
Supplementary information

**A designed photoenzyme for
enantioselective [2+2] cycloadditions**

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Supplementary Information

A Designed Photoenzyme for Enantioselective [2+2]-Cycloadditions

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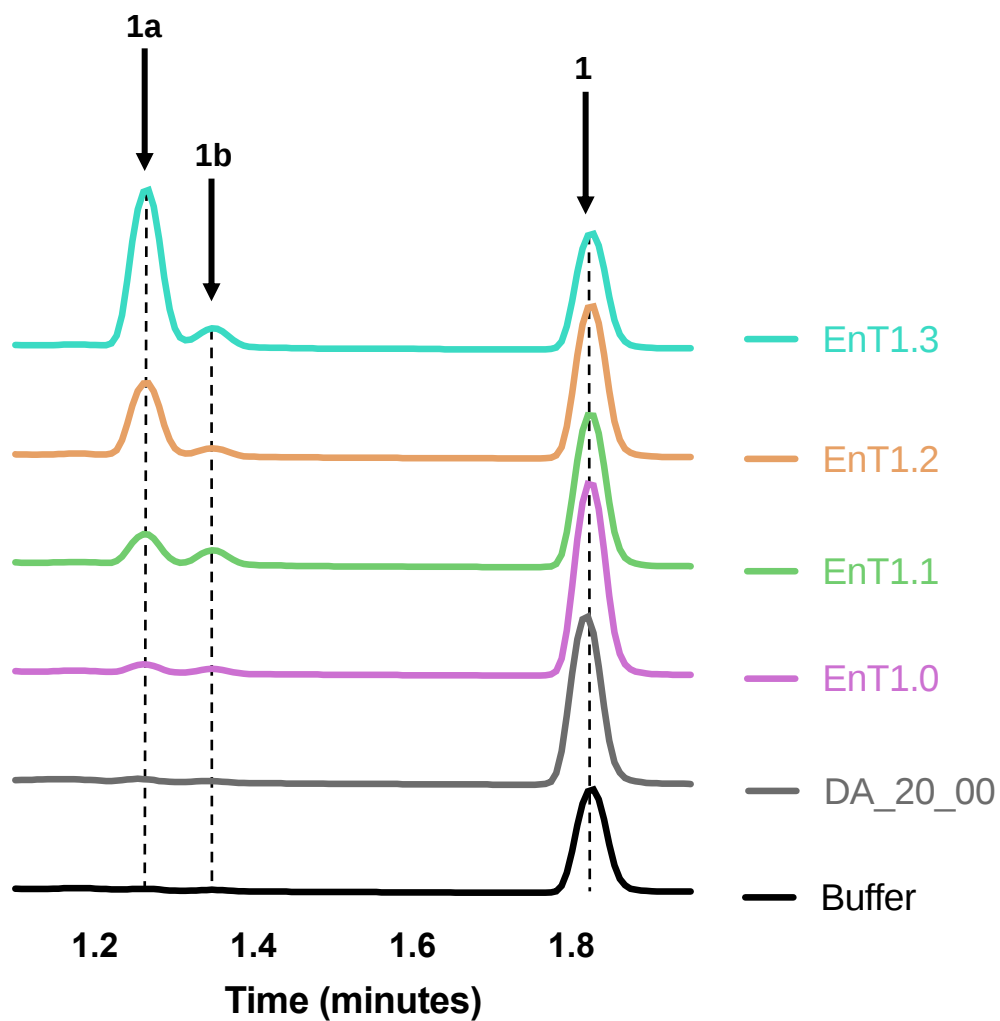
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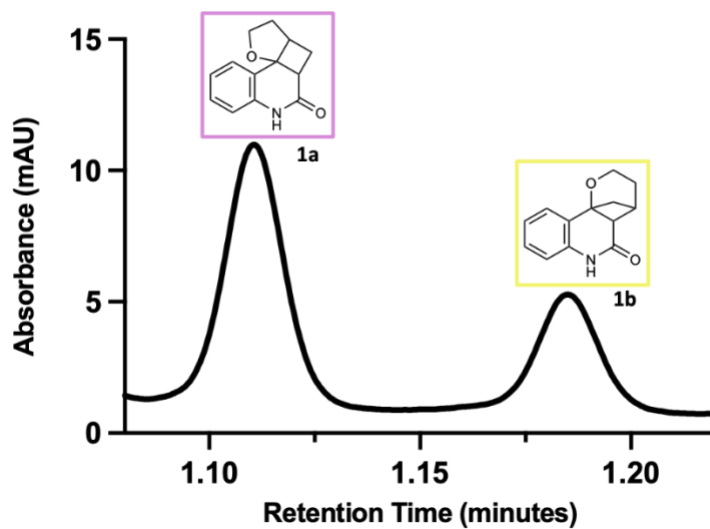
5.213	5.304	5.477	5.330	5.346	5.299	5.326	5.019	5.006	5.040	4.708	4.637
5.309	5.529	5.448	5.233	5.519	5.336	5.217	5.171	5.202	5.134	5.168	5.130
5.488	5.459	5.294	5.555	5.542	5.325	5.290	5.457	5.322	5.124	4.643	4.631
5.354	5.454	5.435	5.306	5.475	5.462	5.452	5.289	5.226	5.185	4.821	4.686
5.366	5.526	5.580	5.456	5.521	5.380	5.420	5.393	5.156	5.178	4.785	4.924
5.397	5.333	5.410	5.451	5.483	5.422	5.431	5.416	5.135	5.121	5.023	5.003
5.250	5.251	5.491	5.467	5.183	5.626	5.428	5.310	5.132	5.175	5.030	4.788
5.213	5.266	5.403	5.405	5.501	5.444	5.230	5.179	5.258	4.863	5.194	5.250

Supplementary Figure 1: Uniformity of light source. Uniformity of light irradiation of a UV curing LED oven chamber (NovaChem) was investigated by monitoring conversions across a 96-well plate. Reaction conditions: 7.5 μM EnT1.0, 400 μM **1**, 30 minutes irradiation (10 seconds on/off pulse) at 365 nm, at 4 $^{\circ}\text{C}$, in 100 μL PBS (pH = 7.4) with 5% DMSO as a cosolvent. Conversions (**1a** and **1b** combined) are shown as a matrix, correlating to their respective plate positions. The relative coefficient of variance is 4.3%.

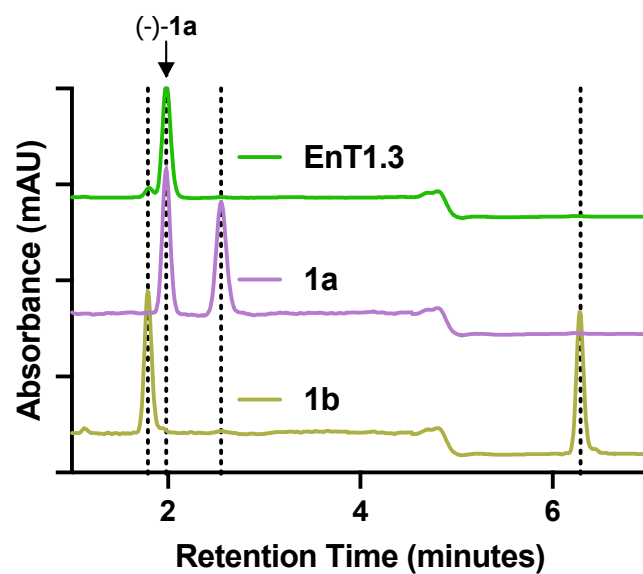


Supplementary Figure 2: UPLC chromatograms of evolution trajectory. UPLC absorbance chromatograms at 260 nm for reactions containing no enzyme, DA_20_00, EnT1.0, EnT1.1, EnT1.2 or EnT1.3. Reaction conditions: 3 μ M catalyst, 400 μ M **1**, 60 minutes irradiation (10 seconds on/off pulse) at 365 nm, at 4 $^{\circ}$ C, in 100 μ L PBS (pH = 7.4) with 5% DMSO as a cosolvent.

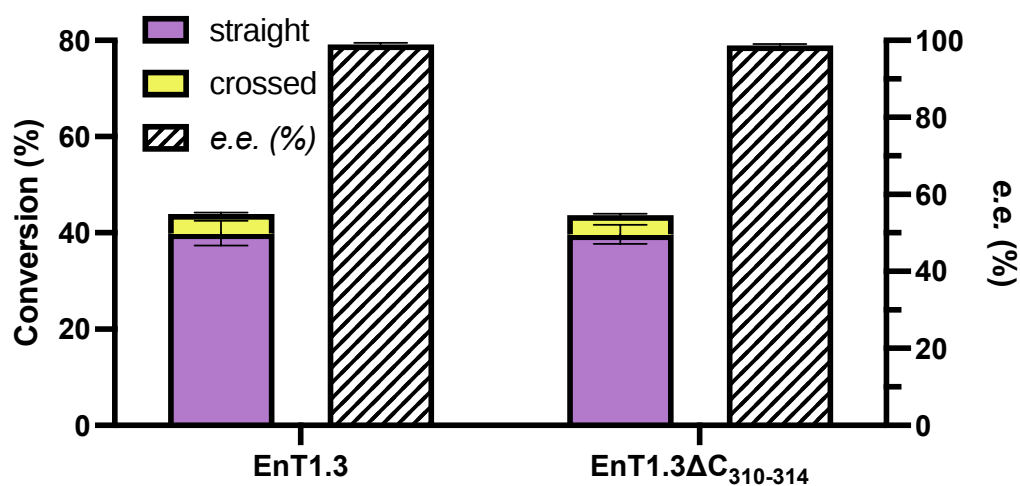
**UPLC trace of EnT1.0 derived
[2+2]-cycloaddition products**



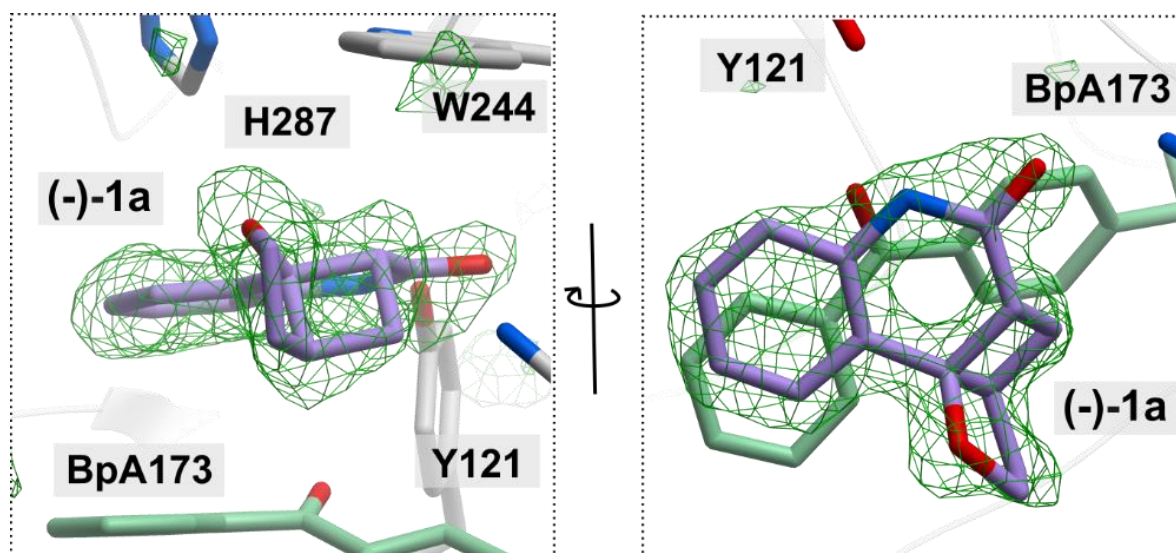
Supplementary Figure 3: UPLC chromatogram showing products derived from reactions with EnT1.0. UPLC chromatogram at 260 nm showing product **1a** and product **1b** (2:1 ratio). Reaction conditions: 7.5 μM EnT1.0, 400 μM **1**, 30 minutes irradiation (10 seconds on/off pulse) at 365 nm, at 4 $^{\circ}\text{C}$, in 100 μL PBS (pH = 7.4) with 5% DMSO as a cosolvent.



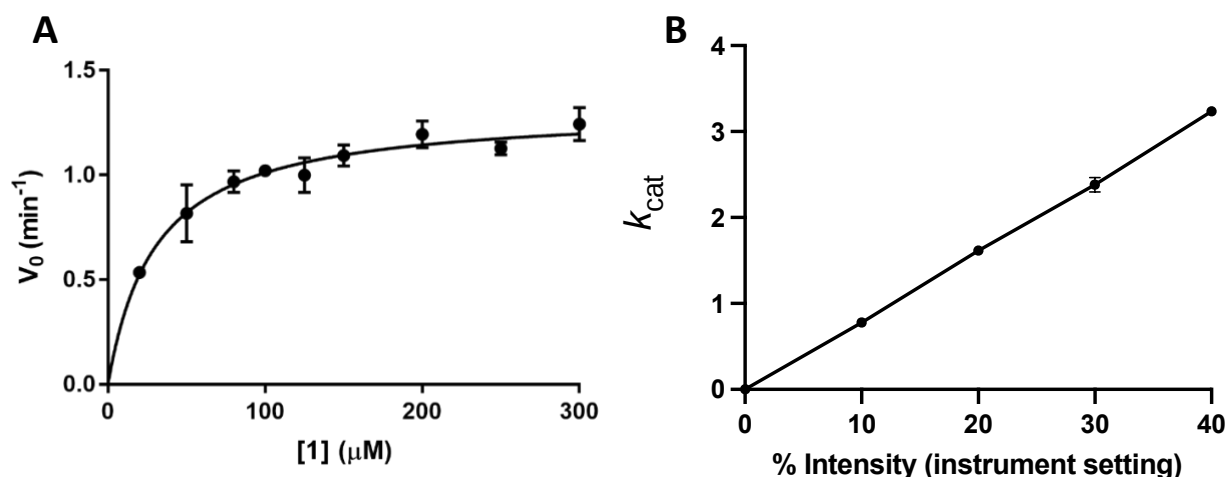
Supplementary Figure 4: SFC chromatograms of racemic product standards and the product of EnT1.3-catalysed reaction using substrate 1. Reaction conditions: 10 μ M EnT1.3, 400 μ M **1**, 60 minutes irradiation (10 seconds on/off pulse) at 365 nm, at 4 $^{\circ}$ C, in 100 μ L PBS (pH = 7.4) with 5% DMSO as a cosolvent. For SFC analysis, reactions were extracted with 3 volumes of ethyl acetate.



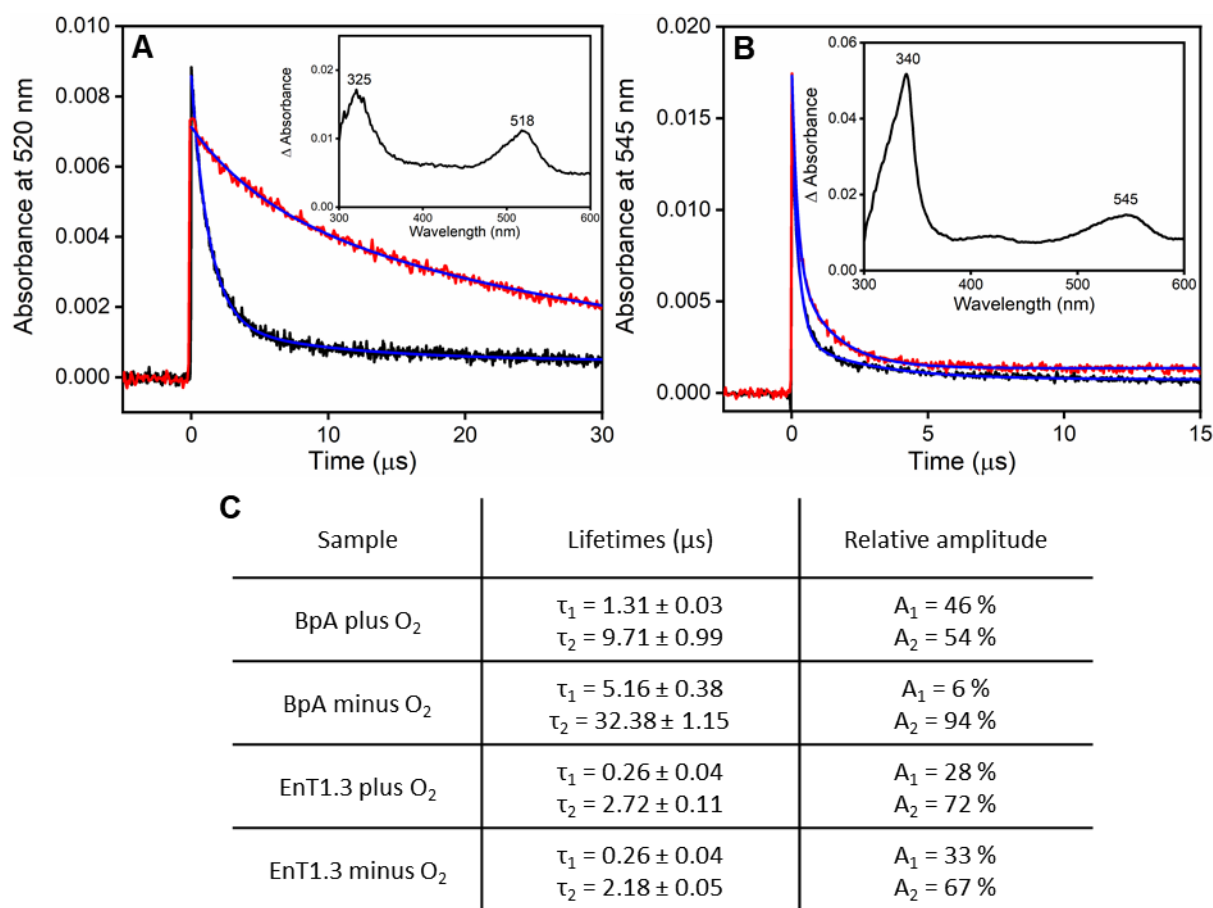
Supplementary Figure 5: A comparison of the activity and selectivity of EnT1.3 and EnT1.3ΔC₃₁₀₋₃₁₄. Reaction conditions: 10 μM catalyst, 400 μM **1**, 15 minutes irradiation (10 seconds on/off pulse) at 365 nm, at 4 °C, PBS (pH = 7.4) with 5% DMSO as a cosolvent. The error bars represent the standard deviation of measurements made in triplicate.



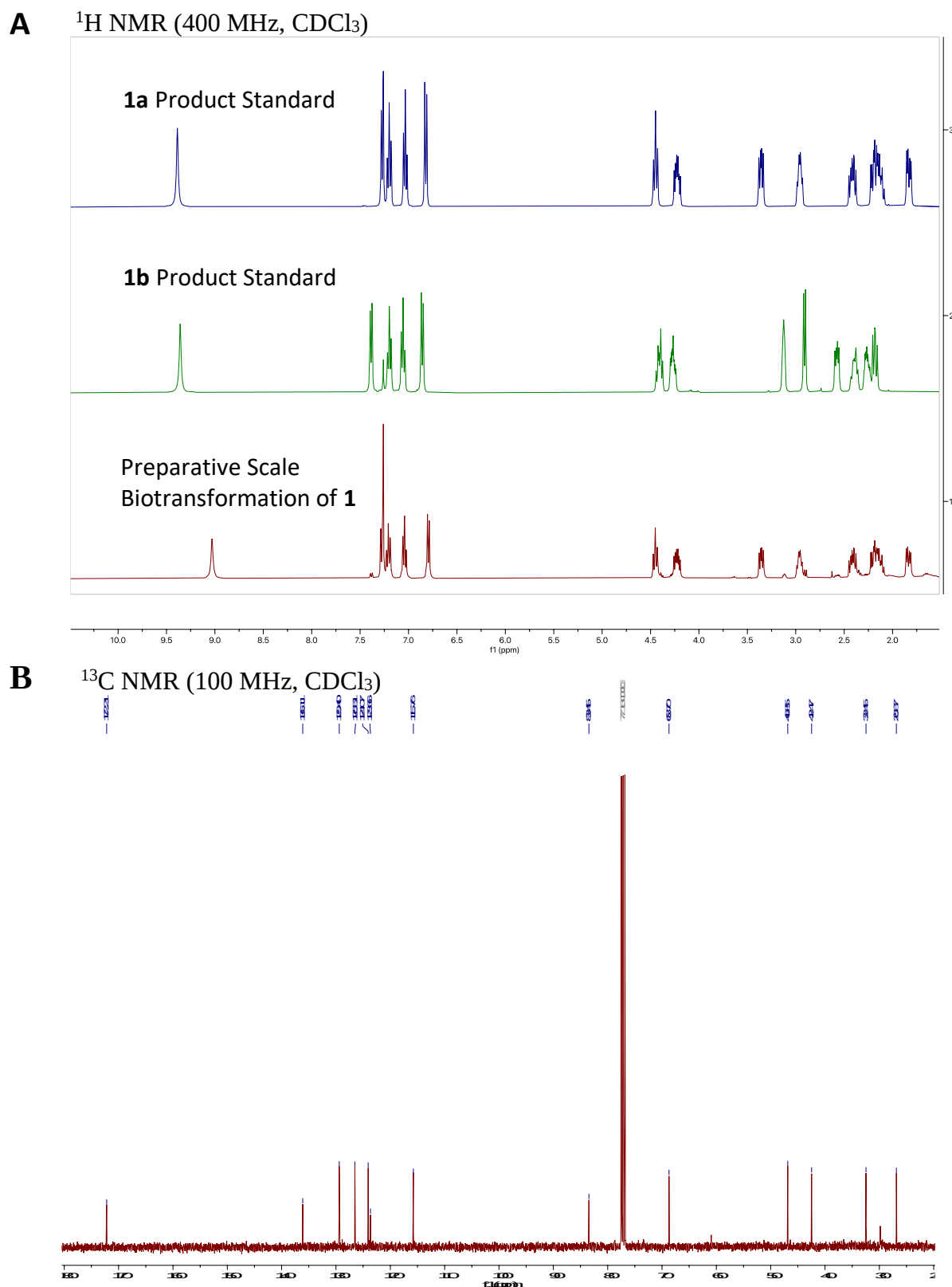
Supplementary Figure 6: The active site of EnT1.3ΔC₃₁₀₋₃₁₄ in complex with the product (-)-1a. The $F_o - F_c$ omit density corresponding to (-)-1a is contoured at 3σ (green mesh). The product, BpA173 and active site residues are shown as atom-coloured sticks (with carbons shown in purple, green, and grey, respectively).



Supplementary Figure 7: Kinetic characterisation of EnT1.3. A) Michaelis-Menten plot for the intramolecular [2+2]-cycloaddition of substrate **1** catalysed by EnT1.3. Kinetic assays were performed at various concentrations of **1**, and $1 \mu\text{M}$ of EnT1.3 in $250 \mu\text{L}$ PBS (pH 7.4) with 5% DMSO as a co-solvent in 96-well plate. The plots show the averaged initial rates which were fitted to the Michaelis-Menten equation using Origin software. Data are a mean $\pm$ S.D. of measurements made in triplicate. B) The dependence of EnT1.3 v_0 on light intensity. Assays were performed with $400 \mu\text{M}$ of substrate **1** and $1 \mu\text{M}$ of EnT1.3 in $500 \mu\text{L}$ PBS (pH 7.4) with 5% DMSO as a co-solvent in 1 mL glass vials. Intensity was varied from 0%-40% (based on instrument settings) with the reactions irradiated 3 cm directly below the LED array.



Supplementary Figure 8: Photophysical properties of BP and EnT1.3. Kinetic transients showing the decay of the benzophenone triplet state on the μ s timescale in the presence (black) and absence (red) of oxygen for A) 50 μ M benzophenone with 5% DMSO as a co-solvent and B) 50 μ M EnT1.3 in PBS (pH 7.4). Transients were measured at 4°C at the respective wavelength and data shown are the average of 50 traces. Data were fitted to a double exponential equation to obtain lifetimes (blue lines). The insets show the absorbance difference spectra of the triplet under the same conditions using a CCD gate width of 100 ns. C) Kinetic parameters determined from double exponential fits shown in A and B.



Supplementary Figure 9: Preparative 12 mg Scale Biotransformation of Substrate 1. A) ^1H NMR spectra (400 MHz, CDCl_3) of **1a** and **1b** chemically synthesised product standards, and crude product extracted from the EnT1.3 (10 μM) catalysed intramolecular [2+2]-cycloaddition of substrate **1** (400 μM) in PBS (pH 7.4) with 5% DMSO as a co-solvent (140 mL reaction volume). B) ^{13}C NMR spectra (100 MHz, CDCl_3) of the crude product of the biotransformation.

Supplementary Information, Table 1: Data summary of conversion and selectivity values of EnT1.0. Reaction conditions: 15 μ M catalyst, 400 μ M **1**, 30 minutes irradiation (10 seconds on/off pulse) at 365 nm, at 4 °C, PBS (pH = 7.4) with 5% DMSO as a cosolvent. Standard deviations are given for measurements in triplicate.

Catalyst	Average Conversion Straight (%)	S.D.	Average Conversion Crossed (%)	S.D.	<i>e.e.</i>	S.D.
-	0.33	0.031	0.19	0.079	-	-
Benzophenone	4.5	0.40	3.1	0.29	-	-
DA_20_00	0.55	0.21	0.32	0.044	-	-
EnT1.0	9.0	0.75	4.4	0.19	46	1.53
EnT1.0 ^y	0.0	-	0.0	-	-	-

[^y] Reaction was performed without exposure to 365 nm light.

Supplementary Information, Table 2: Data summary of conversion values of EnT1.0 and EnT1.3 for Extended Data Figure 4. Reaction conditions: 15 μM catalyst, 400 μM **1**, 10 seconds on/off pulse at 365 nm, 4 $^{\circ}\text{C}$, PBS (pH 7.4) with 5% DMSO as a cosolvent. At each time point, 100 μL of reaction mix was quenched with 1 volume MeCN and analysed by UPLC. Standard deviations are given for measurements in triplicate.

	EnT1.3		EnT1.0	
Time (minutes)	Average Conversion (%)	S.D.	Average Conversion (%)	S.D.
0.0	0.0	-	0.0	-
1.0	4.35	0.07	1.34	0.05
2.0	8.77	0.42	2.38	0.11
3.0	12.76	0.45	3.54	0.17
4.0	17.13	0.55	4.55	0.07
5.0	21.34	0.74	5.49	0.43

Supplementary Information, Table 3: Substrate scope of EnT variants. Reaction conditions for the synthesis of **1a-12a** and **1b, 2b** and **12b**, conversion is displayed as a total of major and minor product where appropriate. All reactions were performed using PBS buffer pH 7.4 and 5% DMSO in triplicate. Standard deviations are given for measurements in triplicate. Reaction conversions are determined based on UPLC calibration curves of authentic standards of starting materials and products. Extinction coefficients and UPLC methods are given in Supplementary Table S8. The *e.e.* values are determined using chiral SFC or HPLC analysis. Chromatograms of substrate standards, product standards, and biotransformations are provided at the end of the Supplementary Information.

Substrate	Product	Variant	Catalyst loading (mol%)	Substrate loading (mM)	Time (hours)	Conversion (%)	S.D.	<i>e.e.</i>
1	1a, 1b	EnT1.3	1.5	0.4	2 ^[1]	94	0.70	99
1	1a, 1b	EnT1.3	2.5	0.4	1 ^[1]	100	0.07	99
1	1a, 1b	EnT1.3 BpA173Ala + BP	2.5	0.4	1 ^[1]	4.2	0.07	12
2	2a, 2b	EnT1.3	10	0.3	4 ^[1]	99	0.29	90
3	3a	EnT1.2	7.5	0.2	3 ^[1]	76	0.95	80
4	4a	EnT1.3	5	0.5	2 ^[1]	98	0.39	92
5	5a	EnT1.3	7.5	0.4	4 ^[2] ^[+]	95	2.37	82
6	6a	EnT1.3	5	0.4	3 ^[2]	100	N.D.	95
7	7a	EnT1.3	10	0.4	3 ^[2]	95	0.31	94
8	8a	EnT1.2	7.5	0.3	4 ^[2]	98	0.5	26
8	8a	EnT1.1 A229S Y37L	7.5	0.3	4 ^[2]	98	0.87	59
9	9a	EnT1.3	5	0.4	3 ^[2]	100	N.D.	97
10	10a	EnT1.3	7.5	0.5	3 ^[2] ^[+]	89	0.46	87
11	11a	EnT1.3	7.5	0.5	3 ^[2] ^[+]	92	0.71	81
12	12a	EnT1.2	7.5	0.5	4 ^[2] ^[+]	89	0.9	48
13	13a, 13b	EnT1.3	7.5	1	3 ^[1] ^[+]	69	0.57	36

13	13a, 13b	EnT1.3 Y121F	7.5	1	3 ^{[1][†]}	94	0.32	78
13	13a, 13b	EnT1.3 Y121F S271C	7.5	1	3 ^{[1][*]}	99	0.05	98
2-Quinolone + methyl vinyl ketone (MVK)	14	EnT1.3	5	2 mM quinolone 50 mM MVK	4 ^[2]	71	0.81	97
2-Quinolone + Ethyl vinyl ketone (EVK)	15	EnT1.3	5	2 mM quinolone 50 mM EVK	4 ^[2]	73	0.1	97

[1] 365nm irradiation wavelength

[2] 395nm irradiation wavelength

[†] LED intensity was set to 50% to minimise background reactions.

[*] LED intensity was set to 40% to minimise background reactions.

Supplementary Information, Table 4: Protein crystallization conditions.

Variant	Mother liquor composition
EnT1.0	0.2 M Ammonium sulfate, 0.1 M BIS-Tris, pH = 5.5, 25% w/v PEG 3350 JCSG condition H7
EnT1.3	0.2 M Ammonium chloride, 0.1 M HEPES, pH = 7.0, 20% w/v PEG 6000 PACT premier Eco condition C8
EnT1.3 Δ C ₃₁₀₋₃₁₄	0.2 M Magnesium formate dihydrate, 20% w/v PEG 3350 JCSG condition A5

Supplementary Information, Table 5: Data collection and refinement statistics

	EnT1.0	EnT1.3	EnT1.3ΔC:product
PDB ascension number	7ZP5	7ZP6	7ZP7
Wavelength (Å)	0.9763	0.9762	0.9763
Resolution range	81.57 - 1.54 (1.595 - 1.54) ^a	54.66 - 1.9 (1.968 - 1.9) ^a	74.59 - 1.7 (1.761 - 1.7) ^a
Space group	C 1 2 1	P 2 ₁ 2 ₁ 2 ₁	P 1 2 ₁ 1
Unit cell dimensions a, b, c, (Å) α, β, γ (°)	173.847, 43.397, 81.7644, 90, 93.9843, 90	50.894, 75.5961, 78.5996 90, 90, 90	50.7517, 73.9581, 74.7216 90, 93.4574, 90
Total reflections	585059 (35012)	329486 (33095)	417082 (35910)
Unique reflections	89549 (8084)	24554 (2405)	60455 (5861)
Multiplicity	6.5 (4.3)	13.4 (13.8)	6.9 (6.1)
Completeness (%)	98.84 (89.62)	99.96 (99.92)	99.64 (97.25)
Mean I/sigma(I)	17.22 (0.67)	8.95 (1.81)	15.85 (2.08)
Wilson B-factor	23.72	18.41	16.81
R-merge	0.05761 (0.8321)	0.2025 (0.873)	0.08916 (0.4921)
R-meas	0.06245 (0.9503)	0.2105 (0.9067)	0.09642 (0.5379)
R-pim	0.02381 (0.448)	0.05688 (0.2423)	0.03641 (0.2143)
CC _{1/2}	0.998 (0.801)	0.996 (0.872)	0.997 (0.876)
CC*	1 (0.943)	0.999 (0.965)	0.999 (0.966)
Reflections used in refinement	89457 (8058)	24547 (2403)	60431 (5860)
Reflections used for R-free	4542 (449)	1239 (123)	3069 (264)
R-work	0.1959 (0.3637)	0.1690 (0.2104)	0.1508 (0.1964)
R-free ^b	0.2231 (0.3768)	0.2133 (0.2776)	0.1807 (0.2400)
CC (work)	0.965 (0.844)	0.962 (0.928)	0.974 (0.922)
CC (free)	0.962 (0.840)	0.942 (0.896)	0.961 (0.874)
Number of non-hydrogen atoms	5359	2840	5621
macromolecules	4901	2566	4883
ligands	10	0	68
solvent	448	274	702
Protein residues	617	321	614
RMS (bonds)	0.010	0.005	0.020
RMS (angles)	1.00	0.90	0.87
Ramachandran favoured (%)	96.71	97.15	96.69
Ramachandran allowed (%)	3.29	2.85	3.31
Ramachandran outliers (%)	0.00	0.00	0.00
Rotamer outliers (%)	0.00	0.37	0.58
Clashscore	2.26	4.93	3.71
Average B-factor	31.98	21.37	21.07
macromolecules	31.42	20.56	19.77
ligands	48.84	-	30.61
Solvent	37.75	28.96	29.67
Number of TLS groups	1	1	1

^aValues in parentheses are for highest resolution shell. ^bR-free was calculated using ~5% of the data separate from the rest

Supplementary Information, Table 6: Cosolvent tolerance of EnT1.3. Reaction conditions: 7.5 μ M EnT1.3, 400 μ M **1**, 10 seconds on/off pulse at 365 nm, 4 °C, PBS (pH 7.4) with the stated cosolvents. Standard deviations are given for measurements in triplicate.

Co-solvent	Average Conversion Straight (%)	S.D.	Average Conversion Crossed (%)	S.D.	<i>e.e.</i>
5% DMSO	33.6	0.5	3.6	0.0	99
10% DMSO	31.1	1.7	3.4	0.2	98
15% DMSO	27.8	0.5	3.1	0.1	98
20% DMSO	24.0	0.3	2.9	0.2	98
25% DMSO	17.8	0.3	2.3	0.0	93
30% DMSO	13.1	0.5	1.8	0.1	90
5% MeCN	28.4	0.8	3.0	0.0	99
10% MeCN	27.8	0.3	3.0	0.1	98
15% MeCN	25.3	0.5	3.0	0.2	94
20% MeCN	16.4	1.8	2.1	0.2	91
25% MeCN	6.3	1.3	1.1	0.1	N/A
30% MeCN	1.4	0.4	0.4	0.1	N/A

Supplementary Information, Table 7: Mass spectrometry data for DA_20_00 and EnT1.0 variants.

