NovaSeq 6000	Illumina	https://www.illumina.com/
RNeasy Mini Kit	Qiagen	Cat#74106
DNA-free DNA Removal kit	Qiagen	Cat#AM1906
RNAClean XP magnetic beads	Beckman Coulter	Cat#A63987
RNA Clean & Concentrator Kit	Zymo Research	Cat#R1017
RiboZero	Illumina	Cat#RZMB12324
Half-area 96-well µClear microtiter plate	Greiner Bio-One	Cat#675096
BREATHseal cover foil	Greiner Bio-One	Cat#676050
LIVE/DEAD® BacLight® Bacterial Viability Kit	Invitrogen	Cat#L7012

Confocal laser scanning microscope SP8	Leica Microsystems	https://www.leica-microsystems.com/
PEI (Polyethylenimine) Cellulose Plates	Merck	Cat#Z122882
Typhoon FLA 9500	GE Healthcare	N/A
Software and Algorithms		
FastQC	https://github.com/s-andrews/FastQC	v0.11.9
bowtie2	Langmead & Salzberg, 2012	v1.3.1
featureCounts	Liao et al., 2014	v2.0.1
edgeR	Chen et al., 2024	N/A
KOBAS-i	https://bioinfo.org/kobas/genelist/	v3.0
pathview	Luo & Brouwer, 2013	N/A
eulerr	DOI: 10.32614/CRAN.package.eulerr	7.0.2
Developer XD 64	Definiens	N/A
Imaris	Bitplane	v7.6
FIJI	Schindelin et al., 2012; DOI 10.1038/nmeth.2019	v1.54f

Supplementary Table 4: List of the barcoded adapter sequences used in the preparation of the RNA-seq library.

Barcode ID	Barcode Sequence 5' --> 3'; 5' phosphorylated
L01	ACCAAGTCGAGATCGGAAGAGCGTCGTGTA
L02	AGCAGCCACAGATCGGAAGAGCGTCGTGTA
L03	AGTAACTGCAGATCGGAAGAGCGTCGTGTA
L04	ATCATCGTGAGATCGGAAGAGCGTCGTGTA
L05	ATTACCACGAGATCGGAAGAGCGTCGTGTA
L06	AAGAATTATAGATCGGAAGAGCGTCGTGTA
L07	AGGCCCAAGAGATCGGAAGAGCGTCGTGTA
L08	ATACAACATAGATCGGAAGAGCGTCGTGTA
L09	ATGAACCAGAGATCGGAAGAGCGTCGTGTA
L10	AATATGGACAGATCGGAAGAGCGTCGTGTA
L11	ACAACTCGCAGATCGGAAGAGCGTCGTGTA
L12	ACGGAGGGCAGATCGGAAGAGCGTCGTGTA
L13	AACATTATTAGATCGGAAGAGCGTCGTGTA
L14	AATCACTTGAGATCGGAAGAGCGTCGTGTA
L15	ACCCGTCTTAGATCGGAAGAGCGTCGTGTA
L16	ACTCGGTACAGATCGGAAGAGCGTCGTGTA
L17	AGTCTGGCGAGATCGGAAGAGCGTCGTGTA
L18	ATCCCGCGGAGATCGGAAGAGCGTCGTGTA

L19	ACGGCACTTAGATCGGAAGAGCGTCGTGTA
L20	AGAGATTGTAGATCGGAAGAGCGTCGTGTA
L21	ATACAGATGAGATCGGAAGAGCGTCGTGTA
L22	ATGGGAGACAGATCGGAAGAGCGTCGTGTA
L23	ACCCTACAGAGATCGGAAGAGCGTCGTGTA
L24	ACTCTAACTAGATCGGAAGAGCGTCGTGTA
L25	AAAGTGTTGAGATCGGAAGAGCGTCGTGTA
L26	AGAGCCATCAGATCGGAAGAGCGTCGTGTA
L27	AGGTCCTCTAGATCGGAAGAGCGTCGTGTA
L28	ATACCGGCCAGATCGGAAGAGCGTCGTGTA
L29	ACCCTCGGCAGATCGGAAGAGCGTCGTGTA
L30	ACTGGATCGAGATCGGAAGAGCGTCGTGTA
L31	ATTTCTAACAGATCGGAAGAGCGTCGTGTA
L32	ACCGGTACCAGATCGGAAGAGCGTCGTGTA

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