

## Supplementary Data

**Table S1:** Full bioinformatic analysis of sequencing data comparing the average of the three replicates for each of the sequencing set-ups to determine the enrichment factors based on the average of the three standard sequencing replicates for NipWT and compared to the standard promethION™ for NipGE.

**Table S2:** Overview of recommend conditions for either of the AS modes.

| AS modes   | Recommended conditions                                                                                                                                                                                                                     |
|------------|--------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| Enrichment | <ul style="list-style-type: none"><li>- Target of interest present at low abundance in the mixture</li><li>- Prior knowledge available on the target(s) of interest, including genetic fingerprint fully known and characterized</li></ul> |
| Depletion  | <ul style="list-style-type: none"><li>- Highly abundant unwanted DNA from the matrix</li><li>- Information on the unwanted matrix ingredients</li><li>- No specific information on the target(s) of interest</li></ul>                     |
| AS modes   | Recommended conditions                                                                                                                                                                                                                     |
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| Depletion  | <ul style="list-style-type: none"><li>- Highly abundant unwanted DNA from the matrix</li><li>- Information on the unwanted matrix ingredients</li><li>- No specific information on the target(s) of interest</li></ul>                     |

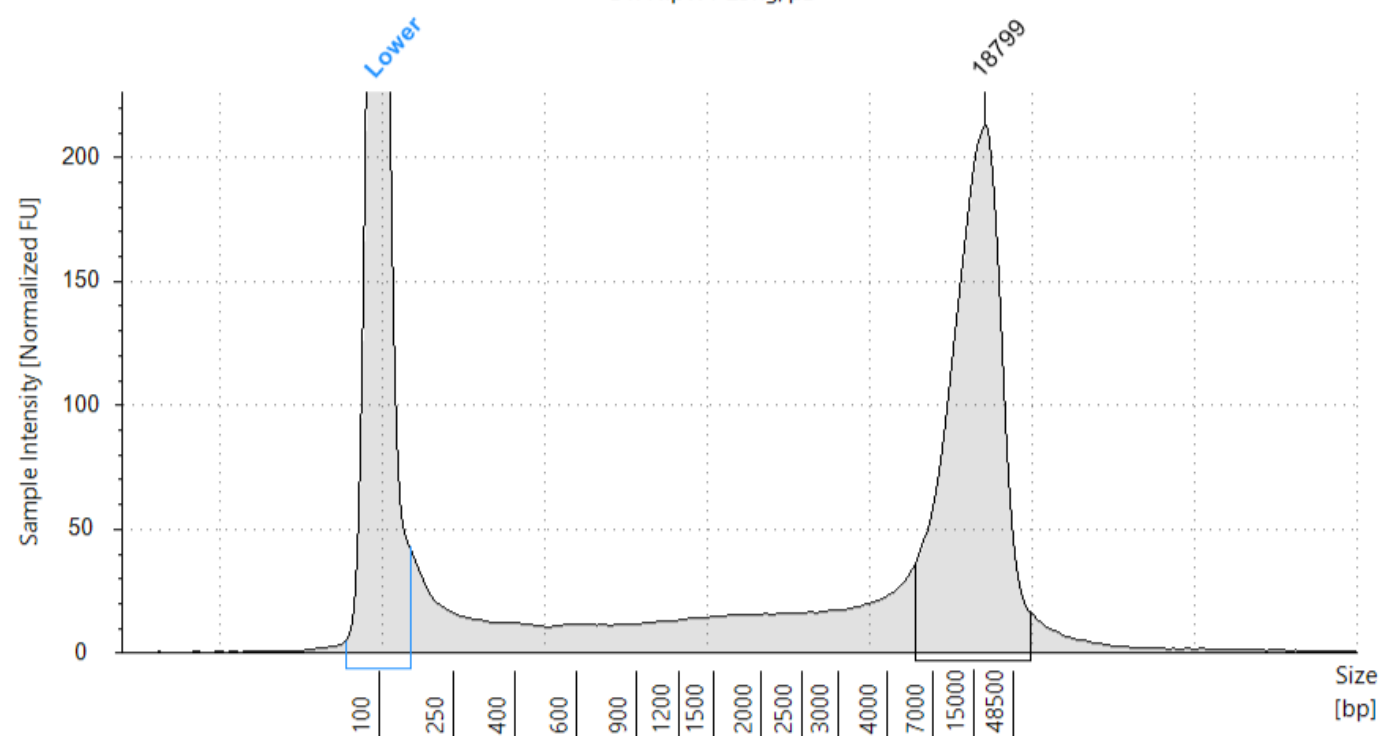
**Table S3:** full list of read depths for all the SNVs related to the cultivar specific marker of NipWT and NipGE for each of the sequencing setups. For each SNV the chromosome ID and position are shown together with the read depth and the reads for either the reference or alternative allele. In all of the cases, there were no reads for the reference allele, indeed confirming the presence of the SNV.