Variant	Expected Mass	Observed Mass
DA_20_00	36012.98	36012.6
DA_20_00 G29BpA	36207.2	36206.4
DA_20_00 D30BpA	36149.17	36148.4
DA_20_00 L148BpA	36151.07	36150.8
DA_20_00 Q195BpA	36136.17	36135.8
EnT1.0	36193.19	36193.2
EnT1.0 Y37A	36101.08	36101.3
EnT1.0 I72A	36151.08	36153.05
EnT1.0 M90A	36133.08	36133.2
EnT1.0 P135A	36167.1	36167.0
EnT1.0 I148A	36151.08	36152.94
EnT1.0 Q195A	36136.18	36137.69
EnT1.0 W244A	36078.08	36080.19
EnT1.0 H287A	36127.08	36128.97
EnT1.1 A229S Y37L	36098.85	36098.57
EnT1.2	36136.08	36135.3
EnT1.3	36196.08	36195.8
EnT1.3 Y37F	36180.08	36178.89
EnT1.3 Y121F	36180.08	36179.17
EnT1.3 Q195A	36139.07	36137.99
EnT1.3 S229A	36180.08	36178.99
EnT1.3 W244A	36080.97	36079.32
EnT1.3 H287A	36129.97	36127.63
EnT1.3 Y121F S271C	36195.86	36194.54
EnT1.3 Y121F Y37L	36129.82	36130.18

Supplementary Information, Table 8: Analytical UPLC methods for products 1a-12a and 1b, 2b 12b.

Substrate	Flow (mL.min ⁻¹)	Mobile Phase (% MeCN in MQ H ₂ O with 0.1% TFA) ^[1]	Run Time (min)	Extinction Coefficient (mM ⁻¹ cm ⁻¹) ^[2]	
				substrate	product(s)
1	1.1	5-40% (1.8 min) 5% (1.81 – 2.2 min)	2.2	747 (1)	718 (1a) 693 (1b)
2	1	20% (2 min) 20-40% (2.1 min) 40% (3 min) 40-20% (3.1 min) 20% (4 min)	4	800 (2)	883 (2a) 491 (2b)
3	1	30%	4	306 (3)	628 (3a)
4	1	17%	5.5	812 (4)	935 (4a)
5	1.2	15% (0.3 min) 15-40% (2.7 min) 15% (1min)	4	212 (5)	310 (5a)
6	1.5	17%	3	804 (6)	1241 (6a)
7	1.5	20%	3.5	297 (7)	929 (7a)
8	1	25%	3.5	592 (8)	1195 (8a)
9	1.8	25%	2.5	522 (9)	963 (9a)
10	1.5	27%	4	452 (10)	671 (10a)
11	1	37%	2.5	796 (11)	1039 (11a)
12	1	30%	3	651 (12)	502 (12a)
13	1	25% (2 min) 25-50% (2.05 min) 50% (3 min) 50-25% (3.01 min) 25% (4 min)	4	283 (12)	496 (13a) 488 (13b)
Quinolone + MVK	1	15%	1.8	775	873 (14)
Quinolone + EVK	1	20%	2	775	918 (15)

^[1] Column temperature was set to 20 °C

^[2] Detector wavelength was set to 260nm

Supplementary Information, Table 9: SFC chiral analytical methods for products 1a-12a and 1b, 2b 12b.

Substrate	Flow (mL.min⁻¹)	Mobile Phase (% MeOH in CO₂)	Run Time (min)	Product(s)	Column Temperature (°C)
1	1	25% (4 min) 25-50% (4.01 min) 50% (8 min) 50-25% (8.01 min) 25% (10 min)	10	1a 1b	20
2	0.8	50%	5	2a 2b	20
3	0.8	50%	4	3a	20
4	0.8	25%	6	4a	30
7	1	30%	5	7a	30
9	0.5	30%	6	9a	20
10	1	20%	6	10a	20
11	1	30%	5	11a	30

Supplementary Information, Table 10: Normal phase HPLC chiral analytical methods

Substrate	Flow (mL.min⁻¹)	Mobile Phase (% IPA in Hexane)	Run Time (min)	Product(s)	Column Temperature (°C)
5^[1]	1	10%	12	5a	20
6^[2]	1	2%	25	6a	20
8^[3]	1	6%	18	8a	20
12^[3]	1	10%	25	12a	20
13^[2]	1	2%	22	13a	20
2-Quinolone^[2]	1	10%	6	14	20
2-Quinolone^[2]	1	10%	6	15	20

^[1] CHIRALPAK ® OJ-H (0.46 cm x 25 cm) column used.

^[2] Diacel 87S82 CHIRALPAK ® IG-3 SFC column (3 mm x 50 mm) column used.

^[3] CHIRALPAK ® AD-H (0.46 cm x 25 cm) column used.

Supplementary Information, Table 11: Primers for the alanine scan, rounds 1 & 2.

Flanking Primers	
XHO_R	ATGCATGCCTCGAGCGATCCGA
NDE_F	CATGCATGCATATGGAGATCCCTGTCA
Alanine Scan	
Y37A_F	TATATTGTGGCTCCAGCAGTGGAGGTAAACGGTAAAC
I72A_F	AACGGCTATGGAGGCCACCAAGCCGGGTG
M90A_F	TTATTTGTAGCTGATGACGTCTGGGCCTTCT
P135A_F	TTATGGATTACGGCCGAGCGGGCGAGGT
I146A_F	CCTGCCGATTTTACCGCATCGTTACAAGAAAAATTCGGG
Q195A_F	CTGATTGTGGCCGAGGACCAACTAAAAAGTTATGG
W244A_F	TTGTTAGTTGCCAATGAGGCTCCTCGCATATC
H287A_F	ATTTTCGTGACGGAGGAGAAAAACAATGCAGTCTGG
Y37A_R	TGGAGCCACAATATAAAAGTC
I72A_R	GCCTCCATAGCCGT
M90A_R	ATCAGCTACAAATAACTGGTTG
P135A_R	GGCCGTAATCCATAAGTT
I146A_R	GGTAAAATCGGCAGGG
Q195A_R	CTCGGCCACAATCAG
W244A_R	ATTGGCAACTAACAAATTGTTG
H287A_R	CTCCGTCACGAAAATGG
Round 1	
A74x_F	TATGGAGGCATACCANNKGGGTGCCAGTGC
M90x_F	TTATTTGTAGCTGATNNKCGTCTGGGCCTTCTT
A120x_F	CGTATGCAGGGTTGCGNNKTACTGCGCTTTTGATTATGA
Y121x_F	ATGCAGGGTTGCGCCNNKTGCGCTTTTGATTATGAAG
P135x_F	TTATGGATTACGGCCNNKGCGGGCGAGG
I146x_F	CCTGCCGATTTTACCGNNKTCGTTACAAGAAAAATTCGGG
L148x_F	GATTTTACCATCTCGNNKCAAGAAAAATTCGGGTCA
Q149x_F	TTTACCATCTCGTTANNKGAAAAATTCGGGTCAATCTATTG
A175x_F	GCATTTAGTAGCCANNKGGTATTGCCGTGCG
Q195x_F	CTGATTGTGGCCGAGNNKCCAACTAAAAAGTTATGGTCC
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Primers used engineering EnT1.3 Y121F towards substrate 12	
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S229x_F	CTCACGAGGGTGGAGCCNNKGGTATGGATTTCGAC
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Protein and DNA sequences:

B = 4-benzoylphenylalanine, BpA

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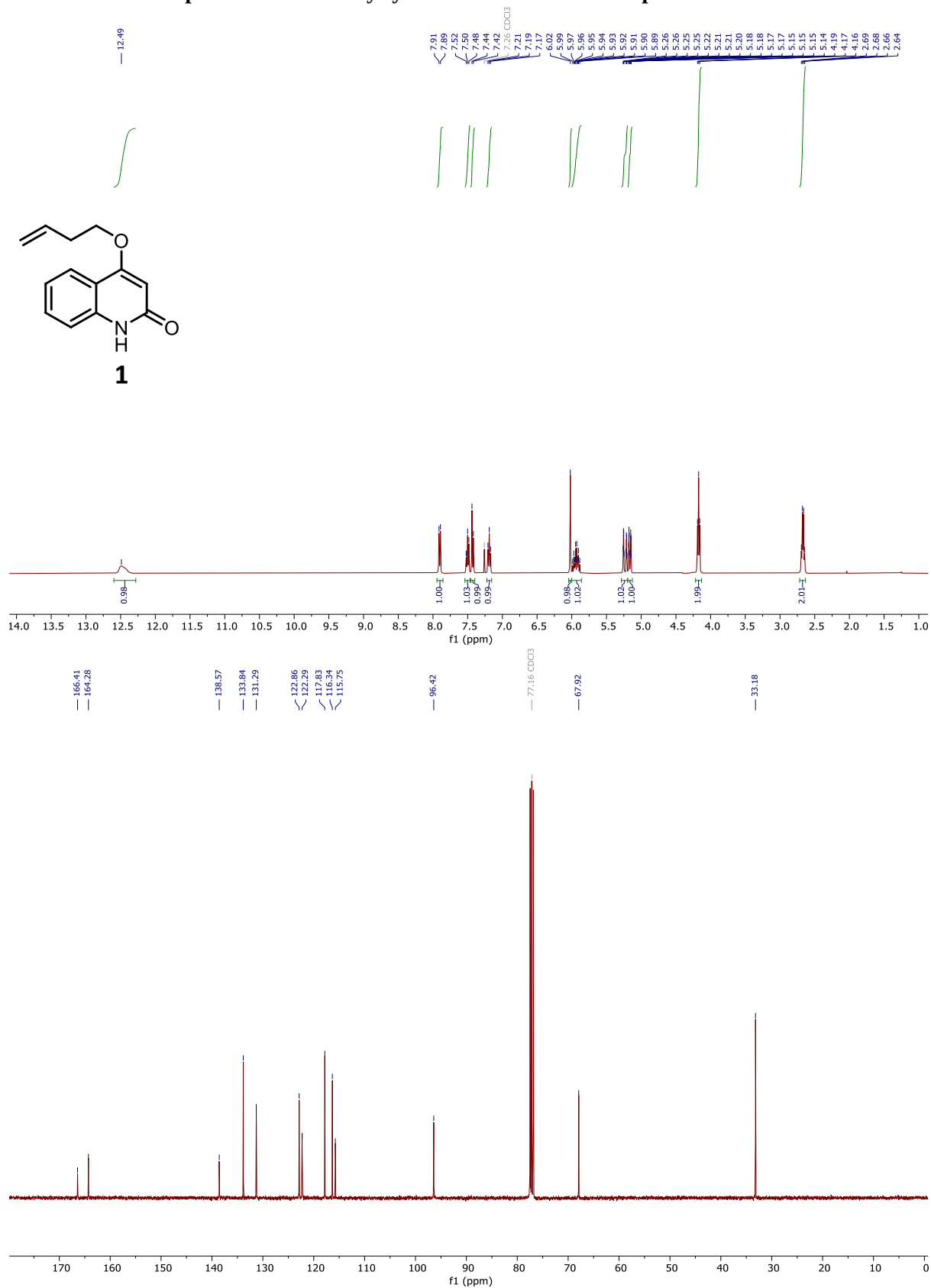
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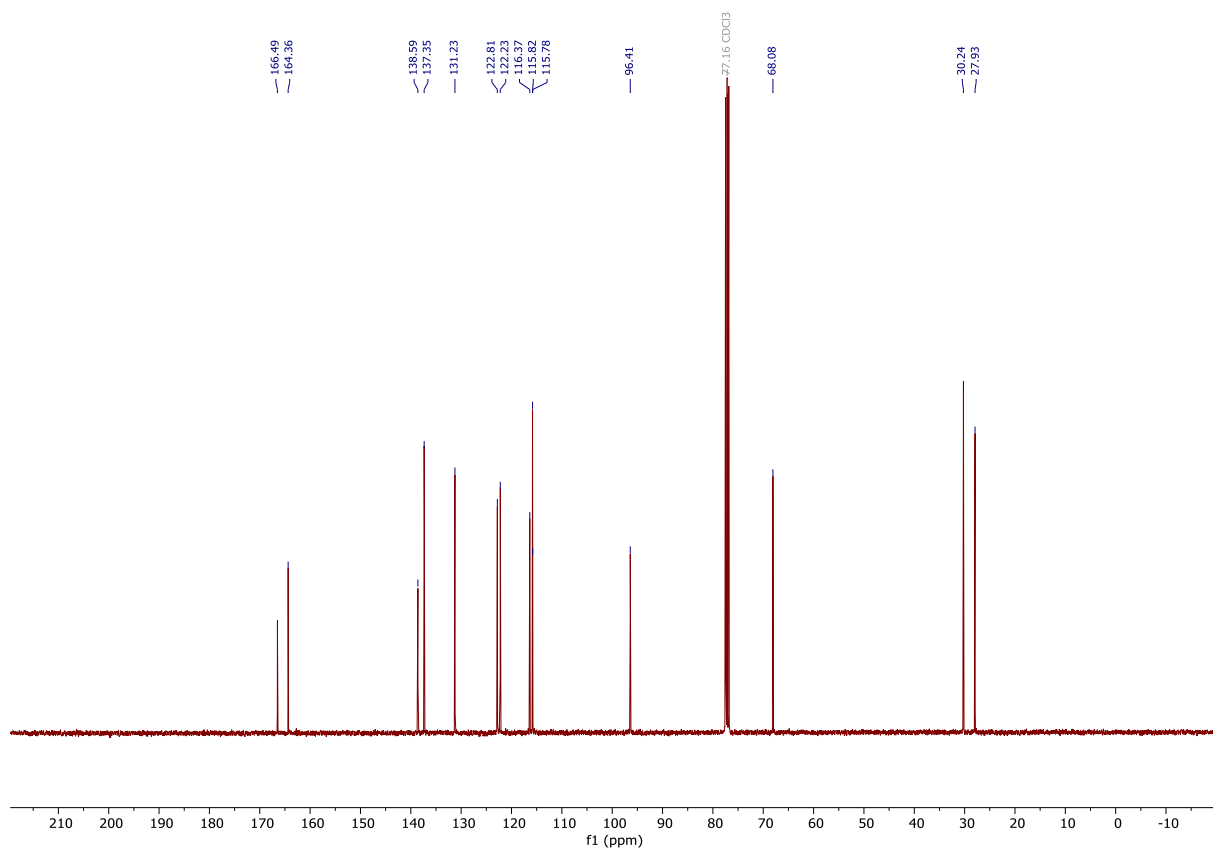
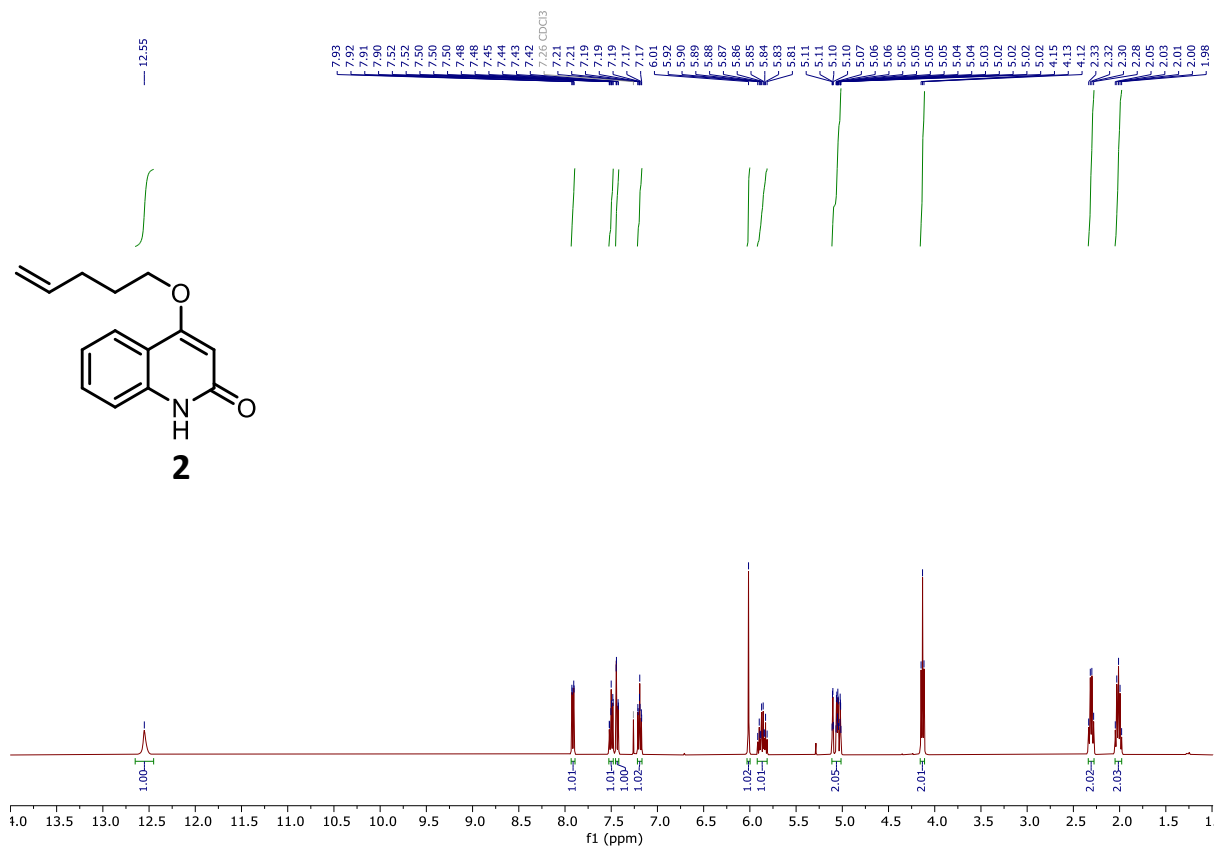
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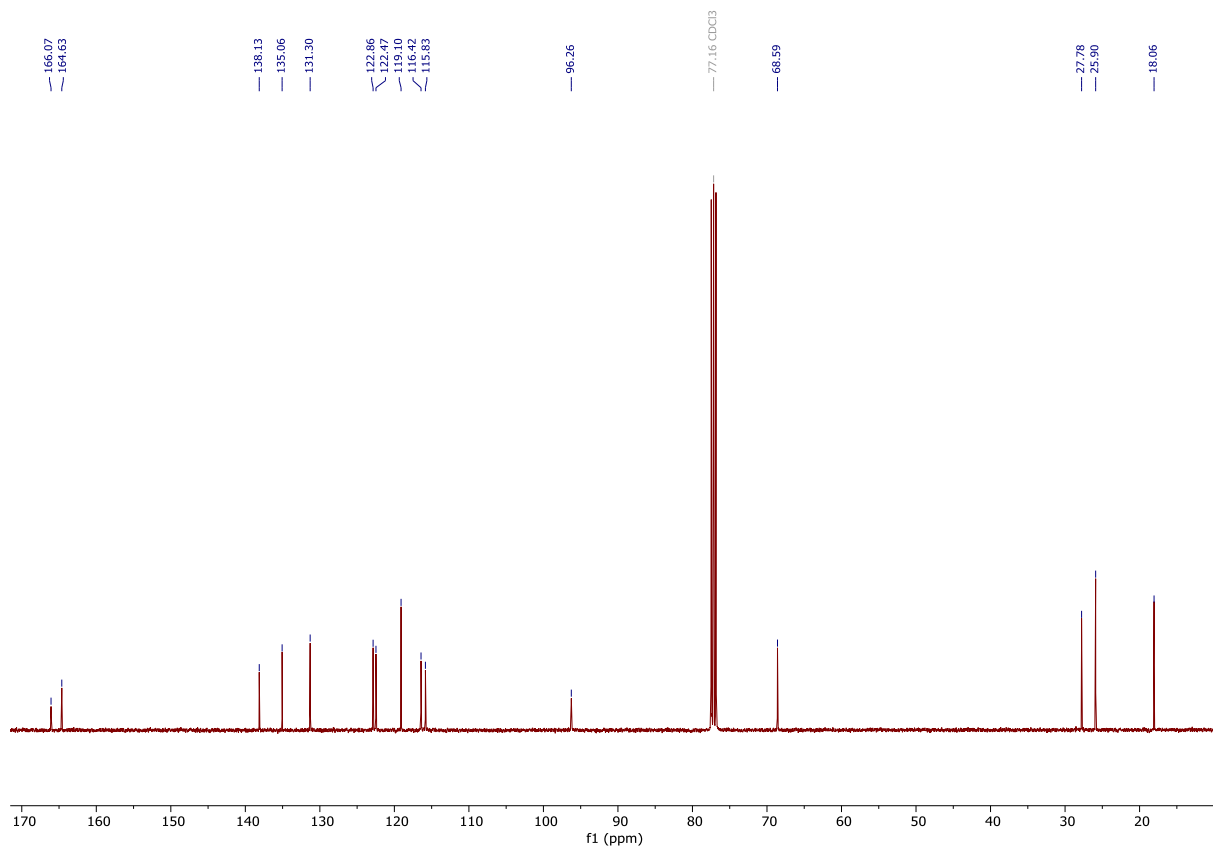
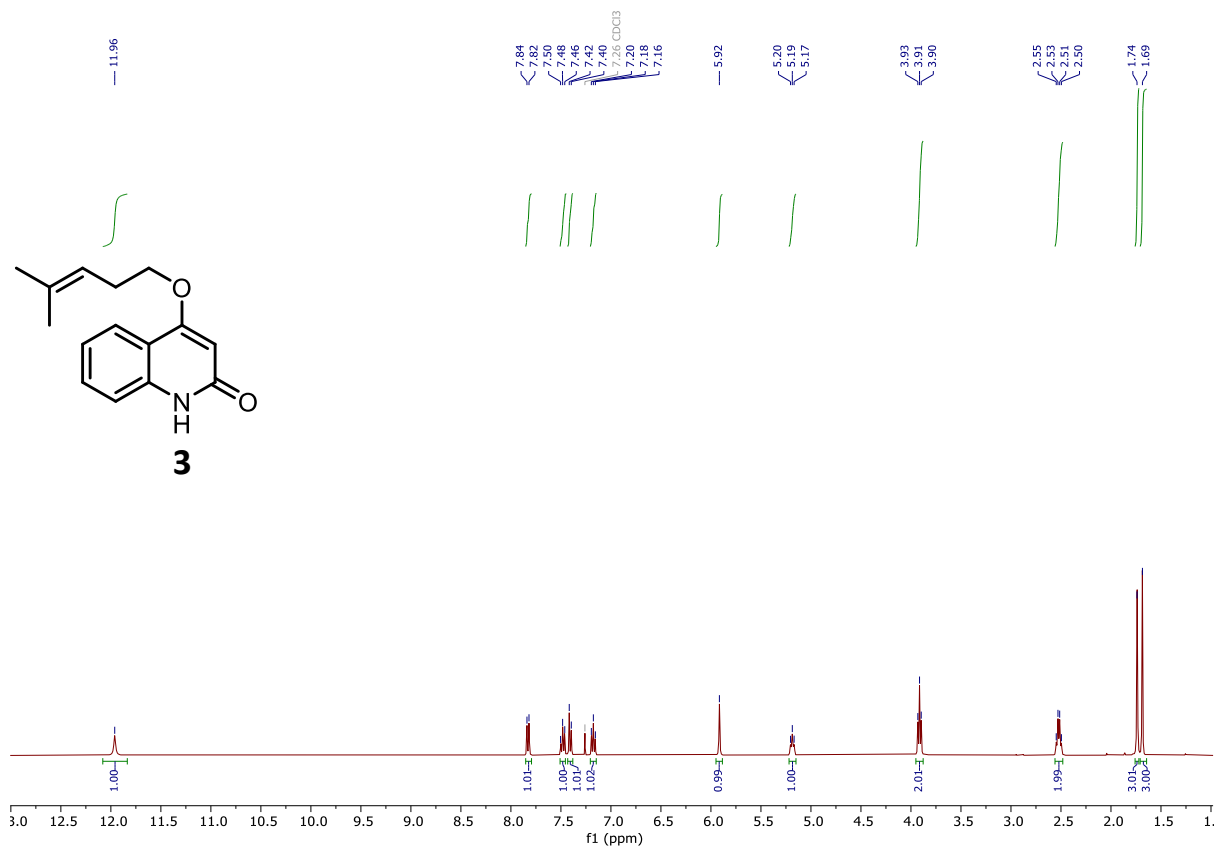
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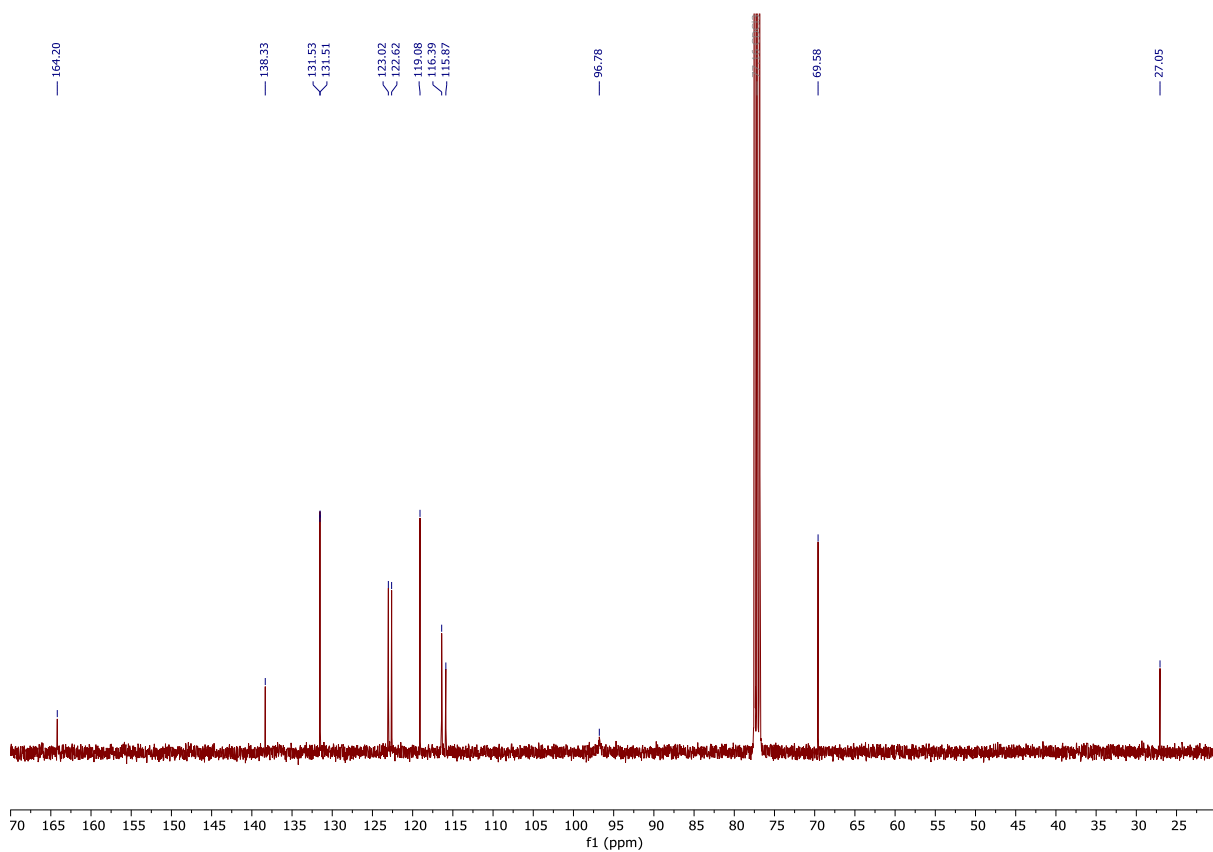
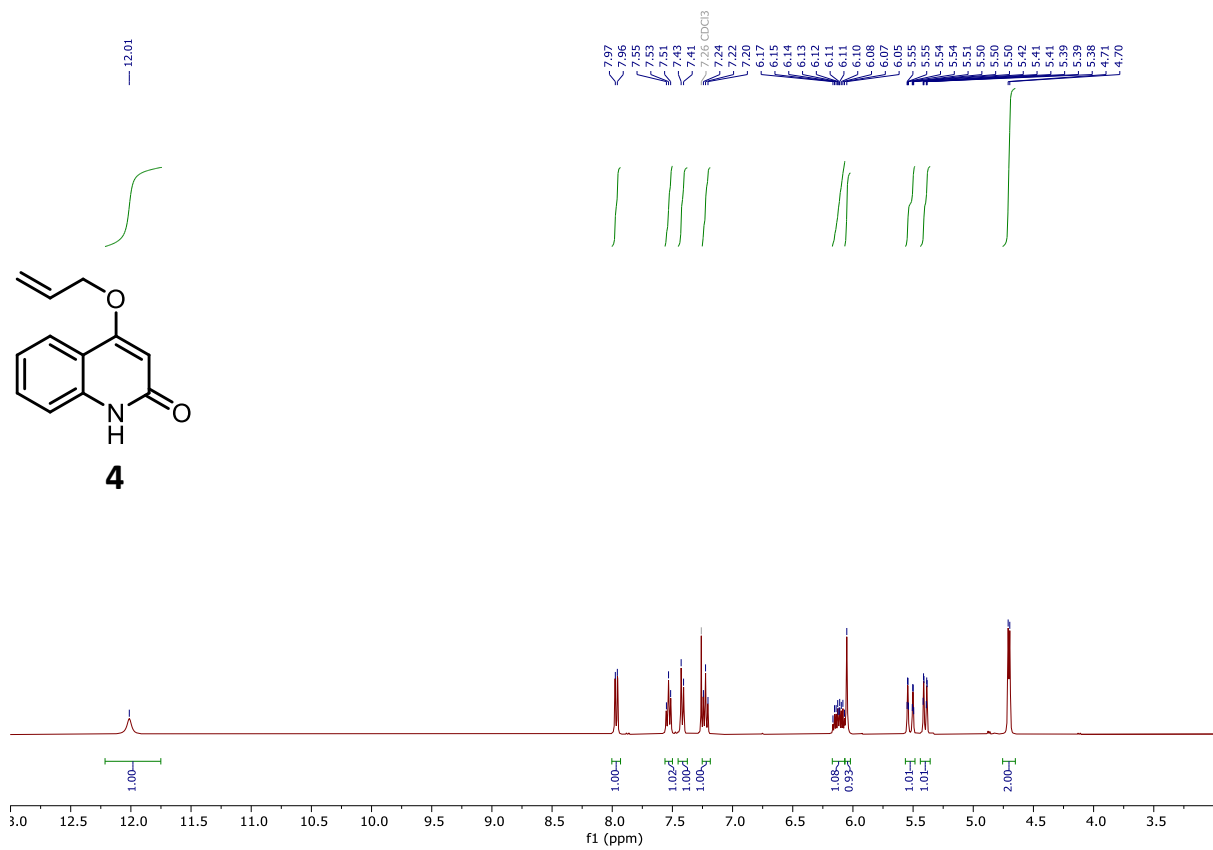
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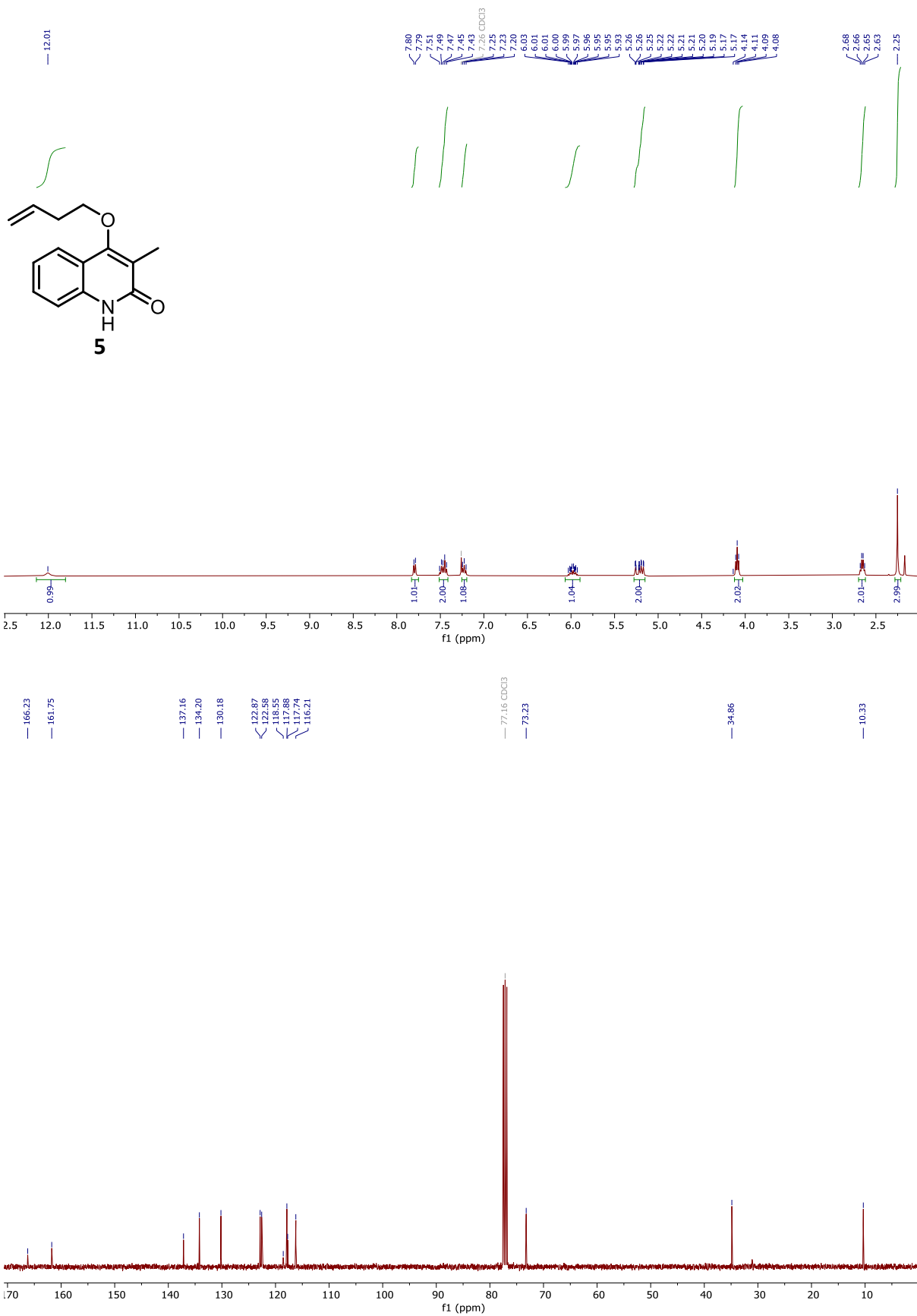
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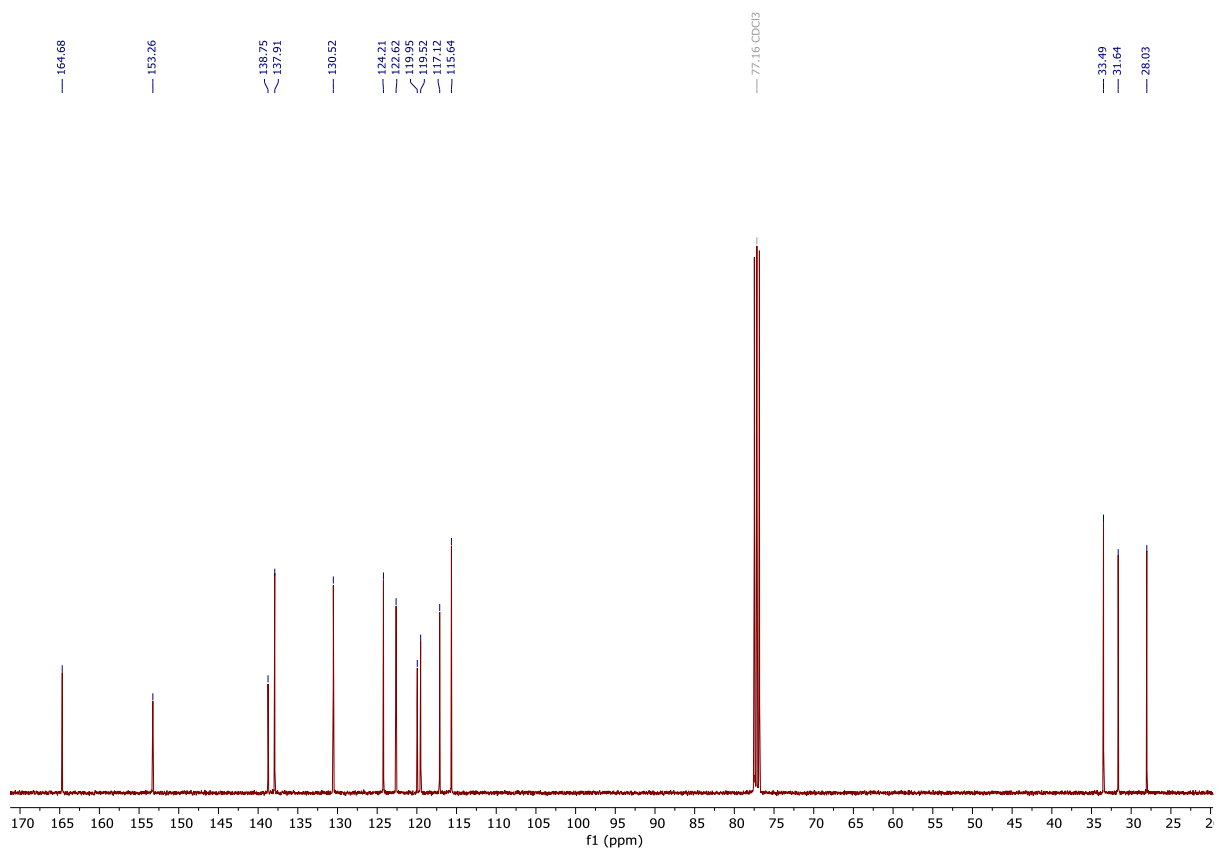
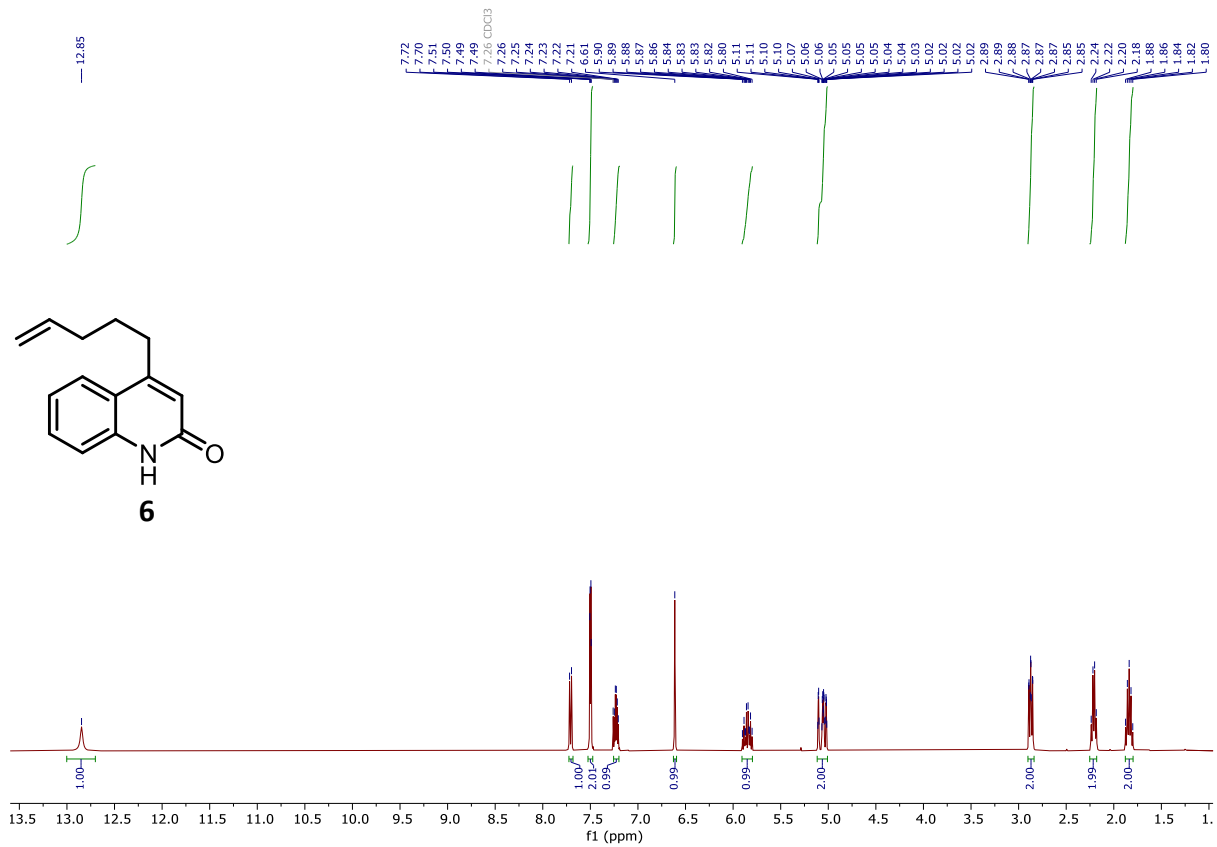


