
Supplementary information

**Transcriptional programmes underlying
cellular identity and microbial
responsiveness in the intestinal epithelium**

In the format provided by the
authors and unedited

SUPPLEMENTAL TABLE 1: SUMMARY OF RECENT KEY FINDINGS

Gene	summary of insight	host organism(s)	references
NF- κ B	Characterizes the localization and expression patterns of TLRs on IECs of both small intestine and colon, and TLR5 induced gene program in Paneth cells and colonic IECs using organoids.	Mouse, mouse organoid	1
	Demonstrates TLR activation is needed to maintain intestinal homeostasis in DSS induced colitis.	Mouse	2
	IEC specific MyD88 knockout shows mice are more susceptible to DSS and have reduced ability to keep microbiota from colonizing colonic mucus. This is associated with reduced plgR, Muc2 and AMP production.	Mouse	3
	Revealed dynamic patterns of NF- κ B activity in vivo in response to microbiota colonization.	Zebrafish	4
	RelA regulates IEC homeostasis and protects against intestinal inflammation	Mouse	5
	Mechanisms through which NEMO prevents intestinal inflammation. Also describes phenotype of RelA, RelB and c-Rel triple intestinal specific knockout mice.	Mouse	6
	Showed TLR4 overexpression in mouse intestine leads to increased susceptibility to colitis.	Mouse	7
	TLR4 overexpression in mouse intestine shapes microbiota, which can transfer increased susceptibility to colitis to wild type mice.	Mouse	8
	TLR5 intestinal specific knockout leads mice to develop metabolic syndrome, increased susceptibility to colitis, and low-grade inflammation.	Mouse	9
	Microbial products activate ROS production in the intestine, which resulted in intestinal damage and early death.	Drosophila	10
	Hyperosmotic shock activates NF- κ B in IECs and mediates inflammatory cytokine production.	Human IECs	11
	NF- κ B is a mediator of intestinal ischemia/reperfusion injury induced cell death.	Rat	12
	Microbiota are important for IL-18 production in IECs, which then activates NF- κ B and its gene program.	Mouse	13
	Mice deficient for NLRP6 develop colitogenic microbiota.	Mouse	14
	NLRP12 serves as a checkpoint for the non-canonical NF- κ B pathway, and suppresses colonic	Mouse	15

	inflammation.		
IFN/IRF	p53 and p21 are ISGs which can be induced by IFN signaling, and promote apoptosis and senescence of IECs.	Mouse	16
	SNPs in IFN receptor genes are associated with IBD.	Human	17
	Describes signaling pathways leading to IRF3 activation.	Human proteins in cell-free assays, human embryonic kidney 293T cells, mouse embryonic fibroblasts, Human bone osteosarcoma (U-2 OS) cells.	18
	IRF3-mediated transcription of Tslp, an important gene for intestinal homeostasis.	Mouse	19
STATs	STAT6 functions independently of IL-4 as a regulator of chromatin compaction.	Mouse	20
	Constitutive activation of STAT6 in IECs effectively combats helminth infections.	Mouse	21
	Long-term activation of STAT3 during chronic inflammation promotes colon tumor formation.	Mouse	22
	STAT3 serves a protective role in IECs against injury and inflammation, promoting mucosal healing.	Mouse	23
	STAT3 activates gene networks that restrict microbial translocation through the intestinal epithelium.	Mouse	24
	Segmented filamentous bacteria (SFB) induce Th17 cell production of IL-22 which signals to IECs.	Mouse	25
	In the lung and gastric epithelia, STAT3 binds the promoter of RegIIIg and drives transcription of the gene.	Mouse	26,27
ISCs	Modelling asymmetric division of mouse ISCs.	Mouse, in silico	28
	Modelling symmetric ISC growth and lineage tracing.	Mice	29
	Optimizing the conditions for mouse, human epithelial organoids and examining their proliferative potential.	Human and mouse organoids	30
	Chromatin accessibility and histone marks in LRG5+ stem cells.	Mouse	31
Sox2	Sox2 expression is required for proper patterning along the anterior gut.		32
	Sox2 can direct anteriorization of the gut, suggests	Mouse	33

	that interplay with Cdx2 important for proper patterning in primitive gut.		
Cdx2	Cdx2 is required for establishing intestinal identity.	Mouse	34
	Cdx2 is required for proper binding of HNF4A and maintains permissive chromatin state.	Mouse	35
	Bile acids regulate Cdx2.	Rat	36
	Stimulation of the intestinal Cdx2 homeobox gene by butyrate in colon cancer cells.	Human adenocarcinoma cell line	37
	Characterizes Cdx2 expression in the posterior gut at E8.5.	Mouse	38
	Cdx2 is expressed in small intestine and colon of adult mice.	Mouse	39
	Cdx2 regulates chromatin remodeling through Brg1-SWI/SNF complex recruitment.	Mouse, human colon cancer (C2bbe1) cells and human embryonic kidney 293 cell lines	40
	Cdx2's role in body segmentation, patterning, and development of intestinal polyps.	Fly	41
	Intestine-specific KO of Cdx2 leads to a loss of intestinal identity.	Mouse	42
	Cdx2 expression in parietal cells of the gastric mucosa develop into intestinal metaplasia.	Mouse	43
	Cdx2 expression driven by Foxa3 in the gastric mucosa led to intestinal neoplasia.	Mouse	44
	Expression of Cdx2 at transitional basal cells leads to Barrett's esophagus-like phenotypes.	Mouse	45
	Transgenic overexpression of Cdx1b using Krt5p promoter induces intestinal metaplasia in the squamous epithelium of the zebrafish esophagus.	Zebrafish	46
	Repressive interaction between Cdx2 and Sox2 at stomach-intestine border.	Mouse	47
	Cdx2 has key, differentiation-specific occupancy of the genome, co-occupies some loci with GATA6 and HNF4A.	Mouse	48
	Cdx2 is vital for specification of intestinal cell types from gut stem cells during development.	Mouse, mouse organoids	49
	Cdx2, in combination with GATA4 and HNF4A, control IEC homeostasis.	Mouse	50
	Cdx2 homologs are important in early intestinal epithelial specification in vertebrates.	Zebrafish	51

Tcf-4	Tcf4 is required for establishment of ISC niche in crypt.	Mouse	52
Sentinel Goblet Cells	Sentinel goblet cells guard the colonic crypt by triggering Nlrp6-dependent Muc2 secretion.	Mouse	53
Secretory Cells	Atoh1/Math1 is required for terminal differentiation of secretory cell.	Mouse	54
	Intestinal Atoh1/Math1 is involved in IEC homeostasis.	Mouse	55
H3K27ac	Differentiating active vs poised enhancers and effects on developmental programs.	Mouse	56
	H3K27ac, a marker for active enhancers, is regulated by microbiota colonization.	Mouse	57
Mediator, Enhancers	Mediator and cohesin connect enhancers and core promoters via DNA looping contributing to maintenance of embryonic stem cell state.	Mouse embryonic stem cells	58
	Defines the enhancer and promoter landscape in IBD.	Human	59
	Sub-classification of human IBDs based on transcriptional profiles.	Human	60
Polycomb, CRRs	Describes the duality of CRRs as enhancers and repressors in general.	Drosophila	61
Polycomb	PRC2 target genes and bivalent domains in the adult somatic intestinal cells.	Mouse	62
	PRC2 contributes to stemness in disease states via Wnt signaling.	Mouse organoids	63
	PRC2 prevents hypomethylated developmental enhancers from becoming reactivated.	Mouse	64
TF Binding	ChIP-seq mapping of binding sites of TFs known to play roles in ES cell development.	Mouse ES cells	65
angptl4, CRRs	Identifies cis-regulatory region for angptl4 expression, under microbial control.	Zebrafish	66
Chromatin accessibility	Microbially associated transcriptional changes are not accompanied by changes in established chromatin landscape along the intestine.	Mouse	67, 57
	Gut microbiome changes gene expression of colonocytes in vitro and for some genes, also the accessible chromatin surrounding TSS.	Human colonocyte cell line	68
	Role of chromatin accessibility during secretory to ISC dedifferentiation.	Mouse	69
Transcripts/Epi genetics/Metabolome	Diurnal oscillations of gut microbiota impact gene expression, histone modifications, and serum metabolites.	Mouse	70
Regionality	Regional gene expression in adult zebrafish gut.	Zebrafish	71

	Conserved regional signature genes and cis-regulatory regions.	Zebrafish, Mouse	72
scRNA-seq	scRNA-seq of IECs.	Mouse	73
	scRNA-seq of enterocytes along crypt-villus axis.	Mouse	74
Gastric identity	Loss of stomach accompanied by loss of pepsinogen and proton pump genes.	Fish	75
	Barx1, a stomach marker, is still expressed in the stomach-less zebrafish.	Zebrafish	76
Intestinal development / Morphogen gradients	Zebrafish intestinal development.	Zebrafish	77
	Intestinal stem cell crypt develops via morphogenic field.	Chicken, Mouse	78
SCFAs	SCFA along the length of the intestine	Pig, Human	79
	Microbially produced SCFA important to regulate inflammation in the intestine.	Mouse	80
	HDACs and SCFAs are drivers of histone crotonylation.	Mouse	81
	Supplementation of SCFAs in germ-free mice can recapitulate histone modification patterns associated with microbial colonization in the proximal colon and liver.	Mouse	82
SCFAs, HDACs	Microbiota inhibits histone de-acetylation via SCFAs.	Human colorectal adenocarcinoma cell line (HT-29)	83
	Butyrate inhibition of HDACs can change depending on metabolic state (e.g., Warburg effect).	Mouse	84
	n-butyrate controls HDAC activity in intestinal macrophages.	Mouse	85
HDACs	HDAC1 and HDAC2 restrain the intestinal inflammatory response by regulating intestinal epithelial cell differentiation.	Mouse	86,87
	HDAC3 is required for Paneth cells in the presence of microbiota.	Mouse	88
DNA methylation	DNA methylation is a determinant of stem cell maintenance and differentiation in small intestine.	Mouse	89
	Differences in DNA methylation and H3K27me3 in embryonic and adult ISCs.	Mouse	90
	Suggests that the microbiota is capable of regulating the postnatal, DNA methylation	Mouse	91

	landscape of ISCs.		
	Describes the effects of microbiota on DNA methylation of IECs.	Mouse	92
	DNA methylation is crucial for intestinal homeostasis.	Zebrafish	93
	DNA methylation regulates intestinal development and establishment of crypts.	Mouse, mouse organoids	94
	Differential DNA methylation associated with IBD.	Human	95
	Dnmt1 and Suv29h1 are required for intestinal terminal differentiation.	Zebrafish	96
	Differential DNA methylation in intestinal fibroblasts from Crohn's disease patients' occurred mostly within introns & intergenic regions, with a few genes showing inversely correlated methylation and transcription.	Human fibroblasts	97
	DNA methylation differences in ISCs vs differentiated IECs.	Mouse	98
	DNA methylation patterns differ across anteroposterior regions of the intestine.	Human patients and organoids	99,100
	DNA methylation examined directly in IECs by genome-wide bisulfite-sequencing, revealed roles for DNA methylation in different aspects of IEC identity.	Human, mouse	58,59
Anti-TNF therapy	Infliximab infusions help manage disrupted barrier function in Crohn's disease patients.	Humans	101
Microbiota and cytokines	Microbiota composition and function impact cytokine expression.	Humans	102
ELF3	Identification of Elf3 as a TF expressed in developing and mature intestinal tract.	Mouse	103,104
	Elf3 is essential for life, intestinal differentiation.	Mouse	105
	Elf3 is the upstream regulator of TGF-betaRII receptor.	Mouse	106
GATA Factors	GATA4 most highly expressed in the duodenum and jejunum, regulates marker of intestinal differentiation, LPH.	Mouse	107
	Gut development demonstrates specific spatio-temporal patterns of TF expression.	Mouse	108
	Gata4 is essential for the maintenance of jejunal-ileal identities in the adult mouse small intestine.	Mouse	109
	GATA4 is essential for jejunal function in mice.	Mouse	110
	GATA4 is sufficient to establish jejunal versus ileal identity in the small intestine.	Mouse	111

