

Unique thermal sensitivity imposes a cold-water energetic barrier for vertical migrators

In the format provided by the
authors and unedited

Supplementary Materials

This PDF file includes:

Supplementary Text

Supplementary Figure 1

Tables S1 and S2

Supplementary References

Supplementary Information

Metabolic Traits and their interrelationships.

α – Physiological oxygen supply capacity ($\mu\text{mol O}_2 \text{ g}^{-1} \text{ h}^{-1} \text{ kPa}^{-1}$): The maximum amount of oxygen that can be supplied per unit time, mass and oxygen pressure. The ratio of oxygen demand to supply (i.e. MR/PO_2) increases as oxygen declines and/or metabolic rate increases until it reaches a critical PO_2 (P_c), at which physiological oxygen supply (e.g. ventilation and cardiac output) is operating at its maximum capacity (α). The α is a species- and temperature- specific constant that quantifies the oxygen dependency of metabolism.

$$\alpha = \text{MR}/P_c$$

MR – Metabolic Rate ($\mu\text{mol O}_2 \text{ g}^{-1} \text{ h}^{-1}$): The rate of aerobic energy usage, estimated from oxygen consumption. The maximum metabolic rate (**MMR**) is the highest achievable MR, typically measured during exercise protocols. At PO_2 less than $P_{c\text{max}}$, MMR is directly oxygen dependent. The standard metabolic rate (**SMR**) refers to the fasted, resting metabolic rate at a specified temperature.

FAS – Factorial Aerobic Scope (no units): The factorial difference between MMR and SMR, which is equivalent to the factorial difference between $P_{c\text{max}}$ and $P_{c\text{SMR}}$. Like MMR, FAS is directly dependent on PO_2 below $P_{c\text{max}}$. FAS is a measure of the aerobic capacity to support activities (e.g., growth, reproduction, locomotion) beyond basic maintenance.

$$\text{FAS} = \text{MMR}/\text{SMR} = P_{c\text{max}}/P_c$$

P_c – The Critical PO_2 (kPa): The PO_2 at which oxygen supply reaches its maximum capacity (α) for any given metabolic rate (see *Eq. 1*). Equivalently, P_c is the PO_2 below which its corresponding metabolic rate becomes oxygen limited. P_c and $P_{c\text{max}}$ refer specifically to the critical PO_2 s for SMR and MMR, respectively.

E – Temperature sensitivity (eV): The temperature (T) dependence of a given physiological metric is parameterized as $\exp(-E/k_B T)$, where E is the temperature dependence and k_B is the Boltzmann constant. The temperature dependence (E) of each metric is the slope of the linear regression of $1/k_B T$ versus the natural log of the metric (i.e. the slope of an Arrhenius plot). The equation below shows the relationships between the temperature sensitivities of each metric.

$$E_{\text{FAS}} = E_{\text{MMR}} - E_{\text{SMR}} = E_{P_{c\text{max}}} - E_{P_c}$$

Fig. S1. Metabolic traits and their interrelationships. A) Maximum metabolic rate (MMR), Standard metabolic rate (SMR), and their critical oxygen pressures (P_{cmax} and P_{cSMR} , respectively) are quantitatively related via the oxygen supply capacity, α . The factorial aerobic scope (FAS) is the factorial difference between MMR and SMR, or between P_{cmax} and P_c , which are quantitatively equivalent. B) Seibel and Deutsch (2020) reported that there is a linear relationship between MMR and PO_2 (constancy of oxygen supply capacity across metabolic states from SMR to MMR at a given temperature). This is evident by the nearly perfect correlation between the oxygen supply capacity derived at SMR and P_c and that derived at MMR and measurement PO_2 (usually 21 kPa). For hypoxic species (red dots), which have a P_{cmax} well below the measurement PO_2 , we divided MMR by the prevailing environmental PO_2 . The slope of the relationship in this figure is 1.0 and the intercept is zero, which means that oxygen supply is the same at the limiting oxygen pressures for any metabolic level between SMR and MMR.