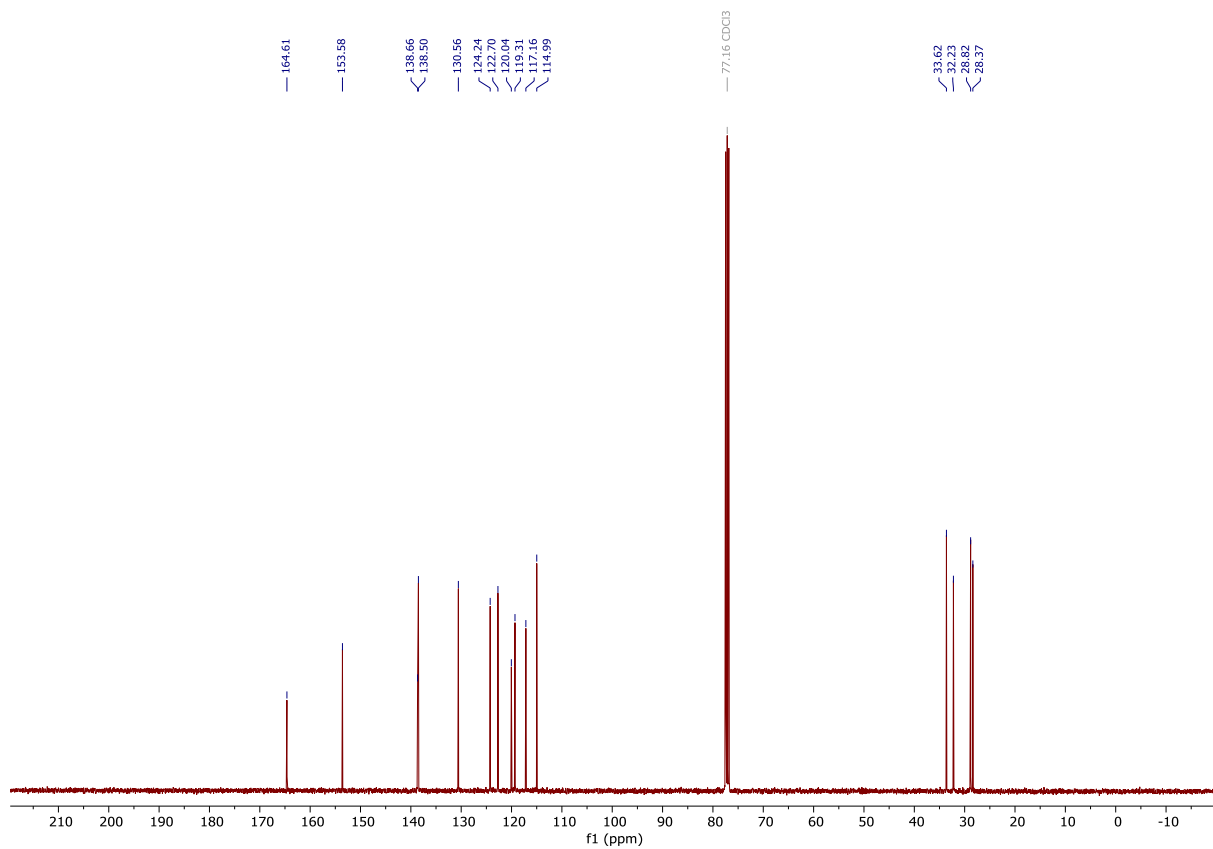
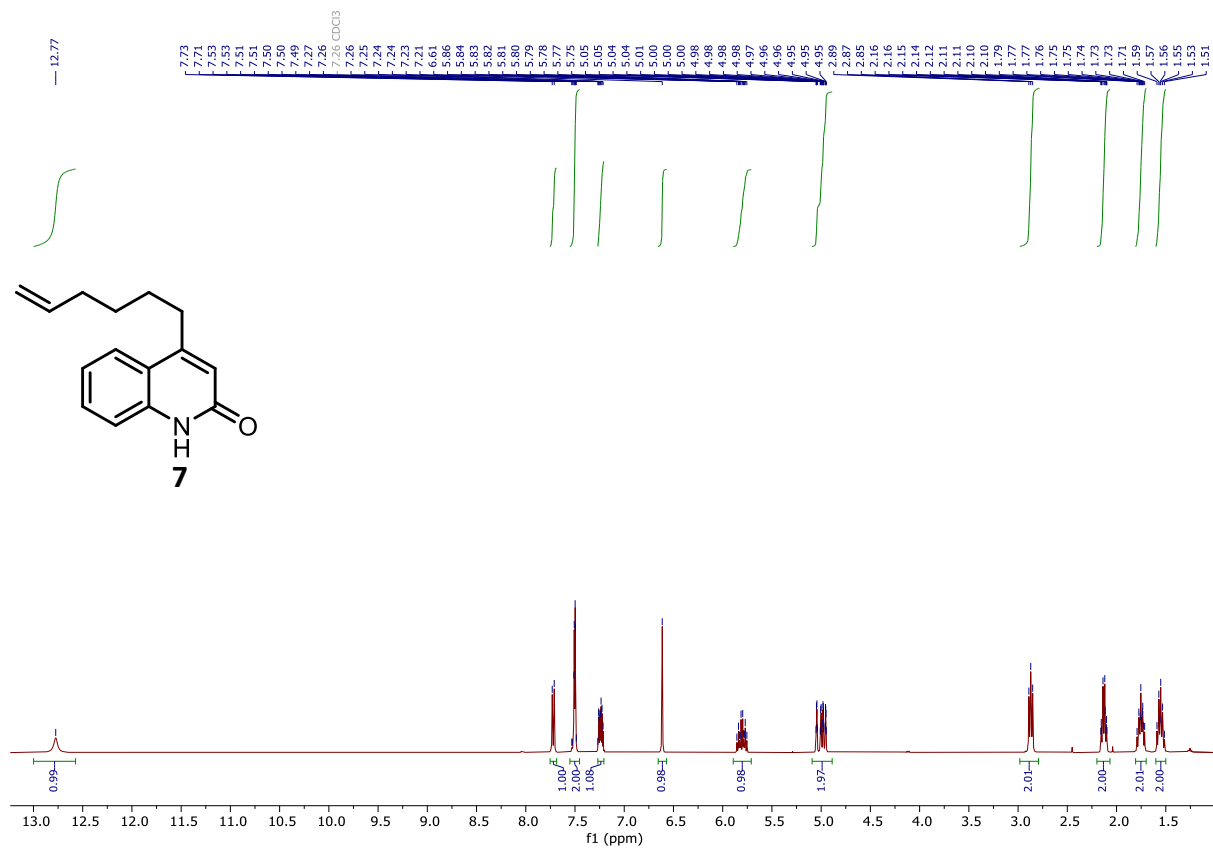


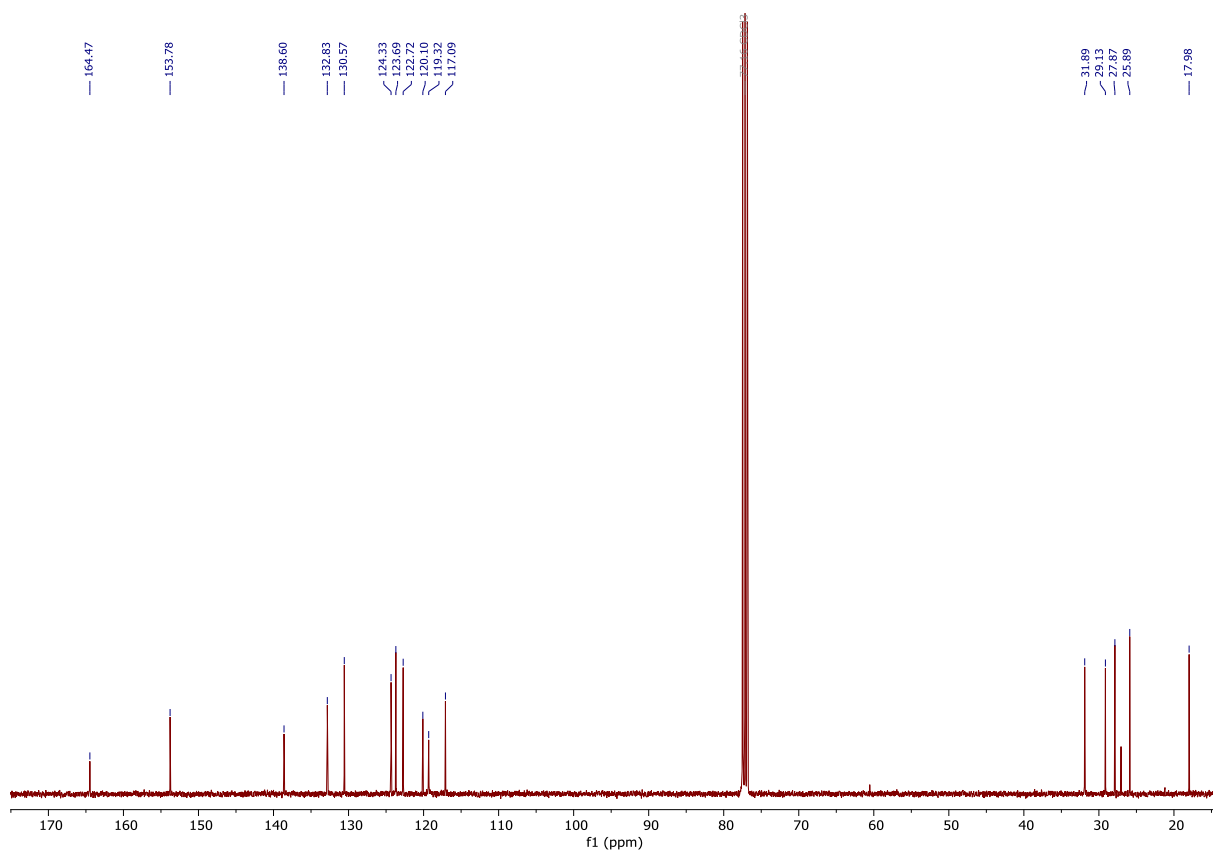
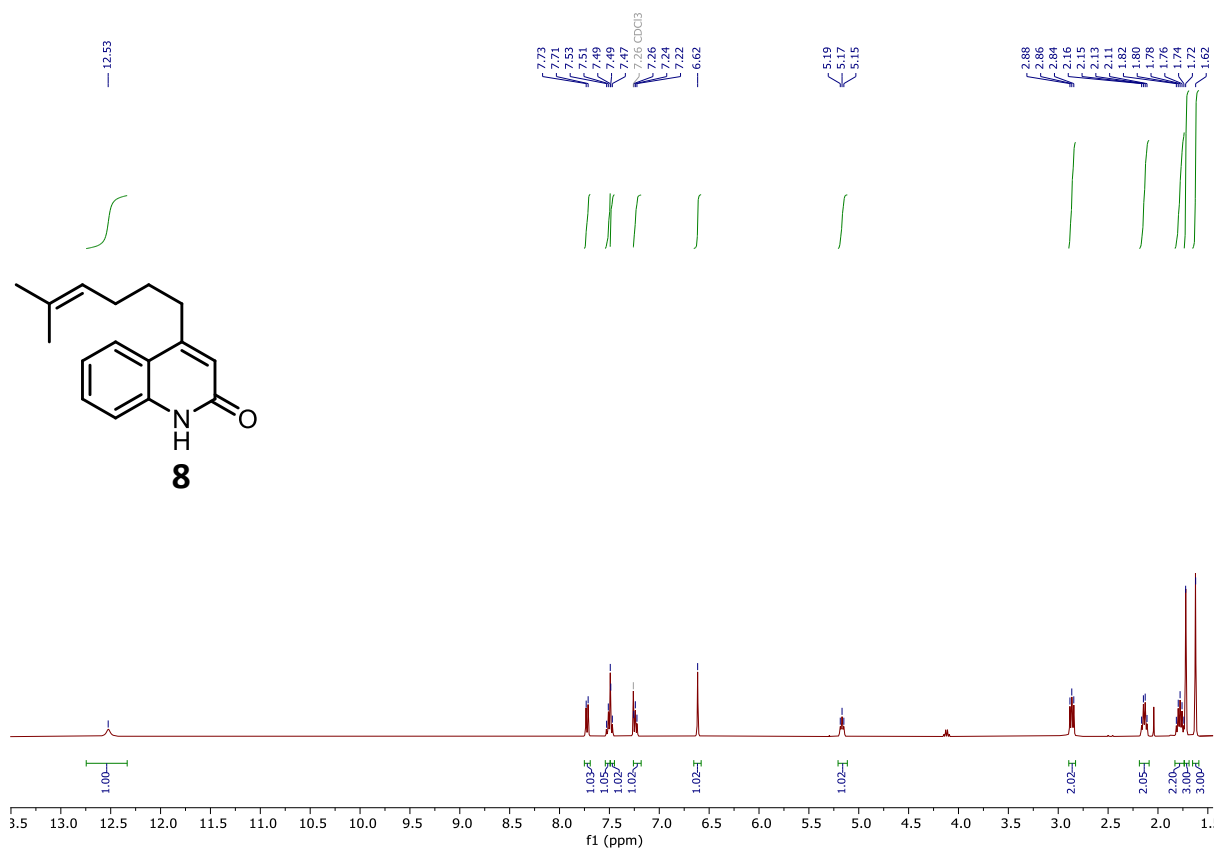


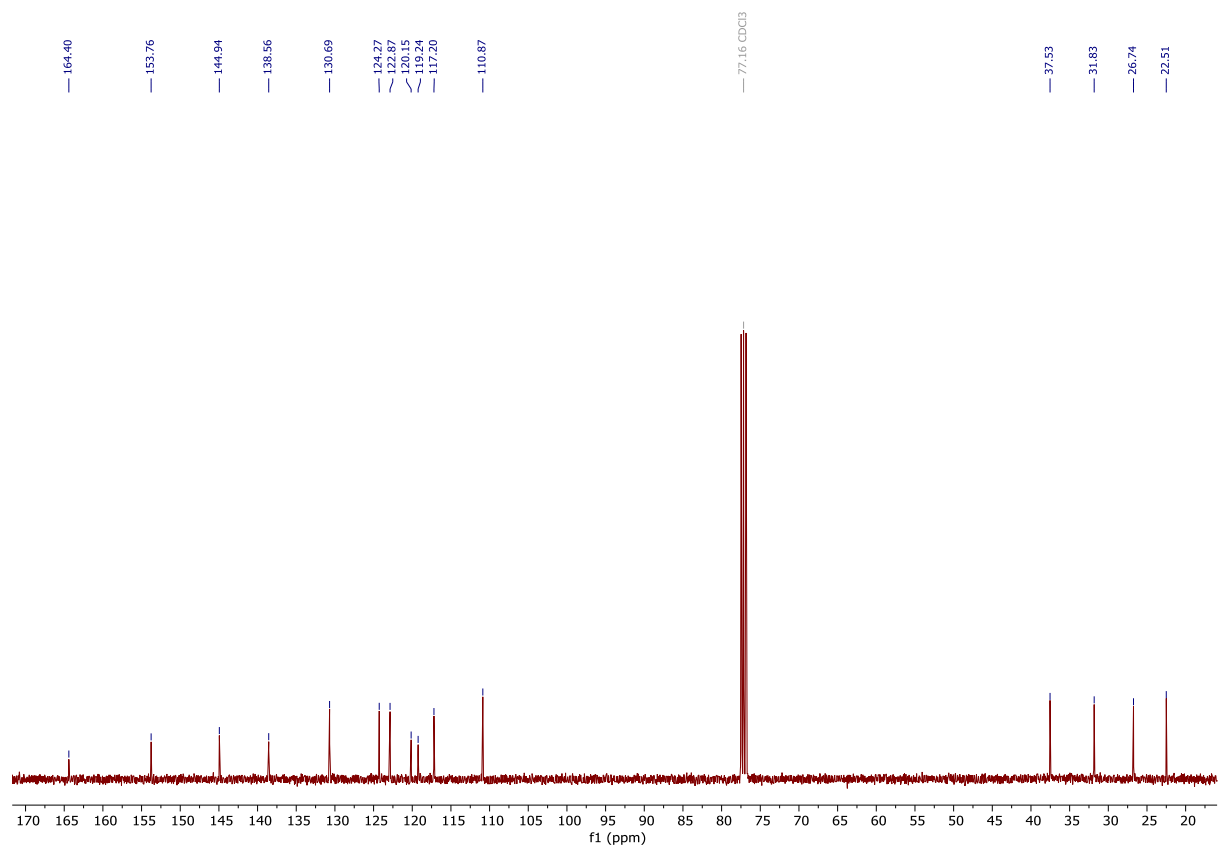
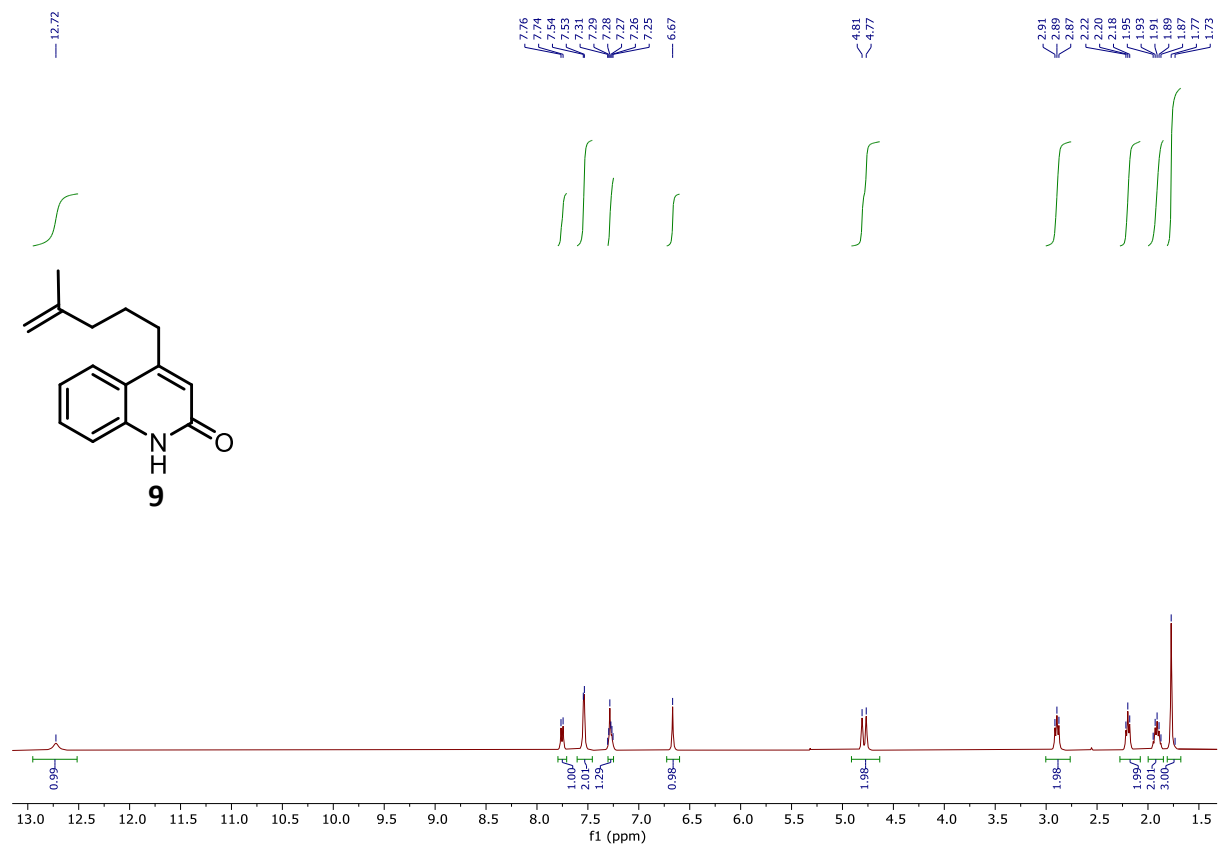


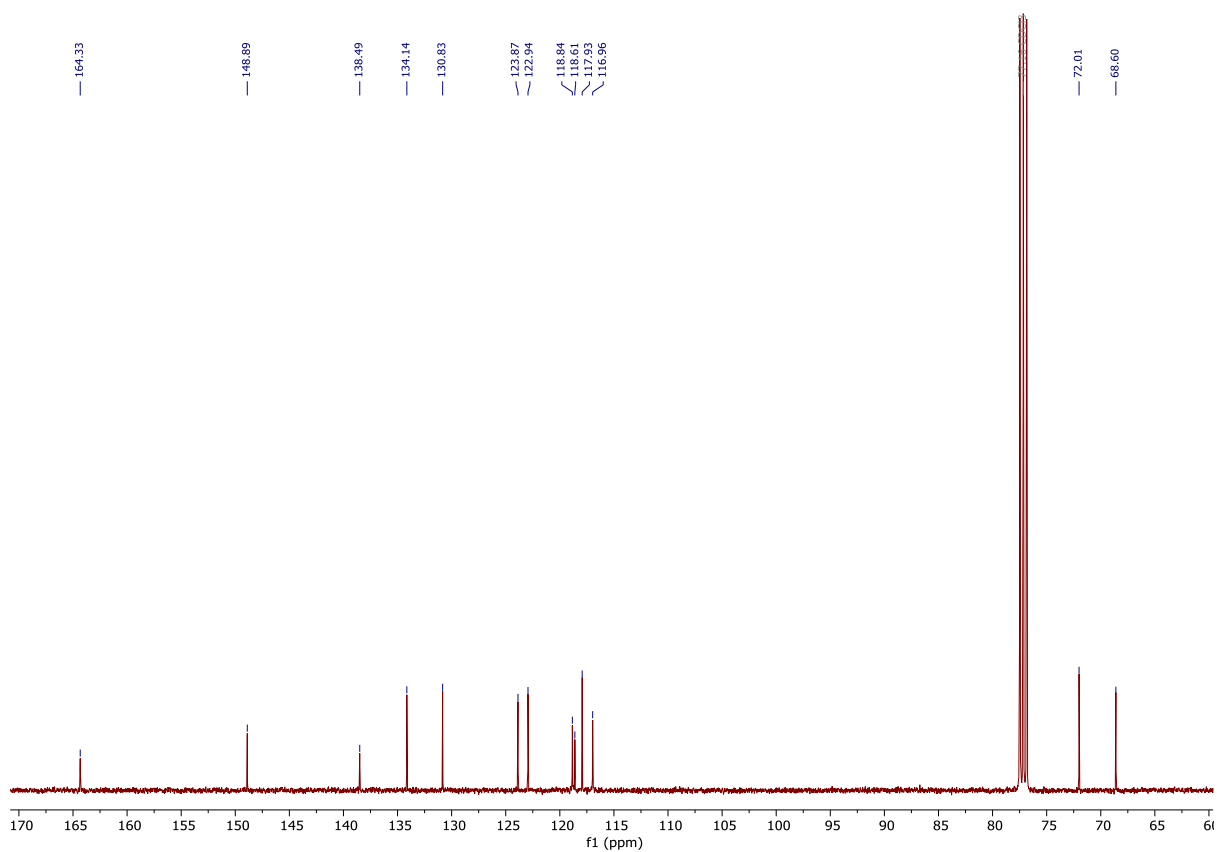
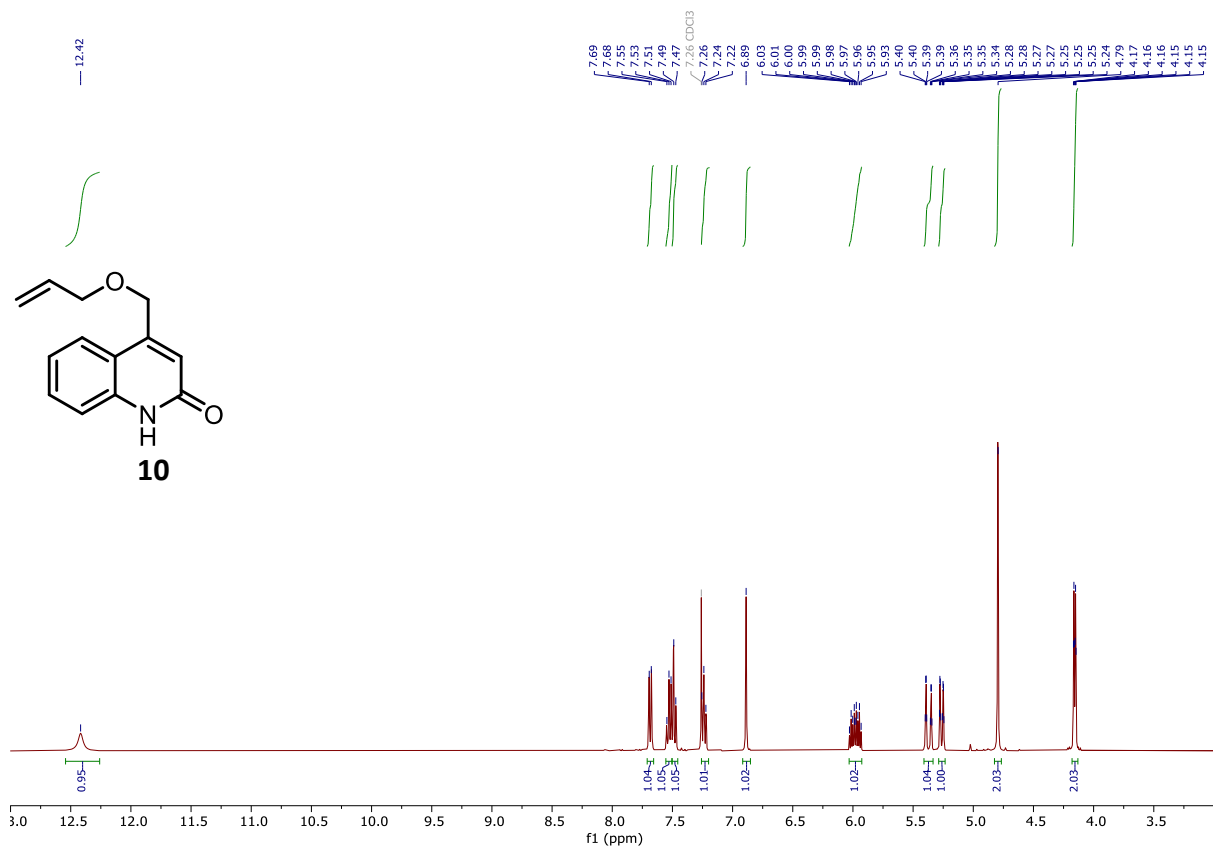