	GATA6 is required for differentiation of colonic epithelium.	Mouse	112
	GATA6 is required for normal morphology and enterocyte gene expression in the ileum but may function redundantly with GATA4 in the jejunum.	Mouse	113
HNF4 (NR2A1, NR2A2, NR2A3)	HNF4 TFs help shape chromatin landscape.	Mouse	114
	Mapping TSS and organ expression of HNF4 gene suggests complex transcriptional regulation.	Mouse	115
	Two alternative promoters for HNF4A produce products with different trans-activation properties.	Human embryonic kidney 293T cells	116
	HNF4A protects against DSS induced colitis.	Mouse	117
	In the liver, knockdown of HNF4A expression induces a proinflammatory feedback circuit that continues to repress HNF4A expression and upregulate proinflammatory genes.	Mouse	118
	HNF4A activity impacts Wnt signaling and cell lineage commitment in the intestine.	Human hepatocyte cell line	119
	Structure, mapping, expression and transcriptional activity of mouse HNF4G gene.	Mouse	120
	HNF4A knockouts fail to complete gastrulation and die early during embryogenesis.	Mouse	121
	HNF4 is required for gastrulation via expression in the visceral endoderm.	Mouse	122
	HNF4A regulates liver development and morphogenesis.	Mouse	123
	HNF4A required for hepatocyte differentiation.	Mouse	124
	HNF4A required for colonic crypt formation.	Mouse	125
	HNF4A protects against DSS-induced colitis.	Mouse	126
	HNF4A may interact with DNA repair machinery.	Human colorectal cancer cells	127
	HNF4A regulates Cldn15 expression and thereby mediates ion transport.	Mouse	128
	SNPs at HNF4A locus associated with UC and CD.	Human	17,129,130
	Differentially active enhancers in IBD are enriched for HNF4A binding sites.	Human	131,132
	HNF4A protects against ROS and promotes gut neoplasia.	Mouse	133
	HNF4A triggers morphological changes in the intestinal epithelium.	Mouse, human pancreatic	134

		adenocarcinoma cell line (MIA PaCa-2), and mouse embryonic fibroblast cell line (NIH-3T3)	
	Human gut microbiota can modify linoleic acid into conjugated linoleic acids.	Bacteria from human colon	135
	SNPs at HNF4G locus associated with UC.	Human	136
	Hnf4g knockout mice have lowered energy expenditure and physical activity during nighttime, and increased weight.	Mouse	137
	HNF4A and HNF4G can heterodimerize.	HepG2 cells	138
	HNF4G has an intestinal expression border at the pyloric boundary.	Mouse	139
	HNF4A has 12 different isoforms with differing N- and C-termini, 11 of which are expressed in the small intestine.	Human	140
	HNF4G is upregulated in intestinal-metaplasia of the stomach.	Human	141
	HNF4G drives enterocyte differentiation.	Mouse organoids	142
	Hnf4g is required for proper balance of cell lineages in the intestinal epithelium, with mutants displaying increased numbers of GLP-1 expressing enteroendocrine cells.	Mouse	143
	Feed-forward loop between HNF4 and SMAD factors contribute to maintenance and stability of enterocyte identity.	Mouse	144
	HNF4 family members bind to long chain fatty acids such as palmitic acid, palmitoyl-CoA, and linoleic acid	Rat, Human, Mouse, Drosophila	145-149
	HNF4A is important for small intestinal absorption of dietary lipids.	Mouse	150
RXR (NR2B1, NR2B2, NR2B3)	Identified vitamin A metabolite, 9-cis retinoic acid as the ligand for RXR	Human	151
	The RXR family of nuclear receptors arose shortly after HNF4-like nuclear receptors evolved.	N/A	152
PPAR (NR1C1, NR1C2, NR1C3)	PPARB/D is highly expressed in enterocytes and regulates expression of fatty acid binding protein in enterocyte cultures.	Mouse	153
	Induction of PPARG activity by chemical ligands,	Mouse	154,155

	rosiglitazone and troglitazone, suppresses colonic inflammation associated with DSS administration in mice.		
	PPARs heterodimerize with RXR and induce the expression of target genes by binding peroxisome proliferator responsive elements (PPREs) throughout the genome.	Human	156
	The SCFA butyrate and PPARG agonists stimulate PPARG dependent expression of Angptl4.	Human colon adenocarcinoma cell line	157,158
	Butyrate-PPARG signaling is important in limiting the availability of luminal nitrate.	Mouse	159
	PPARG regulates beta-defensin in the colonic epithelium, and Pparg deficiency resulted in impaired microbicidal capacity of the colonic mucosa.	Mouse	160
	Genetic variants at human PPARG are associated with increased risk for both UC and CD.	Human	161-163
	UC is associated with decreased PPARG expression.	Human	164,165
	Microbial-TLR4 signaling induces PPARG expression in IECs.	Human Caco2 cell line, primary mouse colonocytes	164
	Mice lacking TLR4, specifically in the intestinal epithelium showed a significant downregulation of PPARG target genes	Mouse	166
	PPARG suppresses TLR signaling by exporting a NF- κ B subunit, RELA, out of the nucleus and inhibiting transcription of proinflammatory genes	Human Caco2 cell line	167
	PPARA is expressed in the intestine.	Mouse	168
	Ppara transcript is reduced by microbiota colonization.	Mouse	169
	Ppara regulates IL-22 production by innate immune cells in response to intestinal microbiota, suppressing pro-inflammatory signaling and promoting expression of antimicrobial peptides and IL10.	Mouse	170
	Pparg activity is dysregulated in the small intestine of mice fed a high-fat diet, and treatment with the PPARG agonist rosiglitazone improved many of the changes in gene expression associated with high-fat diet feeding.	Mouse	171
FXR (NR1H4)	Bile acids serve as an activating ligand for the nuclear receptor FXR.	Mouse, human	172

	Intestinal microbiota modify primary bile acids through bile salt hydrolases (BSH) and other enzymes.	Human iPSC, Mouse	173,174
	Microbial modification of bile acids regulates systemic metabolic processes like fat storage.	Mouse	174-178
	Treatment with caffeic acid phenethyl ester (CAPE), an inhibitor of bacterial BSH, selectively suppressed intestinal FXR signaling by increasing levels of the bile acid, tauro-B-muricholic acid, ultimately leading to decreased gluconeogenesis in the liver.	Mouse	179
	Increasing BSH-retaining species in the microbiota (i.e., Lactobacilli and Bifidobacteria) impacted bile acid metabolism in the intestine and liver.	Mouse	177
	FXR knockouts demonstrating intestinal FXR is required for HFD induced obesity	Mouse	178,180
	Treatment with an FXR antagonist demonstrating intestinal FXR is required for HFD induced obesity.	Mouse	180
	FXR modulates proinflammatory responses by forming a repressor complex with NCOR on the NF- κ B response element on the IL-1B promoter.	Mouse	181
	Loss of FXR function in mice increases their susceptibility to DSS and TNBS induced colitis.	Mouse	182
LXR (NR1H3, NR1H2)	LXR ligands are sterols such as cholesterol and oxysterols.	Human	183
	PPARA is a target of LXR, and they work together in the small intestine to regulate cholesterol homeostasis.	Mouse	184
	Suppresses inflammation in macrophages by inhibiting NF- κ B; microbial ligands repress expression of LXR target genes in immune cells.	Mouse macrophage (RAW 264.7) cell line, mouse peritoneal macrophages	185,186
	LXR deficient mice were more susceptible to DSS-induced colitis.	Mouse	187
	LXR homologs were significantly reduced in the inflamed regions of biopsies from CD and UC patients.	Human	187
Glucocorticoid receptor (GR, NR3C1)	Glucocorticoids suppress NF- κ B and AP-1 activity and increased GR expression is associated with decreased NF- κ B activity.	Rat, human PBMC	188,189
	Glucocorticoids reduce microbially induced TNF inflammation.	Mouse	190
	Genetic variants at human GR, NR3C1 are associated with risk for CD.	Human	191

	Some genetic variants at NR3C1 are associated with hormone resistant therapies in UC and CD.	Human	192,193
Mineralocorticoid receptor (MR, NR3C2)	MR is expressed in the duodenum and colon.	Mouse	194
	MR in the colon is important for sodium balance and blood pressure.	Mouse	195
Vitamin D Receptor (VDR, NR1I1)	VDR knockout mice spontaneously develop colitis.	Mouse	196
	VDR knockout mice have modified microbial communities, greater susceptibility to DSS-induced colitis, and defective barrier and Paneth cell function.	Mouse	197-199
	VDR overexpression improves several genetic models of colitis in mice.	Mouse	200,201
	Low levels of vitamin D are associated with IBD.	Human	202
Constitutive androstane receptor (CAR, NR1I3)	Differences in CAR expression between GF and CV mice.	Mouse	203,204
	CAR mutant mice were more susceptible to DSS-induced colitis.	Mouse	205
	CAR knockout mice spontaneously develop insulin sensitivity, CAR agonist prevented obesity.	Mouse	206
	Mutations in CAR alter intestinal microbiota community and function through bile acid metabolism.	Mouse	207
	CAR was decreased in biopsies from CD and UC patients.	Human	205
Pregnane X Receptor (PXR, NR1I2)	PXR alters intestinal microbiota community and function through bile acid metabolism	Mouse	207
	Indole metabolites induced expression of PXR target genes in jejunal enterocytes.	Mouse	208
	PXR mutants have impaired small intestinal barrier function.	Mouse	208
	PXR agonist attenuated barrier defects observed in DSS-treated animals.	Mouse	209,210
	Decreased PXR expression observed in biopsies from actively inflamed CD tissue.	Human	211
	Other environmental chemicals such as drugs and spices impact PXR activity.	Mouse, human colon cell line	212-214