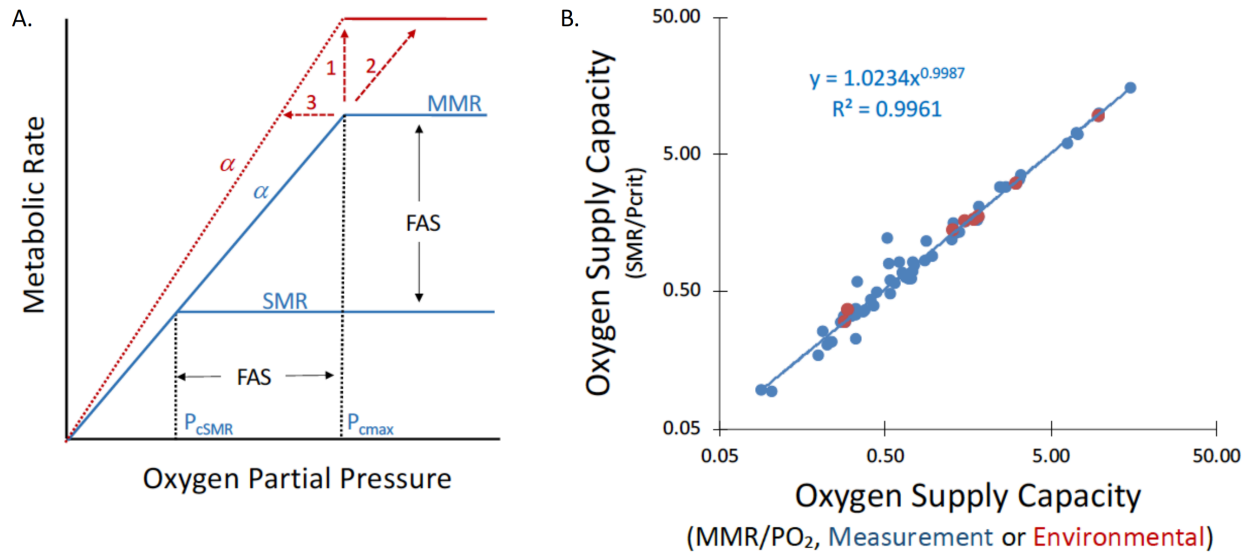


Table S1.

Dosidicus gigas metabolic traits and their temperature sensitivities (E ; Values are means $\pm$ SEM).

Temperature (°C)	SMR ($\mu\text{mol O}_2 \text{ g}^{-1}\text{h}^{-1}$)	MMR ($\mu\text{mol O}_2 \text{ g}^{-1}\text{h}^{-1}$)	P_{cSMR} (kPa)	α ($\mu\text{mol O}_2 \text{ g}^{-1}\text{h}^{-1} \text{ kPa}^{-1}$)
10	8.05 $\pm$ 0.51	17.3 $\pm$ 4.32	1.86 $\pm$ 0.43	4.97
20	10.26 $\pm$ 0.75	23.3 $\pm$ 1.96	3.50 $\pm$ 0.23	3.15
25	16.98 $\pm$ 0.88	69.8 $\pm$ 5.25	4.65 $\pm$ 0.54	3.66

Temperature (°C)	E_{SMR} (eV)	E_{MMR} (eV)	$E_{P_{\text{cSMR}}}$ (eV)	E_{α} (eV)
10-20	0.17	0.21	0.49	-0.33
20-25	0.75	1.65	0.53	0.23
10-25	0.33	0.60	0.51	-0.18

Table S2

Metabolic traits, including the standard metabolic rate (SMR), the critical PO_2 for SMR (P_{cSMR}), and the oxygen supply capacity (α), and their temperature sensitivities for diel vertical migrators.