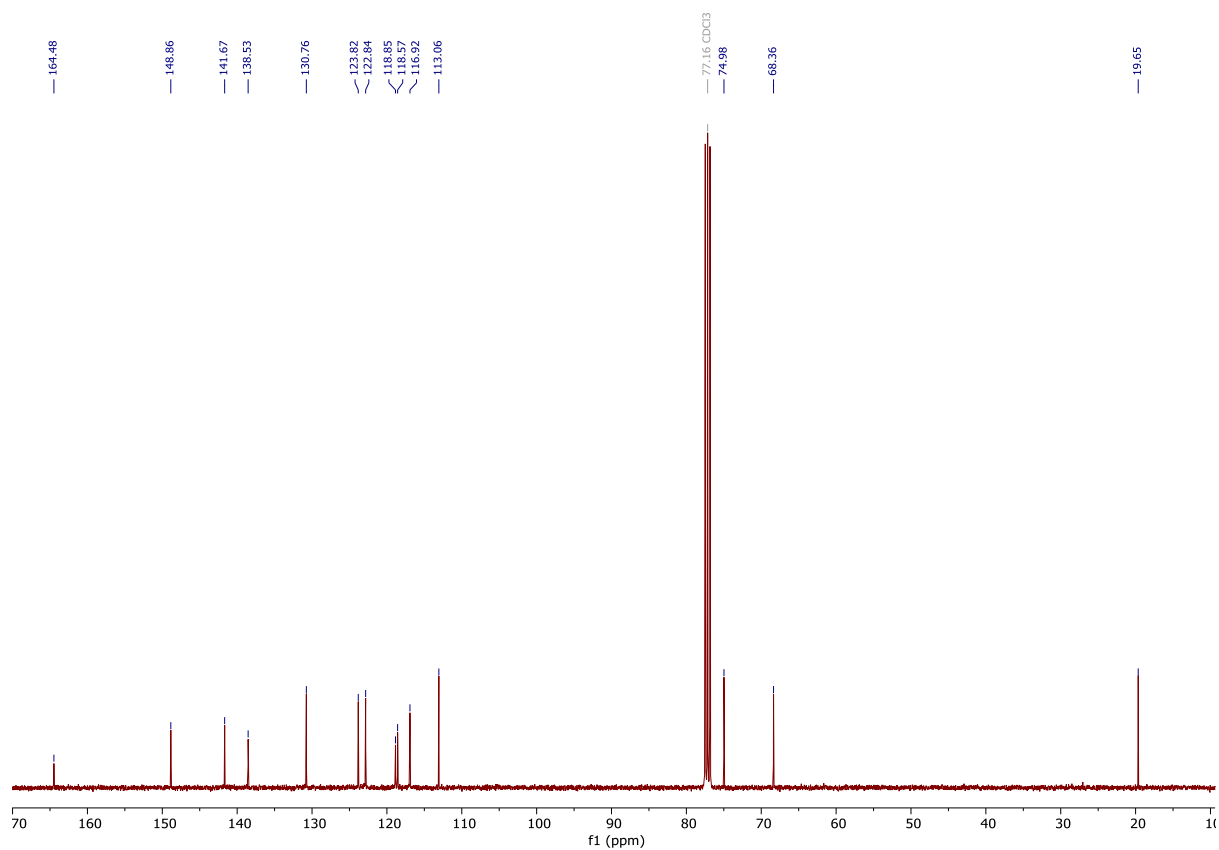
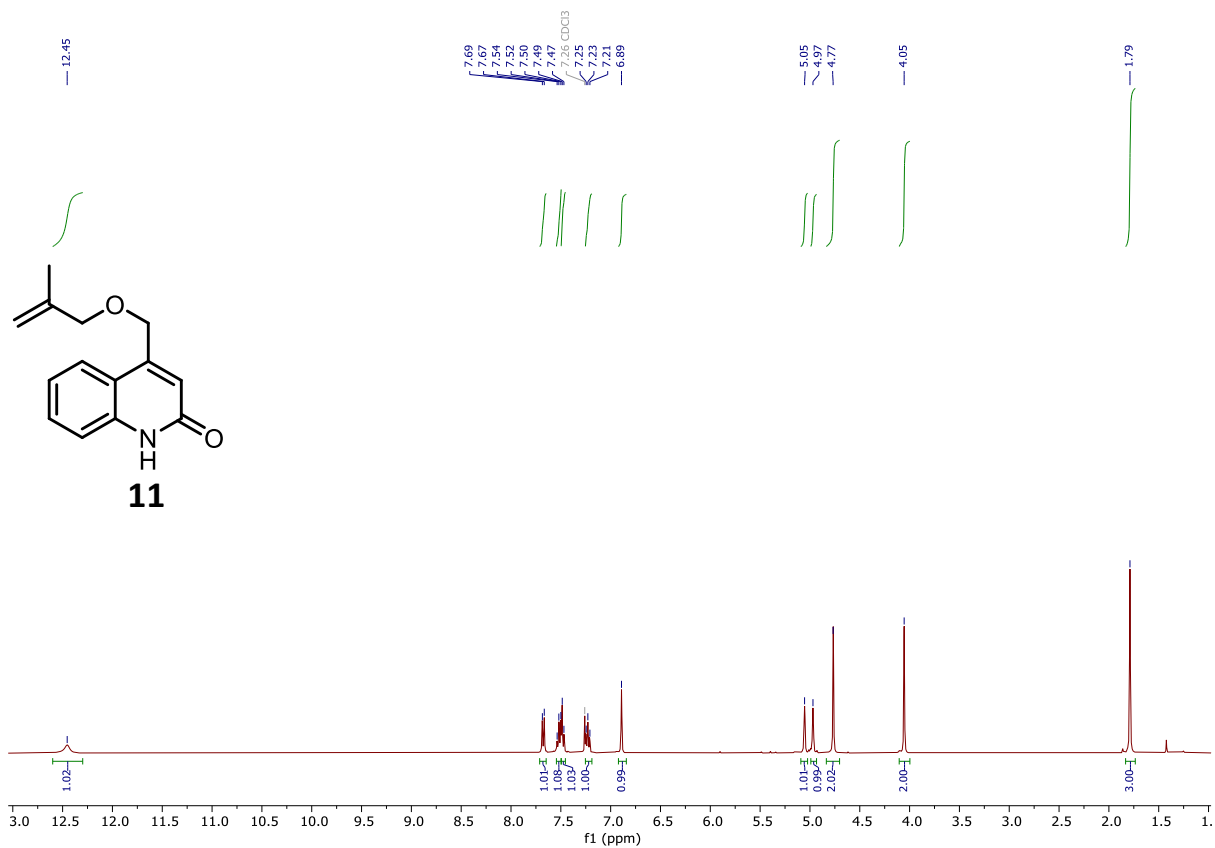


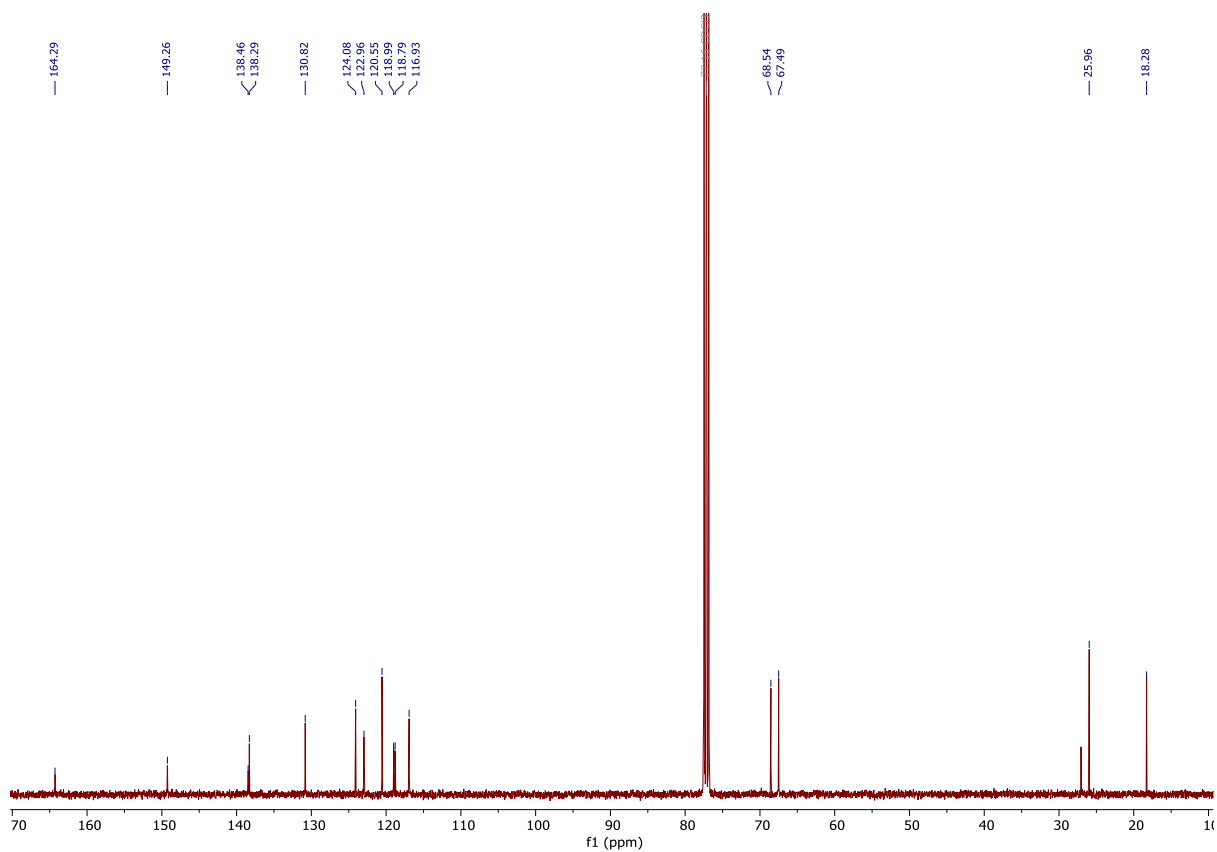
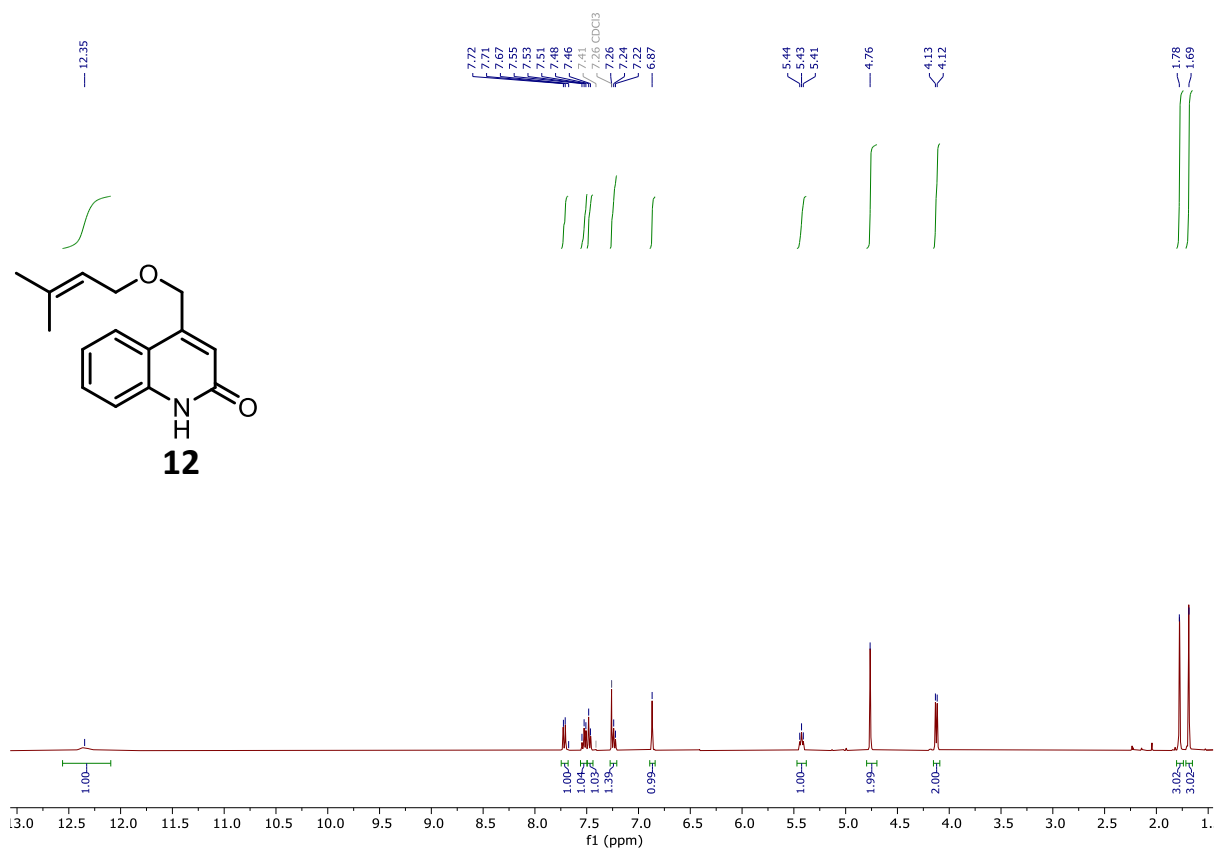


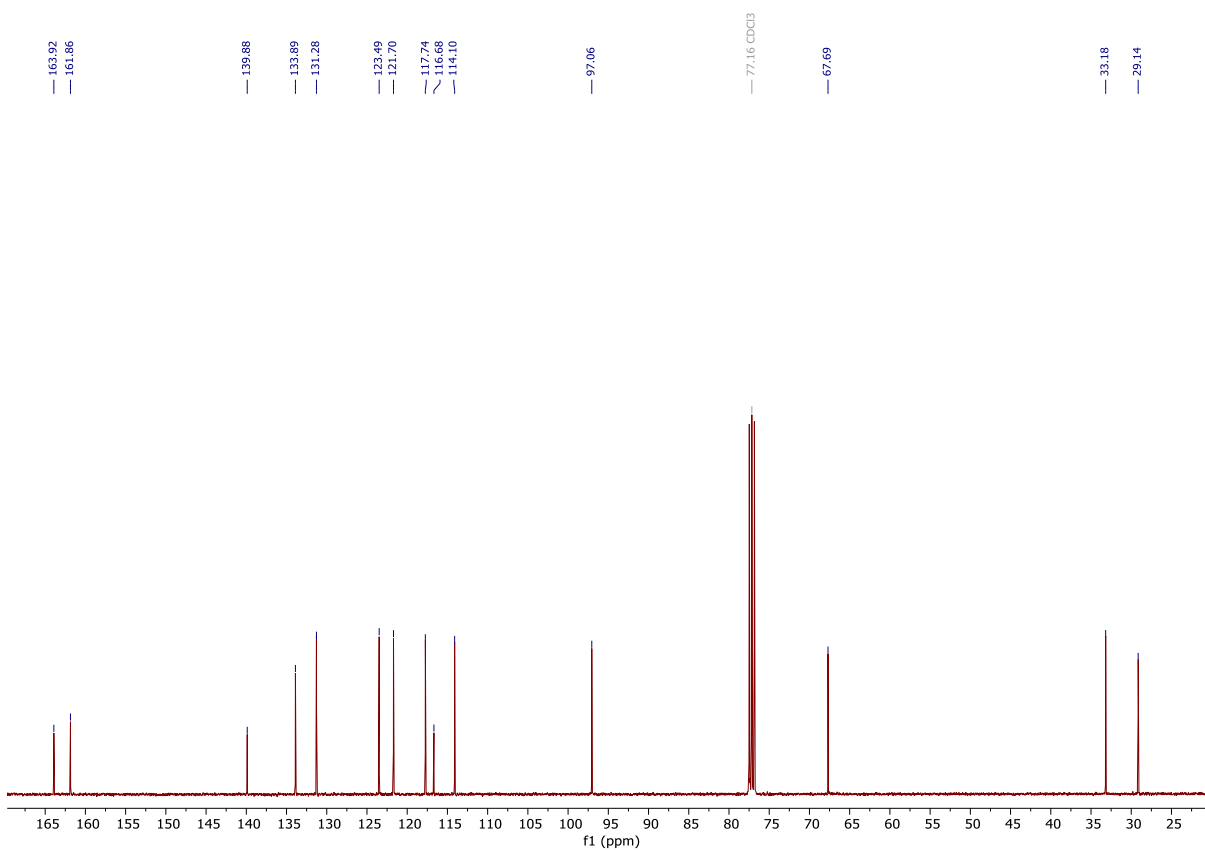
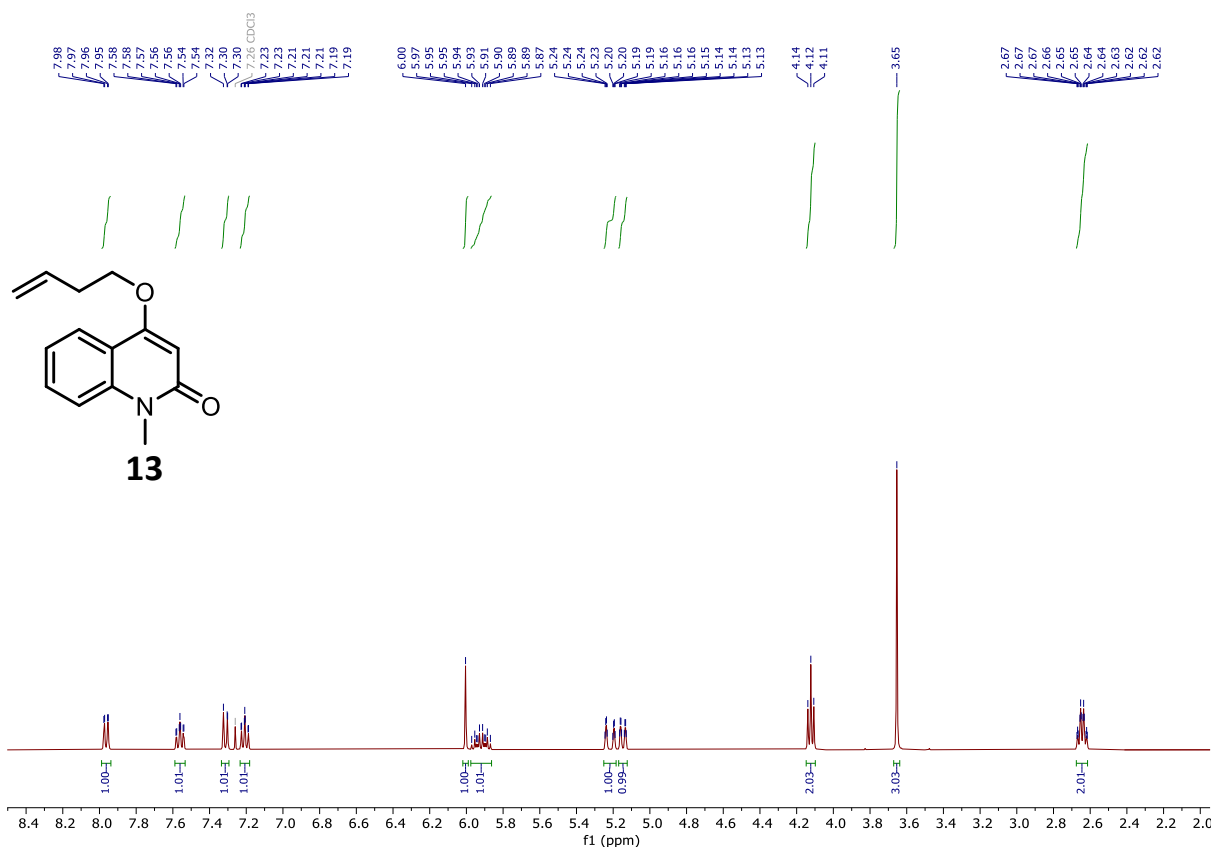


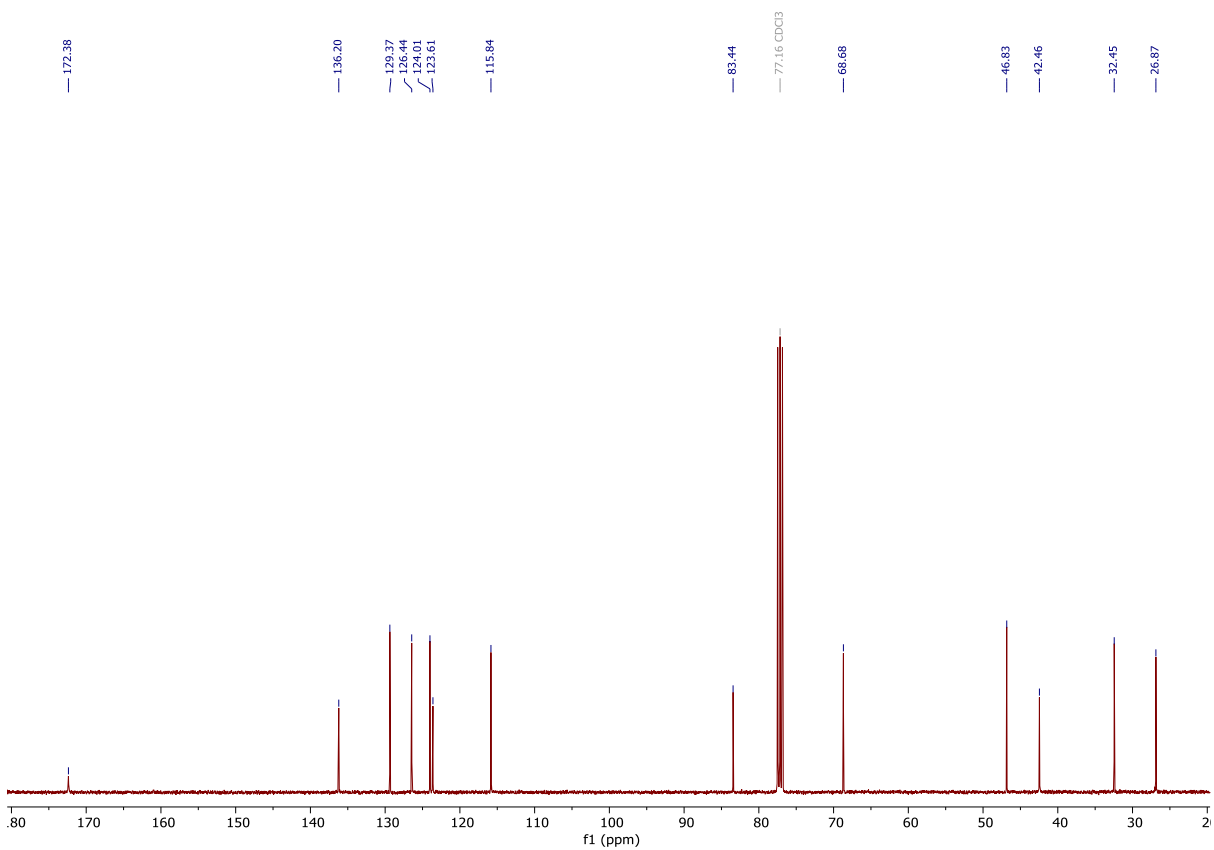
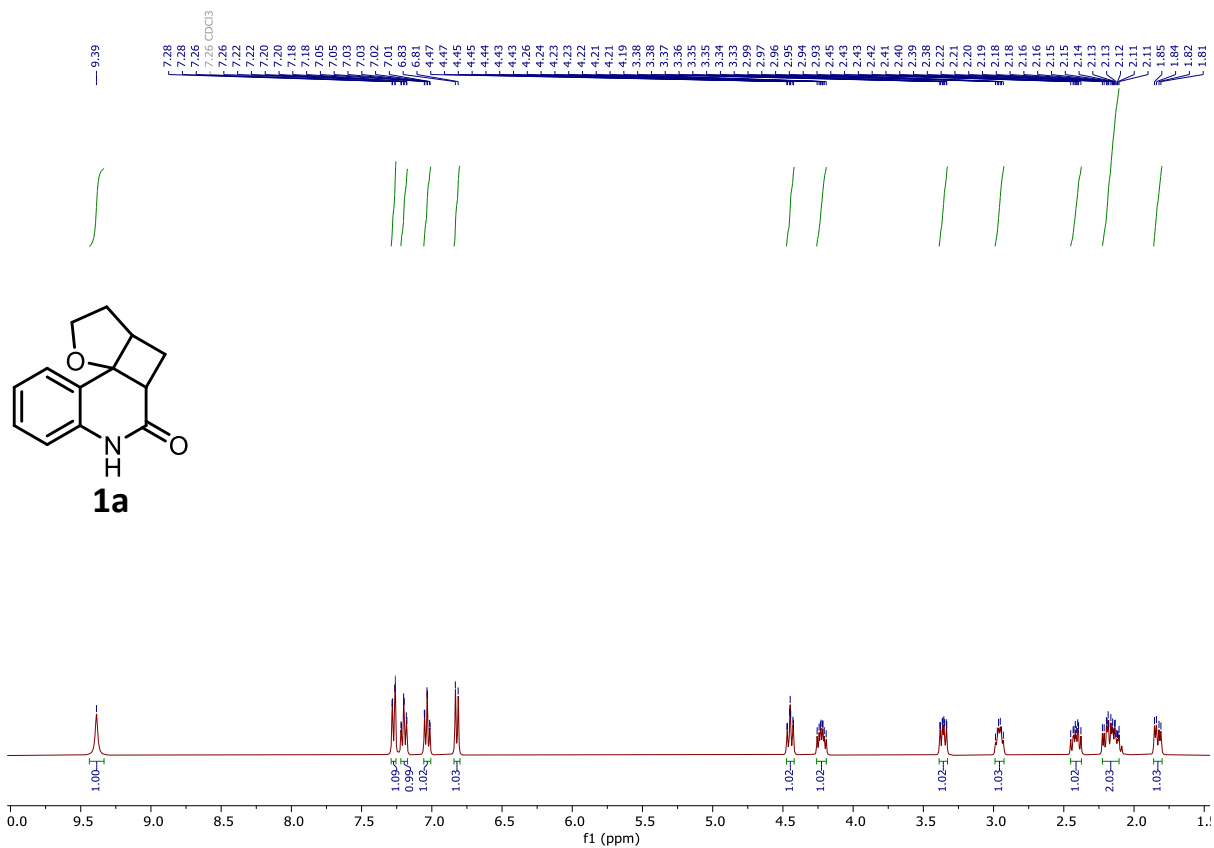


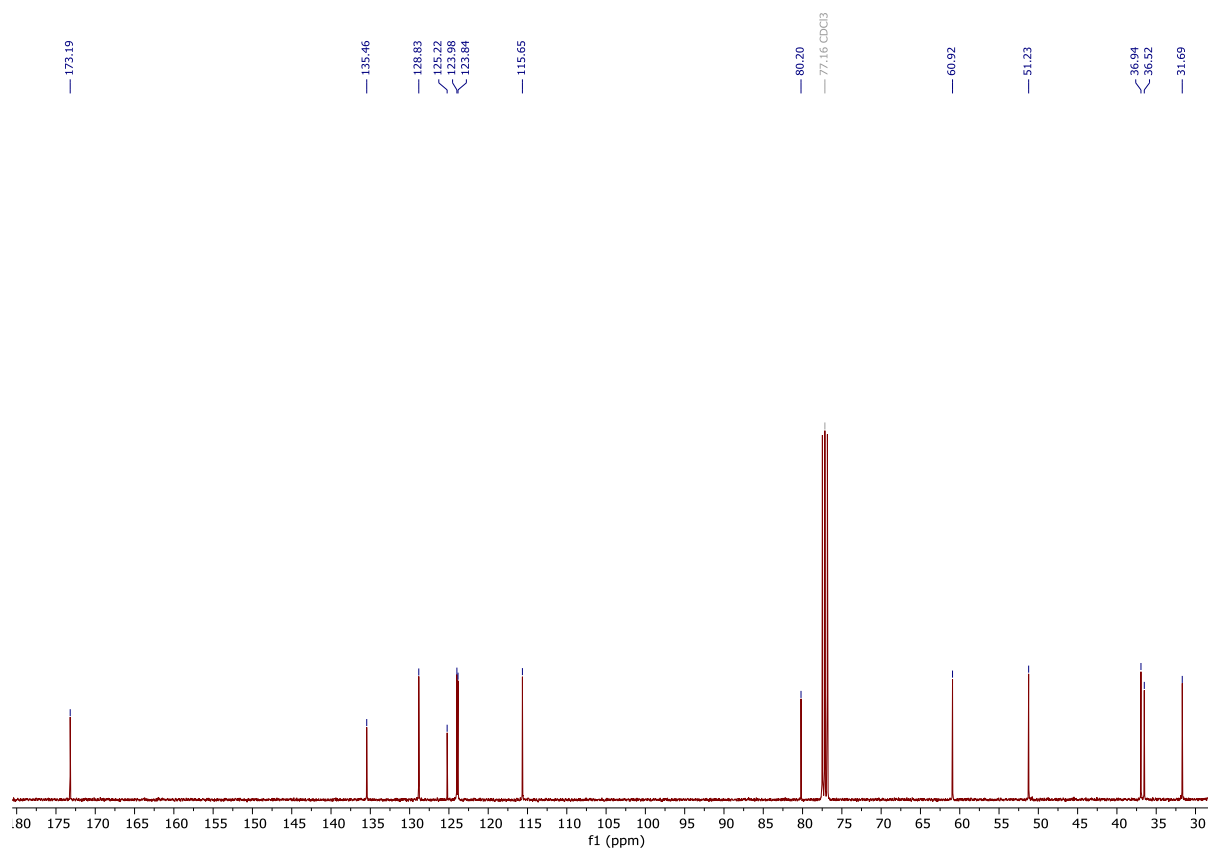
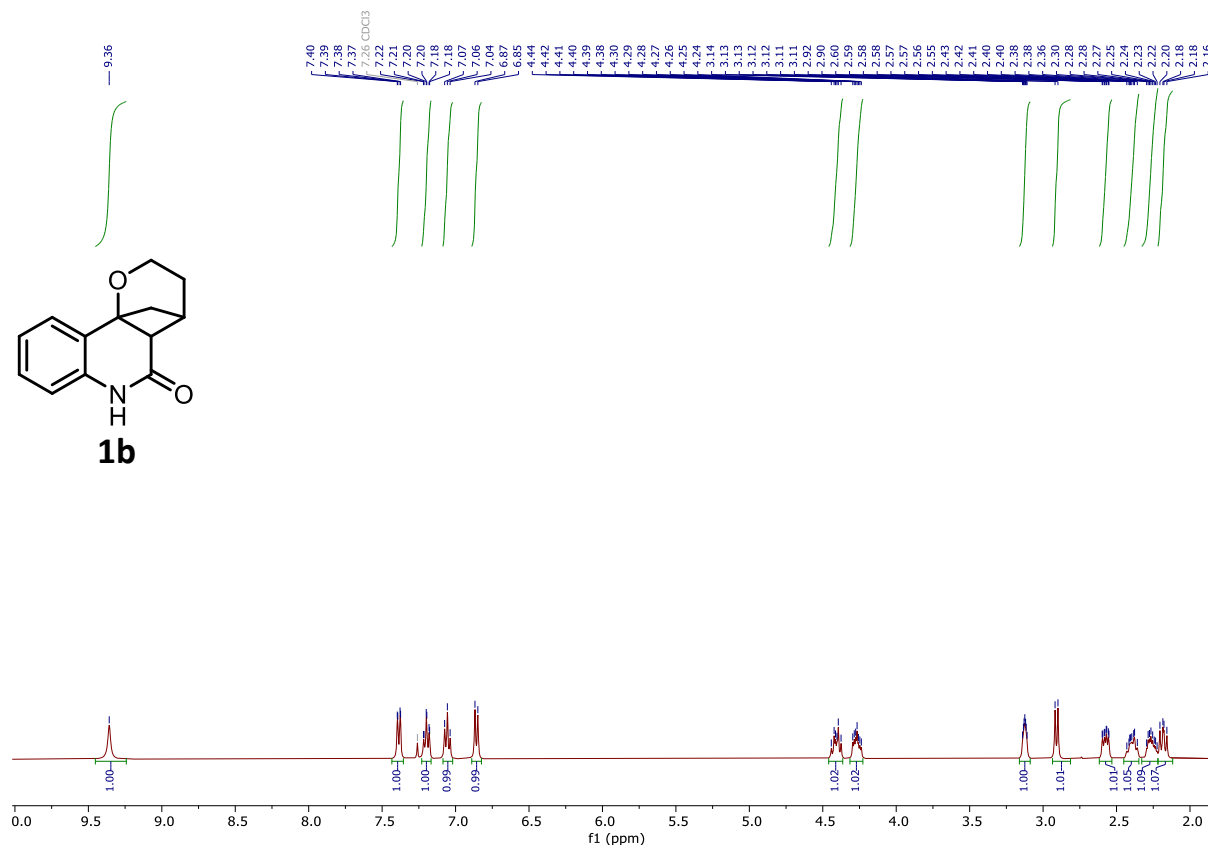
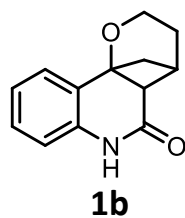


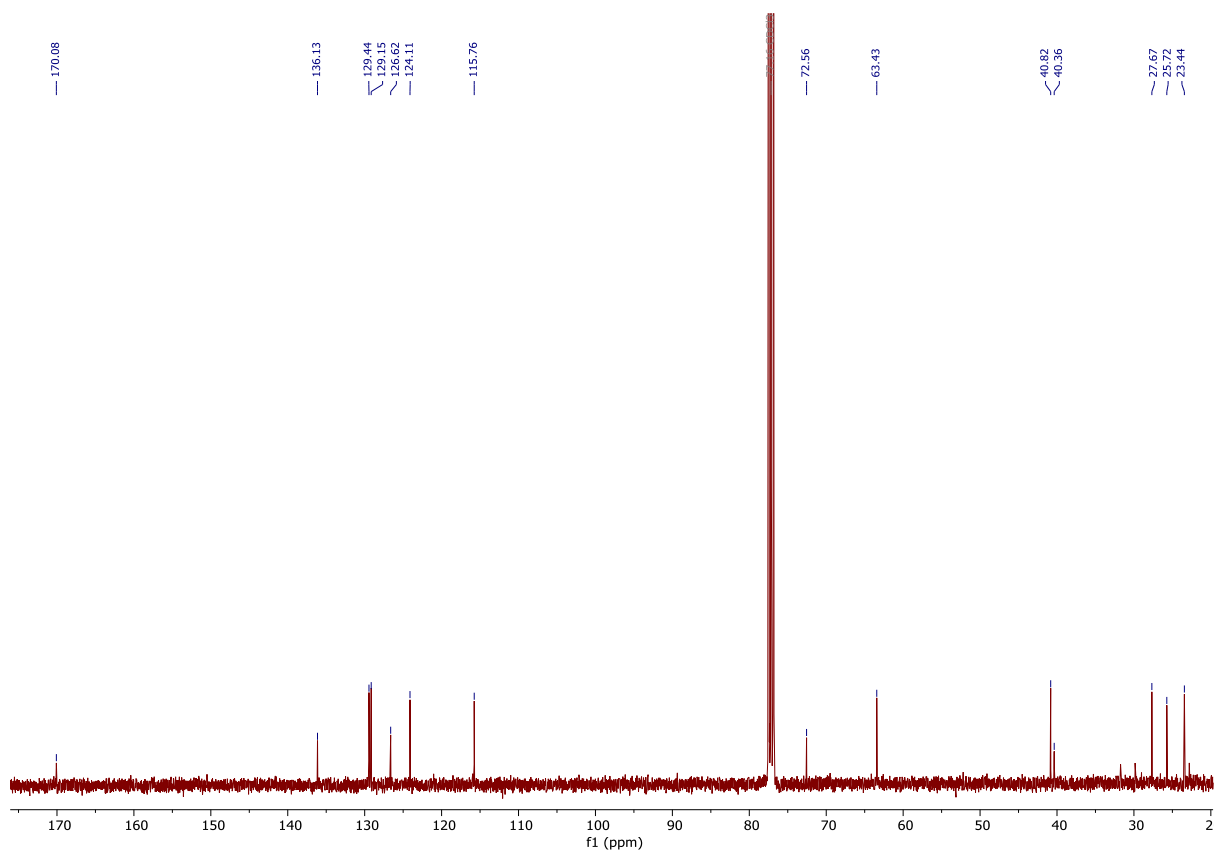
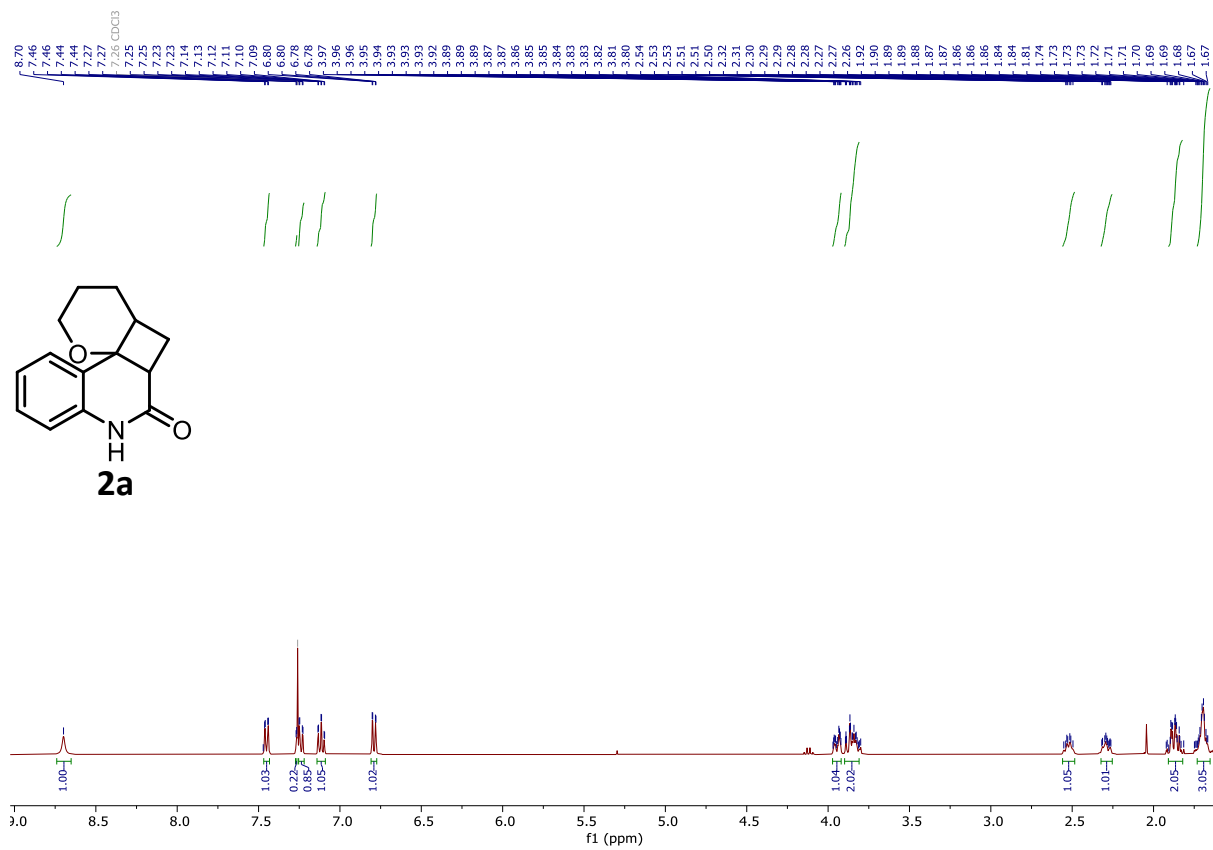