		(LS180), human primary hepatocytes, human hepatic stem cell line (HepaRG)	
HIF	Consumption of SCFA by the colonic epithelia decreases oxygen concentrations colonocytes, activating HIF.	Mouse	215
	HIF activity results in transcriptional activation of Defensin beta, an antimicrobial peptide, MUC3 and ITF, thereby maintaining epithelial barrier function	Mouse	216-218
	HIF induces transcription of the antimicrobial peptide LL77 and protects against <i>Candida albicans</i> infection	Mouse	219
Notch	Microbial repression of RBPJ, an effector transcription factor of Notch signaling is dependent on MyD88.	Zebrafish	220
	Mice lacking RBPJ in their IECs develop a microbiota-dependent and Th17 cell-mediated chronic colitis.	Mouse	221
	Hes1 deficiency in IECs causes an intestinal inflammatory phenotype and dysbiosis.	Mouse	222
	Microbiota colonization results in transient repression of transcripts involved in Notch signaling and a rapid expansion of goblet cells.	Mouse	223,224
	Microbiota colonization results in reduced EEC populations	Mouse	225
	Microbiota colonization induces an expansion of mature goblet cell populations.	Rat	226
	Microbiota colonization promotes goblet cell differentiation.	Zebrafish	227
	NFAT5 suppresses the metabolic regulator mTORC1 and suppresses Notch signaling, which results in an expansion of goblet cells and MUC2 expression.	Human embryonic kidney 293 cells	228,229
WNT	Both GF and antibiotic treated mice have slowed IEC proliferation rates compared to SPF mice, though this was not associated with transcriptional alterations in Wnt pathway target genes that promote proliferation, but rather phosphorylation of ERK and STAT3 activation.	Mouse	230
	GF zebrafish have reduced IEC cytoplasmic beta catenin compared to CV or monoassociated fish,	Zebrafish	231

	correlating with increased proliferation of IECs in CV or monoassociated fish.		
	Describes how Wnt in the intestine can drive both stem and Paneth cell gene programs.	Mouse, human	232
BMP	Genetic profiling of mesenchymal and epithelial compartments of the perinatal mouse intestine.	Mouse	233
	Mutations in BMP signaling pathway members in familial juvenile polyposis.	Humans	234,235
	Mutations in BMP signaling pathway is implicated in colorectal cancer.	Humans	236
	Bmpr1a impacts the proliferation and differentiation of secretory lineage cells.	Mouse	237
	BMP levels regulate enteroendocrine cell EEC hormone expression .	Mouse	238
	SMAD4/HNF4A feedforward loop promotes enterocyte fate.	Mouse	144
	TGF-B1 signaling inhibits of NF-κB activity and IL-6 expression in response to the commensal <i>B. vulgatis</i> .	Rat	239
	TGF-B1/SMAD signaling protects against the colitogenic properties of <i>Enterococcus faecalis</i> .	Mouse	240
AHR	Dioxins lead to AHR activation.	Mammals	241
	Microbial tryptophan metabolites can activate AHR.	Human and Mouse	242-247
	ARNT can form heterodimers with HIF.	Mouse	248
	HIFs roles in intestinal sensitivity to microbes.	Mouse	249
	HIFs roles in synthesis of antimicrobial peptides.	T84 intestinal epithelial cells, Human, Mouse	216-218
	Role of AHR in innate immunity and intestinal barrier integrity.	Review	250
	AHR protects against DSS-induced colitis.	Mouse	251,252
	AHR regulates proliferation and stem cell maintenance and is expressed in Paneth cells.	Mouse	253,254
	AHR loss in colonic stem cells leads to malignancies.	Mouse	255
	Tryptophan metabolites activate IL-10 expression in the colon.	Mouse	256
	AHR is a therapeutic target for IBD.	Human	257-259
NFIL3	Nfil3 is a transcription factor involved in the response of the intestinal epithelium to circadian rhythms and the microbiota.	Mouse	260

ASCL2	Important marker of ISCs; target of Wnt signaling in the intestinal crypt; deletion leads to loss of stem cells; overexpression leads to crypt hyperplasia; part of a direct autoactivation loop that translates the level of Wnt signaling into a “ON/OFF” output and determines stem cell state.	Mouse, mouse organoid	261,262
Reviews			
Stomach loss in fishes			263
ISCs	ISCs and their niche		264
	Regulation of stem cells in intestinal homeostasis and cancer		265
	ISC renewal		266
	Stem cells and intestinal injury		267
Paneth cells			268
Tuft cells			269
Mucins			270
Metagenomics			271
Epigenetics			272
Pioneer transcription factors			273
ETS factors			274
Transcriptional enhancers			275
Transcriptional regulation			276
Histone modifications and modifying enzymes			277-279
Histone acylation			280
Histone tail acetyl- and acylations			281
DNA methylation			282
DNA methylation, miRs in IBD			283

TNF and IBD			284
Anti-TNF therapy in IBD			285,286
Microbial metabolites (SCFAs, etc.)			287
Dietary regulation of host defense peptides			288
Xenobiotics and IECs			289
Inflammasome and pro-inflammatory cytokines			290
Cell death and anoikis			291
NF- κ B family			292
TLRs			293
MyD88			294
ROS production			295
NLRs			296
NLRs in IBD			297
IFN			298
IRFs			299
JAK/STAT			300
PPAR family			301,302
FXR and bile acids			303
RXR			304
VDR			305
AHR			306

BIBLIOGRAPHY

1. Price, A.E., *et al.* A Map of Toll-like Receptor Expression in the Intestinal Epithelium Reveals Distinct Spatial, Cell Type-Specific, and Temporal Patterns. *Immunity* **49**, 560-575 e566 (2018).
2. Rakoff-Nahoum, S., Paglino, J., Eslami-Varzaneh, F., Edberg, S. & Medzhitov, R. Recognition of commensal microflora by toll-like receptors is required for intestinal homeostasis. *Cell* **118**, 229-241 (2004).
3. Frantz, A.L., *et al.* Targeted deletion of MyD88 in intestinal epithelial cells results in compromised antibacterial immunity associated with downregulation of polymeric immunoglobulin receptor, mucin-2, and antibacterial peptides. *Mucosal Immunol* **5**, 501-512 (2012).
4. Kanther, M., *et al.* Microbial colonization induces dynamic temporal and spatial patterns of NF-kappaB activation in the zebrafish digestive tract. *Gastroenterology* **141**, 197-207 (2011).
5. Steinbrecher, K.A., Harmel-Laws, E., Sitcheran, R. & Baldwin, A.S. Loss of epithelial RelA results in deregulated intestinal proliferative/apoptotic homeostasis and susceptibility to inflammation. *J Immunol* **180**, 2588-2599 (2008).
6. Vlantis, K., *et al.* NEMO Prevents RIP Kinase 1-Mediated Epithelial Cell Death and Chronic Intestinal Inflammation by NF-kappaB-Dependent and -Independent Functions. *Immunity* **44**, 553-567 (2016).
7. Fukata, M., *et al.* Constitutive activation of epithelial TLR4 augments inflammatory responses to mucosal injury and drives colitis-associated tumorigenesis. *Inflamm Bowel Dis* **17**, 1464-1473 (2011).
8. Dheer, R., *et al.* Intestinal Epithelial Toll-Like Receptor 4 Signaling Affects Epithelial Function and Colonic Microbiota and Promotes a Risk for Transmissible Colitis. *Infect Immun* **84**, 798-810 (2016).
9. Chassaing, B., Ley, R.E. & Gewirtz, A.T. Intestinal epithelial cell toll-like receptor 5 regulates the intestinal microbiota to prevent low-grade inflammation and metabolic syndrome in mice. *Gastroenterology* **147**, 1363-1377.e1317 (2014).
10. Iatsenko, I., Boquete, J.P. & Lemaitre, B. Microbiota-Derived Lactate Activates Production of Reactive Oxygen Species by the Intestinal NADPH Oxidase Nox and Shortens Drosophila Lifespan. *Immunity* **49**, 929-942 e925 (2018).
11. Nemeth, Z.H., Deitch, E.A., Szabo, C. & Hasko, G. Hyperosmotic stress induces nuclear factor-kappaB activation and interleukin-8 production in human intestinal epithelial cells. *Am J Pathol* **161**, 987-996 (2002).
12. Giakoustidis, A.E., *et al.* Inhibition of intestinal ischemia/reperfusion induced apoptosis and necrosis via down-regulation of the NF-kB, c-Jun and caspase-3 expression by epigallocatechin-3-gallate administration. *Free Radic Res* **42**, 180-188 (2008).
13. Levy, M., *et al.* Microbiota-Modulated Metabolites Shape the Intestinal Microenvironment by Regulating NLRP6 Inflammasome Signaling. *Cell* **163**, 1428-1443 (2015).
14. Elinav, E., *et al.* NLRP6 inflammasome regulates colonic microbial ecology and risk for colitis. *Cell* **145**, 745-757 (2011).
15. Allen, I.C., *et al.* NLRP12 suppresses colon inflammation and tumorigenesis through the negative regulation of noncanonical NF-kappaB signaling. *Immunity* **36**, 742-754 (2012).
16. Katlinskaya, Y.V., *et al.* Type I Interferons Control Proliferation and Function of the Intestinal Epithelium. *Mol Cell Biol* **36**, 1124-1135 (2016).
17. Jostins, L., *et al.* Host-microbe interactions have shaped the genetic architecture of inflammatory bowel disease. *Nature* **491**, 119-124 (2012).

18. Liu, S., *et al.* Phosphorylation of innate immune adaptor proteins MAVS, STING, and TRIF induces IRF3 activation. *Science* **347**, aaa2630 (2015).
19. Negishi, H., *et al.* Essential contribution of IRF3 to intestinal homeostasis and microbiota-mediated Tslp gene induction. *Proc Natl Acad Sci U S A* **109**, 21016-21021 (2012).
20. De Oliveira, T., *et al.* Loss of Stat6 affects chromatin condensation in intestinal epithelial cells causing diverse outcome in murine models of inflammation-associated and sporadic colon carcinogenesis. *Oncogene* **38**, 1787-1801 (2019).
21. Schubart, C., *et al.* Selective expression of constitutively activated STAT6 in intestinal epithelial cells promotes differentiation of secretory cells and protection against helminths. *Mucosal Immunol* **12**, 413-424 (2019).
22. Bollrath, J., *et al.* gp130-mediated Stat3 activation in enterocytes regulates cell survival and cell-cycle progression during colitis-associated tumorigenesis. *Cancer Cell* **15**, 91-102 (2009).
23. Pickert, G., *et al.* STAT3 links IL-22 signaling in intestinal epithelial cells to mucosal wound healing. *J Exp Med* **206**, 1465-1472 (2009).
24. Sugimoto, K., *et al.* IL-22 ameliorates intestinal inflammation in a mouse model of ulcerative colitis. *J Clin Invest* **118**, 534-544 (2008).
25. Ivanov, I., *et al.* Induction of intestinal Th17 cells by segmented filamentous bacteria. *Cell* **139**, 485-498 (2009).
26. Choi, S.M., *et al.* Innate Stat3-mediated induction of the antimicrobial protein Reg3gamma is required for host defense against MRSA pneumonia. *J Exp Med* **210**, 551-561 (2013).
27. Lee, K.S., *et al.* Helicobacter pylori CagA triggers expression of the bactericidal lectin REG3gamma via gastric STAT3 activation. *PLoS One* **7**, e30786 (2012).
28. McHale, P.T. & Lander, A.D. The protective role of symmetric stem cell division on the accumulation of heritable damage. *PLoS Comput Biol* **10**, e1003802 (2014).
29. Snippert, H.J., *et al.* Intestinal crypt homeostasis results from neutral competition between symmetrically dividing Lgr5 stem cells. *Cell* **143**, 134-144 (2010).
30. Sato, T., *et al.* Long-term expansion of epithelial organoids from human colon, adenoma, adenocarcinoma, and Barrett's epithelium. *Gastroenterology* **141**, 1762-1772 (2011).
31. Kim, T.H., *et al.* Broadly permissive intestinal chromatin underlies lateral inhibition and cell plasticity. *Nature* **506**, 511-515 (2014).
32. Que, J., *et al.* Multiple dose-dependent roles for Sox2 in the patterning and differentiation of anterior foregut endoderm. *Development* **134**, 2521-2531 (2007).
33. Raghoebir, L., *et al.* SOX2 redirects the developmental fate of the intestinal epithelium toward a premature gastric phenotype. *J Mol Cell Biol* **4**, 377-385 (2012).
34. Gao, N., White, P. & Kaestner, K.H. Establishment of intestinal identity and epithelial-mesenchymal signaling by Cdx2. *Dev Cell* **16**, 588-599 (2009).
35. Verzi, M.P., Shin, H., San Roman, A.K., Liu, X.S. & Shivdasani, R.A. Intestinal master transcription factor CDX2 controls chromatin access for partner transcription factor binding. *Mol Cell Biol* **33**, 281-292 (2013).
36. Kazumori, H., Ishihara, S., Rumi, M.A., Kadowaki, Y. & Kinoshita, Y. Bile acids directly augment caudal related homeobox gene Cdx2 expression in oesophageal keratinocytes in Barrett's epithelium. *Gut* **55**, 16-25 (2006).
37. Domon-Dell, C., *et al.* Stimulation of the intestinal Cdx2 homeobox gene by butyrate in colon cancer cells. *Gut* **50**, 525-529 (2002).
38. Beck, F., Erler, T., Russell, A. & James, R. Expression of Cdx-2 in the mouse embryo and placenta: possible role in patterning of the extra-embryonic membranes. *Dev Dyn* **204**, 219-227 (1995).