Species (reference)	T (°C)	Mass, g (range)	Mean MO ₂ , μmol O ₂ g ⁻¹ h ⁻¹ (range, n)	Mean P _{crit} , kPa (range, n)	α, μmol O ₂ g ⁻¹ h ⁻¹ kPa ⁻¹	Temperature Sensitivity (E, eV)		
						SMR	P _c	α
Vertical Migrators								
<i>Nyctiphanes simplex</i> * (Euphausiacea)	10	0.037 ± 0.027 (0.007-0.0924)	7.24 ± 1.44 (4.14 – 9.02, 7)	2.12± 0.83 (1.16-3.68, 9)	3.87±1.33 (2.09-5.80; 7)	0.25	0.46	-0.20
	20	0.067 ± 0.043 (0.008-0.128)	10.25 ± 1.85 (9.1-11.98, 4)	4.04 (2.06-6.24; 3)	2.90 (1.6-4.37; 3)			
<i>Tessarabrachion oculatum</i> * (Euphausiacea)	10	0.055 ± 0.014 (0.033-0.076)	7.30 ± 0.90 (6.1 – 8.44, 10)	2.02 ± 0.61 (1.29 – 3.18, 10)	3.88±1.15 (2.49-6.33, 10)	0.35	0.49	-0.12
	20	0.062 ± 0.46 (0.022-0.086)	11.93 ± 2.00 (9.42 – 14.26, 6)	4.04 ± 1.06 (2.20 – 5.01, 6)	3.28 ± 1.62 (2.00-6.00, 6)			
<i>Nematobrachion flexipes</i> * (Euphausiacea)	10	0.089 ± 0.004 (0.07-0.10)	5.17 ± 0.69 (4.26-6.19; 7)	1.25 ± 0.82 (0.85 – 3.12, 7)	4.84±1.38 (1.98-6.16, 7)	0.62	0.76	-0.21
	20	0.045 ± 0.005 (0.045 – 0.112)	12.33 ± 2.25 (10.12-15.33, 6)	3.66 ± 0.93 (2.48-5.10, 6)	3.59±1.18 (1.96-4.83, 6)			
<i>Euphausia mucronata</i> * (Euphausiacea)	10	0.26 ± 0.096 (0.038 – 0.35)	4.84 ± 1.81 (2.21 – 7.87, 20)	1.39 ± 1.44 (0.61 – 2.18, 20)	4.07±2.64 (2.98-5.64)	0.46	.62	-0.23
	20	0.39 ± 0.10	9.22 ± 1.02 (7.2-10.39, 3)	3.30 ± 1.10 (2.48-3.50, 3)	2.93±1.09 (1.4-3.57, 3)			
<i>Dosidicus gigas</i> (Cephalopoda) (56, 57)	10		8.05 ± 0.51	1.86 ± 0.43	4.97	0.33	0.51	-0.20
	20		10.26 ± 0.75	3.50 ± 0.23	3.15			
	25		16.69 ± 0.88	4.65 ± 0.54	3.66			
<i>Euphausia gibboides</i> (58)	13		28.0 (dry mass)	2.25	Dry mass	0.32	0.44	-0.13
	23		43.1 (dry mass)	4.13	Dry mass			
<i>Euphausia hanseni</i> (59)	10		5.19	0.97	5.35	0.71	0.88	-0.17
	20		14.09	3.31	4.26			

