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T cell receptor cross-reactivity expanded by dramatic peptide-MHC adaptability

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Peptide	NCBI Reference Sequence(s)
MLI DRG LGLL	WP_082441782.1
ILI DRG LGIM	YP_009336482.1
QLM DRG LGVL	APQ42521.1
ILN DRG LGML	XP_012938420.1 XP_022519643.1 XP_019768580.1 KZS15328.1 EFX89764.1
LLF DRG TGLM	WP_061021764.1
LLW DRG IGLL	OYT65050.1
MML DRG LVLM	WP_067737417.1 WP_008829097.1 WP_043869125.1
MML DRG LVMM	WP_008332046.1 WP_090062244.1 WP_074641131.1
MMW DRG TLIL	WP_014126739.1
IMF DRG LGLL	WP_062805772.1
TLF DRG LGL	BAC79679.1
MIF DRG VWL	WP_011053120.1
MVF DRG HPL	CAD41203.1
RLW DRG EIM	OAP17390.1
LLW DRG QNL	WP_000380056.1
ILW DRG REL	WP_084163317.1
DIW DRG IHL	WP_103068111.1
MML DRG QRL	OUT77584.1
MML DRG KEL	WP_011095018.1 WP_103523545.1
MMV DRG TTL	XP_021274615.1

Supplementary Table 1. Naturally occurring peptides resembling the DRG class of peptides found in various genomes. A non-comprehensive sampling of naturally occurring peptides from various proteomes with a DRG motif predicted to be compatible with the DMF5 TCR. BLAST searches required ideal anchor residues in positions 2 and/or 9 for nonamers. Decamers required an ideal anchor in position 2 and a large hydrophobic anchor in position 10 to facilitate the register shift.

Peptide	K_D (μM) ^a	ΔG° (kcal/mol) ^b	$\Delta\Delta G^\circ$ (kcal/mol) ^c	T_m ($^\circ\text{C}$) ^d
ELAGIGILTV (MART-1)	5.5 $\pm$ 0.5	-7.17 $\pm$ 0.05	--	65.0 $\pm$ 0.1
SMAGIGIVDV	202 $\pm$ 24	-5.04 $\pm$ 0.07	2.13 $\pm$ 0.09	65.8 $\pm$ 0.1
SMLGIGIVPV	43 $\pm$ 13	-5.96 $\pm$ 0.18	1.21 $\pm$ 0.19	66.1 $\pm$ 0.2
NMGGLGIMPV	7.3 $\pm$ 0.9	-7.00 $\pm$ 0.07	0.17 $\pm$ 0.09	58.6 $\pm$ 0.2
NLSNLGILV	11 $\pm$ 1.4	-6.76 $\pm$ 0.08	0.41 $\pm$ 0.06	66.3 $\pm$ 0.1
IMEDVGWLVN	121 $\pm$ 12	-5.34 $\pm$ 0.06	1.83 $\pm$ 0.08	55.1 $\pm$ 0.3
ILEDRGFNQV	99 $\pm$ 9	-5.46 $\pm$ 0.05	1.71 $\pm$ 0.07	46.1 $\pm$ 0.2
LMFDRGMSLL	123 $\pm$ 16	-5.33 $\pm$ 0.08	1.84 $\pm$ 0.09	50.0 $\pm$ 0.4
MMWDRGMGLL	144 $\pm$ 20	-5.24 $\pm$ 0.08	1.93 $\pm$ 0.09	45.4 $\pm$ 0.1
MMWDRGLGMM	32 $\pm$ 2	-6.13 $\pm$ 0.04	1.04 $\pm$ 0.06	45.2 $\pm$ 0.1

^aError in K_D is standard error from non-linear curve fitting of replicated datasets to a 1:1 equilibrium binding model as described in Methods.

^bBinding free energy changes ($\Delta G^\circ = RT\ln[K_D]$); errors reflect propagated K_D error. T = 298K (25 $^\circ\text{C}$).

^cDifference in binding free energy relative to MART-1 ($\Delta\Delta G^\circ = \Delta G^\circ_{\text{MART-1}} - \Delta G^\circ_{\text{pep}}$); errors are propagated from errors in ΔG° .

^dError in T_m is standard error from non-linear curve fit of temperature derivative data to a bi-Gaussian function as described in Methods.