39. James, R., Erler, T. & Kazenwadel, J. Structure of the murine homeobox gene *cdx-2*. Expression in embryonic and adult intestinal epithelium. *J Biol Chem* **269**, 15229-15237 (1994).
40. Nguyen, T.T., *et al.* Cdx2 Regulates Gene Expression through Recruitment of Brg1-associated Switch-Sucrose Non-fermentable (SWI-SNF) Chromatin Remodeling Activity. *J Biol Chem* **292**, 3389-3399 (2017).
41. Chawengsaksophak, K., James, R., Hammond, V.E., Kontgen, F. & Beck, F. Homeosis and intestinal tumours in Cdx2 mutant mice. *Nature* **386**, 84-87 (1997).
42. Grainger, S., Savory, J.G. & Lohnes, D. Cdx2 regulates patterning of the intestinal epithelium. *Dev Biol* **339**, 155-165 (2010).
43. Mutoh, H., *et al.* Conversion of gastric mucosa to intestinal metaplasia in Cdx2-expressing transgenic mice. *Biochem Biophys Res Commun* **294**, 470-479 (2002).
44. Silberg, D.G., *et al.* Cdx2 ectopic expression induces gastric intestinal metaplasia in transgenic mice. *Gastroenterology* **122**, 689-696 (2002).
45. Jiang, M., *et al.* Transitional basal cells at the squamous-columnar junction generate Barrett's oesophagus. *Nature* **550**, 529-533 (2017).
46. Hu, B., *et al.* Transgenic overexpression of *cdx1b* induces metaplastic changes of gene expression in zebrafish esophageal squamous epithelium. *Zebrafish* **10**, 218-227 (2013).
47. Sherwood, R.I., Chen, T.Y. & Melton, D.A. Transcriptional dynamics of endodermal organ formation. *Dev Dyn* **238**, 29-42 (2009).
48. Verzi, M.P., *et al.* Differentiation-specific histone modifications reveal dynamic chromatin interactions and partners for the intestinal transcription factor CDX2. *Dev Cell* **19**, 713-726 (2010).
49. Stringer, E.J., *et al.* Cdx2 determines the fate of postnatal intestinal endoderm. *Development* **139**, 465-474 (2012).
50. San Roman, A.K., Aronson, B.E., Krasinski, S.D., Shivdasani, R.A. & Verzi, M.P. Transcription factors GATA4 and HNF4A control distinct aspects of intestinal homeostasis in conjunction with transcription factor CDX2. *J Biol Chem* **290**, 1850-1860 (2015).
51. Flores, M.V., *et al.* Intestinal differentiation in zebrafish requires Cdx1b, a functional equivalent of mammalian Cdx2. *Gastroenterology* **135**, 1665-1675 (2008).
52. Korinek, V., *et al.* Depletion of epithelial stem-cell compartments in the small intestine of mice lacking Tcf-4. *Nat Genet* **19**, 379-383 (1998).
53. Birchenough, G.M., Nystrom, E.E., Johansson, M.E. & Hansson, G.C. A sentinel goblet cell guards the colonic crypt by triggering Nlrp6-dependent Muc2 secretion. *Science* **352**, 1535-1542 (2016).
54. Yang, Q., Bermingham, N.A., Finegold, M.J. & Zoghbi, H.Y. Requirement of Math1 for secretory cell lineage commitment in the mouse intestine. *Science* **294**, 2155-2158 (2001).
55. Shroyer, N.F., *et al.* Intestine-specific ablation of mouse atonal homolog 1 (Math1) reveals a role in cellular homeostasis. *Gastroenterology* **132**, 2478-2488 (2007).
56. Creighton, M.P., *et al.* Histone H3K27ac separates active from poised enhancers and predicts developmental state. *Proc Natl Acad Sci U S A* **107**, 21931-21936 (2010).
57. Davison, J.M., *et al.* Microbiota regulate intestinal epithelial gene expression by suppressing the transcription factor Hepatocyte nuclear factor 4 alpha. *Genome Res* **27**, 1195-1206 (2017).
58. Kagey, M.H., *et al.* Mediator and cohesin connect gene expression and chromatin architecture. *Nature* **467**, 430-435 (2010).
59. Boyd, M., *et al.* Characterization of the enhancer and promoter landscape of inflammatory bowel disease from human colon biopsies. *Nat Commun* **9**, 1661 (2018).

60. Weiser, M., *et al.* Molecular classification of Crohn's disease reveals two clinically relevant subtypes. *Gut* **67**, 36-42 (2018).
61. Erceg, J., *et al.* Dual functionality of cis-regulatory elements as developmental enhancers and Polycomb response elements. *Genes Dev* (2017).
62. Jadhav, U., *et al.* Acquired Tissue-Specific Promoter Bivalency Is a Basis for PRC2 Necessity in Adult Cells. *Cell* **165**, 1389-1400 (2016).
63. Oittinen, M., *et al.* Polycomb Repressive Complex 2 Enacts Wnt Signaling in Intestinal Homeostasis and Contributes to the Instigation of Stemness in Diseases Entailing Epithelial Hyperplasia or Neoplasia. *Stem Cells* **35**, 445-457 (2017).
64. Jadhav, U., *et al.* Extensive Recovery of Embryonic Enhancer and Gene Memory Stored in Hypomethylated Enhancer DNA. *Mol Cell* **74**, 542-554 e545 (2019).
65. Chen, X., *et al.* Integration of external signaling pathways with the core transcriptional network in embryonic stem cells. *Cell* **133**, 1106-1117 (2008).
66. Camp, J.G., Jazwa, A.L., Trent, C.M. & Rawls, J.F. Intronic cis-regulatory modules mediate tissue-specific and microbial control of angptl4/fiaf transcription. *PLoS Genet* **8**, e1002585 (2012).
67. Camp, J.G., *et al.* Microbiota modulate transcription in the intestinal epithelium without remodeling the accessible chromatin landscape. *Genome Res* **24**, 1504-1516 (2014).
68. Richards, A.L., *et al.* Gut Microbiota Has a Widespread and Modifiable Effect on Host Gene Regulation. *mSystems* **4**(2019).
69. Jadhav, U., *et al.* Dynamic Reorganization of Chromatin Accessibility Signatures during Dedifferentiation of Secretory Precursors into Lgr5+ Intestinal Stem Cells. *Cell Stem Cell* **21**, 65-77 e65 (2017).
70. Thaïss, C.A., *et al.* Microbiota Diurnal Rhythmicity Programs Host Transcriptome Oscillations. *Cell* **167**, 1495-1510 e1412 (2016).
71. Wang, Z., *et al.* Morphological and molecular evidence for functional organization along the rostrocaudal axis of the adult zebrafish intestine. *BMC Genomics* **11**, 392 (2010).
72. Lickwar, C.R., *et al.* Genomic dissection of conserved transcriptional regulation in intestinal epithelial cells. *PLoS Biol* **15**, e2002054 (2017).
73. Haber, A.L., *et al.* A single-cell survey of the small intestinal epithelium. *Nature* **551**, 333-339 (2017).
74. Moor, A.E., *et al.* Spatial Reconstruction of Single Enterocytes Uncovers Broad Zonation along the Intestinal Villus Axis. *Cell* **175**, 1156-1167 e1115 (2018).
75. Castro, L.F., *et al.* Recurrent gene loss correlates with the evolution of stomach phenotypes in gnathostome history. *Proc Biol Sci* **281**, 20132669 (2014).
76. Muncan, V., *et al.* T-cell factor 4 (Tcf7l2) maintains proliferative compartments in zebrafish intestine. *EMBO Rep* **8**, 966-973 (2007).
77. Li, J., Prochaska, M., Maney, L. & Wallace, K.N. Development and organization of the zebrafish intestinal epithelial stem cell niche. *Dev Dyn* (2019).
78. Shyer, A.E., Huycke, T.R., Lee, C., Mahadevan, L. & Tabin, C.J. Bending gradients: how the intestinal stem cell gets its home. *Cell* **161**, 569-580 (2015).
79. Cummings, J.H., Pomare, E.W., Branch, W.J., Naylor, C.P. & Macfarlane, G.T. Short chain fatty acids in human large intestine, portal, hepatic and venous blood. *Gut* **28**, 1221-1227 (1987).
80. Maslowski, K.M., *et al.* Regulation of inflammatory responses by gut microbiota and chemoattractant receptor GPR43. *Nature* **461**, 1282-1286 (2009).
81. Fellows, R., *et al.* Microbiota derived short chain fatty acids promote histone crotonylation in the colon through histone deacetylases. *Nat Commun* **9**, 105 (2018).
82. Krautkramer, K.A., *et al.* Diet-Microbiota Interactions Mediate Global Epigenetic Programming in Multiple Host Tissues. *Mol Cell* **64**, 982-992 (2016).