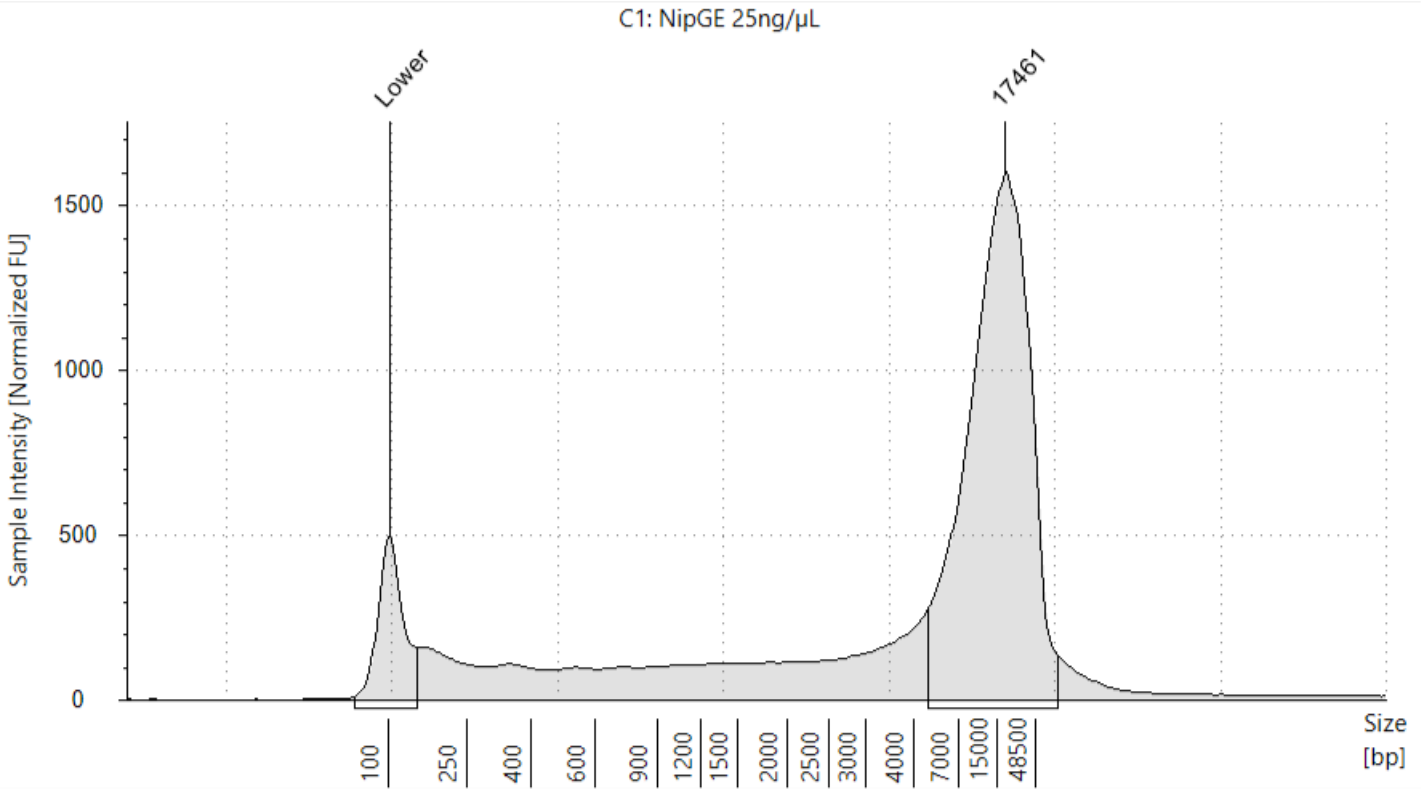
**Table S4: Ratio verification of each DNA mixture using ddPCR. The ratio is calculated based on the total genome length of each crop species times the measured amount of copies to ensure that the ratio incorporates the smaller length of the rice haploid (Arumuganathan & Earle, 1991; Shang et al., 2023).**