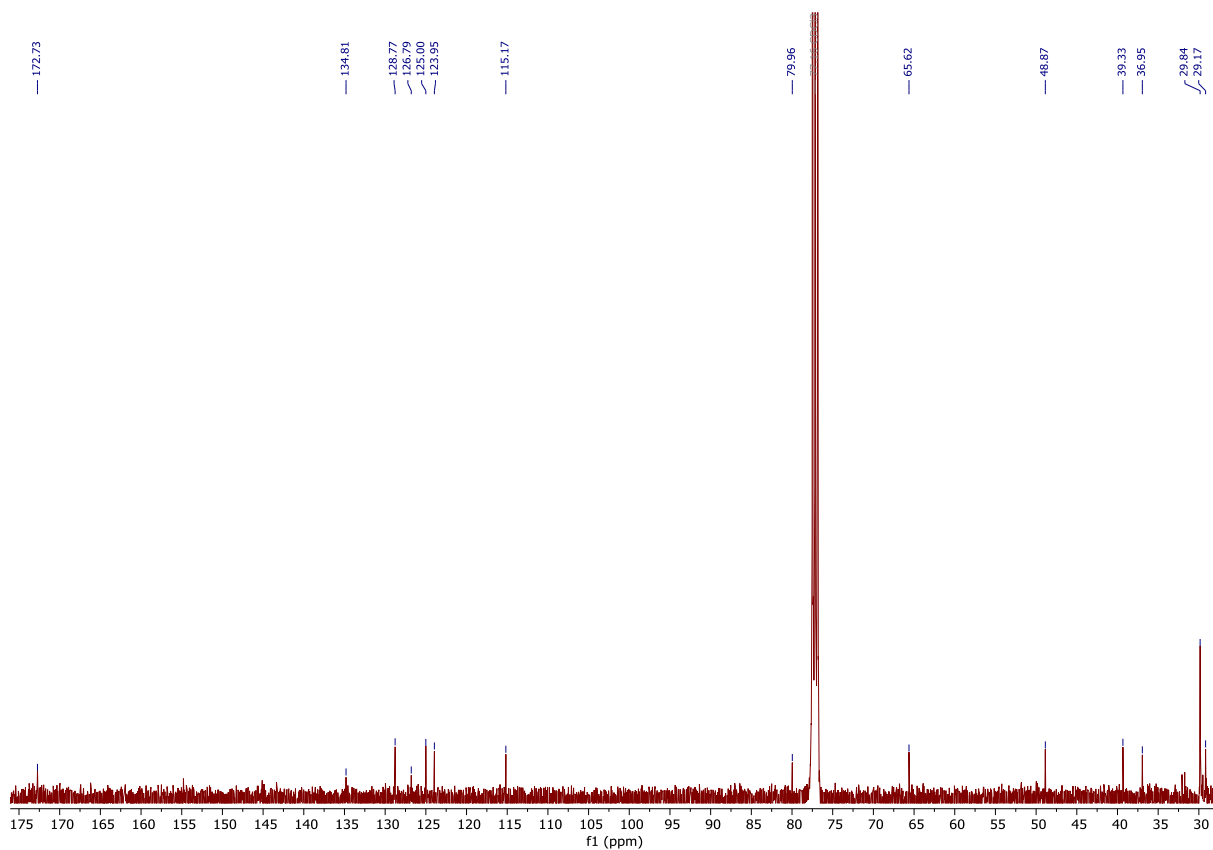
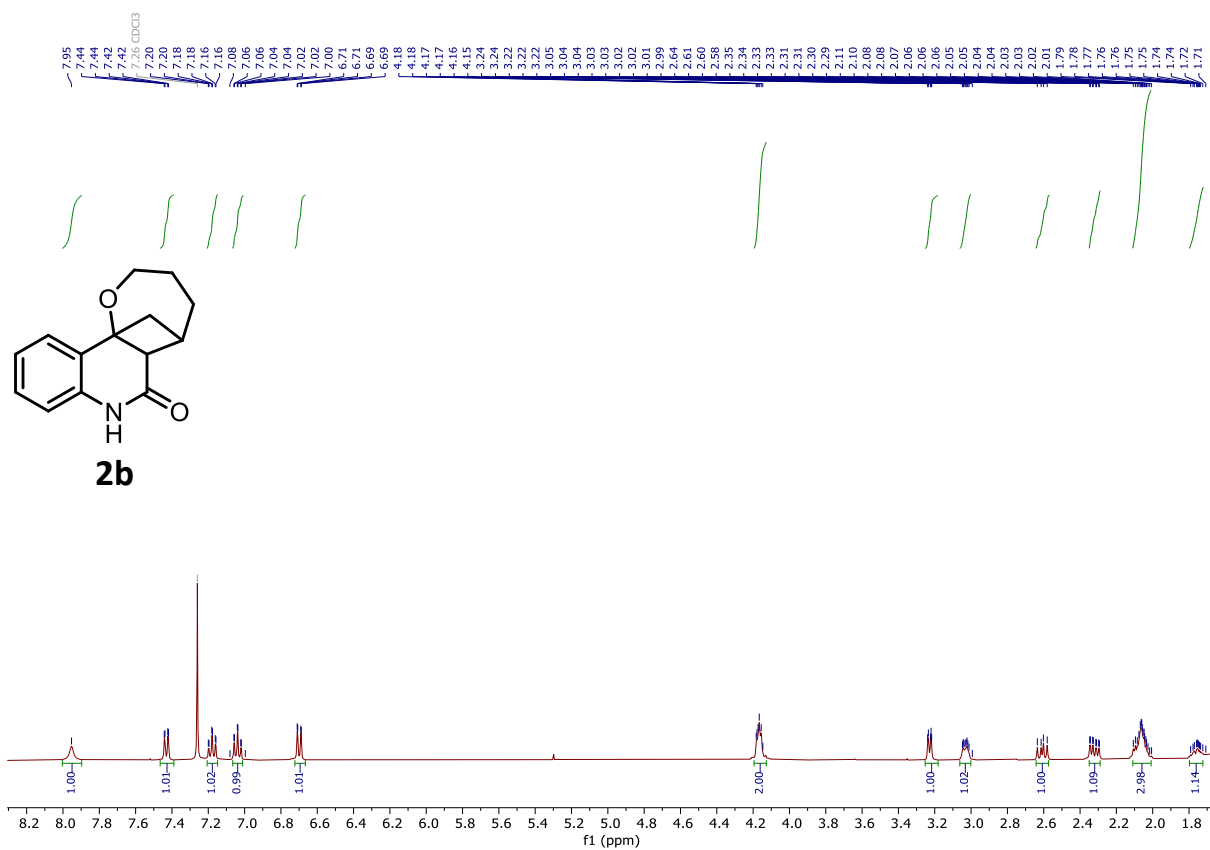


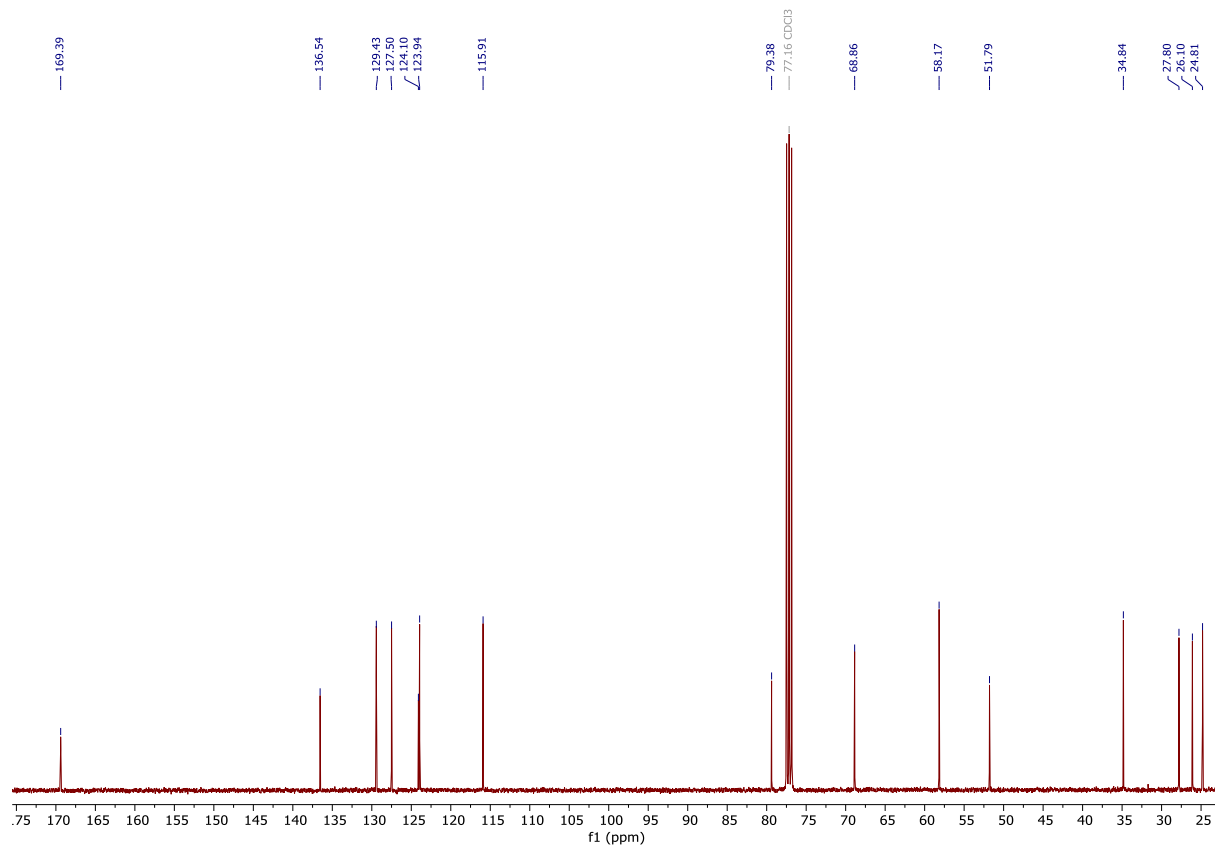
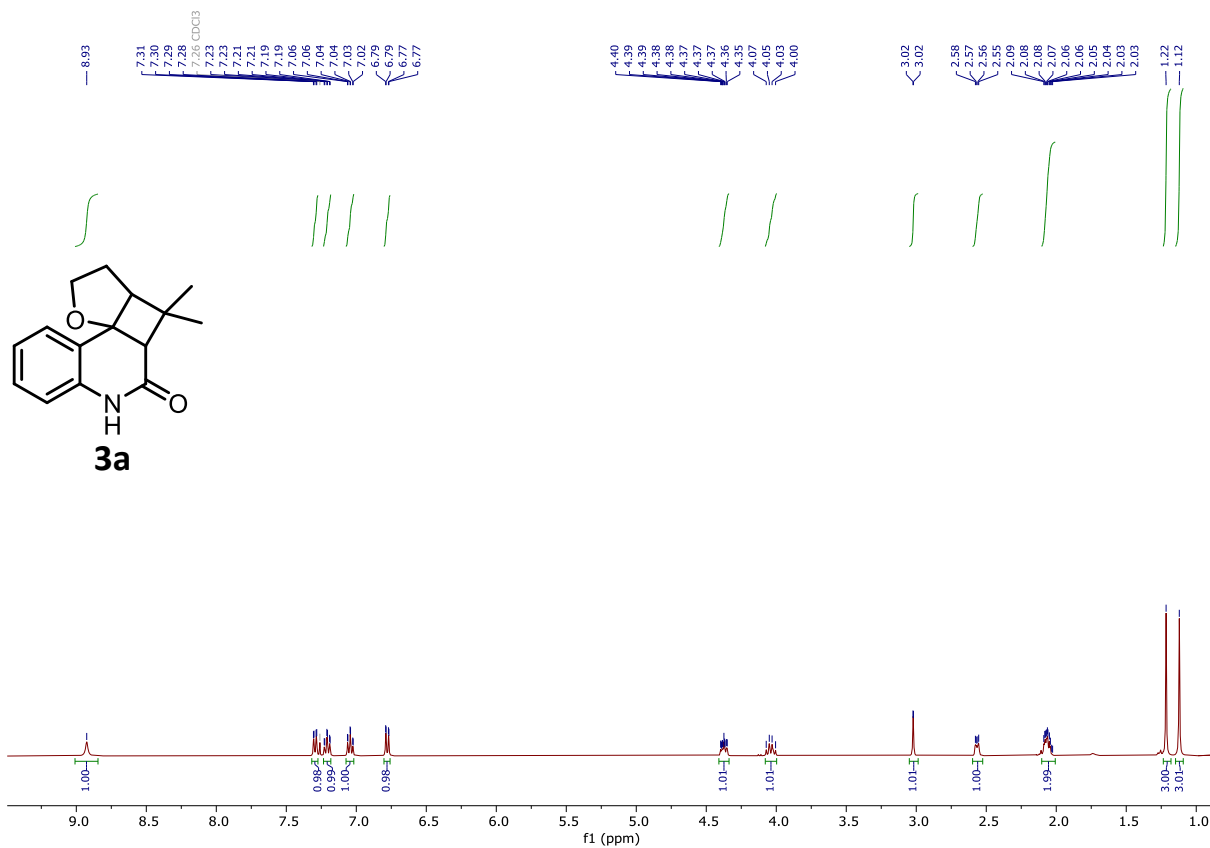


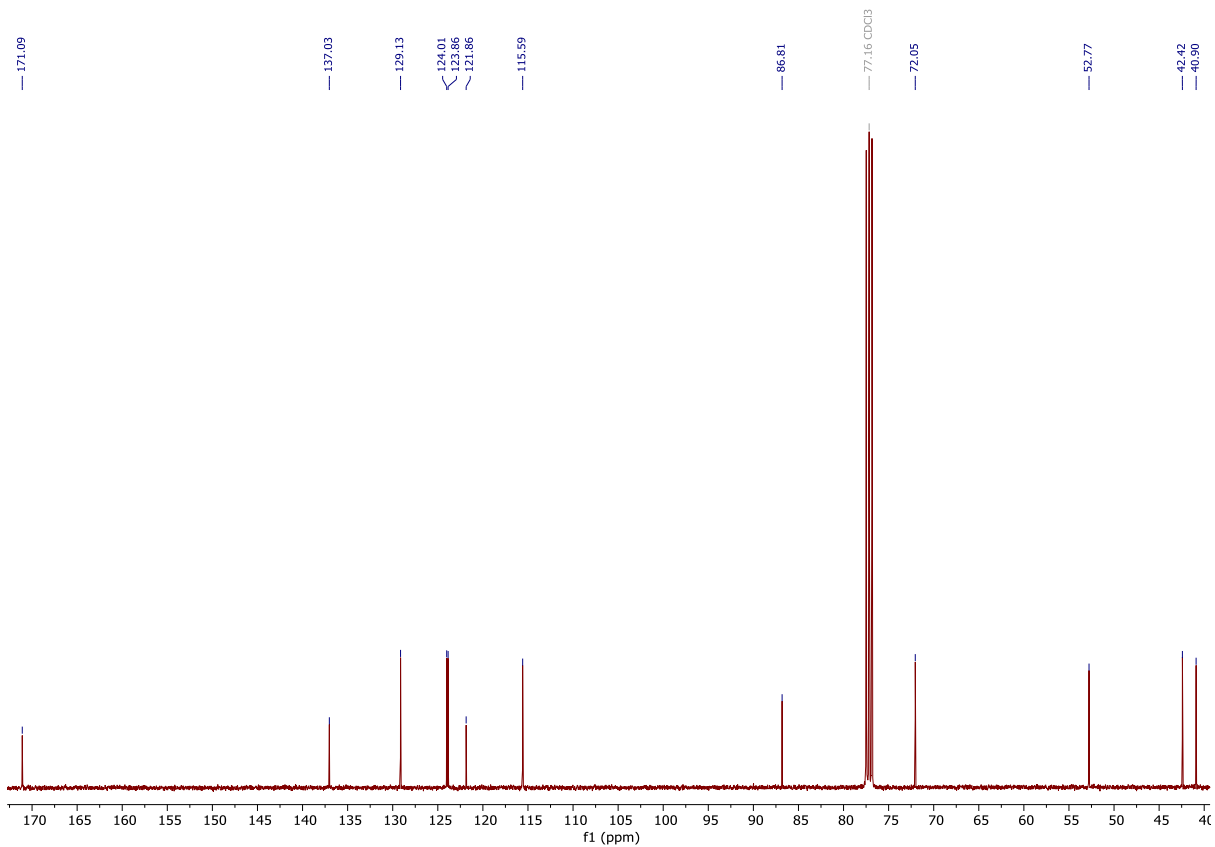
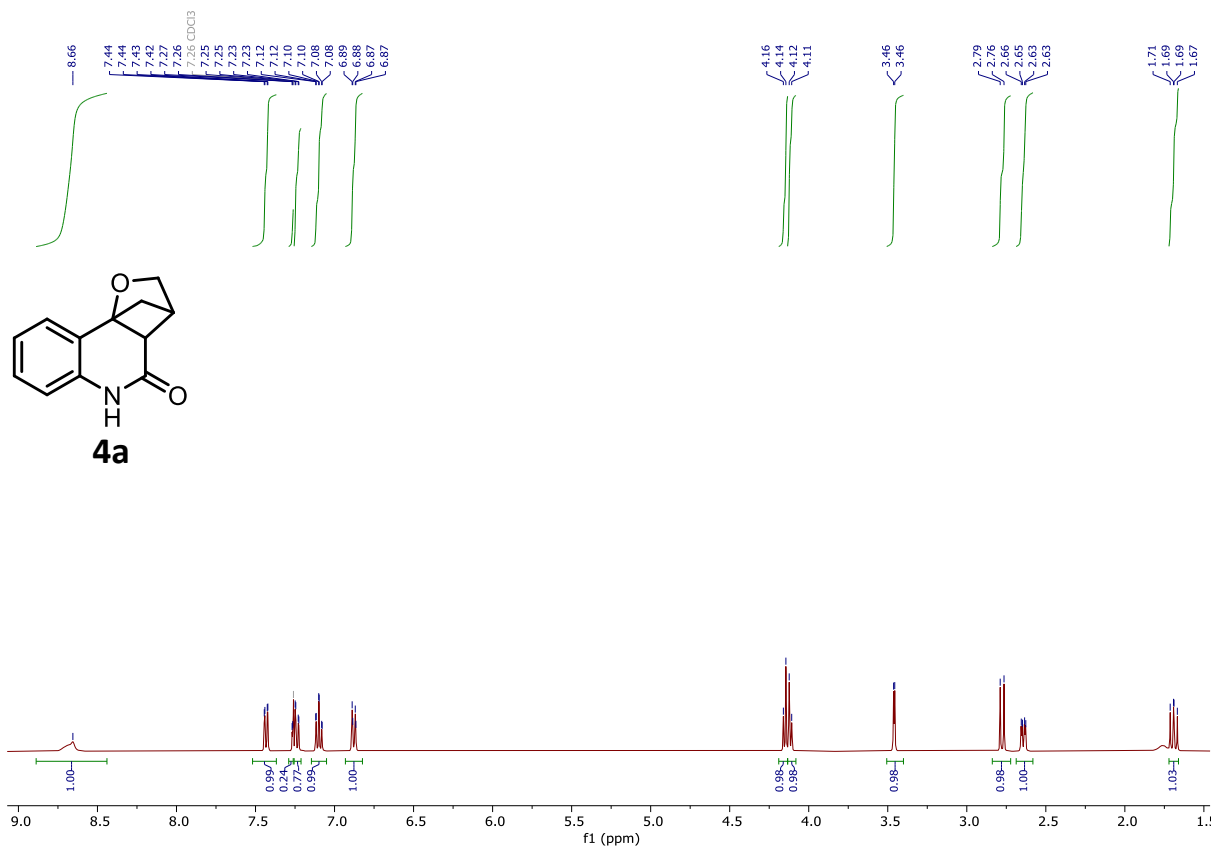


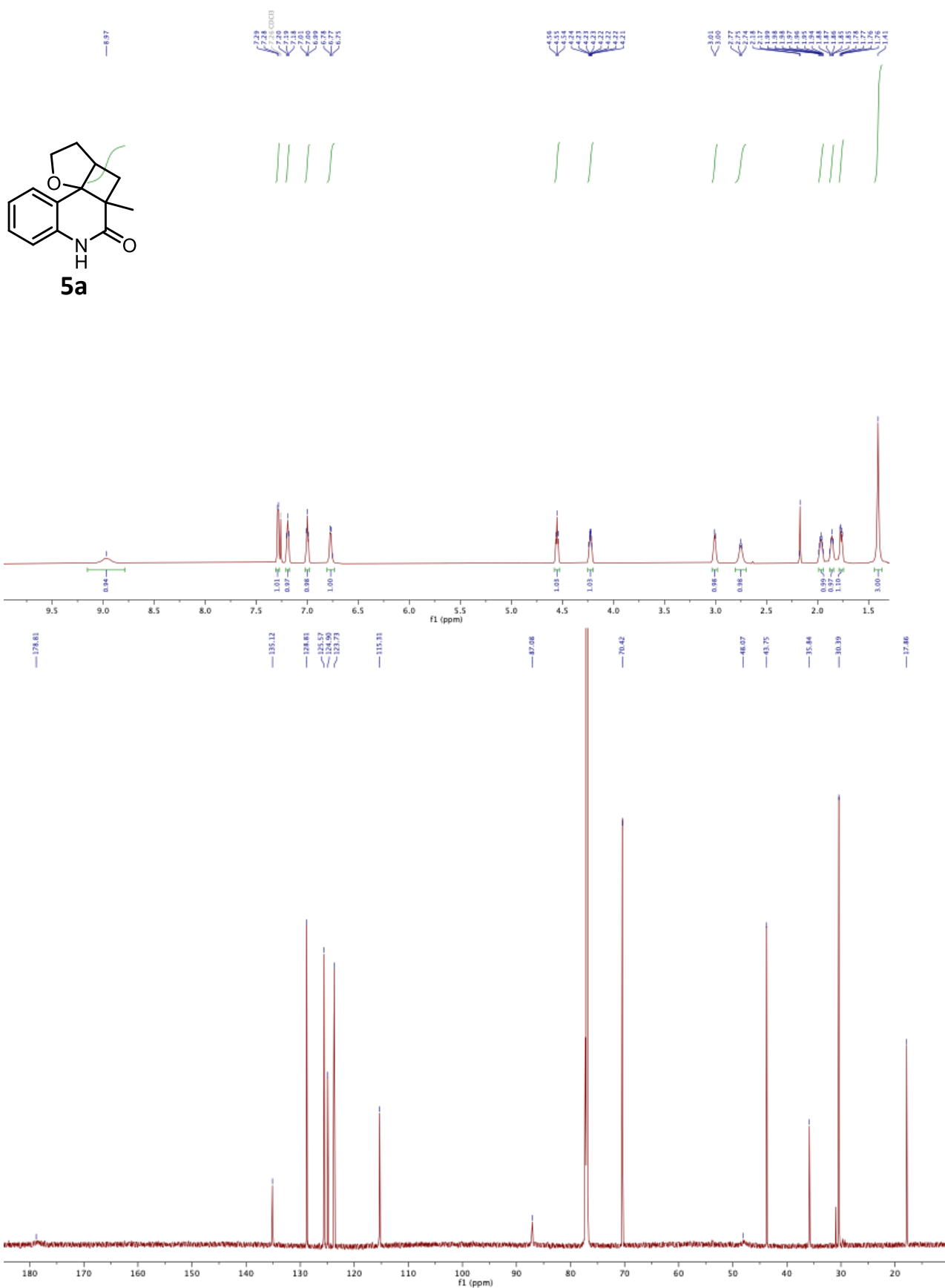


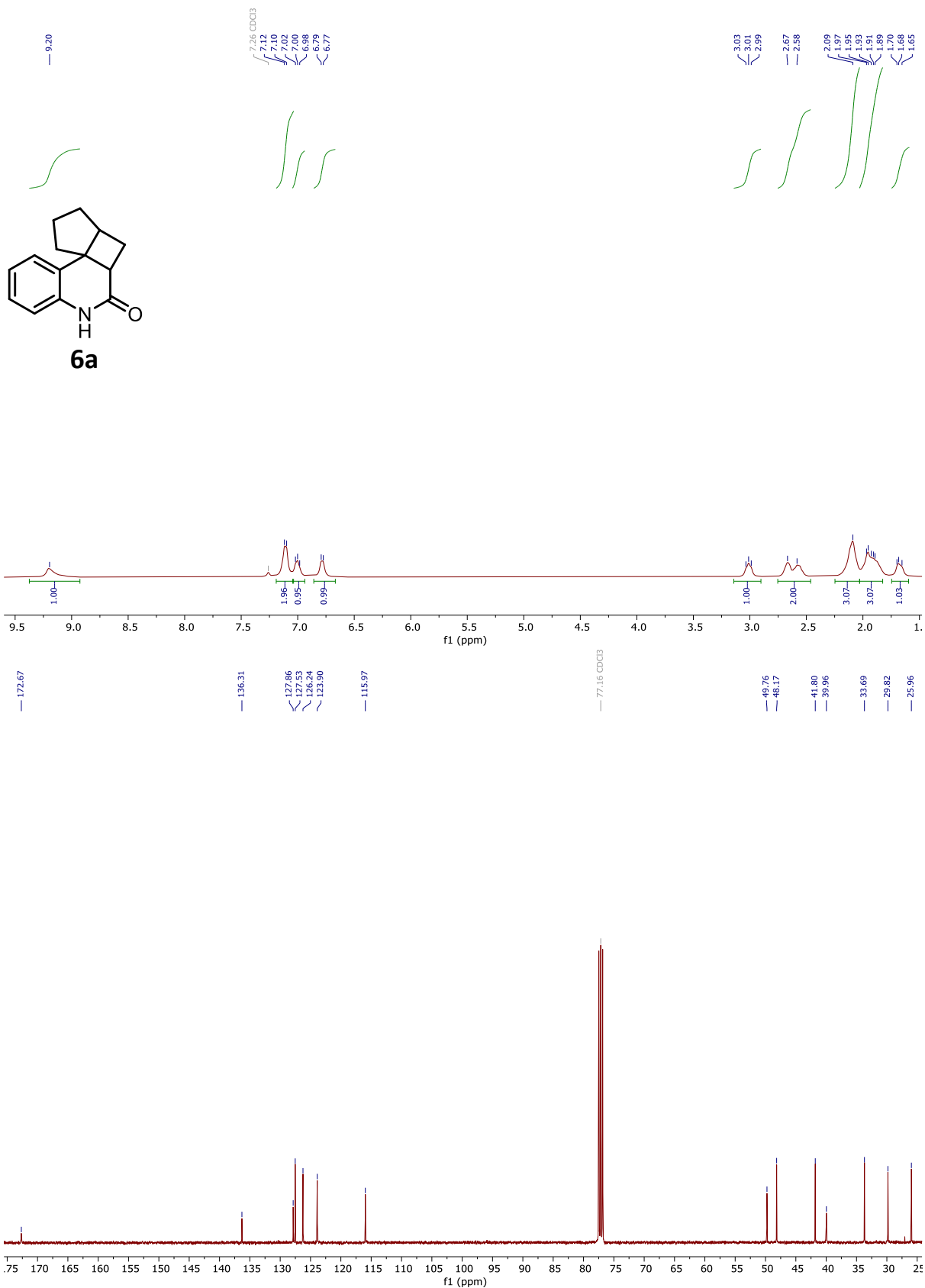


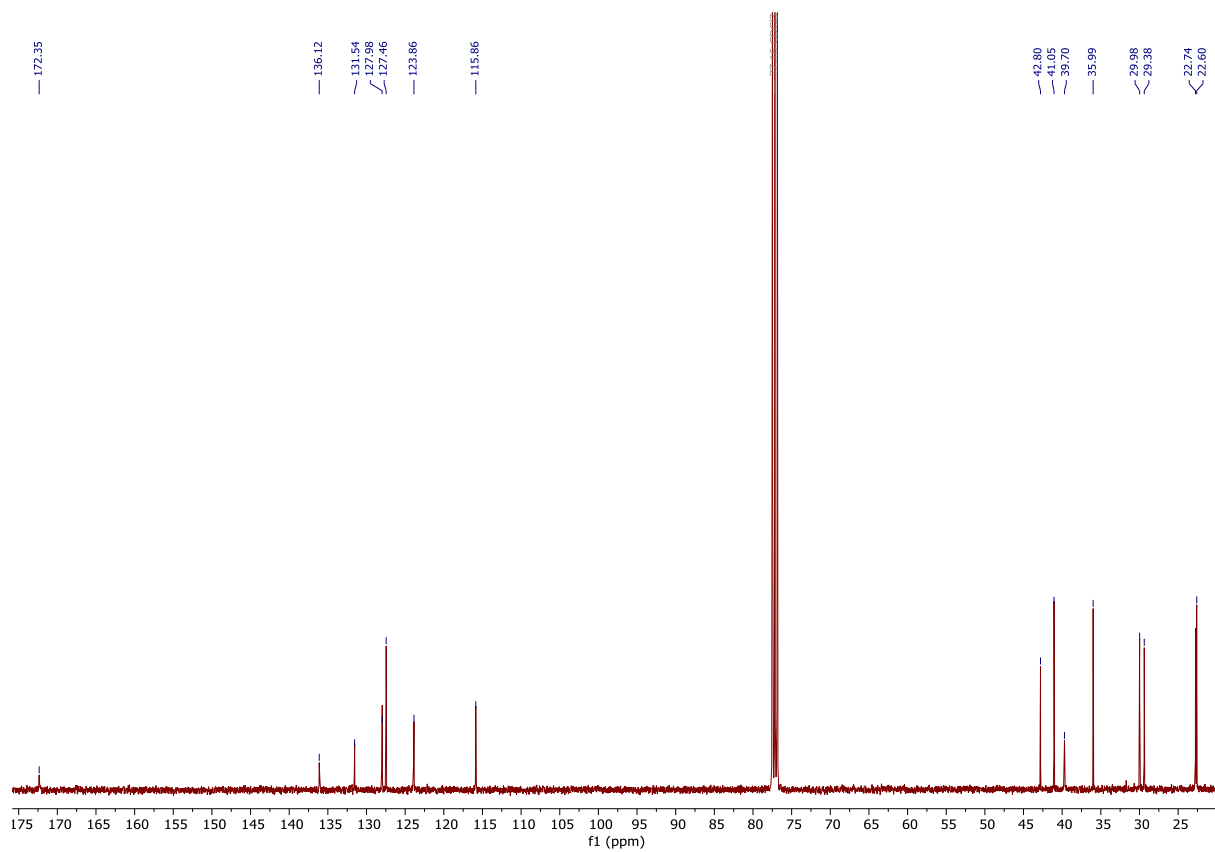
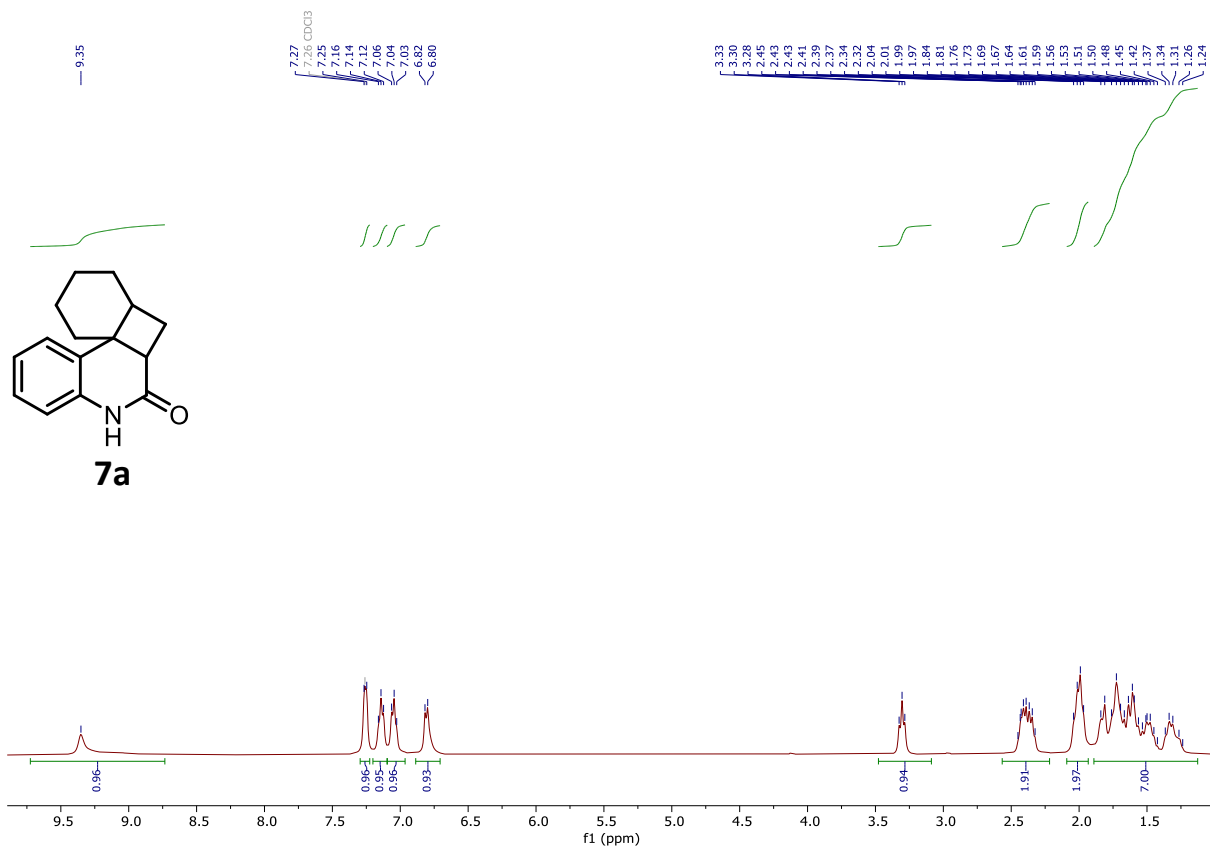