83. Yuille, S., Reichardt, N., Panda, S., Dunbar, H. & Mulder, I.E. Human gut bacteria as potent class I histone deacetylase inhibitors in vitro through production of butyric acid and valeric acid. *PLoS One* **13**, e0201073 (2018).
84. Donohoe, D.R., *et al.* The Warburg effect dictates the mechanism of butyrate-mediated histone acetylation and cell proliferation. *Mol Cell* **48**, 612-626 (2012).
85. Chang, P.V., Hao, L., Offermanns, S. & Medzhitov, R. The microbial metabolite butyrate regulates intestinal macrophage function via histone deacetylase inhibition. *Proc Natl Acad Sci U S A* **111**, 2247-2252 (2014).
86. Turgeon, N., *et al.* HDAC1 and HDAC2 restrain the intestinal inflammatory response by regulating intestinal epithelial cell differentiation. *PLoS One* **8**, e73785 (2013).
87. Zimmerlin, C.D., *et al.* HDAC1 and HDAC2 collectively regulate intestinal stem cell homeostasis. *FASEB J* **29**, 2070-2080 (2015).
88. Alenghat, T., *et al.* Histone deacetylase 3 coordinates commensal-bacteria-dependent intestinal homeostasis. *Nature* **504**, 153-157 (2013).
89. Sheaffer, K.L., *et al.* DNA methylation is required for the control of stem cell differentiation in the small intestine. *Genes Dev* **28**, 652-664 (2014).
90. Kazakevych, J., Sayols, S., Messner, B., Krienke, C. & Soshnikova, N. Dynamic changes in chromatin states during specification and differentiation of adult intestinal stem cells. *Nucleic Acids Res* **45**, 5770-5784 (2017).
91. Yu, D.H., *et al.* Postnatal epigenetic regulation of intestinal stem cells requires DNA methylation and is guided by the microbiome. *Genome Biol* **16**, 211 (2015).
92. Pan, W.H., *et al.* Exposure to the gut microbiota drives distinct methylome and transcriptome changes in intestinal epithelial cells during postnatal development. *Genome Med* **10**, 27 (2018).
93. Marjoram, L., *et al.* Epigenetic control of intestinal barrier function and inflammation in zebrafish. *Proc Natl Acad Sci U S A* **112**, 2770-2775 (2015).
94. Elliott, E.N., Sheaffer, K.L., Schug, J., Stappenbeck, T.S. & Kaestner, K.H. Dnmt1 is essential to maintain progenitors in the perinatal intestinal epithelium. *Development* **142**, 2163-2172 (2015).
95. Lin, Z., *et al.* Identification of disease-associated DNA methylation in intestinal tissues from patients with inflammatory bowel disease. *Clin Genet* **80**, 59-67 (2011).
96. Rai, K., *et al.* Zebra fish Dnmt1 and Suv39h1 regulate organ-specific terminal differentiation during development. *Mol Cell Biol* **26**, 7077-7085 (2006).
97. Sadler, T., *et al.* Genome-wide analysis of DNA methylation and gene expression defines molecular characteristics of Crohn's disease-associated fibrosis. *Clin Epigenetics* **8**, 30 (2016).
98. Kaaij, L.T., *et al.* DNA methylation dynamics during intestinal stem cell differentiation reveals enhancers driving gene expression in the villus. *Genome Biol* **14**, R50 (2013).
99. Kraiczy, J., *et al.* Assessing DNA methylation in the developing human intestinal epithelium: potential link to inflammatory bowel disease. *Mucosal Immunol* **9**, 647-658 (2016).
100. Howell, K.J., *et al.* DNA Methylation and Transcription Patterns in Intestinal Epithelial Cells From Pediatric Patients With Inflammatory Bowel Diseases Differentiate Disease Subtypes and Associate With Outcome. *Gastroenterology* **154**, 585-598 (2018).
101. Noth, R., *et al.* Anti-TNF-alpha antibodies improve intestinal barrier function in Crohn's disease. *J Crohns Colitis* **6**, 464-469 (2012).
102. Schirmer, M., *et al.* Linking the Human Gut Microbiome to Inflammatory Cytokine Production Capacity. *Cell* **167**, 1125-1136 e1128 (2016).
103. Tymms, M.J., *et al.* A novel epithelial-expressed ETS gene, ELF3: human and murine cDNA sequences, murine genomic organization, human mapping to 1q32.2 and expression in tissues and cancer. *Oncogene* **15**, 2449-2462 (1997).

104. Choi, M.Y., *et al.* A dynamic expression survey identifies transcription factors relevant in mouse digestive tract development. *Development* **133**, 4119-4129 (2006).
105. Ng, A.Y., *et al.* Inactivation of the transcription factor Elf3 in mice results in dysmorphogenesis and altered differentiation of intestinal epithelium. *Gastroenterology* **122**, 1455-1466 (2002).
106. Flentjar, N., *et al.* TGF-betaRII rescues development of small intestinal epithelial cells in Elf3-deficient mice. *Gastroenterology* **132**, 1410-1419 (2007).
107. van Wering, H.M., *et al.* Complex regulation of the lactase-phlorizin hydrolase promoter by GATA-4. *Am J Physiol Gastrointest Liver Physiol* **287**, G899-909 (2004).
108. Fang, R., Olds, L.C. & Sibley, E. Spatio-temporal patterns of intestine-specific transcription factor expression during postnatal mouse gut development. *Gene Expr Patterns* **6**, 426-432 (2006).
109. Bosse, T., *et al.* Gata4 is essential for the maintenance of jejunal-ileal identities in the adult mouse small intestine. *Mol Cell Biol* **26**, 9060-9070 (2006).
110. Battle, M.A., *et al.* GATA4 is essential for jejunal function in mice. *Gastroenterology* **135**, 1676-1686 e1671 (2008).
111. Thompson, C.A., *et al.* GATA4 Is Sufficient to Establish Jejunal Versus Ileal Identity in the Small Intestine. *Cell Mol Gastroenterol Hepatol* **3**, 422-446 (2017).
112. Beuling, E., *et al.* GATA6 is required for proliferation, migration, secretory cell maturation, and gene expression in the mature mouse colon. *Mol Cell Biol* **32**, 3392-3402 (2012).
113. Walker, E.M., Thompson, C.A. & Battle, M.A. GATA4 and GATA6 regulate intestinal epithelial cytodifferentiation during development. *Dev Biol* **392**, 283-294 (2014).
114. Chen, L., *et al.* HNF4 factors control chromatin accessibility and are redundantly required for maturation of the fetal intestine. *bioRxiv*, 610428 (2019).
115. Taraviras, S., Monaghan, A.P., Schutz, G. & Kelsey, G. Characterization of the mouse HNF-4 gene and its expression during mouse embryogenesis. *Mech Dev* **48**, 67-79 (1994).
116. Torres-Padilla, M.E. & Weiss, M.C. Effects of interactions of hepatocyte nuclear factor 4alpha isoforms with coactivators and corepressors are promoter-specific. *FEBS Lett* **539**, 19-23 (2003).
117. Chahar, S., *et al.* Chromatin profiling reveals regulatory network shifts and a protective role for hepatocyte nuclear factor 4alpha during colitis. *Mol Cell Biol* **34**, 3291-3304 (2014).
118. Hatzia Apostolou, M., *et al.* An HNF4alpha-miRNA inflammatory feedback circuit regulates hepatocellular oncogenesis. *Cell* **147**, 1233-1247 (2011).
119. Cattin, A.L., *et al.* Hepatocyte nuclear factor 4alpha, a key factor for homeostasis, cell architecture, and barrier function of the adult intestinal epithelium. *Mol Cell Biol* **29**, 6294-6308 (2009).
120. Taraviras, S., *et al.* Primary structure, chromosomal mapping, expression and transcriptional activity of murine hepatocyte nuclear factor 4gamma. *Biochim Biophys Acta* **1490**, 21-32 (2000).
121. Chen, W.S., *et al.* Disruption of the HNF-4 gene, expressed in visceral endoderm, leads to cell death in embryonic ectoderm and impaired gastrulation of mouse embryos. *Genes Dev* **8**, 2466-2477 (1994).
122. Duncan, S.A., Nagy, A. & Chan, W. Murine gastrulation requires HNF-4 regulated gene expression in the visceral endoderm: tetraploid rescue of Hnf-4(-/-) embryos. *Development* **124**, 279-287 (1997).
123. Parviz, F., *et al.* Hepatocyte nuclear factor 4alpha controls the development of a hepatic epithelium and liver morphogenesis. *Nat Genet* **34**, 292-296 (2003).

124. Li, J., Ning, G. & Duncan, S.A. Mammalian hepatocyte differentiation requires the transcription factor HNF-4alpha. *Genes Dev* **14**, 464-474 (2000).
125. Garrison, W.D., *et al.* Hepatocyte nuclear factor 4alpha is essential for embryonic development of the mouse colon. *Gastroenterology* **130**, 1207-1220 (2006).
126. Ahn, S.H., *et al.* Hepatocyte nuclear factor 4alpha in the intestinal epithelial cells protects against inflammatory bowel disease. *Inflamm Bowel Dis* **14**, 908-920 (2008).
127. Babeu, J.P., *et al.* Quantitative Proteomics Identifies DNA Repair as a Novel Biological Function for Hepatocyte Nuclear Factor 4alpha in Colorectal Cancer Cells. *Cancers (Basel)* **11**(2019).
128. Darsigny, M., *et al.* Loss of hepatocyte-nuclear-factor-4alpha affects colonic ion transport and causes chronic inflammation resembling inflammatory bowel disease in mice. *PLoS One* **4**, e7609 (2009).
129. Barrett, J.C., *et al.* Genome-wide association study of ulcerative colitis identifies three new susceptibility loci, including the HNF4A region. *Nat Genet* **41**, 1330-1334 (2009).
130. Marcil, V., *et al.* Association between genetic variants in the HNF4A gene and childhood-onset Crohn's disease. *Genes Immun* **13**, 556-565 (2012).
131. Mokry, M., *et al.* Many inflammatory bowel disease risk loci include regions that regulate gene expression in immune cells and the intestinal epithelium. *Gastroenterology* **146**, 1040-1047 (2014).
132. Meddens, C.A., *et al.* Systematic analysis of chromatin interactions at disease associated loci links novel candidate genes to inflammatory bowel disease. *Genome Biol* **17**, 247 (2016).
133. Darsigny, M., *et al.* Hepatocyte nuclear factor-4alpha promotes gut neoplasia in mice and protects against the production of reactive oxygen species. *Cancer Res* **70**, 9423-9433 (2010).
134. Babeu, J.P., Darsigny, M., Lussier, C.R. & Boudreau, F. Hepatocyte nuclear factor 4alpha contributes to an intestinal epithelial phenotype in vitro and plays a partial role in mouse intestinal epithelium differentiation. *Am J Physiol Gastrointest Liver Physiol* **297**, G124-134 (2009).
135. Devillard, E., McIntosh, F.M., Duncan, S.H. & Wallace, R.J. Metabolism of linoleic acid by human gut bacteria: different routes for biosynthesis of conjugated linoleic acid. *J Bacteriol* **189**, 2566-2570 (2007).
136. Franke, A., *et al.* Systematic association mapping identifies NELL1 as a novel IBD disease gene. *PLoS One* **2**, e691 (2007).
137. Gerdin, A.K., *et al.* Phenotypic screening of hepatocyte nuclear factor (HNF) 4-gamma receptor knockout mice. *Biochem Biophys Res Commun* **349**, 825-832 (2006).
138. Daigo, K., *et al.* Proteomic analysis of native hepatocyte nuclear factor-4alpha (HNF4alpha) isoforms, phosphorylation status, and interactive cofactors. *J Biol Chem* **286**, 674-686 (2011).
139. Li, X., *et al.* Dynamic patterning at the pylorus: formation of an epithelial intestine-stomach boundary in late fetal life. *Dev Dyn* **238**, 3205-3217 (2009).
140. Ko, H.L., Zhuo, Z. & Ren, E.C. HNF4alpha Combinatorial Isoform Heterodimers Activate Distinct Gene Targets that Differ from Their Corresponding Homodimers. *Cell Rep* **26**, 2549-2557 e2543 (2019).
141. Sousa, J.F., *et al.* miR-30-HNF4gamma and miR-194-NR2F2 regulatory networks contribute to the upregulation of metaplasia markers in the stomach. *Gut* **65**, 914-924 (2016).
142. Lindeboom, R.G., *et al.* Integrative multi-omics analysis of intestinal organoid differentiation. *Mol Syst Biol* **14**, e8227 (2018).