| Mixture                | Composition | Copies per reaction (20 µl) |          | Genome size (Mbp) | Total bases (Mbp) |          |
|------------------------|-------------|-----------------------------|----------|-------------------|-------------------|----------|
|                        |             | Expected                    | Observed |                   | Expected          | Observed |
| 1% NipWT + 99% Soybean | NipWT rice  | 55                          | 54       | 385.7             | 21213.5           | 20719    |
|                        | Soybean     | 2100                        | 1860     | 1013.2            | 2127720           | 1884633  |
|                        | Ratio       |                             |          |                   | 0.99%             | 1.09%    |
| 1% NipGE + 99% Soybean | NipGE rice  | 55                          | 63       | 385.7             | 21213.5           | 24376    |
|                        | Soybean     | 2100                        | 2463     | 1013.2            | 2127720           | 2495512  |
|                        | Ratio       |                             |          |                   | 0.99%             | 0.97%    |

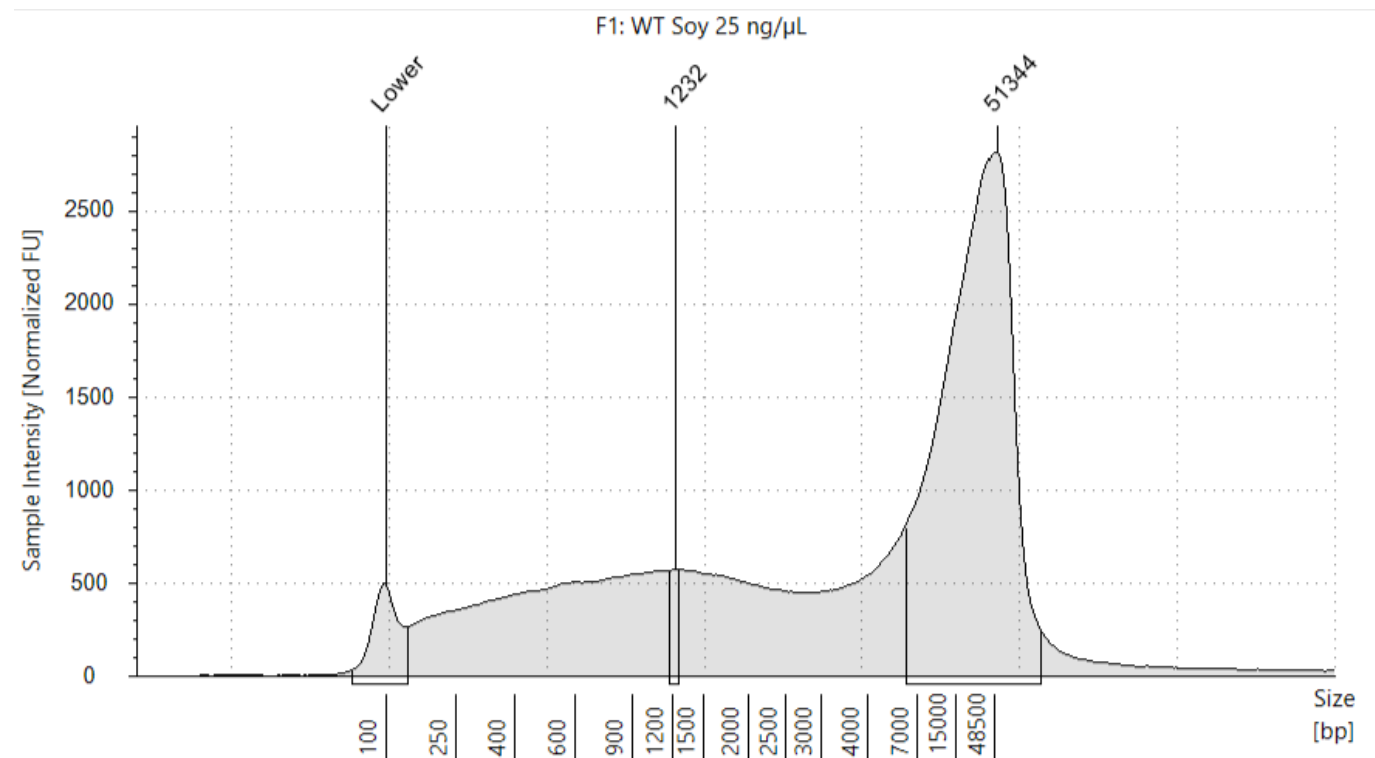
A: NipWT

B1: NipWT 25ng/μL



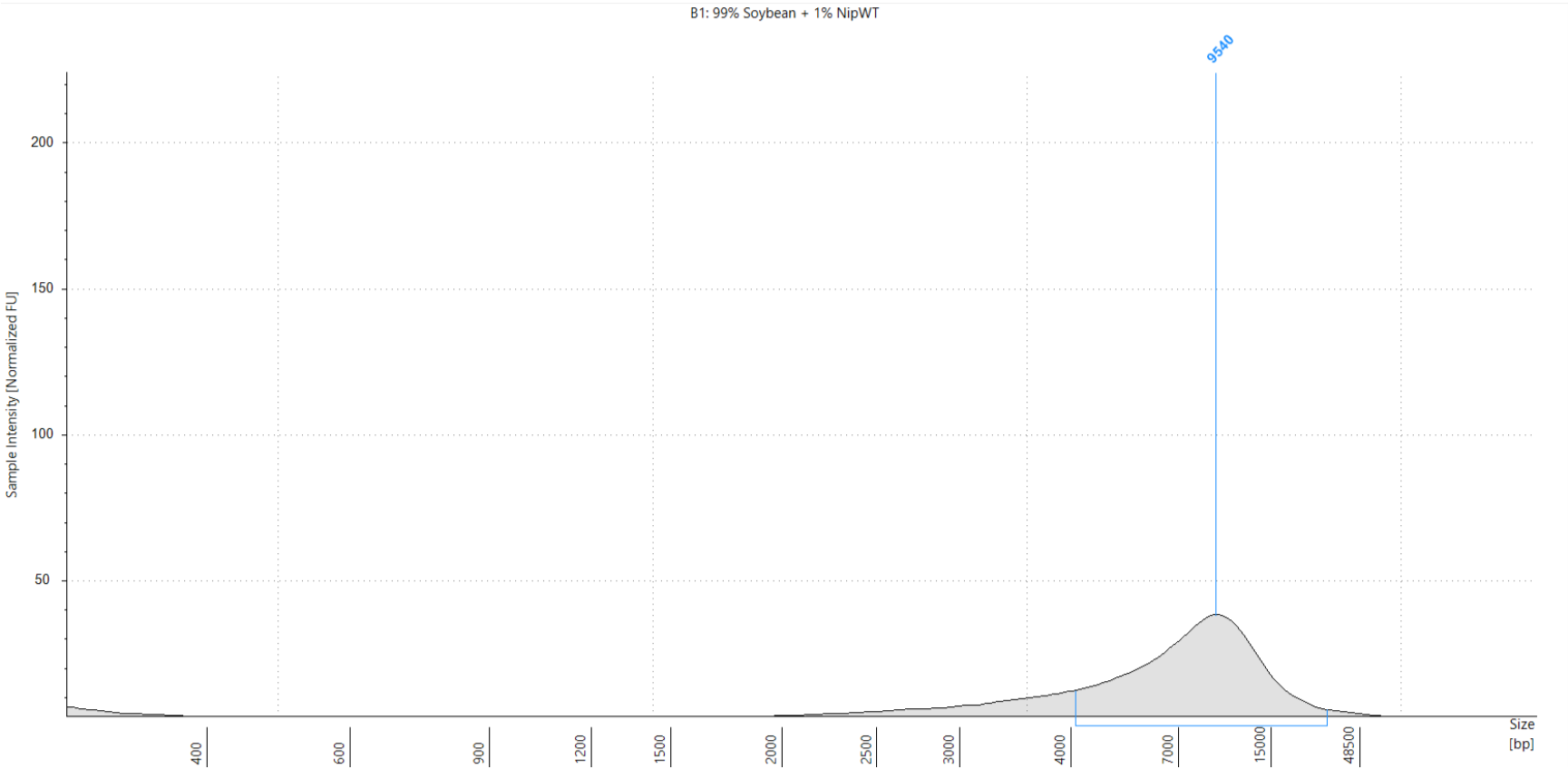


C: Soybean

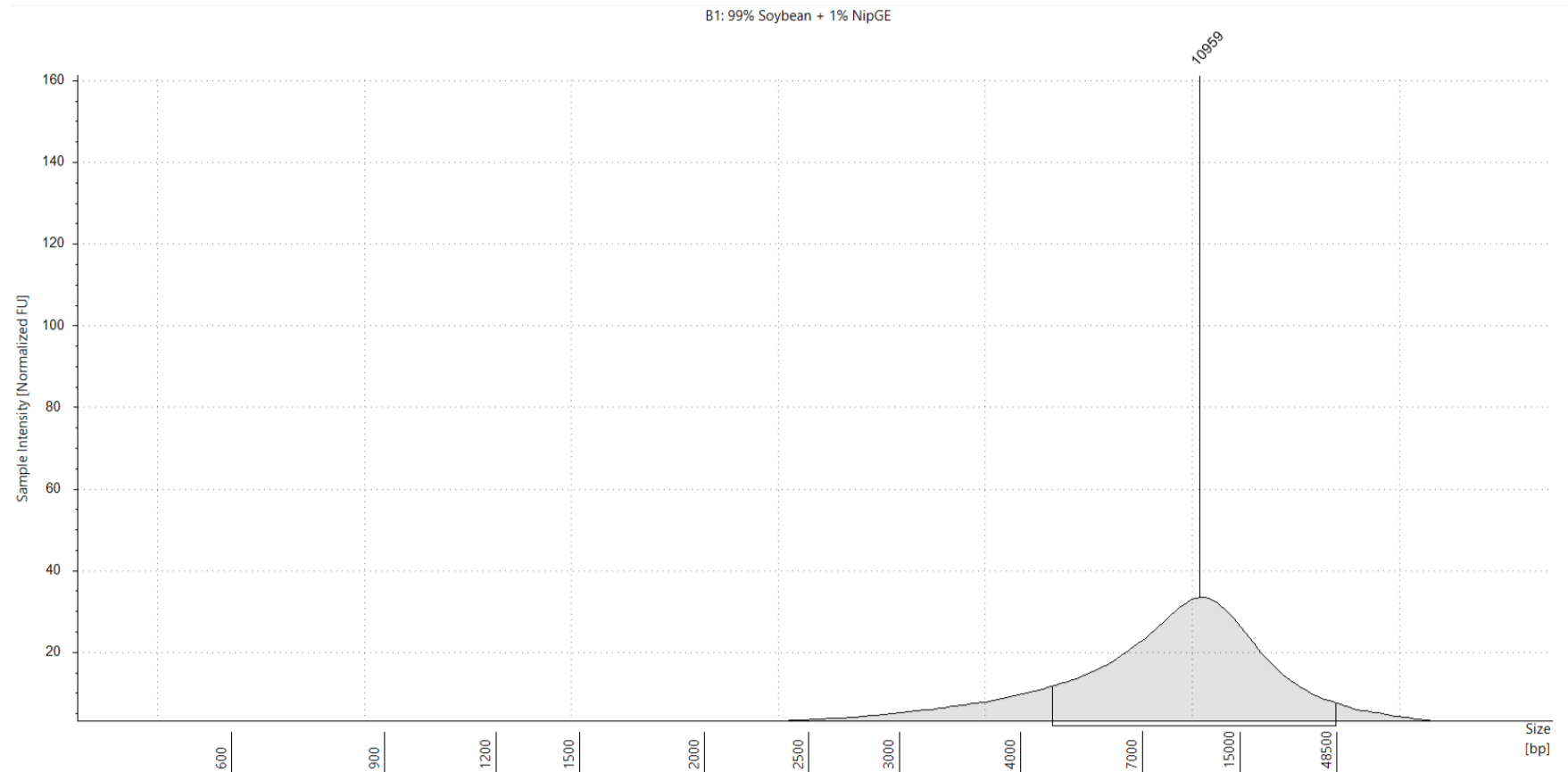


**Figure S1: Genomic profiles of ingredients using a TapeStation 4200 device with the associated Genomic Screen Tape and reagents (Agilent Technologies, Santa Clara, California, USA). NipWT (A), NipGE (B) and Soybean (C).**