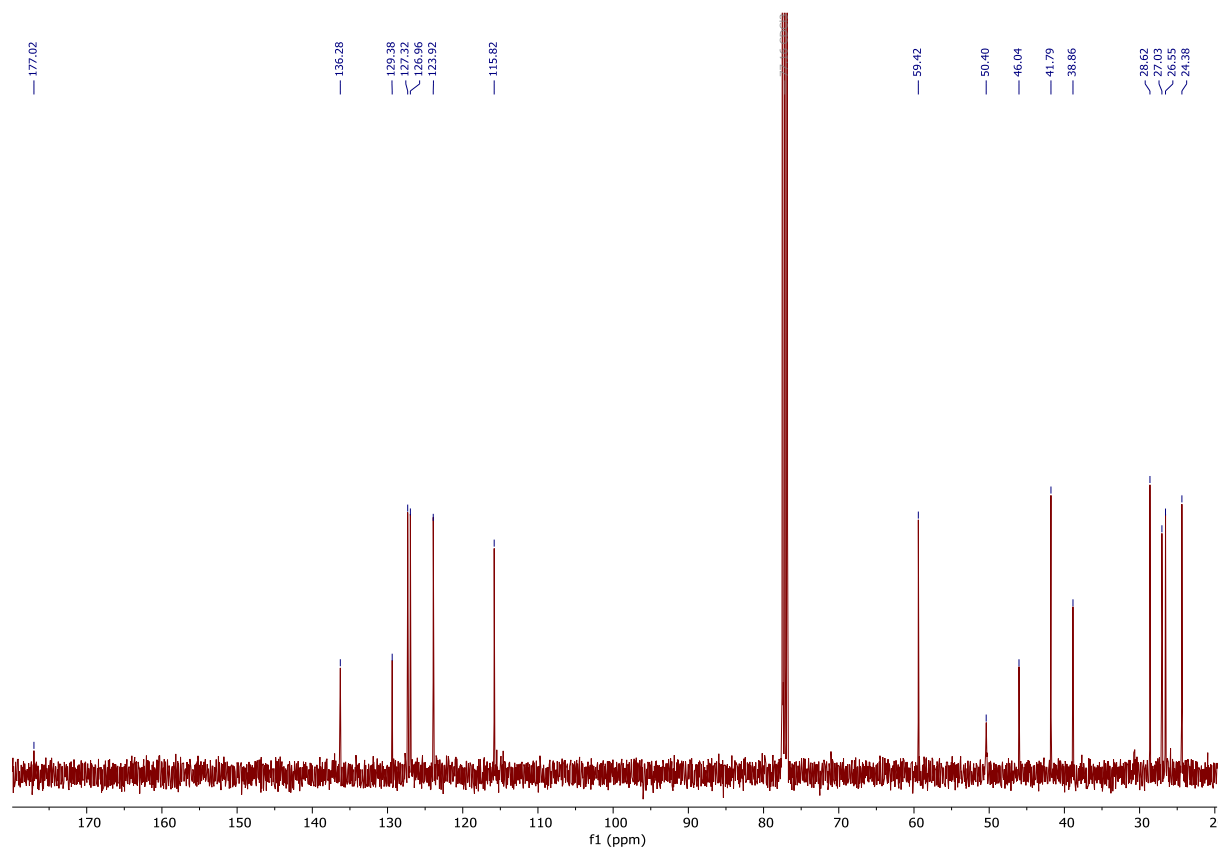
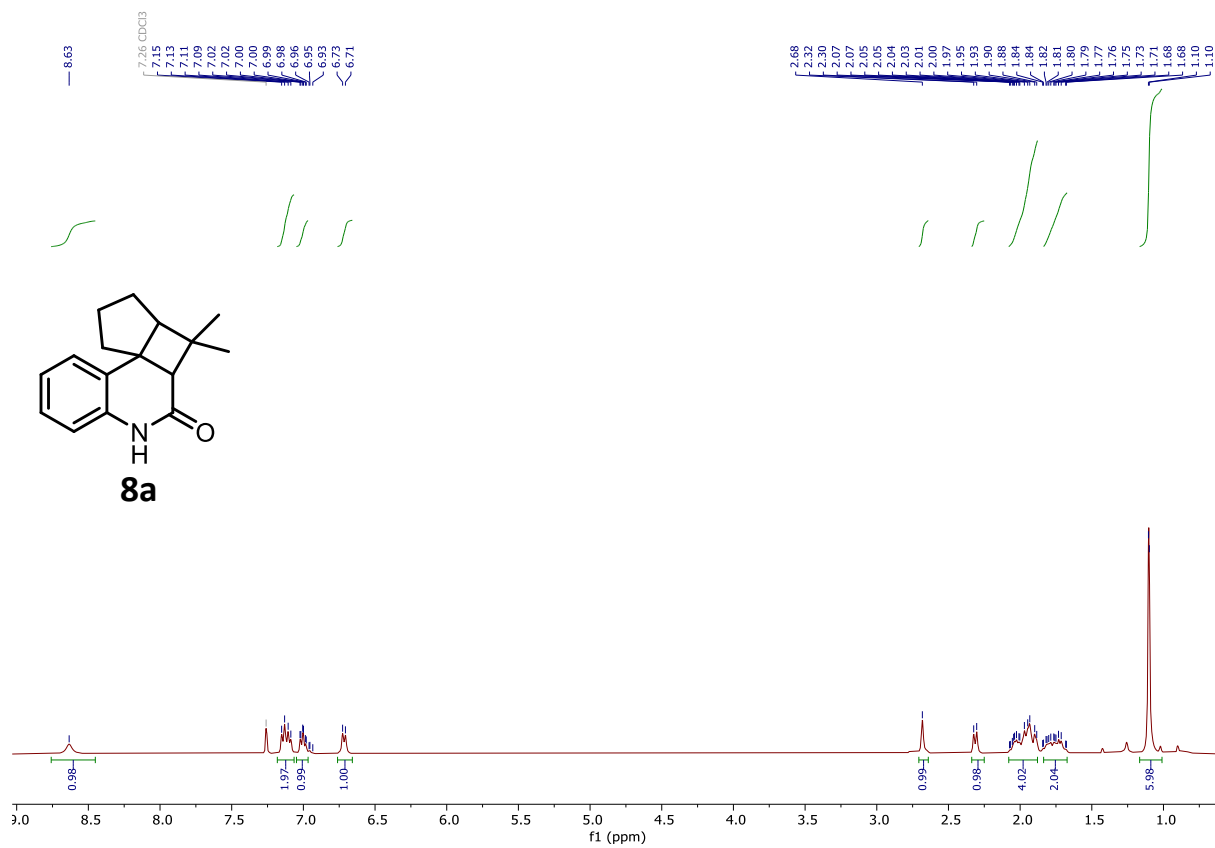


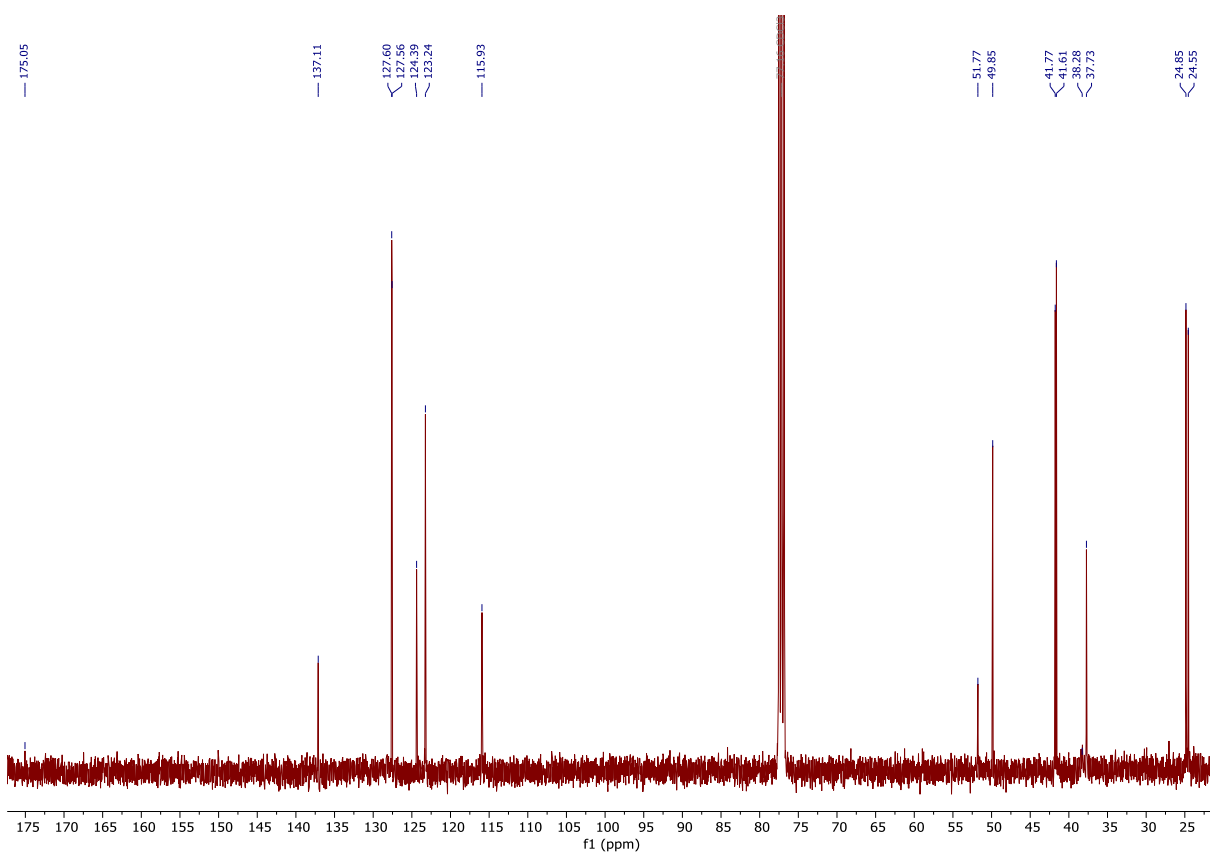
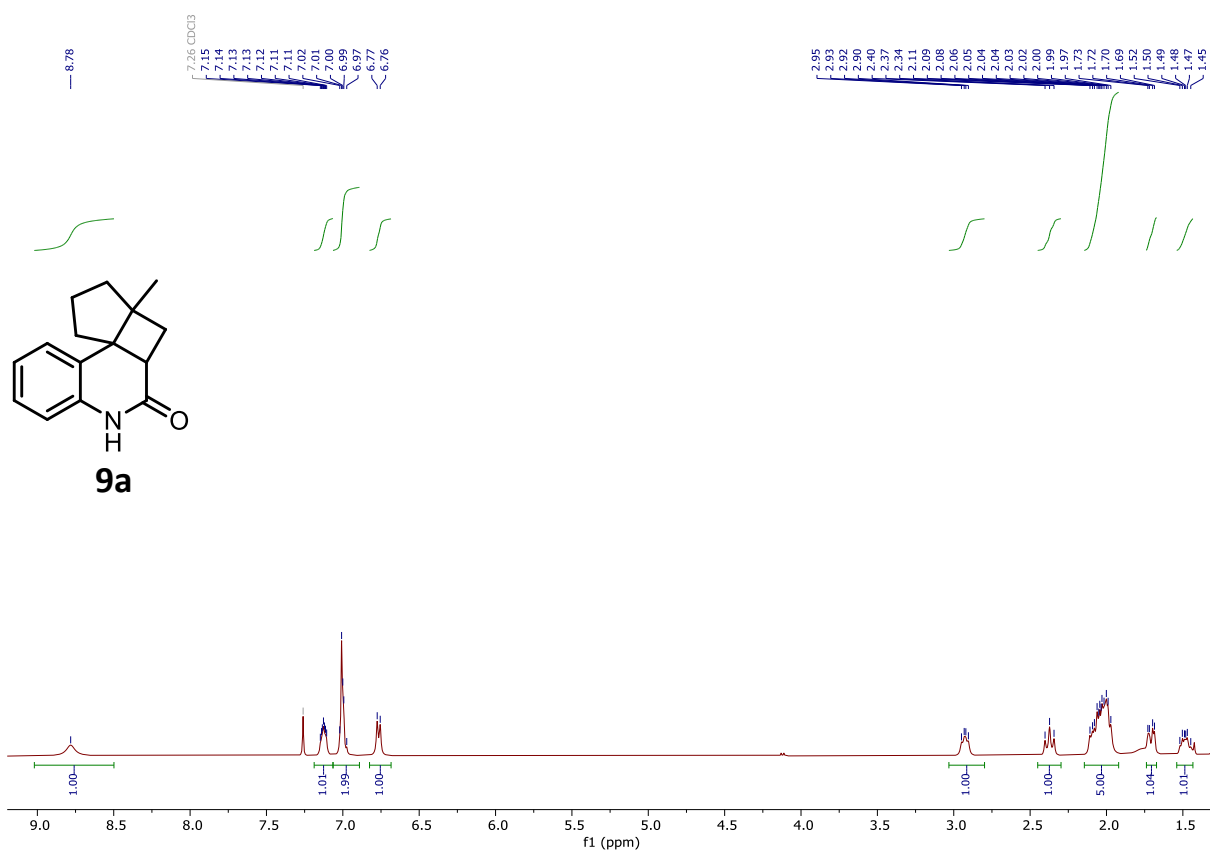


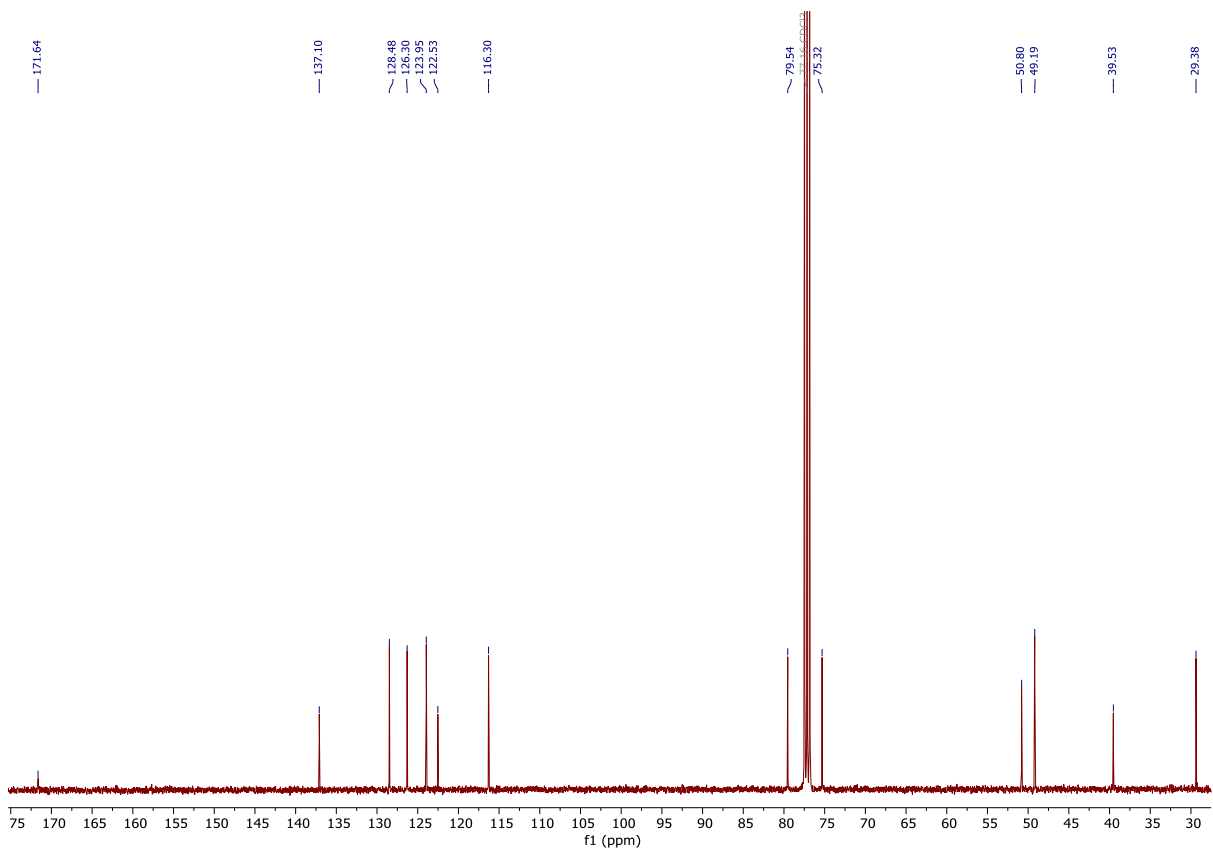
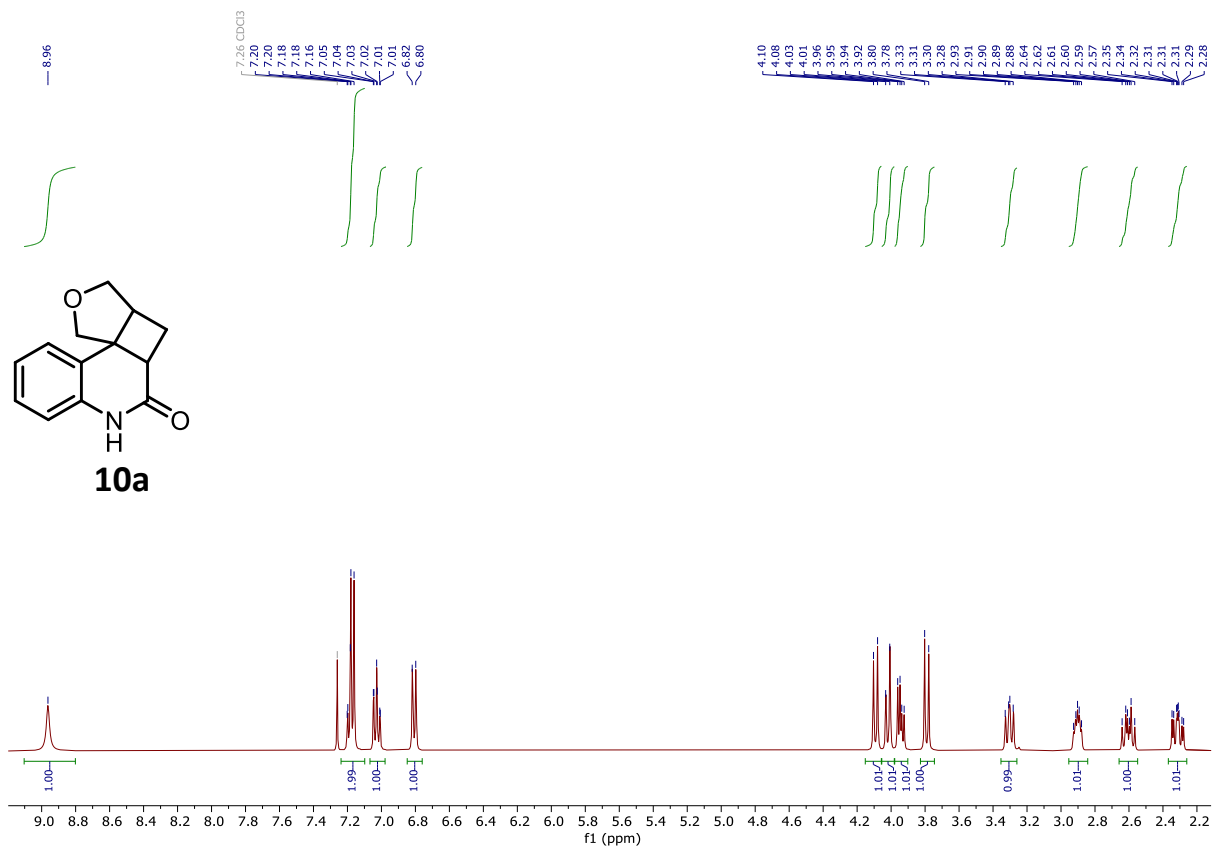


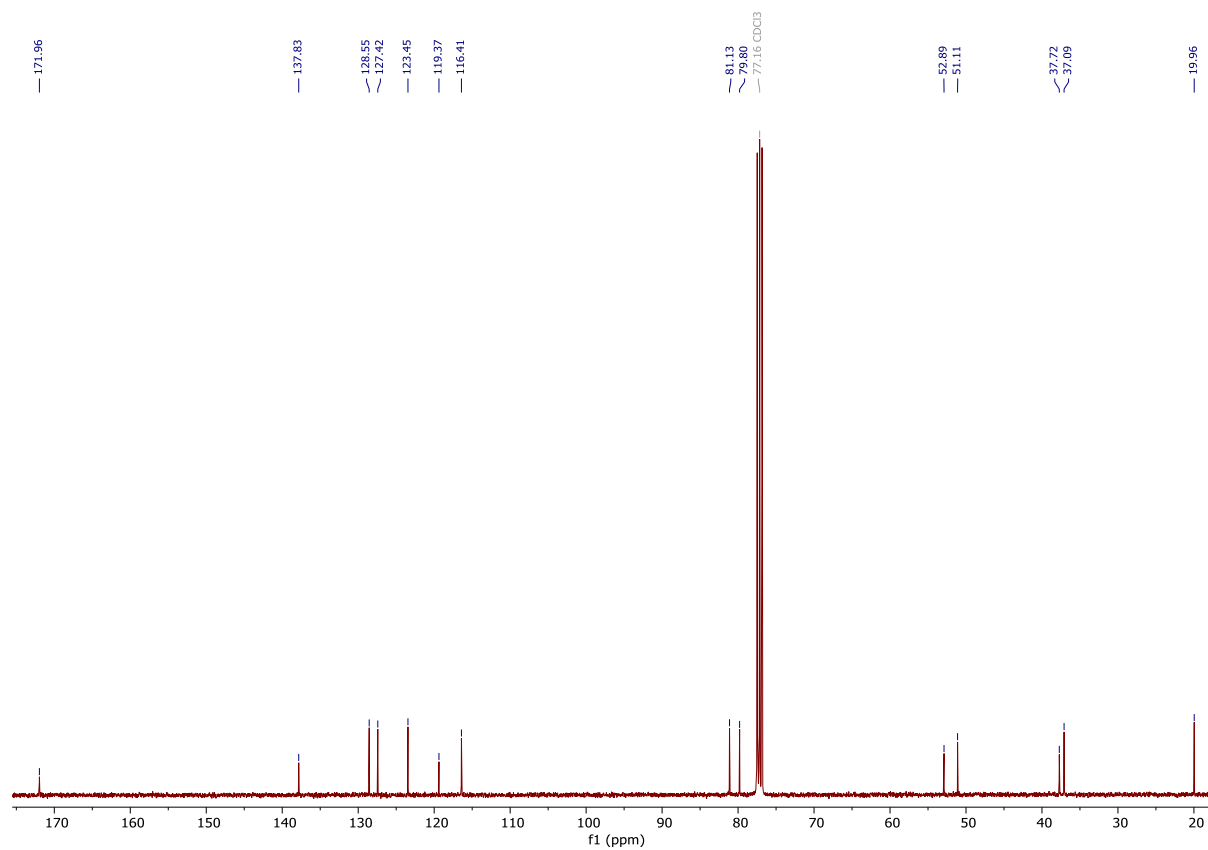
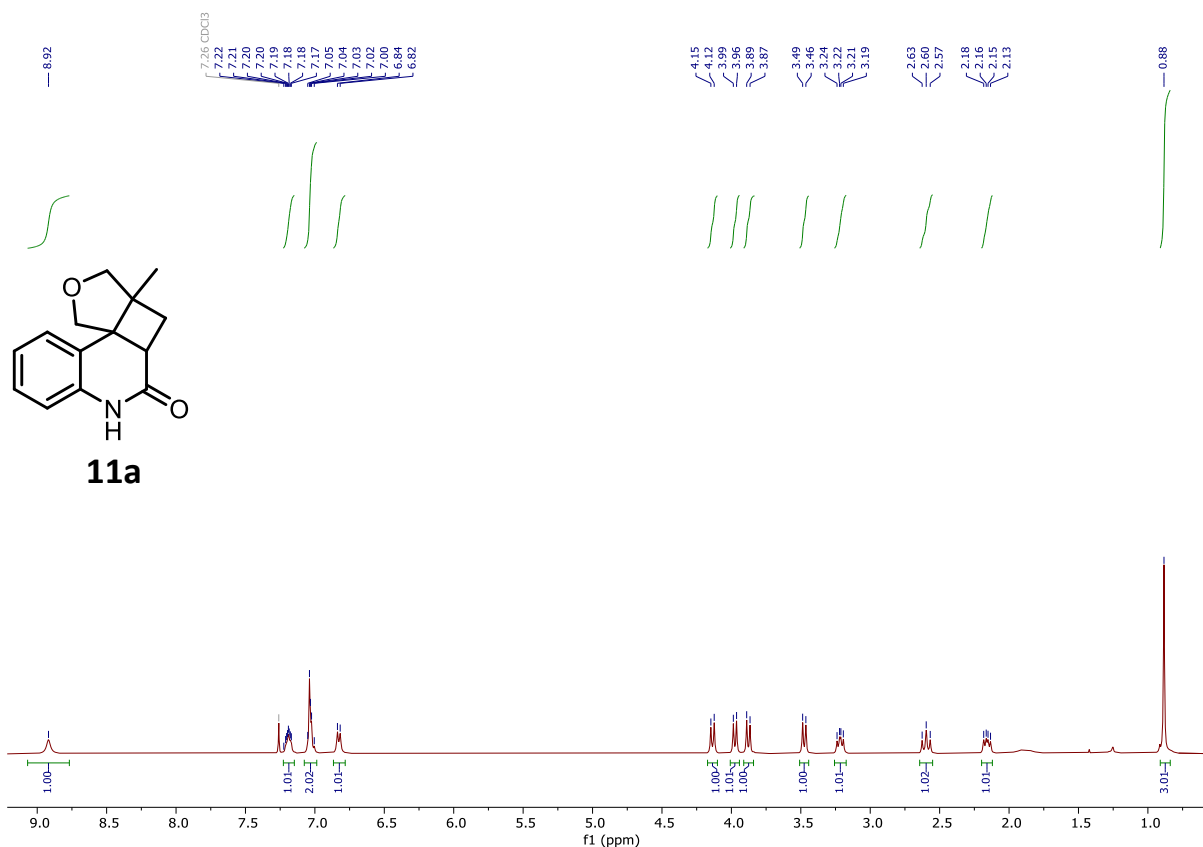


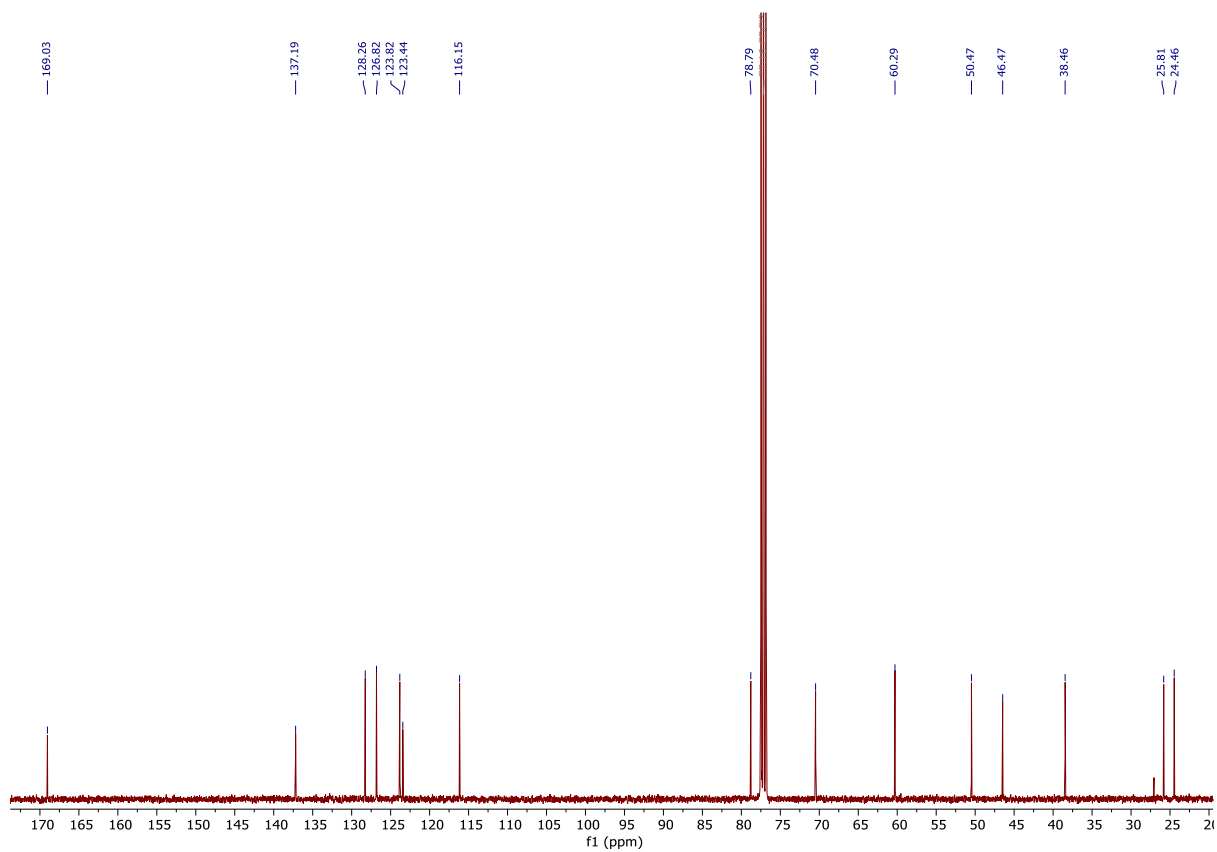
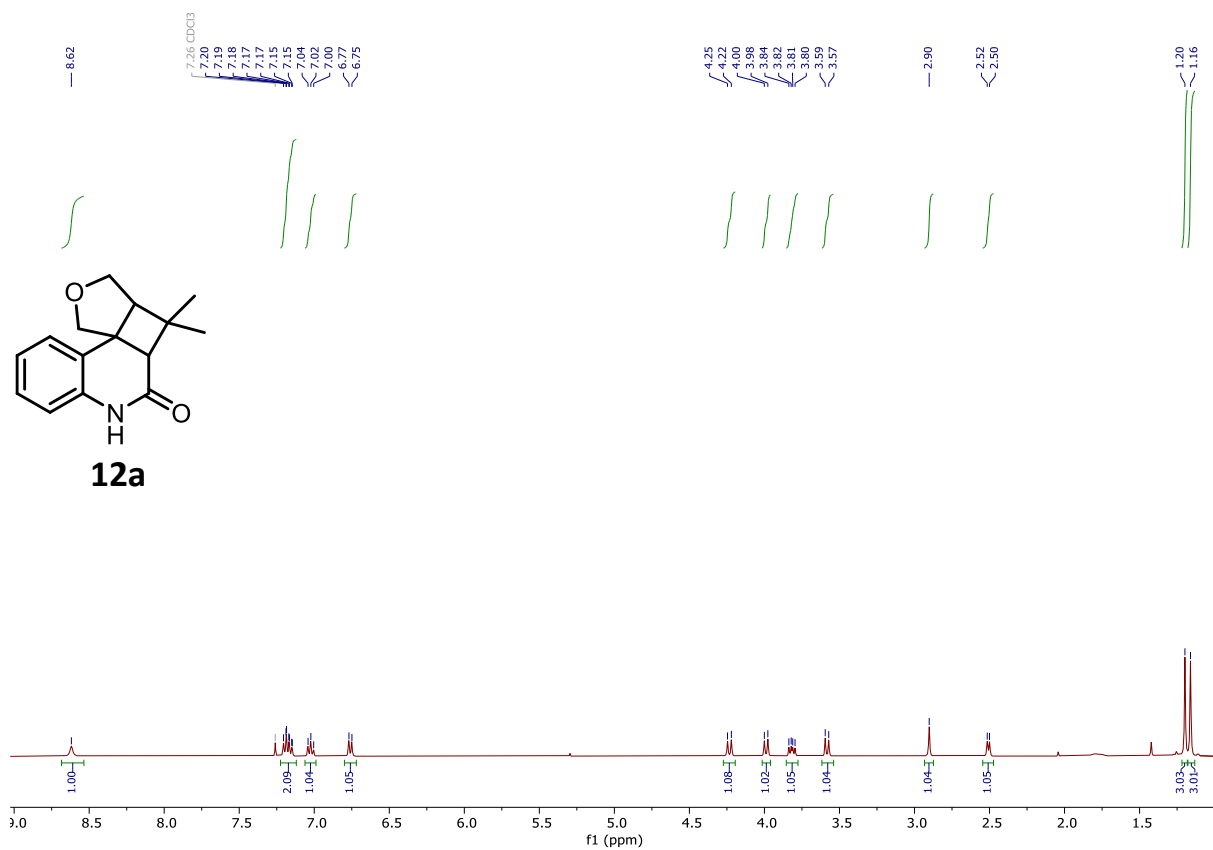