143. Baraille, F., *et al.* Glucose Tolerance Is Improved in Mice Invalidated for the Nuclear Receptor HNF-4gamma: A Critical Role for Enteroendocrine Cell Lineage. *Diabetes* **64**, 2744-2756 (2015).
144. Chen, L., *et al.* A reinforcing HNF4-SMAD4 feed-forward module stabilizes enterocyte identity. *Nat Genet* **51**, 777-785 (2019).
145. Hertz, R., Magenheimer, J., Berman, I. & Bar-Tana, J. Fatty acyl-CoA thioesters are ligands of hepatic nuclear factor-4alpha. *Nature* **392**, 512-516 (1998).
146. Dhe-Paganon, S., Duda, K., Iwamoto, M., Chi, Y.I. & Shoelson, S.E. Crystal structure of the HNF4 alpha ligand binding domain in complex with endogenous fatty acid ligand. *J Biol Chem* **277**, 37973-37976 (2002).
147. Wisely, G.B., *et al.* Hepatocyte nuclear factor 4 is a transcription factor that constitutively binds fatty acids. *Structure* **10**, 1225-1234 (2002).
148. Yuan, X., *et al.* Identification of an endogenous ligand bound to a native orphan nuclear receptor. *PLoS One* **4**, e5609 (2009).
149. Palanker, L., Tennessen, J.M., Lam, G. & Thummel, C.S. Drosophila HNF4 regulates lipid mobilization and beta-oxidation. *Cell Metab* **9**, 228-239 (2009).
150. Frochot, V., *et al.* The transcription factor HNF-4alpha: a key factor of the intestinal uptake of fatty acids in mouse. *Am J Physiol Gastrointest Liver Physiol* **302**, G1253-1263 (2012).
151. Mangelsdorf, D.J., Ong, E.S., Dyck, J.A. & Evans, R.M. Nuclear receptor that identifies a novel retinoic acid response pathway. *Nature* **345**, 224-229 (1990).
152. Bridgham, J.T., *et al.* Protein evolution by molecular tinkering: diversification of the nuclear receptor superfamily from a ligand-dependent ancestor. *PLoS Biol* **8**(2010).
153. Poirier, H., *et al.* Differential involvement of peroxisome-proliferator-activated receptors alpha and delta in fibrates and fatty-acid-mediated inductions of the gene encoding liver fatty-acid-binding protein in the liver and the small intestine. *Biochem J* **355**, 481-488 (2001).
154. Su, C.G., *et al.* A novel therapy for colitis utilizing PPAR-gamma ligands to inhibit the epithelial inflammatory response. *J Clin Invest* **104**, 383-389 (1999).
155. Celinski, K., *et al.* Comparison of the anti-inflammatory and therapeutic actions of PPAR-gamma agonists rosiglitazone and troglitazone in experimental colitis. *J Physiol Pharmacol* **63**, 631-640 (2012).
156. Kliewer, S.A., Umesono, K., Noonan, D.J., Heyman, R.A. & Evans, R.M. Convergence of 9-cis retinoic acid and peroxisome proliferator signalling pathways through heterodimer formation of their receptors. *Nature* **358**, 771-774 (1992).
157. Alex, S., *et al.* Short-chain fatty acids stimulate angiopoietin-like 4 synthesis in human colon adenocarcinoma cells by activating peroxisome proliferator-activated receptor gamma. *Mol Cell Biol* **33**, 1303-1316 (2013).
158. Korecka, A., *et al.* ANGPTL4 expression induced by butyrate and rosiglitazone in human intestinal epithelial cells utilizes independent pathways. *Am J Physiol Gastrointest Liver Physiol* **304**, G1025-1037 (2013).
159. Byndloss, M.X., *et al.* Microbiota-activated PPAR-gamma signaling inhibits dysbiotic Enterobacteriaceae expansion. *Science* **357**, 570-575 (2017).
160. Peyrin-Biroulet, L., *et al.* Peroxisome proliferator-activated receptor gamma activation is required for maintenance of innate antimicrobial immunity in the colon. *Proc Natl Acad Sci U S A* **107**, 8772-8777 (2010).
161. Glas, J., *et al.* Role of PPARG gene variants in inflammatory bowel disease. *Inflamm Bowel Dis* **17**, 1057-1058 (2011).
162. Andersen, V., *et al.* Polymorphisms in NF-kappaB, PXR, LXR, PPARgamma and risk of inflammatory bowel disease. *World J Gastroenterol* **17**, 197-206 (2011).

163. Bank, S., *et al.* Polymorphisms in the inflammatory pathway genes TLR2, TLR4, TLR9, LY96, NFKBIA, NFKB1, TNFA, TNFRSF1A, IL6R, IL10, IL23R, PTPN22, and PPARG are associated with susceptibility of inflammatory bowel disease in a Danish cohort. *PLoS One* **9**, e98815 (2014).
164. Dubuquoy, L., *et al.* Impaired expression of peroxisome proliferator-activated receptor gamma in ulcerative colitis. *Gastroenterology* **124**, 1265-1276 (2003).
165. Dou, X., Xiao, J., Jin, Z. & Zheng, P. Peroxisome proliferator-activated receptor-gamma is downregulated in ulcerative colitis and is involved in experimental colitis-associated neoplasia. *Oncol Lett* **10**, 1259-1266 (2015).
166. Lu, P., *et al.* Intestinal epithelial Toll-like receptor 4 prevents metabolic syndrome by regulating interactions between microbes and intestinal epithelial cells in mice. *Mucosal Immunol* **11**, 727-740 (2018).
167. Kelly, D., *et al.* Commensal anaerobic gut bacteria attenuate inflammation by regulating nuclear-cytoplasmic shuttling of PPAR-gamma and RelA. *Nat. Immunol.* **5**, 104-112 (2004).
168. Braissant, O., Fufelle, F., Scotto, C., Dauca, M. & Wahli, W. Differential expression of peroxisome proliferator-activated receptors (PPARs): tissue distribution of PPAR-alpha, -beta, and -gamma in the adult rat. *Endocrinology* **137**, 354-366 (1996).
169. Bäckhed, F., *et al.* The gut microbiota as an environmental factor that regulates fat storage. *Proc Natl Acad Sci U S A* **101**, 15718-15723 (2004).
170. Manoharan, I., *et al.* Homeostatic PPARalpha Signaling Limits Inflammatory Responses to Commensal Microbiota in the Intestine. *J Immunol* **196**, 4739-4749 (2016).
171. Tomas, J., *et al.* High-fat diet modifies the PPAR-gamma pathway leading to disruption of microbial and physiological ecosystem in murine small intestine. *Proc Natl Acad Sci U S A* **113**, E5934-E5943 (2016).
172. Makishima, M., *et al.* Identification of a Nuclear Receptor for Bile Acids. *Science* **284**, 1362-1365 (1999).
173. Devlin, A.S. & Fischbach, M.A. A biosynthetic pathway for a prominent class of microbiota-derived bile acids. *Nat Chem Biol* **11**, 685-690 (2015).
174. Sayin, S.I., *et al.* Gut microbiota regulates bile acid metabolism by reducing the levels of tauro-beta-muricholic acid, a naturally occurring FXR antagonist. *Cell Metab* **17**, 225-235 (2013).
175. Wahlstrom, A., *et al.* Induction of farnesoid X receptor signaling in germ-free mice colonized with a human microbiota. *J Lipid Res* **58**, 412-419 (2017).
176. Li, F., *et al.* Microbiome remodelling leads to inhibition of intestinal farnesoid X receptor signalling and decreased obesity. *Nat Commun* **4**, 2384 (2013).
177. Degirolamo, C., Rainaldi, S., Bovenga, F., Murzilli, S. & Moschetta, A. Microbiota modification with probiotics induces hepatic bile acid synthesis via downregulation of the Fxr-Fgf15 axis in mice. *Cell Rep* **7**, 12-18 (2014).
178. Parseus, A., *et al.* Microbiota-induced obesity requires farnesoid X receptor. *Gut* **66**, 429-437 (2017).
179. Xie, C., *et al.* An Intestinal Farnesoid X Receptor-Ceramide Signaling Axis Modulates Hepatic Gluconeogenesis in Mice. *Diabetes* **66**, 613-626 (2017).
180. Zhang, L., *et al.* Farnesoid X Receptor Signaling Shapes the Gut Microbiota and Controls Hepatic Lipid Metabolism. *mSystems* **1**(2016).
181. Vavassori, P., Mencarelli, A., Renga, B., Distrutti, E. & Fiorucci, S. The bile acid receptor FXR is a modulator of intestinal innate immunity. *J Immunol* **183**, 6251-6261 (2009).
182. Gadaleta, R.M., *et al.* Farnesoid X receptor activation inhibits inflammation and preserves the intestinal barrier in inflammatory bowel disease. *Gut* **60**, 463-472 (2011).

183. Janowski, B.A., Willy, P.J., Devi, T.R., Falck, J.R. & Mangelsdorf, D.J. An oxysterol signalling pathway mediated by the nuclear receptor LXR alpha. *Nature* **383**, 728-731 (1996).
184. Colin, S., *et al.* Intestine-specific regulation of PPARalpha gene transcription by liver X receptors. *Endocrinology* **149**, 5128-5135 (2008).
185. Joseph, S.B., Castrillo, A., Laffitte, B.A., Mangelsdorf, D.J. & Tontonoz, P. Reciprocal regulation of inflammation and lipid metabolism by liver X receptors. *Nat Med* **9**, 213-219 (2003).
186. Castrillo, A., Joseph, S.B., Marathe, C., Mangelsdorf, D.J. & Tontonoz, P. Liver X receptor-dependent repression of matrix metalloproteinase-9 expression in macrophages. *J Biol Chem* **278**, 10443-10449 (2003).
187. Jakobsson, T., *et al.* The oxysterol receptor LXRbeta protects against DSS- and TNBS-induced colitis in mice. *Mucosal Immunol* **7**, 1416-1428 (2014).
188. Sakurai, H., *et al.* Suppression of NF-kappa B and AP-1 activation by glucocorticoids in experimental glomerulonephritis in rats: molecular mechanisms of anti-nephritic action. *Biochim Biophys Acta* **1362**, 252-262 (1997).
189. Mandrekar, P., Bellerose, G. & Szabo, G. Inhibition of NF-kappa B binding correlates with increased nuclear glucocorticoid receptor levels in acute alcohol-treated human monocytes. *Alcohol Clin Exp Res* **26**, 1872-1879 (2002).
190. Ballegeer, M., *et al.* Glucocorticoid receptor dimers control intestinal STAT1 and TNF-induced inflammation in mice. *J Clin Invest* **128**, 3265-3279 (2018).
191. Franke, A., *et al.* Genome-wide meta-analysis increases to 71 the number of confirmed Crohn's disease susceptibility loci. *Nat Genet* **42**, 1118-1125 (2010).
192. Mwinyi, J., Wenger, C., Eloranta, J.J. & Kullak-Ublick, G.A. Glucocorticoid receptor gene haplotype structure and steroid therapy outcome in IBD patients. *World J Gastroenterol* **16**, 3888-3896 (2010).
193. Krupoves, A., Mack, D., Deslandres, C., Seidman, E. & Amre, D.K. Variation in the glucocorticoid receptor gene (NR3C1) may be associated with corticosteroid dependency and resistance in children with Crohn's disease. *Pharmacogenet Genomics* **21**, 454-460 (2011).
194. Fuller, P.J. & Verity, K. Mineralocorticoid receptor gene expression in the gastrointestinal tract: distribution and ontogeny. *J Steroid Biochem* **36**, 263-267 (1990).
195. Nakamura, T., *et al.* Intestinal Mineralocorticoid Receptor Contributes to Epithelial Sodium Channel-Mediated Intestinal Sodium Absorption and Blood Pressure Regulation. *J Am Heart Assoc* **7**(2018).
196. Froicu, M., Zhu, Y. & Cantorna, M.T. Vitamin D receptor is required to control gastrointestinal immunity in IL-10 knockout mice. *Immunology* **117**, 310-318 (2006).
197. Kong, J., *et al.* Novel role of the vitamin D receptor in maintaining the integrity of the intestinal mucosal barrier. *Am J Physiol Gastrointest Liver Physiol* **294**, G208-216 (2008).
198. Wu, S., *et al.* Intestinal epithelial vitamin D receptor deletion leads to defective autophagy in colitis. *Gut* **64**, 1082-1094 (2015).
199. Wang, J., *et al.* Genome-wide association analysis identifies variation in vitamin D receptor and other host factors influencing the gut microbiota. *Nat Genet* **48**, 1396-1406 (2016).
200. Liu, W., *et al.* Intestinal epithelial vitamin D receptor signaling inhibits experimental colitis. *J Clin Invest* **123**, 3983-3996 (2013).
201. Golan, M.A., *et al.* Transgenic Expression of Vitamin D Receptor in Gut Epithelial Cells Ameliorates Spontaneous Colitis Caused by Interleukin-10 Deficiency. *Dig Dis Sci* **60**, 1941-1947 (2015).