Table S2: Coatal Marine Animals continued

Table S2: Coatal Marine Animals continued								
<i>Gadus morhua</i> (60)	18		3.84	6.42	0.60	0.71	0.34	0.37
	5		0.9	3.49	0.26			
	10		1.66	4.91	0.34			
	15		2.1	6.52	0.322			
	2		0.75	3.2	0.23			
<i>Sciaenops ocellatus</i> (61)	24		5.78	4.2	1.38	0.43	0.24	0.19
	30		8.09	5.05	1.60			
<i>Cyclopterus lumpus</i> (61)	10		2.5	7.13	0.35	0.42	0.26	0.16
	16		3.57	8.83	0.40			
<i>Chromis atripectoralis</i> (62)	35		7.56	4.88	1.55	1.10	0.75	0.35
	33		5.72	4.8	1.19			
	31		5.15	4.31	1.19			
	29		3.21	3.29	0.98			
<i>Salmo salar</i> (63)	18		6.37	7.56	0.84	0.33	0.19	0.14
	22		9.17	9.45	0.97			
<i>Morone saxatilis</i> (64)	20		2.27	5.5	0.41	0.80	0.38	0.42
	26		4.28	7.43	0.58			
<i>Carcharhinus plumbeus</i> (65)	24		4.38	8.13	0.54	0.37	0.31	0.06
	28		5.10	9.69	0.53			
	32		6.41	11.25	0.57			
<i>Centropristis striata</i> (66)	12		1.31	3.36	0.39	0.62	0.23	0.39
	17		1.89	3.99	0.47			
	22		2.72	4.73	0.58			
	24		3.15	5.07	0.62			
	27		3.93	5.62	0.70			
<i>Dicentrarchus labrax</i> (67)	10		1.13	4.46	0.25	0.64	0.16	0.48
	15		2.03	4.95	0.41			
	20		2.78	5.47	0.51			
<i>Gobiodon erythrospilus</i> (68)	29		8.11	5.48	1.48	0.72	0.63	0.10
	31		9.10	6.15	1.48			
	32		9.94	6.41	1.55			
	33		11.58	7.91	1.46			
<i>Gobiodon histrio</i> (68)	29		6.29	4.78	1.32	1.23	0.84	0.38
	31		9.31	5.63	1.65			
	32		9.62	5.89	1.63			
	33		12.04	8.04	1.50			
<i>Paralichthys dentatus</i> (69)	22		1.41	5.67	0.25	0.69	0.35	0.33
	30		2.88	8.19	0.35			
<i>Scyliorhinus canicular</i> (70)	7		0.80	7.66	0.10	0.64	0.23	0.41
	12		1.13	8.58	0.13			
	17		1.90	10.83	0.18			
<i>Squalus acanthias</i> (71)	10		0.79	3.33	0.24	0.61	0.21	0.40
	13		1.26	3.24	0.39			
	17		1.65	3.90	0.42			
	21		2.03	4.37	0.46			
	23		2.22	4.42	0.50			
<i>Callinectes sapidus</i> (72)	17		1.20	3.57	0.34	1.04	0.45	0.59
	23		1.80	4.20	0.43			
	28		4.20	7.98	0.53			

Table S2: Coastal Marine Animals continued								
<i>Carcinus maenas</i> (73)	7		0.73	2.85	0.26	0.61	0.40	0.21
	15		1.74	3.76	0.46			
	22		2.93	8.03	0.36			
<i>Chrysoblephus laticeps</i> (74)	16		2.42	4.62	0.52	0.66	0.42	0.24
	20		3.56	5.90	0.60			
	24		4.93	7.27	0.68			
<i>Octopus vulgaris</i> (75)	14		2.16	4.57	0.47	0.77	0.30	0.47
	16		2.80	5.10	0.55			
	18		3.45	5.63	0.61			
	20		4.20	6.14	0.68			
<i>Lates calcarifer</i> (76)	26		2.81	3.36	0.84	0.58	0.22	0.36
	36		5.81	4.41	1.32			
<i>Tautoglabrus adspersus</i> (77)	1		0.66	3.55	0.19	0.38	0.22	0.16
	8		0.99	4.49	0.22			

Supplementary References

55. Matthew A. Birk. (2022). matthewabirk/DVM_temp_sensitivity: v1.0 (v1.0). Zenodo. <https://doi.org/10.5281/zenodo.7015724>
56. Birk, M. A., McLean, E. L. & Seibel, B. A., Ocean acidification does not limit squid metabolism via blood oxygen supply. *J. Exp. Biol.* 221, (2018).
57. Trueblood, L. A. & Seibel, B. A., The jumbo squid, *Dosidicus gigas* (Ommastrephidae), living in oxygen minimum zones I: Oxygen consumption rates and critical oxygen partial pressures. *Deep-sea Res.* 195, 218-224 (2013).
58. Kiko, R., Hauss, H., Bucholz, F. & Melzner, F., Ammonium excretion and oxygen respiration of copepods and euphausiids exposed to oxygen minimum zone conditions. *Biogeosciences* 13, 2241-2255 (2016).
59. Tremblay, N., Hunerlage, K. & Werner, T., Hypoxia tolerance of 10 Euphausiid species in relation to vertical temperature and oxygen gradients. *Front. Physiol.* 24 (2020).
60. Chabot, D. & Claireaux, G., Environmental hypoxia as a metabolic constraint on fish: The case of Atlantic cod, *Gadus morhua*. *Mar. Poll. Bull.* 57, 287-294 (2008).
61. Ern, R., Norin, T., Gamperl, A. K. & Esbaugh, A. J., Oxygen dependence of upper thermal limits in fishes. *J Exp Biol* 219, 3376-3383 (2016).
62. Ern, R., Johansen, J. L., Rummer, J. L. & Esbaugh, A. J., Effects of hypoxia and ocean acidification on the upper thermal niche boundaries of coral reef fishes. *Biol Lett* 13, (2017).
63. Barnes, R. K., King, H. & Carter, C. G., Hypoxia tolerance and oxygen regulation in Atlantic salmon, *Salmo salar*, from a Tasmanian population. *Aquaculture* 318, 397-401 (2011).
64. Lapointe, D., Vogelbein, W. K., Fabrizio, M. C., Gauthier, D. T. & Brill, R. W., Temperature, hypoxia, and mycobacteriosis: effects on adult striped bass *Morone saxatilis* metabolic performance. *Dis Aquat Organ* 108, 113-127 (2014).
65. Crear, D. P. *et al.*, The impacts of warming and hypoxia on the performance of an obligate ram ventilator. *. Conservation Physiology*, 7, coz026 (2019). <https://doi.org/10.1093/conphys/coz026>
66. Slesinger, E. *et al.*, The effect of ocean warming on black sea bass (*Centropristis striata*) aerobic scope and hypoxia tolerance. *PLOS One* 14, e021839 (2019).
67. Claireaux, G. & Lagardere, J.-P., Influence of temperature, oxygen and salinity on the