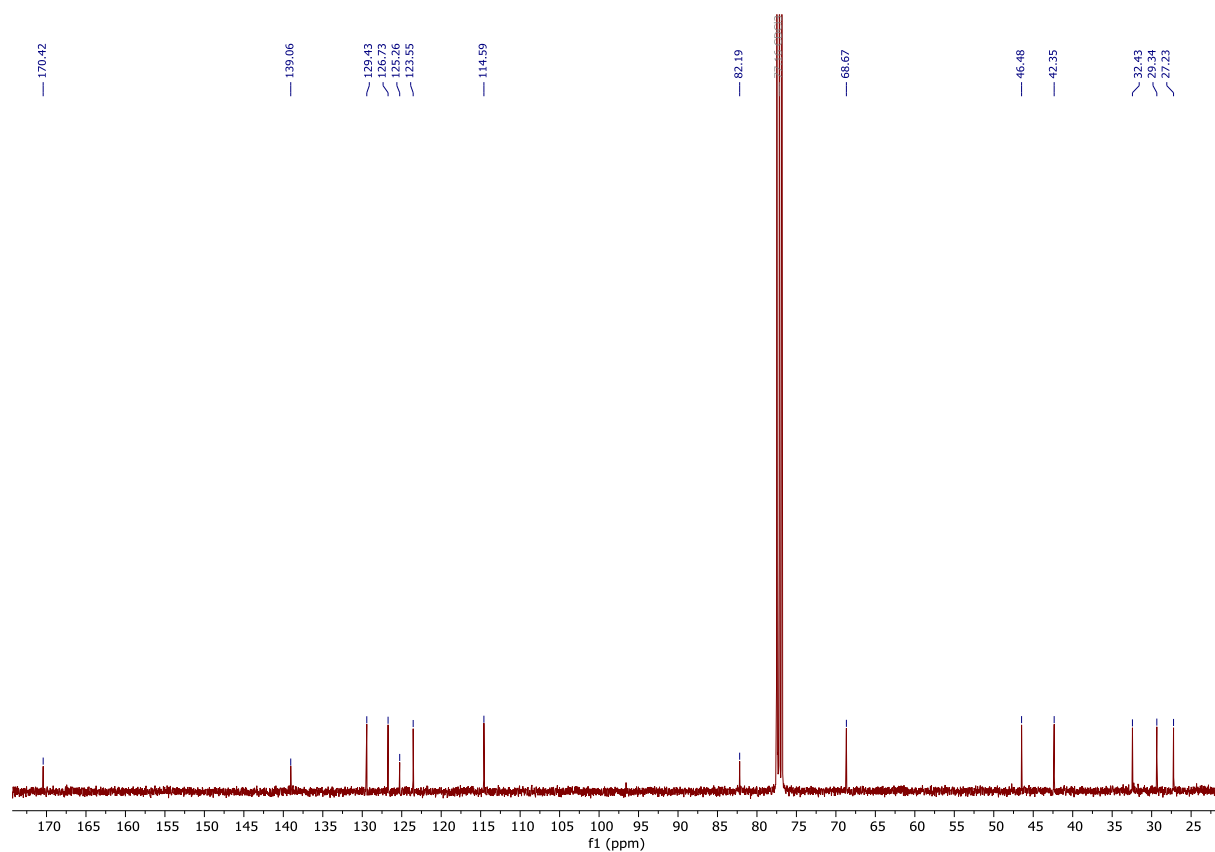
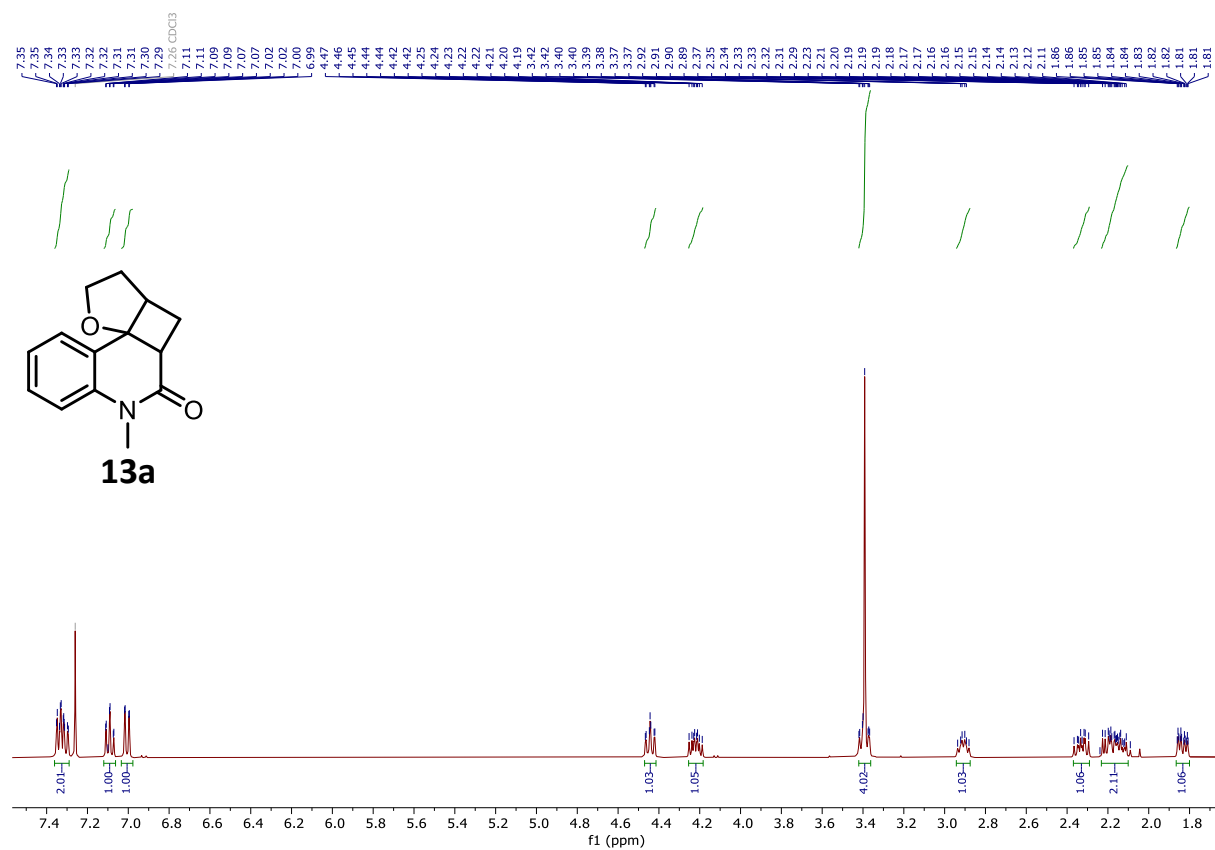


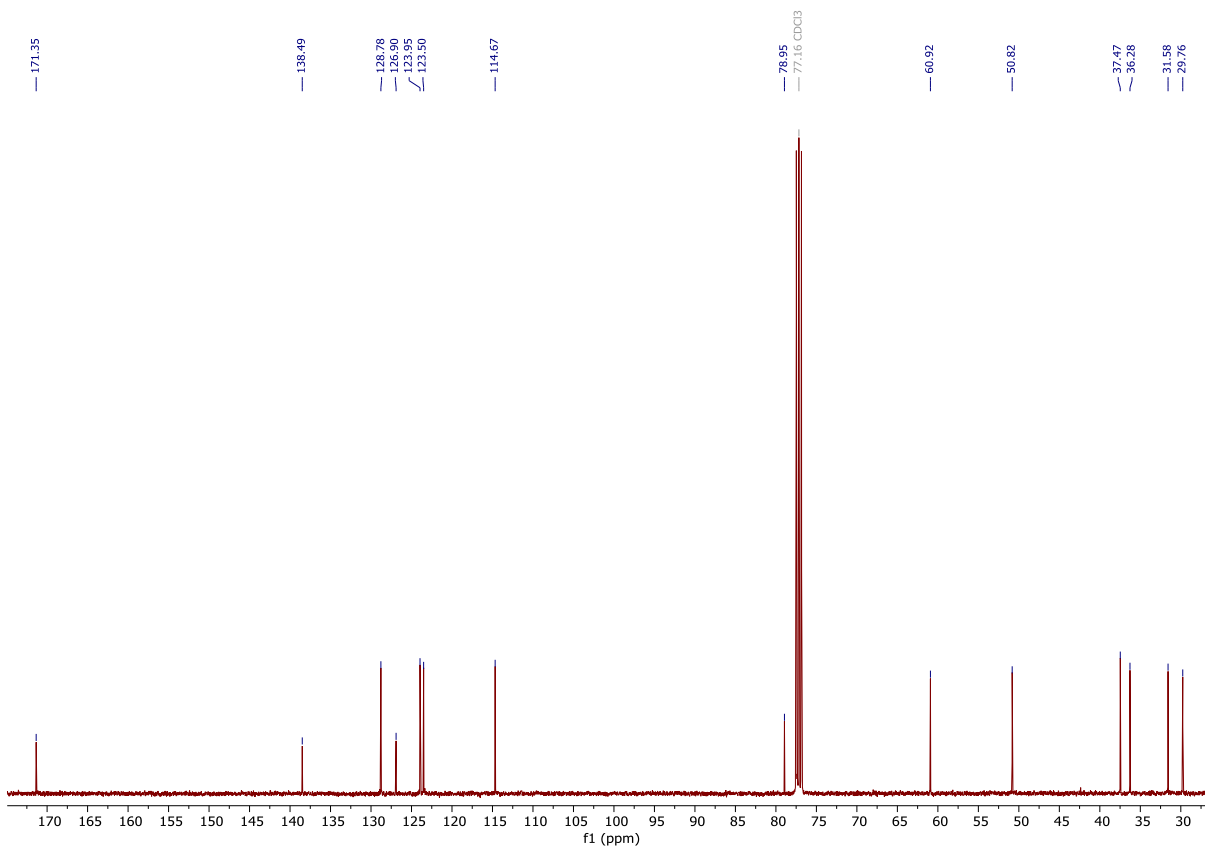
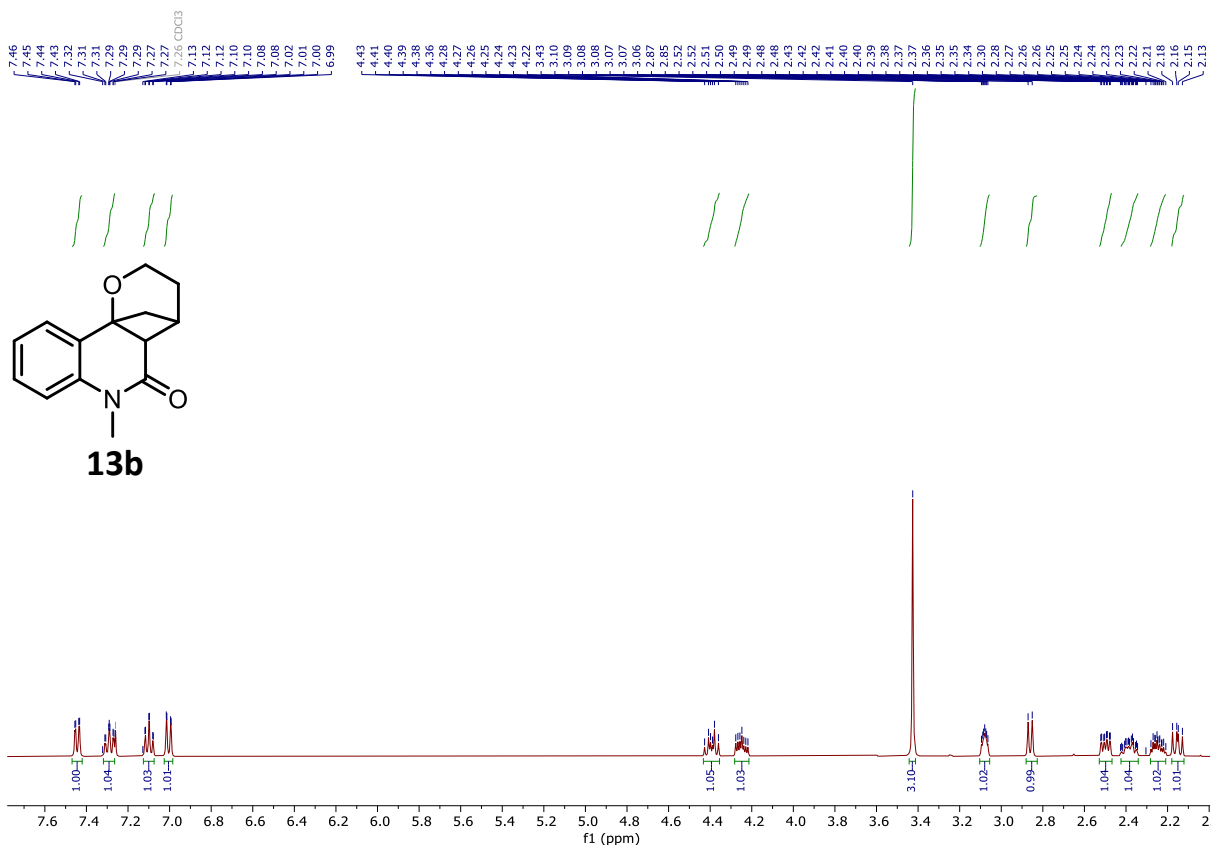


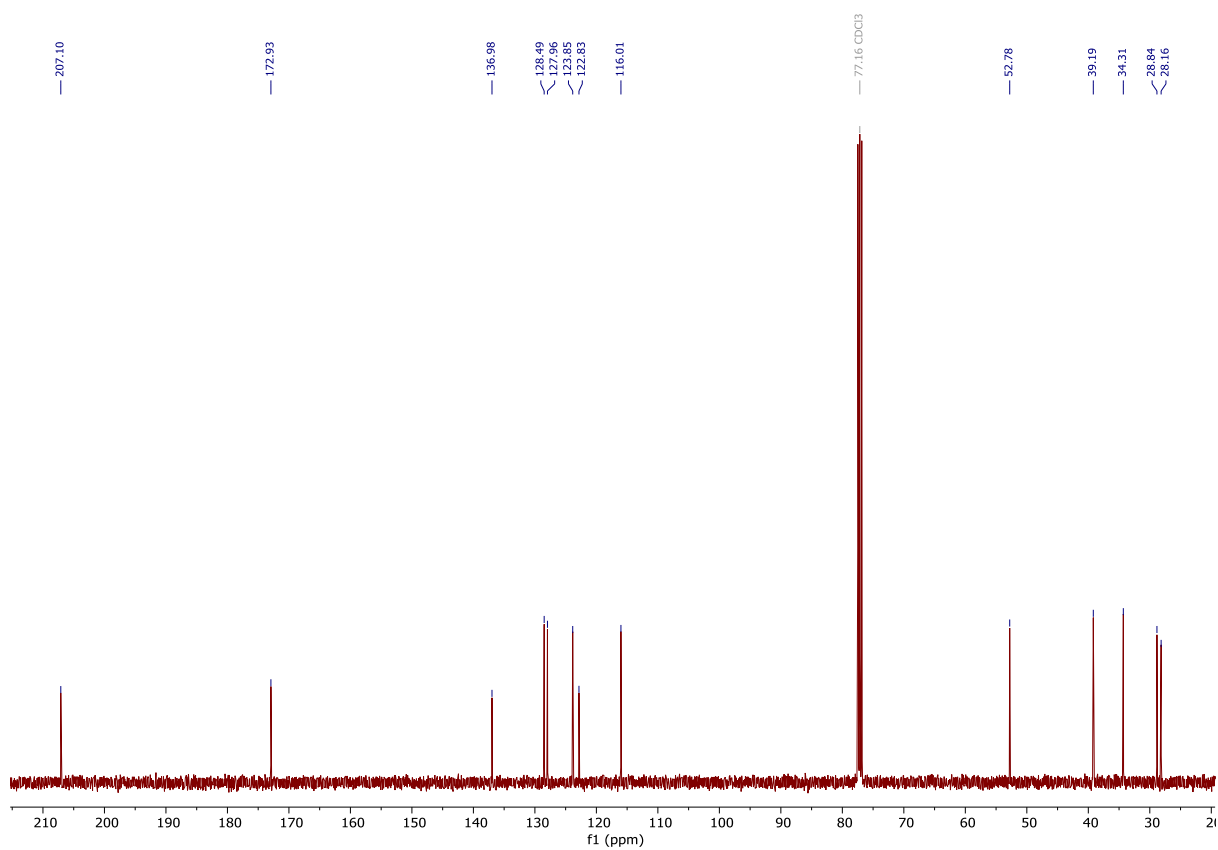
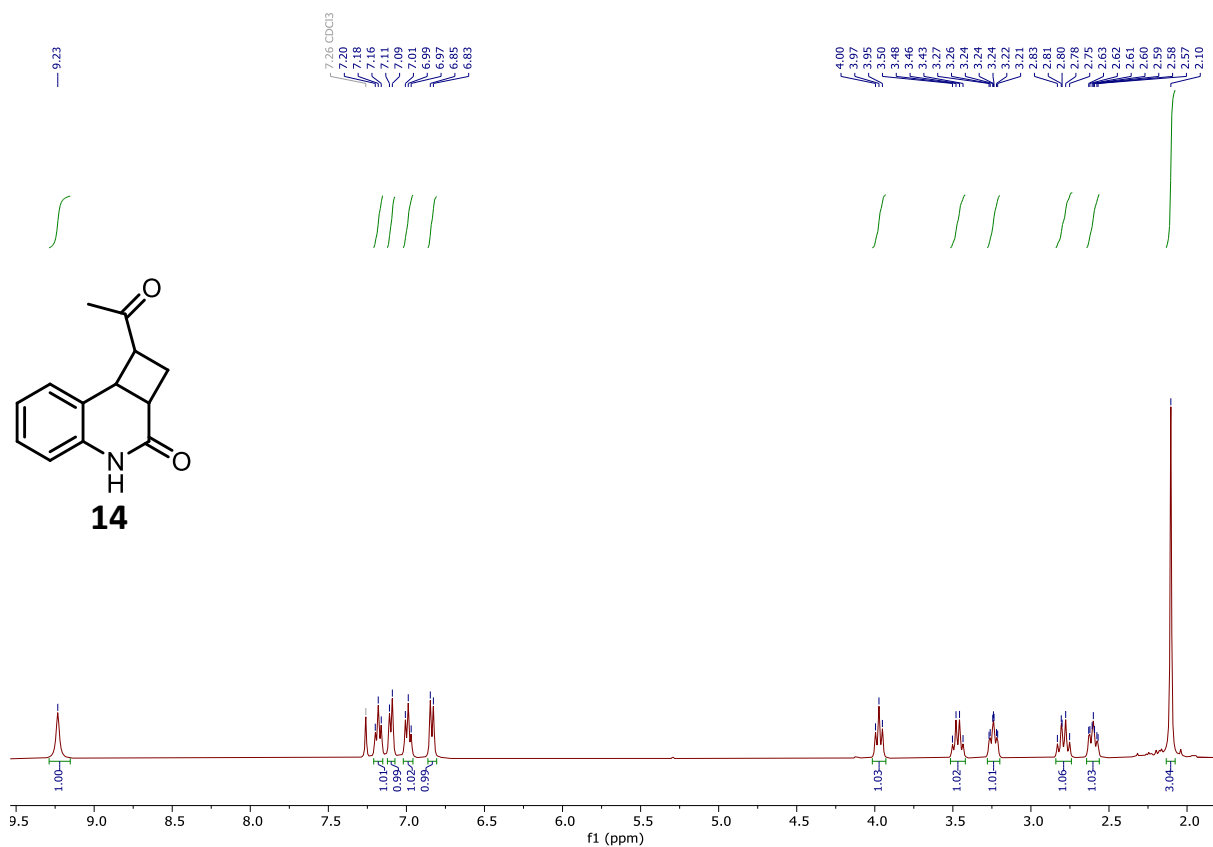


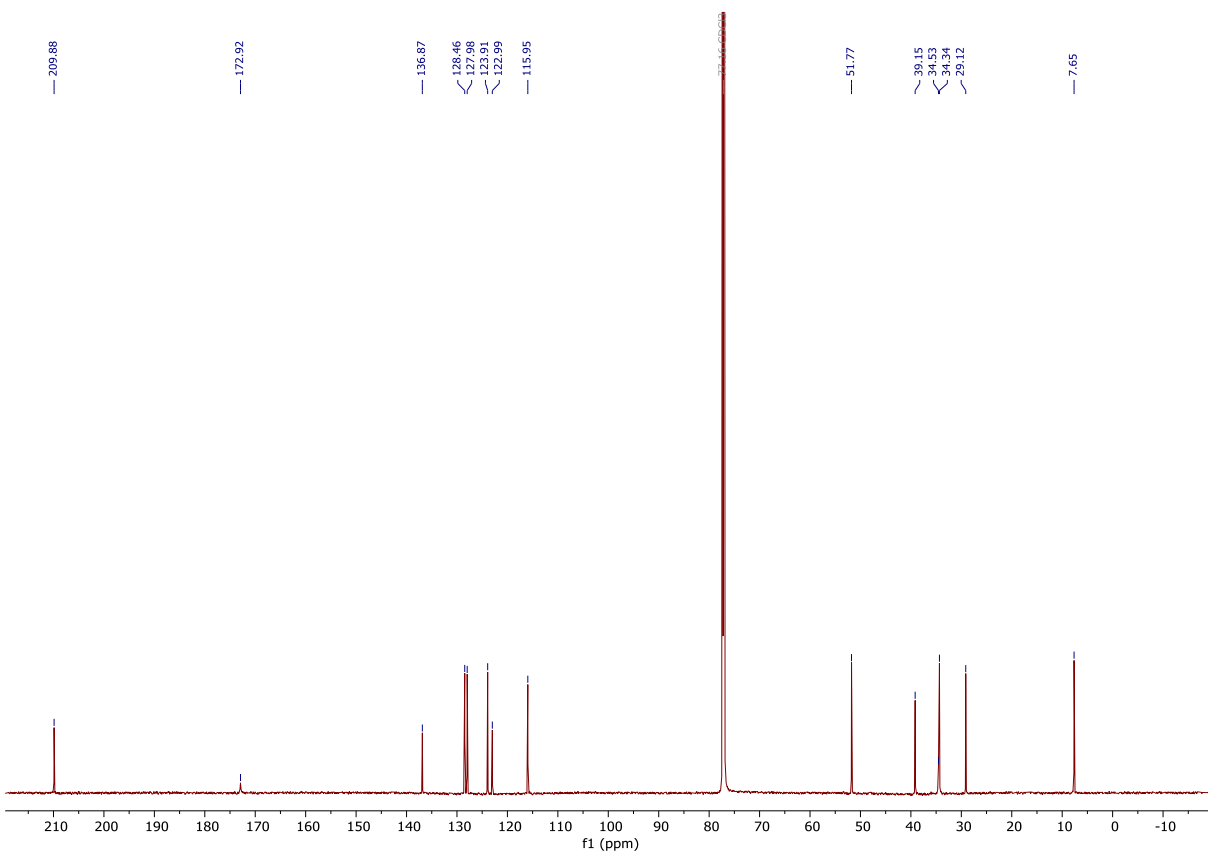
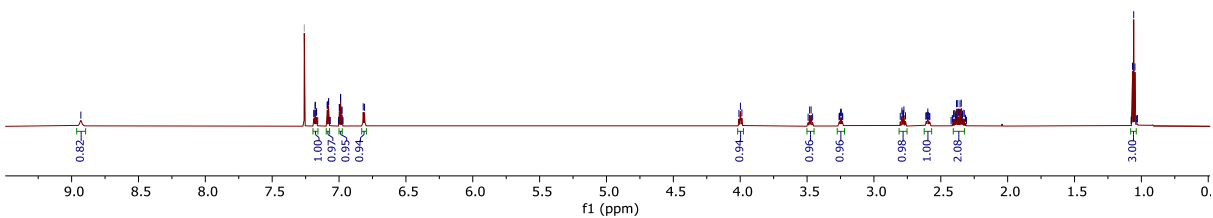




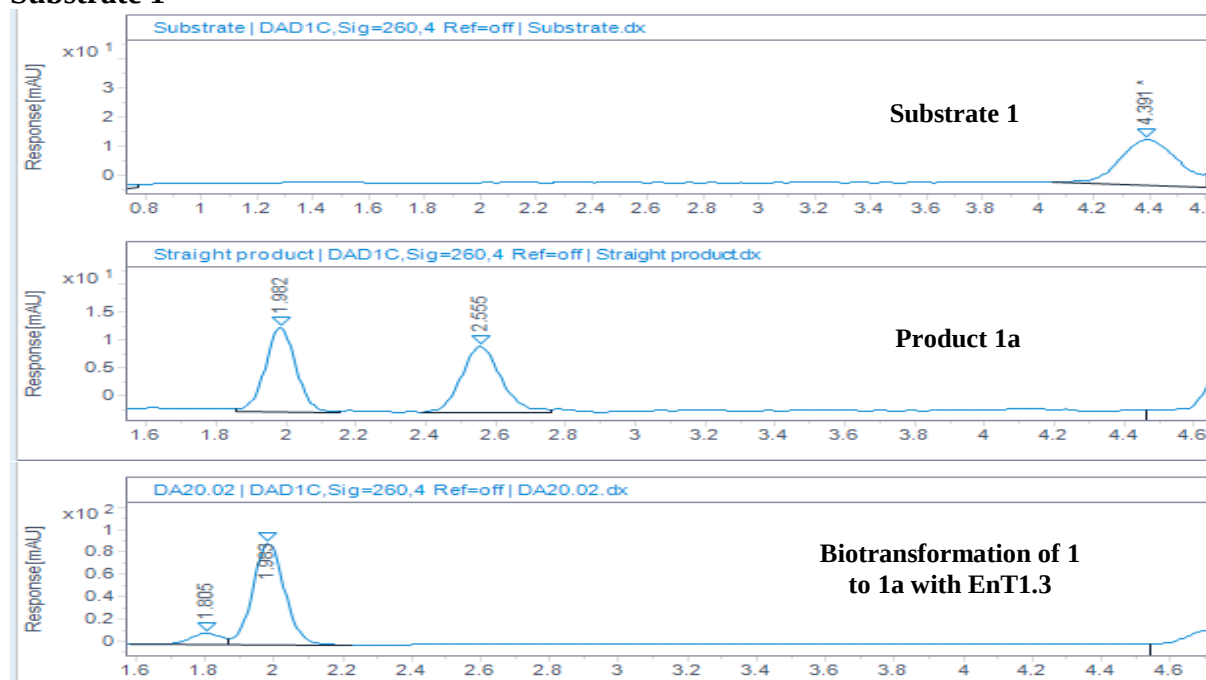






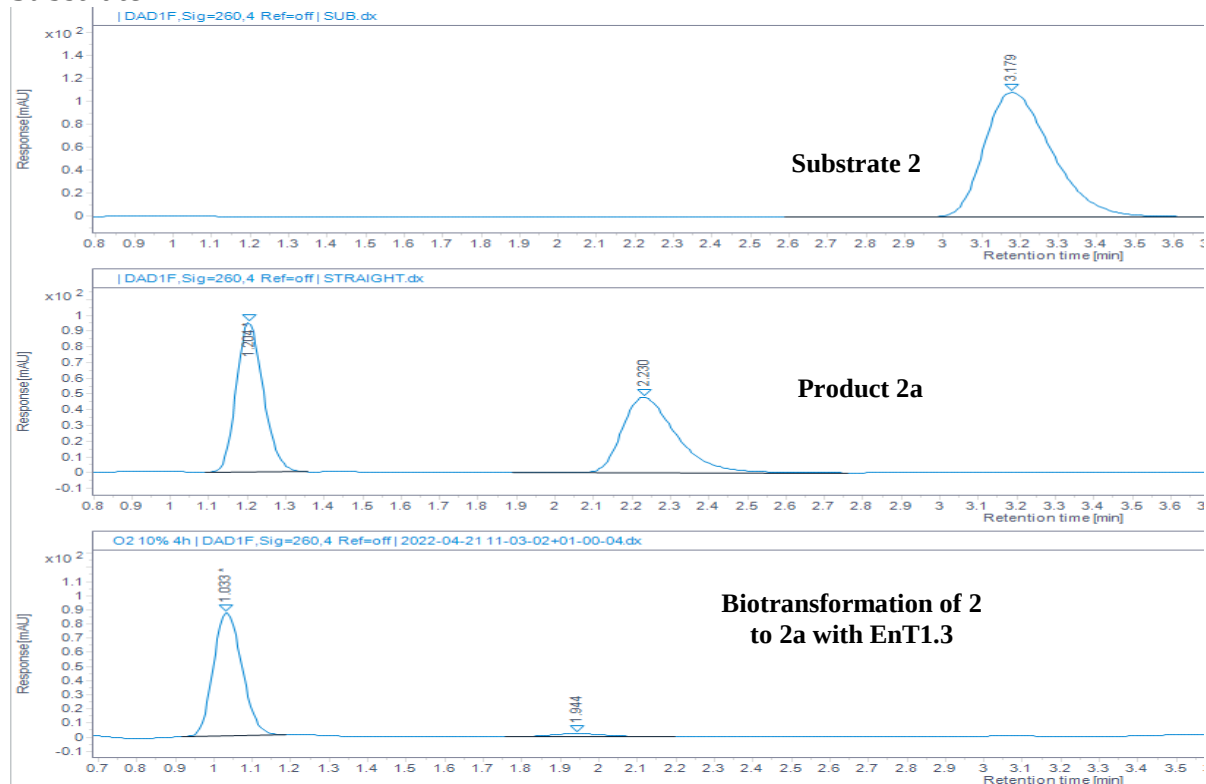


Substrate 1



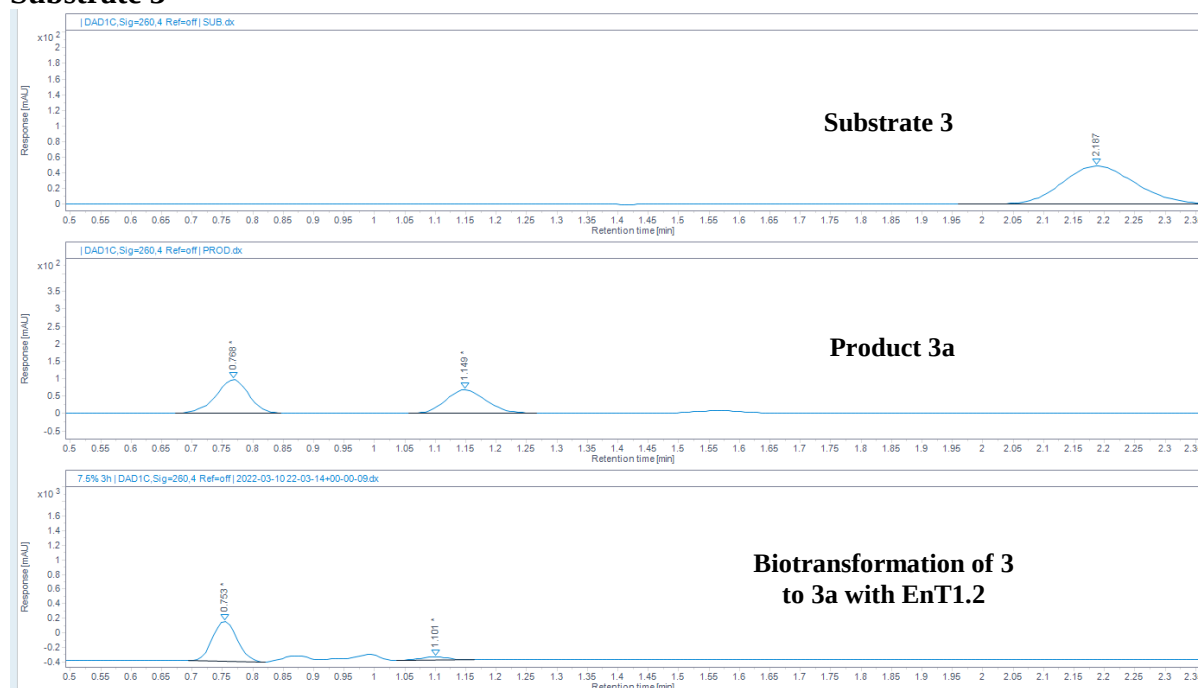
HPLC (IG-3 SFC column, 3 mm x 50 mm, CO₂/methanol=75:25)= **1**= 4.4 min; (-)-**1a**= 1.9 min; (+)-**1a**=2.5 min. (Note: the peak at 1.8 min is one enantiomer of product **1b**).

Substrate 2



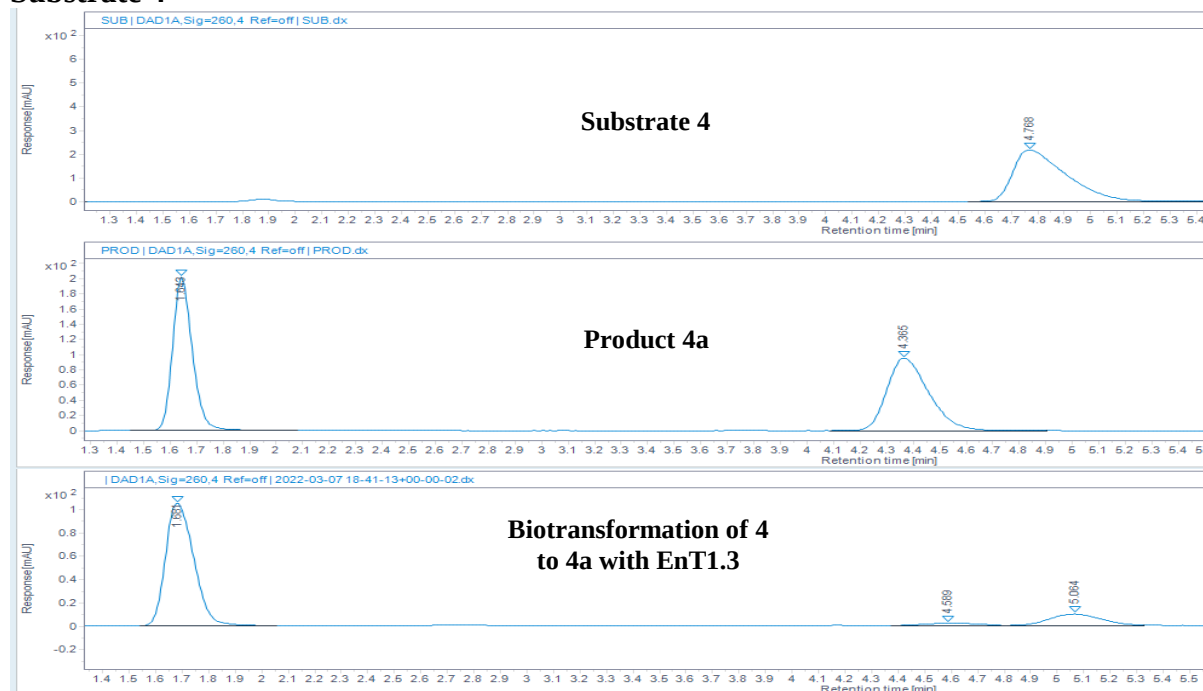
HPLC (IG-3 SFC column, 3 mm x 50 mm, CO₂/methanol=50:50)= **2**= 3.1 min; (-)-**2a**= 1.0 min; (+)-**2a**=1.9 min.