202. Gubatan, J. & Moss, A.C. Vitamin D in inflammatory bowel disease: more than just a supplement. *Curr Opin Gastroenterol* **34**, 217-225 (2018).
203. Lundin, A., et al. Gut flora, Toll-like receptors and nuclear receptors: a tripartite communication that tunes innate immunity in large intestine. *Cell Microbiol* **10**, 1093-1103 (2008).
204. Bjorkholm, B., et al. Intestinal microbiota regulate xenobiotic metabolism in the liver. *PLoS One* **4**, e6958 (2009).
205. Hudson, G.M., et al. Constitutive androstane receptor regulates the intestinal mucosal response to injury. *Br J Pharmacol* **174**, 1857-1871 (2017).
206. Gao, J., He, J., Zhai, Y., Wada, T. & Xie, W. The constitutive androstane receptor is an anti-obesity nuclear receptor that improves insulin sensitivity. *J Biol Chem* **284**, 25984-25992 (2009).
207. Dempsey, J.L., et al. Pharmacological Activation of PXR and CAR Downregulates Distinct Bile Acid-Metabolizing Intestinal Bacteria and Alters Bile Acid Homeostasis. *Toxicol Sci* **168**, 40-60 (2019).
208. Venkatesh, M., et al. Symbiotic bacterial metabolites regulate gastrointestinal barrier function via the xenobiotic sensor PXR and Toll-like receptor 4. *Immunity* **41**, 296-310 (2014).
209. Terc, J., Hansen, A., Alston, L. & Hirota, S.A. Pregnane X receptor agonists enhance intestinal epithelial wound healing and repair of the intestinal barrier following the induction of experimental colitis. *Eur J Pharm Sci* **55**, 12-19 (2014).
210. Garg, A., et al. Pregnane X Receptor Activation Attenuates Inflammation-Associated Intestinal Epithelial Barrier Dysfunction by Inhibiting Cytokine-Induced Myosin Light-Chain Kinase Expression and c-Jun N-Terminal Kinase 1/2 Activation. *J Pharmacol Exp Ther* **359**, 91-101 (2016).
211. Shakhnovich, V., et al. Decreased Pregnane X Receptor Expression in Children with Active Crohn's Disease. *Drug Metab Dispos* **44**, 1066-1069 (2016).
212. Meng, Z., et al. The atypical antipsychotic quetiapine induces hyperlipidemia by activating intestinal PXR signaling. *JCI Insight* **4**(2019).
213. Sui, Y., et al. Intestinal pregnane X receptor links xenobiotic exposure and hypercholesterolemia. *Mol Endocrinol* **29**, 765-776 (2015).
214. Bartonkova, I. & Dvorak, Z. Essential oils of culinary herbs and spices activate PXR and induce CYP3A4 in human intestinal and hepatic in vitro models. *Toxicol Lett* **296**, 1-9 (2018).
215. Kelly, C.J., et al. Crosstalk between Microbiota-Derived Short-Chain Fatty Acids and Intestinal Epithelial HIF Augments Tissue Barrier Function. *Cell Host Microbe* **17**, 662-671 (2015).
216. Louis, N.A., et al. Selective induction of mucin-3 by hypoxia in intestinal epithelia. *J Cell Biochem* **99**, 1616-1627 (2006).
217. Kelly, C.J., et al. Fundamental role for HIF-1alpha in constitutive expression of human beta defensin-1. *Mucosal Immunol* **6**, 1110-1118 (2013).
218. Furuta, G.T., et al. Hypoxia-inducible factor 1-dependent induction of intestinal trefoil factor protects barrier function during hypoxia. *J Exp Med* **193**, 1027-1034 (2001).
219. Fan, D., et al. Activation of HIF-1alpha and LL-37 by commensal bacteria inhibits *Candida albicans* colonization. *Nat Med* **21**, 808-814 (2015).
220. Troll, J.V., et al. Microbiota promote secretory cell determination in the intestinal epithelium by modulating host Notch signaling. *Development* **145**(2018).
221. Obata, Y., et al. Epithelial cell-intrinsic Notch signaling plays an essential role in the maintenance of gut immune homeostasis. *J Immunol* **188**, 2427-2436 (2012).
222. Guo, X.K., Ou, J., Liang, S., Zhou, X. & Hu, X. Epithelial Hes1 maintains gut homeostasis by preventing microbial dysbiosis. *Mucosal Immunol* **11**, 716-726 (2018).

223. El Aidy, S., *et al.* Temporal and spatial interplay of microbiota and intestinal mucosa drive establishment of immune homeostasis in conventionalized mice. *Mucosal Immunol* **5**, 567-579 (2012).
224. El Aidy, S., *et al.* The gut microbiota elicits a profound metabolic reorientation in the mouse jejunal mucosa during conventionalisation. *Gut* **62**, 1306-1314 (2013).
225. Peck, B.C., *et al.* Functional Transcriptomics in Diverse Intestinal Epithelial Cell Types Reveals Robust MicroRNA Sensitivity in Intestinal Stem Cells to Microbial Status. *J Biol Chem* **292**, 2586-2600 (2017).
226. Sharma, R., Schumacher, U., Ronaasen, V. & Coates, M. Rat intestinal mucosal responses to a microbial flora and different diets. *Gut* **36**, 209-214 (1995).
227. Bates, J.M., *et al.* Distinct signals from the microbiota promote different aspects of zebrafish gut differentiation. *Dev. Biol.* **297**, 374-386 (2006).
228. Zhou, Y., Wang, Q., Weiss, H.L. & Evers, B.M. Nuclear factor of activated T-cells 5 increases intestinal goblet cell differentiation through an mTOR/Notch signaling pathway. *Mol Biol Cell* **25**, 2882-2890 (2014).
229. Zhou, Y., Rychahou, P., Wang, Q., Weiss, H.L. & Evers, B.M. TSC2/mTORC1 signaling controls Paneth and goblet cell differentiation in the intestinal epithelium. *Cell death & disease* **6**, e1631 (2015).
230. Park, J.H., *et al.* Promotion of Intestinal Epithelial Cell Turnover by Commensal Bacteria: Role of Short-Chain Fatty Acids. *PLoS One* **11**, e0156334 (2016).
231. Cheesman, S.E., Neal, J.T., Mittge, E., Seredick, B.M. & Guillemin, K. Epithelial cell proliferation in the developing zebrafish intestine is regulated by the Wnt pathway and microbial signaling via Myd88. *Proc Natl Acad Sci U S A* **108 Suppl 1**, 4570-4577 (2011).
232. van Es, J.H., *et al.* Wnt signalling induces maturation of Paneth cells in intestinal crypts. *Nat Cell Biol* **7**, 381-386 (2005).
233. Li, X., *et al.* Deconvoluting the intestine: molecular evidence for a major role of the mesenchyme in the modulation of signaling cross talk. *Physiol Genomics* **29**, 290-301 (2007).
234. Howe, J.R., *et al.* Mutations in the SMAD4/DPC4 gene in juvenile polyposis. *Science* **280**, 1086-1088 (1998).
235. Howe, J.R., *et al.* Germline mutations of the gene encoding bone morphogenetic protein receptor 1A in juvenile polyposis. *Nat Genet* **28**, 184-187 (2001).
236. Sancho, E., Battle, E. & Clevers, H. Signaling pathways in intestinal development and cancer. *Annu Rev Cell Dev Biol* **20**, 695-723 (2004).
237. Auclair, B.A., Benoit, Y.D., Rivard, N., Mishina, Y. & Perreault, N. Bone morphogenetic protein signaling is essential for terminal differentiation of the intestinal secretory cell lineage. *Gastroenterology* **133**, 887-896 (2007).
238. Beumer, J., *et al.* Enteroendocrine cells switch hormone expression along the crypt-to-villus BMP signalling gradient. *Nat Cell Biol* **20**, 909-916 (2018).
239. Haller, D., *et al.* Transforming growth factor-beta 1 inhibits non-pathogenic Gram negative bacteria-induced NF-kappa B recruitment to the interleukin-6 gene promoter in intestinal epithelial cells through modulation of histone acetylation. *J Biol Chem* **278**, 23851-23860 (2003).
240. Ruiz, P.A., Shkoda, A., Kim, S.C., Sartor, R.B. & Haller, D. IL-10 gene-deficient mice lack TGF-beta/Smad signaling and fail to inhibit proinflammatory gene expression in intestinal epithelial cells after the colonization with colitogenic *Enterococcus faecalis*. *J Immunol* **174**, 2990-2999 (2005).
241. Kawajiri, K. & Fujii-Kuriyama, Y. The aryl hydrocarbon receptor: a multifunctional chemical sensor for host defense and homeostatic maintenance. *Exp Anim* **66**, 75-89 (2017).