**A : 99% Soybean + 1% NipWT**

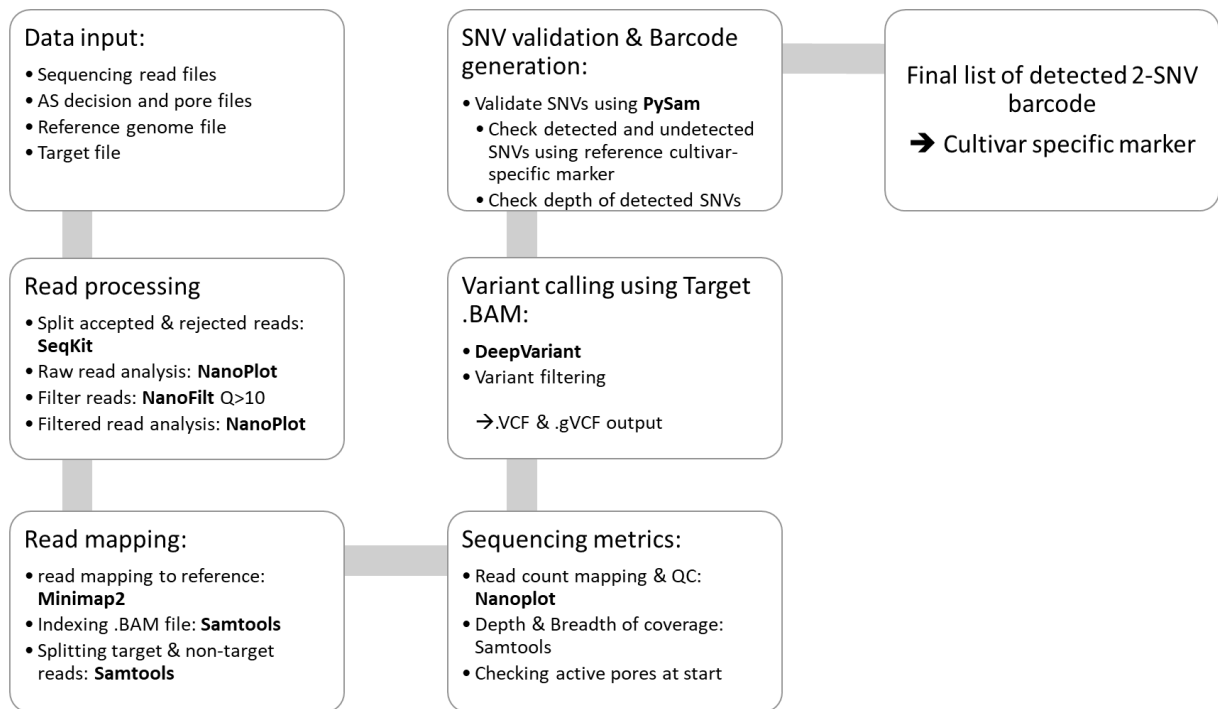


**B : 99% Soybean + 1% NipGE**

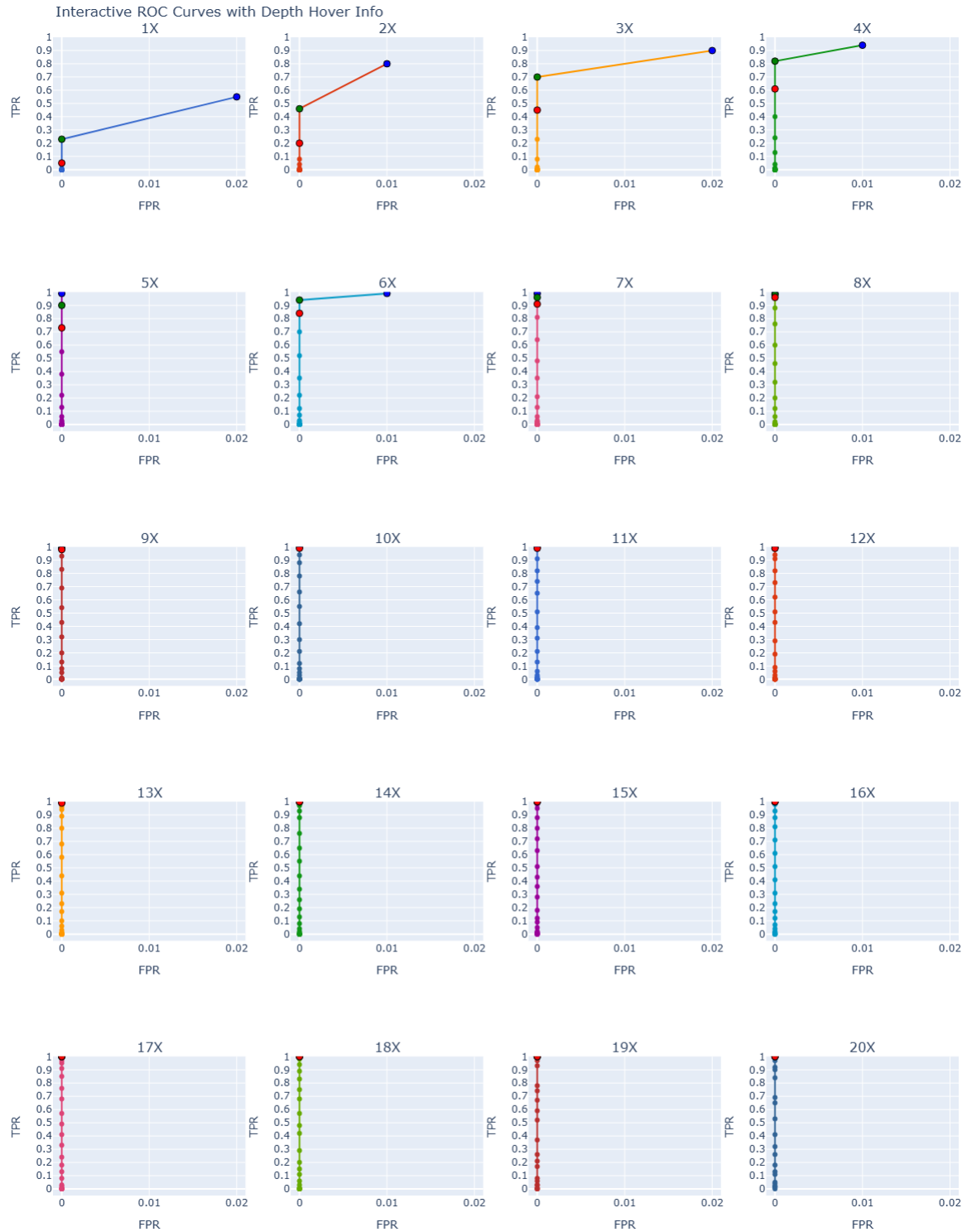


**Figure S2: Verification of sequencing mix fragmentation length using a Tapestation 4200 device with the associated Genomic Screen Tape and reagents (Agilent Technologies, Santa Clara, California, USA).**

The ideal length of fragments for Adaptive sampling libraries is between 6 and 8 kb (Oxford Nanopore Technologies, 2020) for NipWT (A) and NipGE (B) mixture. The verification of the fragment length was performed directly after the fragmentation step, prior to further library preparation.



**Figure S3: Schematic overview of read processing pipeline from raw sequencing reads to cultivar-specific 2-SNV barcode generation**



**Figure S4: Receiver Operating Characteristic (ROC) curves for SNV detection within the Nipponbare-cultivar marker.** Each subplot represents an average coverage depth, calculated as the mean true positive (TP) and false positive (FP) rates across replicates. Different points within each subplot indicate varying variant filtering cut-offs. The depth cut-offs of 1X, 2X, and 3X are shown in blue, green, and red, respectively, as the false positive rate (FPR) approaches zero beyond 3X across all depths. For extra safety, a minimum depth of 5X was therefore chosen.