Supplementary Table 2. DMF5 binding affinities and pMHC T_m values for GIG and DRG-class peptides.

For the DMF5 TCR binding HLA-A2 presenting each peptide in the DRG and GIG class, the table shows binding affinity and binding free energy values as determined by SPR. The right-most column shows the thermal stability of each peptide/HLA-A2 complex as determined by DSF.

Peptide	K_D (μM) ^a	ΔG° (kcal/mol) ^a	$\Delta\Delta G^\circ$ (kcal/mol) ^a	T_m ($^\circ\text{C}$) ^a
MMWDRGLG A M	> 500 ^a	< 4.5	> 1.6	43.5 $\pm$ 0.5
MMWDRGLG M V	NBD ^b	< 5.0	> 2.0	56.7 $\pm$ 0.3
MMWDRGLG M	30 $\pm$ 2	-6.17 $\pm$ 0.04	0.04 $\pm$ 0.06 ^c	49.1 $\pm$ 0.3
MMWDRG A GMM	> 500 ^d	< 4.5	> 1.6	42.1 $\pm$ 0.3
MMWD A GLGMM	> 500 ^d	< 4.5	> 1.6	44.5 $\pm$ 0.2

^a K_D , ΔG° , $\Delta\Delta G^\circ$, and T_m errors defined as in Supplementary Table 2.

^bNo binding detected; $\Delta\Delta G^\circ$ value is an estimated lower limit.

^c $\Delta\Delta G^\circ$ relative to MMWDRGLGMM (data in Supplementary Table 2).

^dBinding detected but too weak to quantify; K_D and $\Delta\Delta G^\circ$ values are estimated upper/lower limits.

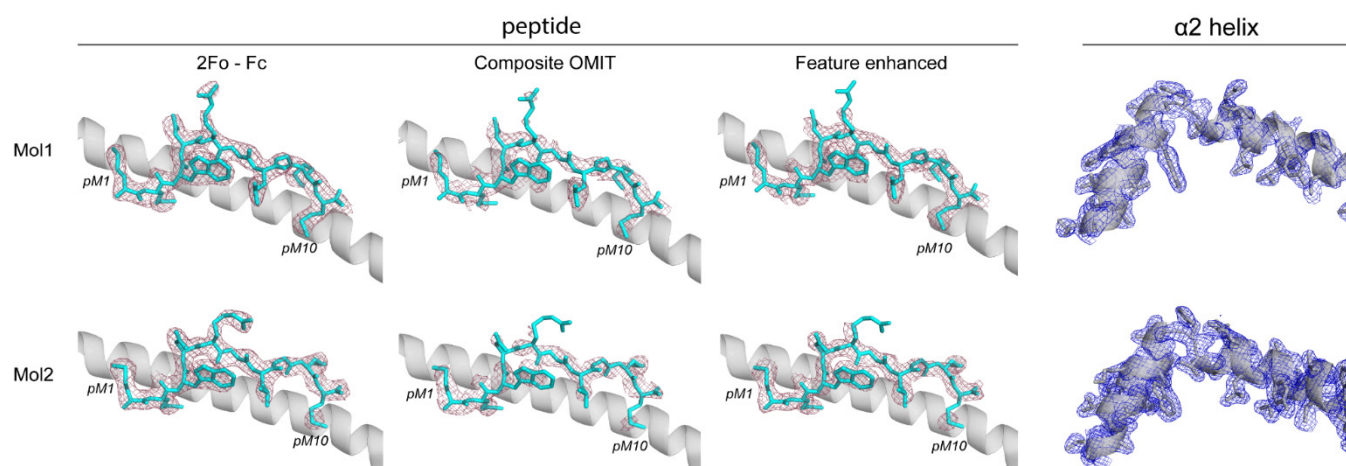
Supplementary Table 3. DMF5 binding affinities and pMHC T_m values for MMWDRGLGMM variants.

For the DMF5 TCR binding HLA-A2 presenting each variant of the MMWDRGLGMM peptide, the table shows binding affinity and binding free energy values as determined by SPR. The right-most column shows the thermal stability of each peptide/HLA-A2 complex as determined by DSF.