242. Moura-Alves, P., *et al.* AhR sensing of bacterial pigments regulates antibacterial defence. *Nature* **512**, 387-392 (2014).
243. Hubbard, T.D., *et al.* Adaptation of the human aryl hydrocarbon receptor to sense microbiota-derived indoles. *Sci Rep* **5**, 12689 (2015).
244. Lamas, B., *et al.* CARD9 impacts colitis by altering gut microbiota metabolism of tryptophan into aryl hydrocarbon receptor ligands. *Nat Med* **22**, 598-605 (2016).
245. Sun, M., Ma, N., He, T., Johnston, L.J. & Ma, X. Tryptophan (Trp) modulates gut homeostasis via aryl hydrocarbon receptor (AhR). *Crit Rev Food Sci Nutr*, 1-9 (2019).
246. Marinelli, L., *et al.* Identification of the novel role of butyrate as AhR ligand in human intestinal epithelial cells. *Sci Rep* **9**, 643 (2019).
247. Fukumoto, S., *et al.* Identification of a probiotic bacteria-derived activator of the aryl hydrocarbon receptor that inhibits colitis. *Immunol Cell Biol* **92**, 460-465 (2014).
248. Lisy, K. & Peet, D.J. Turn me on: regulating HIF transcriptional activity. *Cell Death Differ* **15**, 642-649 (2008).
249. Albenberg, L., *et al.* Correlation Between Intraluminal Oxygen Gradient and Radial Partitioning of Intestinal Microbiota. *Gastroenterology* **147**, 1055-1063.e1058 (2014).
250. Lamas, B., Natividad, J.M. & Sokol, H. Aryl hydrocarbon receptor and intestinal immunity. *Mucosal Immunol* **11**, 1024-1038 (2018).
251. Furumatsu, K., *et al.* A role of the aryl hydrocarbon receptor in attenuation of colitis. *Dig Dis Sci* **56**, 2532-2544 (2011).
252. Yu, M., *et al.* Aryl Hydrocarbon Receptor Activation Modulates Intestinal Epithelial Barrier Function by Maintaining Tight Junction Integrity. *Int J Biol Sci* **14**, 69-77 (2018).
253. Kawajiri, K., *et al.* Aryl hydrocarbon receptor suppresses intestinal carcinogenesis in ApcMin/+ mice with natural ligands. *Proc Natl Acad Sci U S A* **106**, 13481-13486 (2009).
254. Park, J.H., Choi, A.J., Kim, S.J., Cheong, S.W. & Jeong, S.Y. AhR activation by 6-formylindolo[3,2-b]carbazole and 2,3,7,8-tetrachlorodibenzo-p-dioxin inhibit the development of mouse intestinal epithelial cells. *Environ Toxicol Pharmacol* **43**, 44-53 (2016).
255. Metidji, A., *et al.* The Environmental Sensor AHR Protects from Inflammatory Damage by Maintaining Intestinal Stem Cell Homeostasis and Barrier Integrity. *Immunity* **50**, 1542 (2019).
256. Lanis, J.M., *et al.* Tryptophan metabolite activation of the aryl hydrocarbon receptor regulates IL-10 receptor expression on intestinal epithelia. *Mucosal Immunol* **10**, 1133-1144 (2017).
257. Kawai, S., *et al.* Indigo Naturalis ameliorates murine dextran sodium sulfate-induced colitis via aryl hydrocarbon receptor activation. *J Gastroenterol* **52**, 904-919 (2017).
258. Yoshimatsu, Y., *et al.* Development of an Indigo Naturalis Suppository for Topical Induction Therapy in Patients with Ulcerative Colitis. *Digestion*, 1-7 (2019).
259. Marafini, I., *et al.* NPD-0414-2 and NPD-0414-24, Two Chemical Entities Designed as Aryl Hydrocarbon Receptor (AhR) Ligands, Inhibit Gut Inflammatory Signals. *Front Pharmacol* **10**, 380 (2019).
260. Wang, Y., *et al.* The intestinal microbiota regulates body composition through NFIL3 and the circadian clock. *Science* **357**, 912-916 (2017).
261. van der Flier, L.G., *et al.* Transcription factor achaete scute-like 2 controls intestinal stem cell fate. *Cell* **136**, 903-912 (2009).
262. Schuijers, J., *et al.* Ascl2 acts as an R-spondin/Wnt-responsive switch to control stemness in intestinal crypts. *Cell Stem Cell* **16**, 158-170 (2015).
263. Wilson, J.M. & Castro, L.F.C. 1 - Morphological diversity of the gastrointestinal tract in fishes. in *Fish Physiology*, Vol. 30 (eds. Grosell, M., Farrell, A.P. & Brauner, C.J.) 1-55 (Academic Press, 2010).

264. Tan, D.W. & Barker, N. Intestinal stem cells and their defining niche. *Current topics in developmental biology* **107**, 77-107 (2014).
265. Medema, J.P. & Vermeulen, L. Microenvironmental regulation of stem cells in intestinal homeostasis and cancer. *Nature* **474**, 318-326 (2011).
266. Umar, S. Intestinal stem cells. *Curr Gastroenterol Rep* **12**, 340-348 (2010).
267. Bankaitis, E.D., Ha, A., Kuo, C.J. & Magness, S.T. Reserve Stem Cells in Intestinal Homeostasis and Injury. *Gastroenterology* **155**, 1348-1361 (2018).
268. Zhang, Z. & Liu, Z. Paneth cells: the hub for sensing and regulating intestinal flora. *Sci China Life Sci* **59**, 463-467 (2016).
269. Gerbe, F., Legraverend, C. & Jay, P. The intestinal epithelium tuft cells: specification and function. *Cell Mol Life Sci* **69**, 2907-2917 (2012).
270. Shirazi, T., Longman, R.J., Corfield, A.P. & Probert, C.S. Mucins and inflammatory bowel disease. *Postgrad Med J* **76**, 473-478 (2000).
271. van den Bogert, B., Leimena, M.M., de Vos, W.M., Zoetendal, E.G. & Kleerebezem, M. Functional Intestinal Metagenomics. *Handbook of Molecular Microbial Ecology II*, 175-190 (2011).
272. Elliott, E.N. & Kaestner, K.H. Epigenetic regulation of the intestinal epithelium. *Cell Mol Life Sci* **72**, 4139-4156 (2015).
273. Iwafuchi-Doi, M. & Zaret, K.S. Pioneer transcription factors in cell reprogramming. *Genes Dev* **28**, 2679-2692 (2014).
274. Jedlicka, P. & Gutierrez-Hartmann, A. Ets transcription factors in intestinal morphogenesis, homeostasis and disease. *Histol Histopathol* **23**, 1417-1424 (2008).
275. Bulger, M. & Groudine, M. Functional and mechanistic diversity of distal transcription enhancers. *Cell* **144**, 327-339 (2011).
276. Lee, T.I. & Young, R.A. Transcriptional regulation and its misregulation in disease. *Cell* **152**, 1237-1251 (2013).
277. Bannister, A.J. & Kouzarides, T. Regulation of chromatin by histone modifications. *Cell Res* **21**, 381-395 (2011).
278. Shlyueva, D., Stampfel, G. & Stark, A. Transcriptional enhancers: from properties to genome-wide predictions. *Nat Rev Genet* **15**, 272-286 (2014).
279. Lawrence, M., Daujat, S. & Schneider, R. Lateral Thinking: How Histone Modifications Regulate Gene Expression. *Trends Genet* **32**, 42-56 (2016).
280. Sabari, B.R., Zhang, D., Allis, C.D. & Zhao, Y. Metabolic regulation of gene expression through histone acylations. *Nat Rev Mol Cell Biol* **18**, 90-101 (2017).
281. Barnes, C.E., English, D.M. & Cowley, S.M. Acetylation & Co: an expanding repertoire of histone acylations regulates chromatin and transcription. *Essays Biochem* **63**, 97-107 (2019).
282. Edwards, J.R., Yarychivska, O., Boulard, M. & Bestor, T.H. DNA methylation and DNA methyltransferases. *Epigenetics Chromatin* **10**, 23 (2017).
283. Furey, T.S., Sethupathy, P. & Sheikh, S.Z. Redefining the IBDs using genome-scale molecular phenotyping. *Nat Rev Gastroenterol Hepatol* **16**, 296-311 (2019).
284. Bamias, G., Jia, L.G. & Cominelli, F. The tumor necrosis factor-like cytokine 1A/death receptor 3 cytokine system in intestinal inflammation. *Curr Opin Gastroenterol* **29**, 597-602 (2013).
285. Adegbola, S.O., Sahnun, K., Warusavitarne, J., Hart, A. & Tozer, P. Anti-TNF Therapy in Crohn's Disease. *Int J Mol Sci* **19**(2018).
286. Kopylov, U. & Seidman, E. Predicting durable response or resistance to antitumor necrosis factor therapy in inflammatory bowel disease. *Therap Adv Gastroenterol* **9**, 513-526 (2016).
287. Dalile, B., Van Oudenhove, L., Vervliet, B. & Verbeke, K. The role of short-chain fatty acids in microbiota-gut-brain communication. *Nat Rev Gastroenterol Hepatol* (2019).

288. Wu, J., Ma, N., Johnston, L.J. & Ma, X. Dietary Nutrients Mediate Intestinal Host Defense Peptide Expression. *Adv Nutr* (2019).
289. Sellers, R.S. & Morton, D. The colon: from banal to brilliant. *Toxicol Pathol* **42**, 67-81 (2014).
290. Wang, J., Dong, R. & Zheng, S. Roles of the inflammasome in the gut-liver axis (Review). *Mol Med Rep* **19**, 3-14 (2019).
291. Gilmore, A.P. Anoikis. *Cell Death Differ* **12 Suppl 2**, 1473-1477 (2005).
292. Oeckinghaus, A. & Ghosh, S. The NF-kappaB family of transcription factors and its regulation. *Cold Spring Harb Perspect Biol* **1**, a000034 (2009).
293. Kawasaki, T. & Kawai, T. Toll-like receptor signaling pathways. *Front Immunol* **5**, 461 (2014).
294. Jain, A., Kaczanowska, S. & Davila, E. IL-1 Receptor-Associated Kinase Signaling and Its Role in Inflammation, Cancer Progression, and Therapy Resistance. *Front Immunol* **5**, 553 (2014).
295. Jones, R.M., Mercante, J.W. & Neish, A.S. Reactive oxygen production induced by the gut microbiota: pharmacotherapeutic implications. *Curr Med Chem* **19**, 1519-1529 (2012).
296. Lei-Leston, A.C., Murphy, A.G. & Maloy, K.J. Epithelial Cell Inflammasomes in Intestinal Immunity and Inflammation. *Front Immunol* **8**, 1168 (2017).
297. Davis, B.K., *et al.* Emerging significance of NLRs in inflammatory bowel disease. *Inflamm Bowel Dis* **20**, 2412-2432 (2014).
298. Pott, J. & Stockinger, S. Type I and III Interferon in the Gut: Tight Balance between Host Protection and Immunopathology. *Front Immunol* **8**, 258 (2017).
299. Tamura, T., Yanai, H., Savitsky, D. & Taniguchi, T. The IRF family transcription factors in immunity and oncogenesis. *Annual review of immunology* **26**, 535-584 (2008).
300. Kisseleva, T., Bhattacharya, S., Braunstein, J. & Schindler, C.W. Signaling through the JAK/STAT pathway, recent advances and future challenges. *Gene* **285**, 1-24 (2002).
301. Chinetti, G., Fruchart, J.C. & Staels, B. Peroxisome proliferator-activated receptors (PPARs): nuclear receptors at the crossroads between lipid metabolism and inflammation. *Inflamm Res* **49**, 497-505 (2000).
302. Hasan, A.U., Rahman, A. & Kobori, H. Interactions between Host PPARs and Gut Microbiota in Health and Disease. *Int J Mol Sci* **20**(2019).
303. Gonzalez, F.J., Jiang, C. & Patterson, A.D. An Intestinal Microbiota-Farnesoid X Receptor Axis Modulates Metabolic Disease. *Gastroenterology* **151**, 845-859 (2016).
304. Evans, R.M. & Mangelsdorf, D.J. Nuclear Receptors, RXR, and the Big Bang. *Cell* **157**, 255-266 (2014).
305. Li, Y.C., Chen, Y. & Du, J. Critical roles of intestinal epithelial vitamin D receptor signaling in controlling gut mucosal inflammation. *J Steroid Biochem Mol Biol* **148**, 179-183 (2015).
306. Bock, K.W. Human and rodent aryl hydrocarbon receptor (AHR): from mediator of dioxin toxicity to physiologic AHR functions and therapeutic options. *Biol Chem* **398**, 455-464 (2017).