Substrate 3



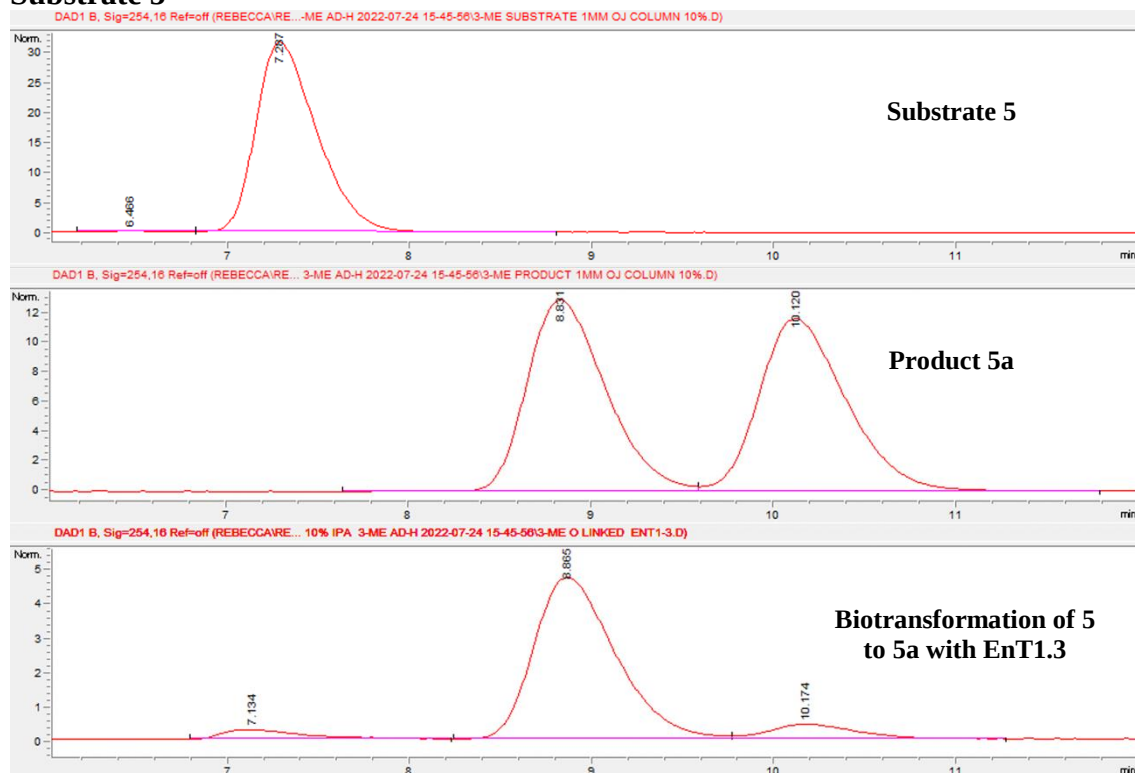
HPLC (IG-3 SFC column, 3 mm x 50 mm, $\text{CO}_2/\text{methanol}=50:50$)= **3**= 2.1 min; (-)-**3a**= 0.7 min; (+)-**3a**=1.1 min. With this substrate there are minor additional impurity peaks observed between 0.8-1 min that arise from a non-enzymatic background reaction.

Substrate 4



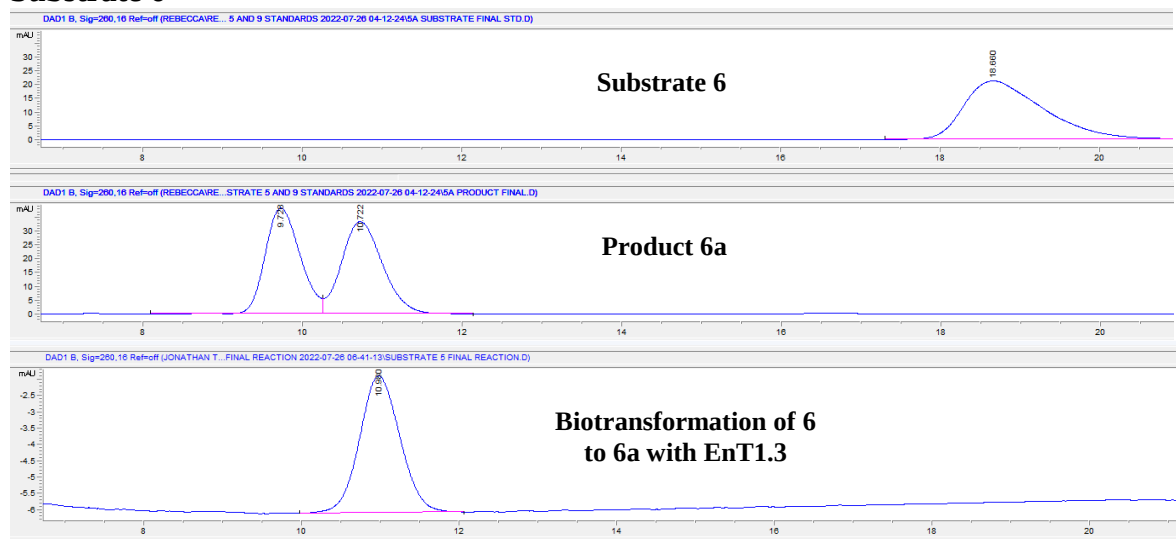
HPLC (IG-3 SFC column, 3 mm x 50 mm, $\text{CO}_2/\text{methanol}=75:25$)= **4**= 4.7 min; (-)-**4a**= 1.6 min; (+)-**4a**=4.5 min.

Substrate 5



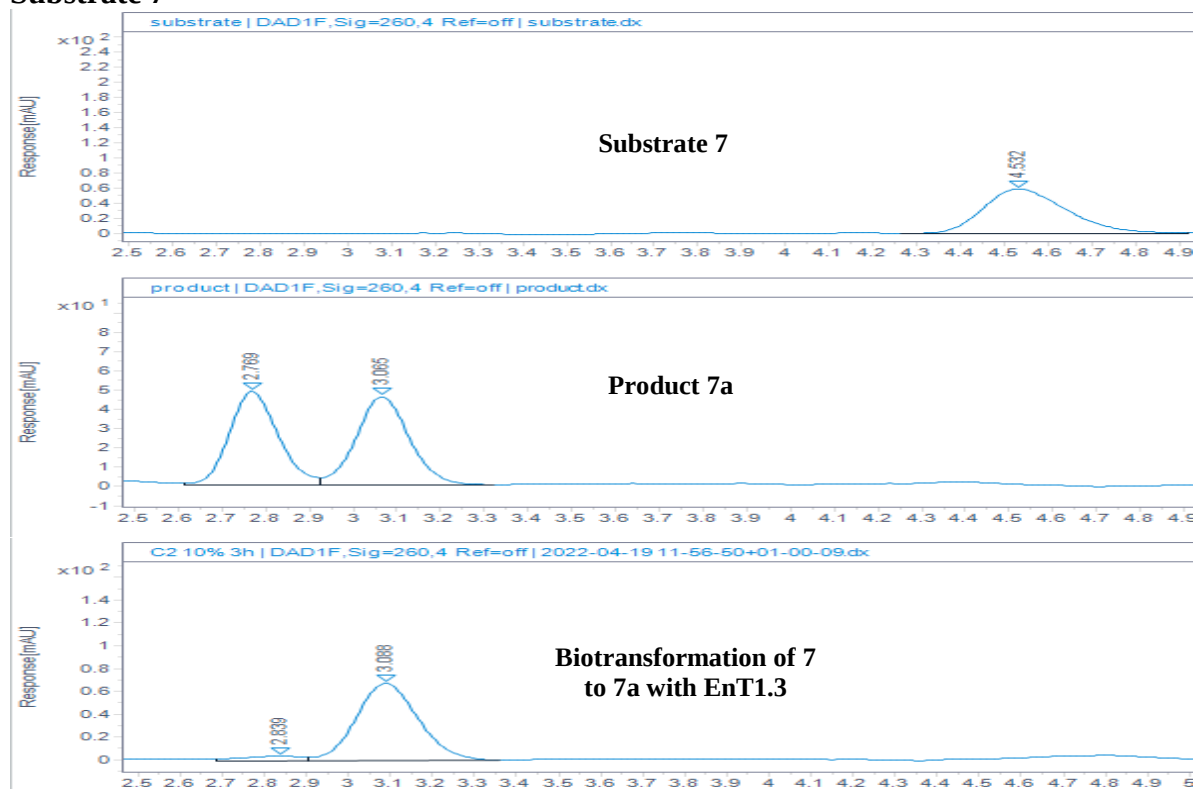
HPLC (AD-H, 0.46cm x 25cm, n-hexane/iso-propanol=90:10)= 5= 7.1 min; (-)-5a= 8.9 min; (+)-5a=10.2 min.

Substrate 6



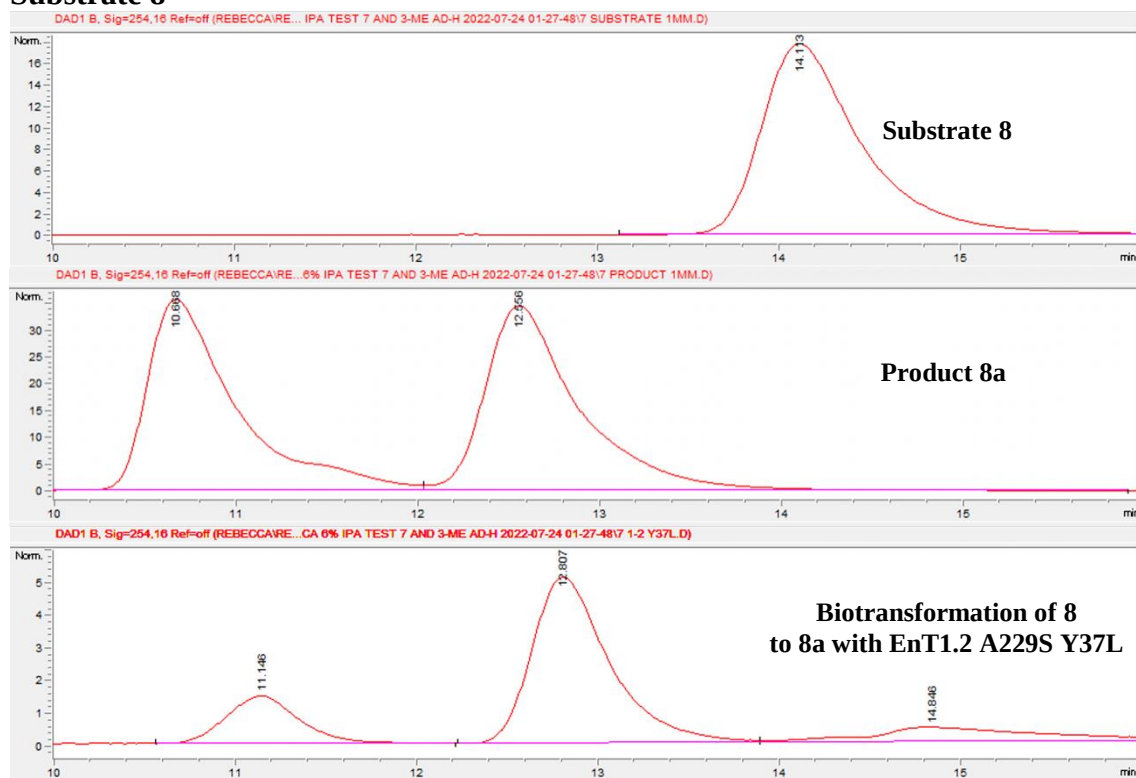
HPLC (IG-3 SFC column, 3 mm x 50 mm, n-hexane/iso-propanol=98:2)= 6= 18.6 min; (-)-6a= 10.7 min; (+)-6a= 9.7 min.

Substrate 7



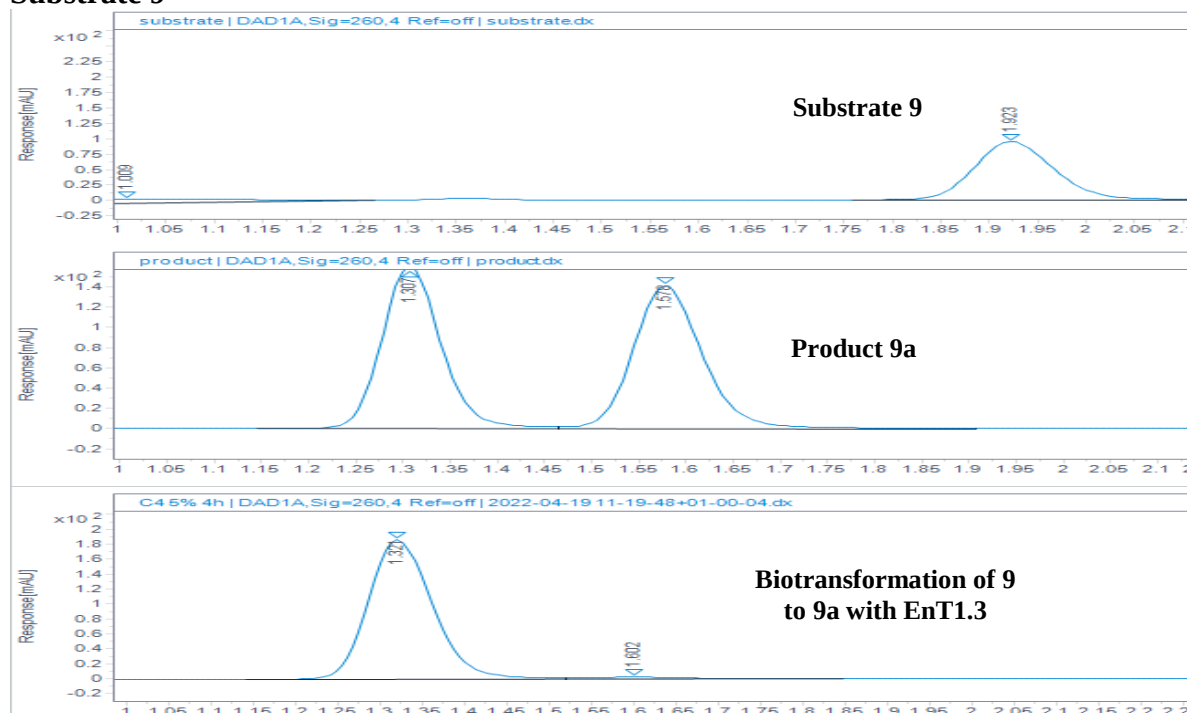
HPLC (IG-3 SFC column, 3 mm x 50 mm, CO₂/methanol=80:20)= 7= 4.5 min; (-)-7a= 3.0 min; (+)-7a=2.8 min.

Substrate 8



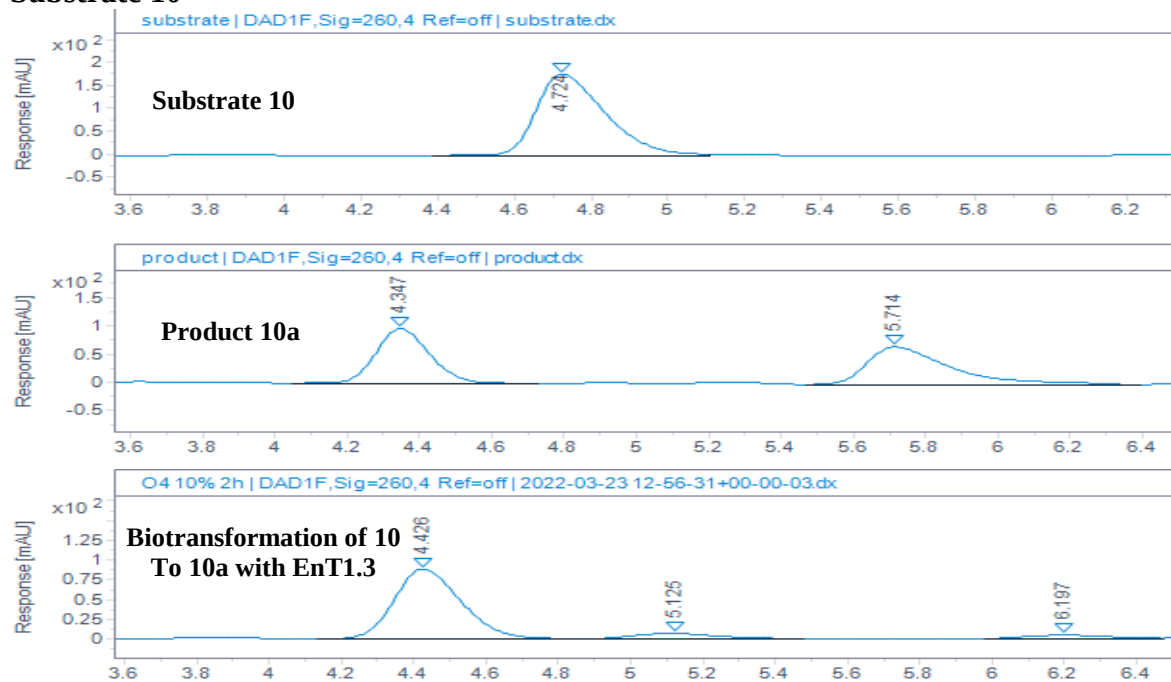
HPLC (AD-H, 0.46cm x 25cm, n-hexane/iso-propanol=94:6)= 7= 14.8 min; (-)-7= 12.8 min; (+)-7=11.1 min.

Substrate 9



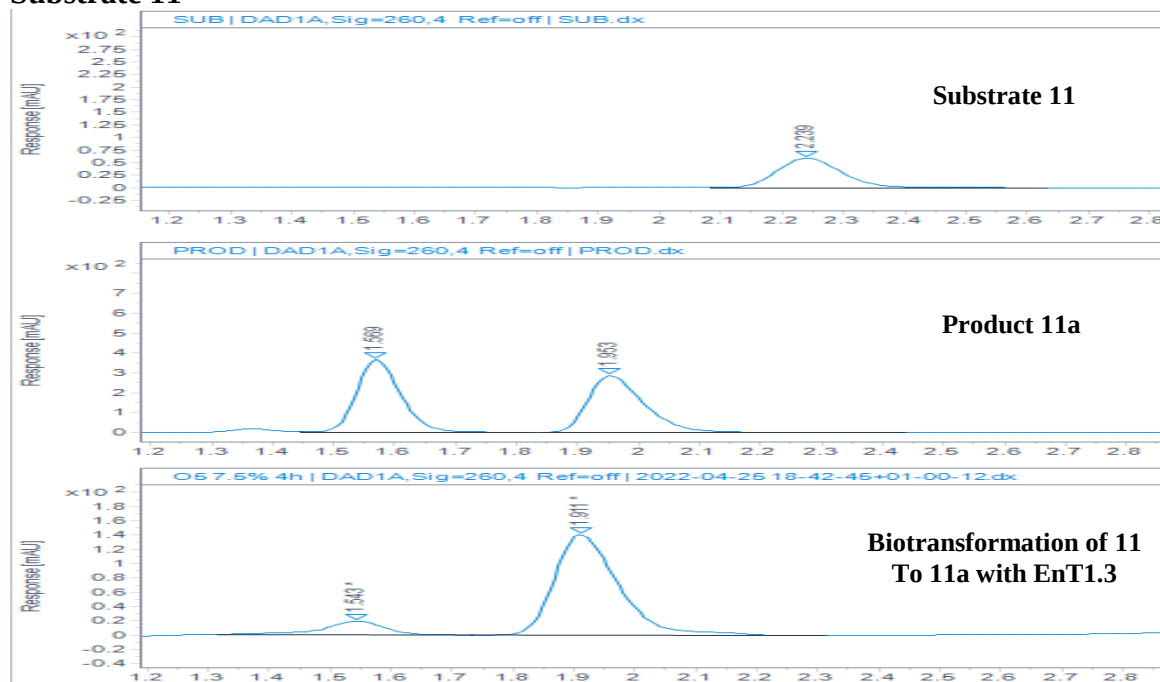
HPLC (IG-3 SFC column, 3 mm x 50 mm, CO₂/methanol=70:30)= **9**= 1.9 min; (-)-**9a**= 1.3 min; (+)-**9a**=1.6 min.

Substrate 10



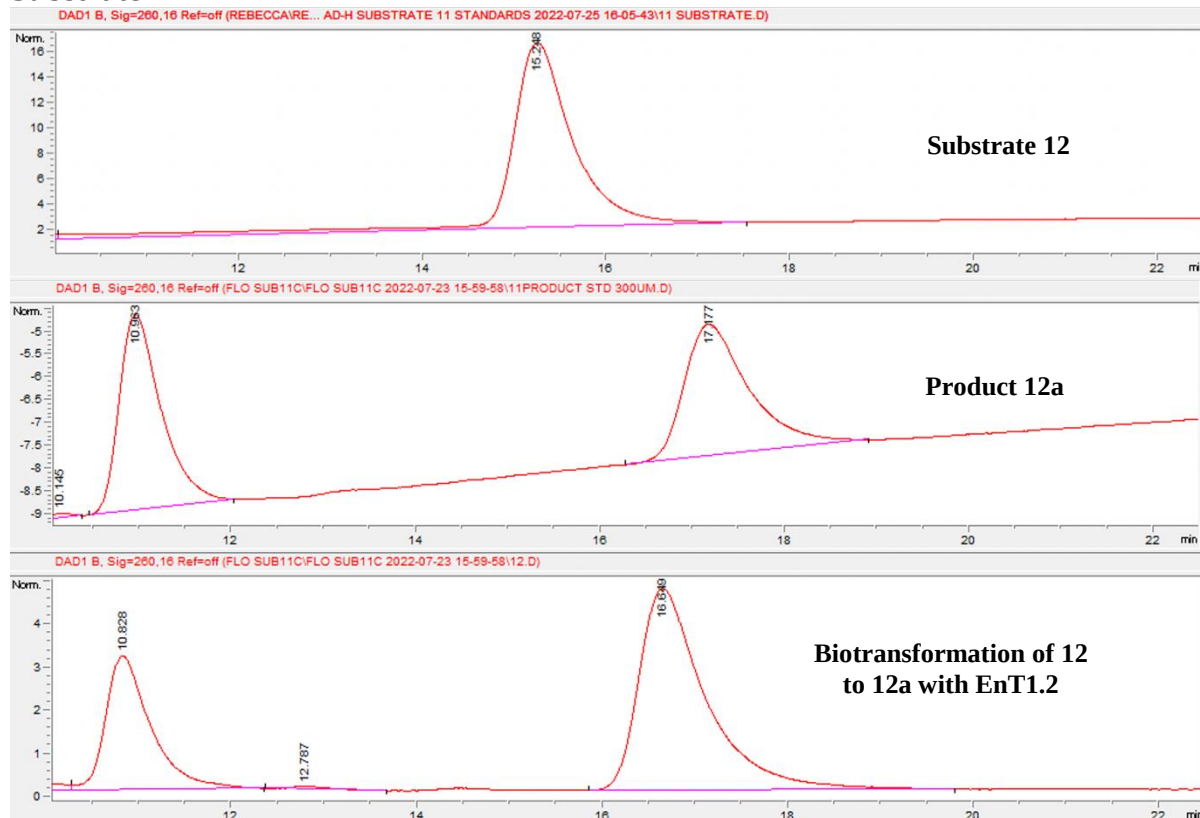
HPLC (IG-3 SFC column, 3 mm x 50 mm, CO₂/methanol=85:15)= **10**= 4.7 min; (-)-**10a**= 4.4 min; (+)-**10a**=5.7 min.

Substrate 11



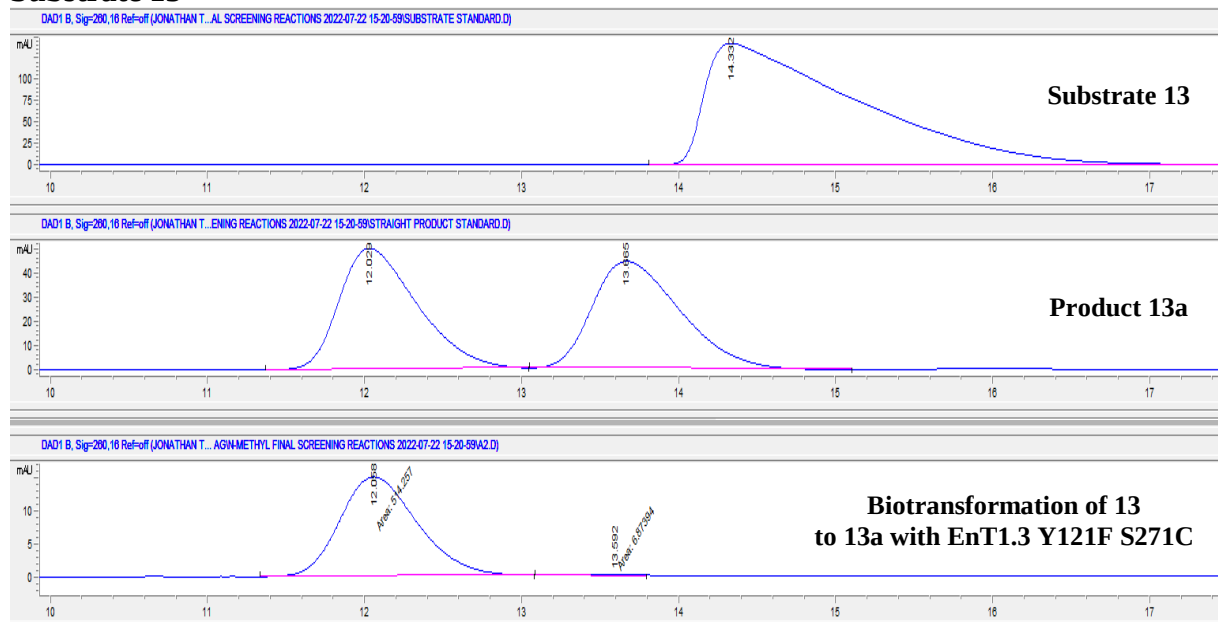
HPLC (IG-3 SFC column, 3 mm x 50 mm, CO₂/methanol=70:30)= **11**= 2.2 min; (-)-**11a**= 1.9 min; (+)-**11a**=1.5 min.

Substrate 12



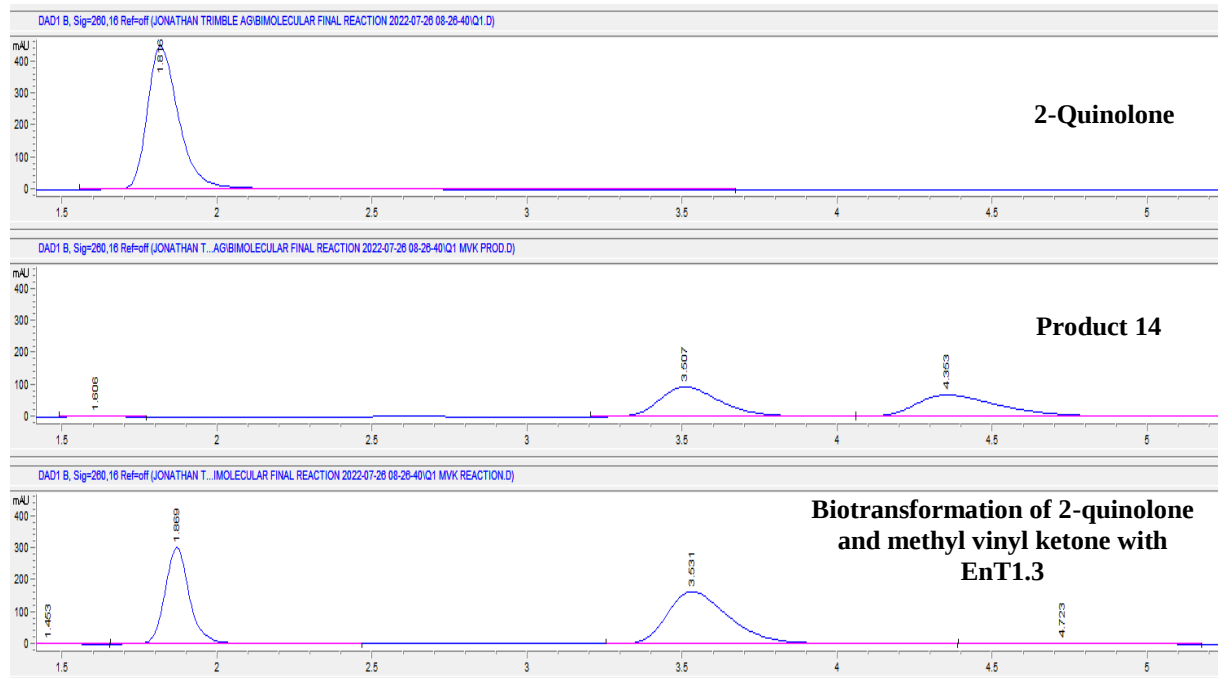
HPLC (AD-H, 0.46cm x 25cm, n-hexane/iso-propanol=90:10)= **12**= 15.2 min; (-)-**12**= 17 min; (+)-**12**=10.9 min.

Substrate 13



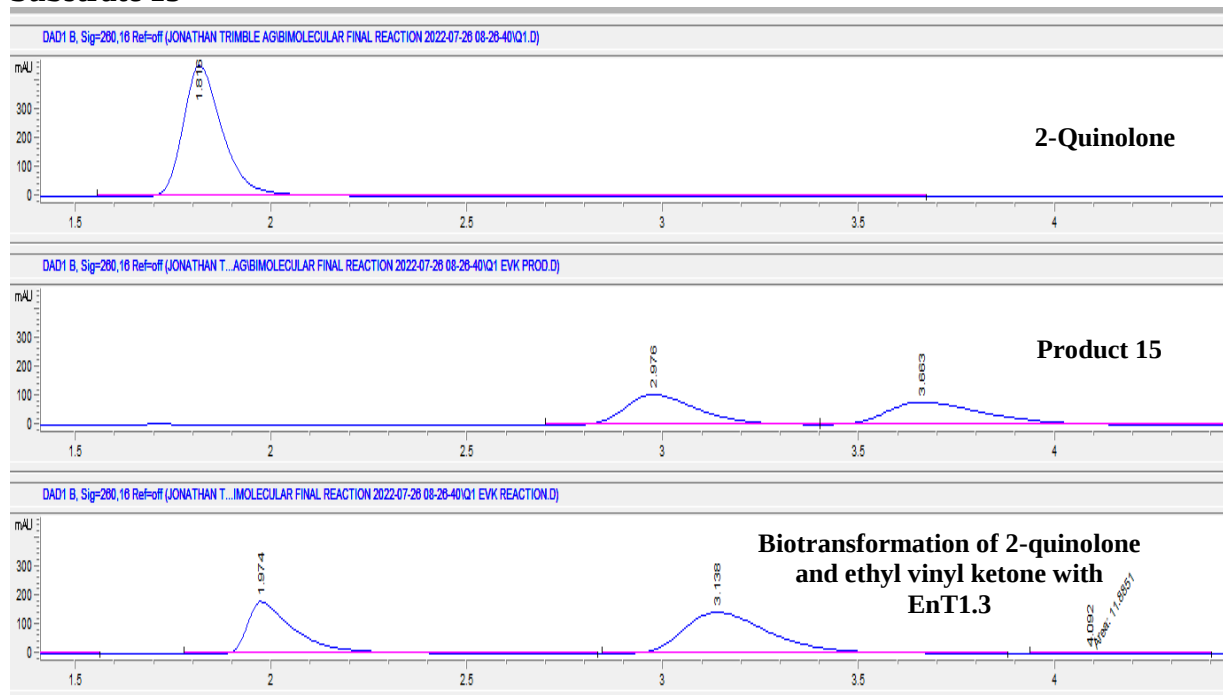
HPLC (IG-3 SFC column, 3 mm x 50 mm, n-hexane/iso-propanol=98:2)= **13**= 14.3 min; (-)-**13a**= 12.0 min; (+)-**13a**= 13.6 min.

Substrate 14



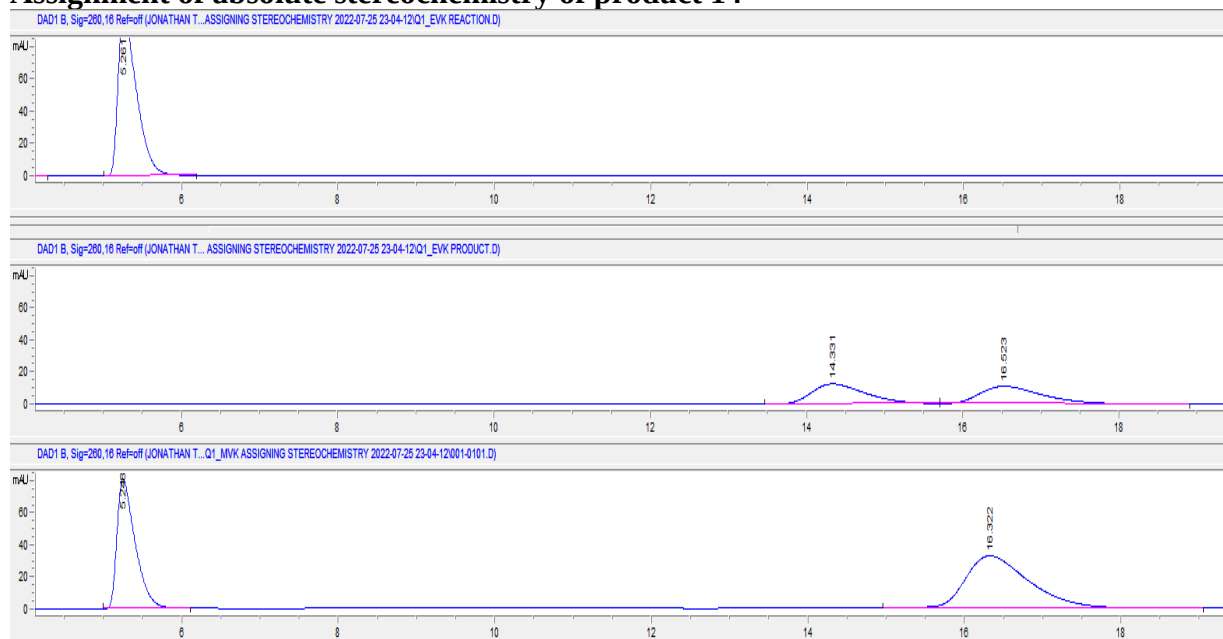
HPLC (IG-3 SFC column, 3 mm x 50 mm, n-hexane/iso-propanol=90:10)= **2-quinolone**= 1.8 min; (-)-**14**= 3.5 min; (+)-**14**= 4.4 min.

Substrate 15



HPLC (IG-3 SFC column, 3 mm x 50 mm, n-hexane/iso-propanol=90:10)= **2-quinolone**= 1.9 min; (-)-**15**= 3.1 min; (+)-**15**= 4.0 min.

Assignment of absolute stereochemistry of product 14



HPLC (OJ-H, 0.46cm x 25cm, n-hexane/iso-propanol=80:20)= **2-quinolone**= 5.2 min; (-)-**14**= 16.3 min; (+)-**14**=14.3 min. This HPLC method is identical to that described in reference 37 in order to assign the absolute stereochemistry of product **14** based on comparison of retention times.