

Supplementary Table legends

- Supplementary Table 1:** Demographic and clinical characteristics of participants included in the spatial transcriptomics study are provided.
- Supplementary Table 2:** Demographic and clinical characteristics of single-nucleus sequencing samples used for dataset integration are summarized.
- Supplementary Table 3:** Differentially expressed genes between annotated cell types in the CosMx dataset are listed.
- Supplementary Table 4:** Differentially expressed genes between annotated cell types in the Xenium dataset are listed.
- Supplementary Table 5:** Differentially expressed genes between annotated cell types in the imputed dataset are provided.
- Supplementary Table 6:** Differentially expressed genes between spatial niches in the CosMx dataset are presented.
- Supplementary Table 7:** Differentially expressed genes between spatial niches in the Xenium dataset are presented.
- Supplementary Table 8:** Differentially expressed genes between spatial niches in the imputed dataset are presented.
- Supplementary Table 9:** Differentially expressed genes between DKD and control samples in the imputed dataset are listed.
- Supplementary Table 10:** Cell–cell interactions identified within PT, IPT, LOH, and iLOH niches are detailed.
- Supplementary Table 11:** Cell–cell interactions identified within DCT, glomerular, vascular, fibroblast, immune, CD, and CNT niches are detailed.
- Supplementary Table 12:** Differentially expressed genes between center-cell IPT microenvironments in the CosMx dataset are listed.
- Supplementary Table 13:** Differentially expressed genes between center-cell iLOH microenvironments in the CosMx dataset are listed.
- Supplementary Table 14:** Differentially expressed genes between center-cell IPT microenvironments in the Xenium dataset are listed.
- Supplementary Table 15:** Differentially expressed genes between center-cell iLOH microenvironments in the Xenium dataset are listed.
- Supplementary Table 16:** Differentially expressed genes between injured tubular microenvironments within a 20 μm surrounding region in the CosMx dataset are presented.
- Supplementary Table 17:** Differentially expressed genes between injured tubular microenvironments within a 20 μm surrounding region in the Xenium dataset are presented.
- Supplementary Table 18:** Differentially expressed genes between injured tubular microenvironments within a 20 μm surrounding region in the imputed dataset are presented.
- Supplementary Table 19:** All identified cell–cell interactions within the injured tubular microenvironment are provided.
- Supplementary Table 20:** All identified cell–cell interactions within the injured tubular immune microenvironment are provided.
- Supplementary Table 21:** All identified cell–cell interactions within the glomerular immune microenvironment are provided.
- Supplementary Table 22:** Spatially variable genes identified across niches are listed.
- Supplementary Table 23:** Spatially variable genes identified in the B cell–predominant immune microenvironment and the profibrotic injured tubular microenvironment are listed.
- Supplementary Table 24:** Clinical characteristics of B[−] and B⁺ DKD patient groups are summarized.
- Supplementary Table 25:** Differentially expressed genes between B⁺ and B[−] groups in bulk sequencing data from the TRIDENT cohort are listed.
- Supplementary Table 26:** Elastic net modeling results from the TRIDENT cohort are presented.
- Supplementary Table 27:** Plasma proteomics validation results in the UK Biobank cohort are provided.
- Supplementary Table 28:** Genes assayed in the Xenium and CosMx spatial transcriptomics panels are listed.

Additional References:

66. Suchanek, O. *et al.* Tissue-resident B cells orchestrate macrophage polarisation and function. *Nat. Commun.* **14**, 7081 (2023).
67. Fleig, S. V. *et al.* Long-term B cell depletion associates with regeneration of kidney function. *Immun. Inflamm. Dis.* **9**, 1479–1488 (2021).
68. Sato, Y. *et al.* Developmental stages of tertiary lymphoid tissue reflect local injury and inflammation in mouse and human kidneys. *Kidney Int.* **98**, 448–463 (2020).
69. Sato, Y., Silina, K., van den Broek, M., Hirahara, K. & Yanagita, M. The roles of tertiary lymphoid structures in chronic diseases. *Nat. Rev. Nephrol.* **19**, 525–537 (2023).
70. Chang, A. *et al.* In situ B cell-mediated immune responses and tubulointerstitial inflammation in human lupus nephritis. *J. Immunol. Baltim. Md 1950* **186**, 1849–1860 (2011).
71. Steinmetz, O. M. *et al.* BCA-1/CXCL13 expression is associated with CXCR5-positive B-cell cluster formation in acute renal transplant rejection. *Kidney Int.* **67**, 1616–1621 (2005).