- metabolism of the European sea bass. *J. Sea Res.* 42, 157-168 (1999).
68. Sorensen, C., Munday, P. L. & Nilsson, G. E., Aerobic vs anaerobic scope: sibling species of fish indicate that temperature dependence of hypoxia tolerance can predict future survival. *Global Change Biol.* 20, 724-729 (2014).
 69. Capossela, K. M., Brill, B. W., Fabrizio, M. C. & Bushnell, P. G., Metabolic and cardiorespiratory responses of summer flounder *Paralichthys dentatus* to hypoxia at two temperatures. *J. Fish Biol.* 81, 1043-1058 (2012).
 70. Butler, P. J. & Taylor, E. W., The effect of progressive hypoxia on respiration in the dogfish (*Scyliorhinus canicula*) at different seasonal temperatures. *J. Exp. Biol.* 63, 117-130 (1975).
 71. Andres, A., The effects of temperature and oxygen availability on aerobic performance in three coastal shark species; *Squalus acanthias*, *Carcharhinus limbatus*, and *Carcharhinus leucas*. Dissertation. University of South Florida (2022).
 72. Batterson, C. V. & Cameron, J. N., Characteristics of Resting Ventilation and Response to Hypoxia, Hypercapnia, and Emersion in the Blue Crab, *Callinectes sapidus* (Rathbun). *J. Exp. Zool.* 203, 403-418 (1978).
 73. Robertson, R. F. & Taylor, M. E. W., Specific dynamic action in the shore crab, *Carcinus maenas* (L.), in relation to acclimation temperature and to the onset of emersion response. *Physiol. Biochem. Zool.* 75, 350-359 (2002).
 74. Duncan, M. I., James, N. C., Potts, W. M. & Bates, A. E., Different drivers, common mechanism; the distribution of a reef fish is restricted by local -scale oxygen and temperature constraints on aerobic metabolism. *Conserv. Physiol.* 8, (2020).
 75. Valverde, J. C. & Garcia, B. G., Suitable dissolved oxygen levels for common octopus (*Octopus vulgaris* cuvier, 1797) at different weights and temperatures: analysis of respiratory behavior. *Aquaculture* 244, 303-314 (2004).
 76. Collins, G. M., Clark, T. D., Rummer, J. L. & Carton, A. G., Hypoxia tolerance is conserved across genetically distinct sub-populations of an iconic, tropical Australian teleost (*Lates calcarifer*). *Conserv Physiol* 1, cot029 (2013).
 77. Corkum, C. P. & Gamperl, A. K., Does the ability to metabolically downregulate alter the hypoxia tolerance of fishes?: A comparative study using Cunner (*T. adspersus*) and Greenland Cod (*G. ogac*). *J. Exp. Zool.* 311A, 231-239 (2009).