	MMWDRGLGMM/HLA-A2	DMF5-MMWDRGLGMM/ HLA-A2	DMF5-SMLGIGIVPV/ HLA-A2
Data collection			
Space group	P2 ₁	C2	C2
Cell dimensions			
<i>a</i> , <i>b</i> , <i>c</i> (Å)	58.30, 84.93, 84.32	232.08, 50.33, 84.36	228.11, 49.90, 92.45
α , β , γ (°)	90.00, 89.97, 90.00	90.00, 99.52, 90.00	90.00, 95.72, 90.00
Resolution (Å) *	19.95-2.50 (2.59-2.50)	19.94-2.15 (2.23-2.15)	38.75-2.39 (2.48-2.39)
<i>R</i> _{sym} or <i>R</i> _{merge}	0.16 (0.69)	0.12 (0.59)	0.07 (0.62)
<i>I</i> / σ <i>I</i>	8.52 (2.64)	10.98 (2.20)	24.63 (2.12)
Completeness (%)	98.5 (97.6)	98.8 (96.4)	98.0 (98.5)
Redundancy	3.0 (3.0)	2.7 (2.2)	3.8 (3.7)
Refinement			
Resolution (Å)	19.95-2.50	19.94-2.15	38.75-2.39
No. reflections	28114	49053	38730
<i>R</i> _{work} / <i>R</i> _{free}	0.211/0.249	0.180/0.226	0.201/0.244
No. atoms	6373	7007	6696
Protein	6288	6527	6484
Ligand/ion	0	0	0
Water	85	480	212
<i>B</i> -factors	54.0	39.2	52.31
Protein	54.6	39.4	52.95
Ligand/ion	0	0	0
Water	40.8	36.57	32.62
R.m.s. deviations			
Bond lengths (Å)	0.002	0.003	0.002
Bond angles (°)	0.45	0.60	0.58

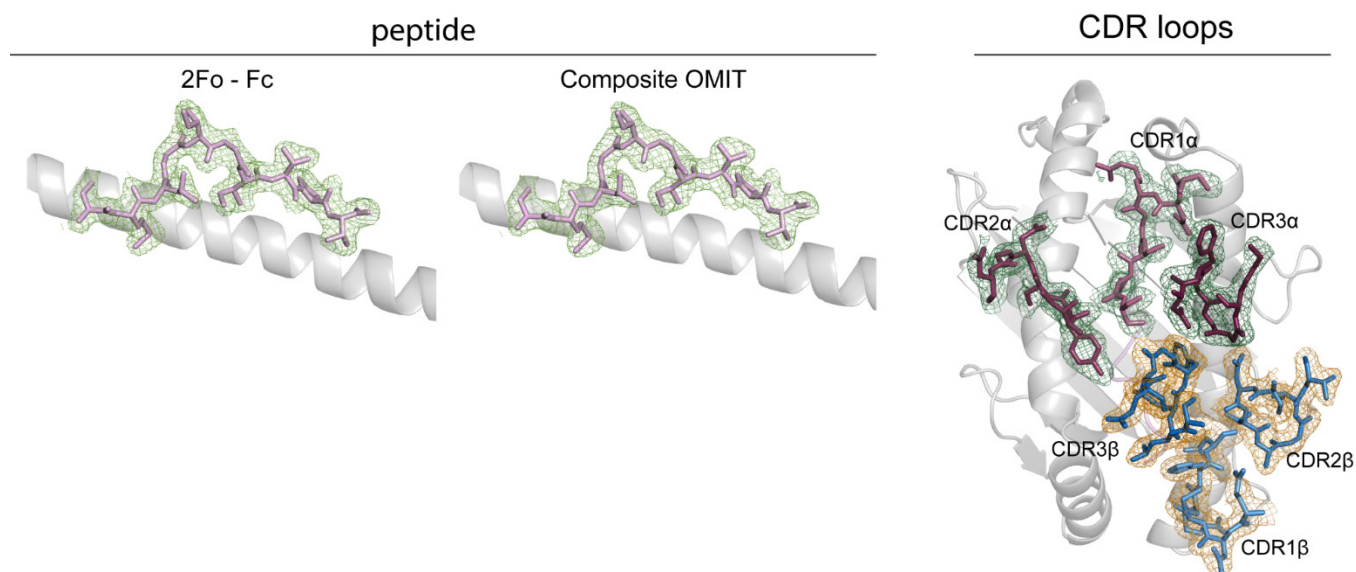
*Highest-resolution shell is shown in parentheses.

Supplementary Table 4. X-ray data collection and refinement statistics for the indicated structures.

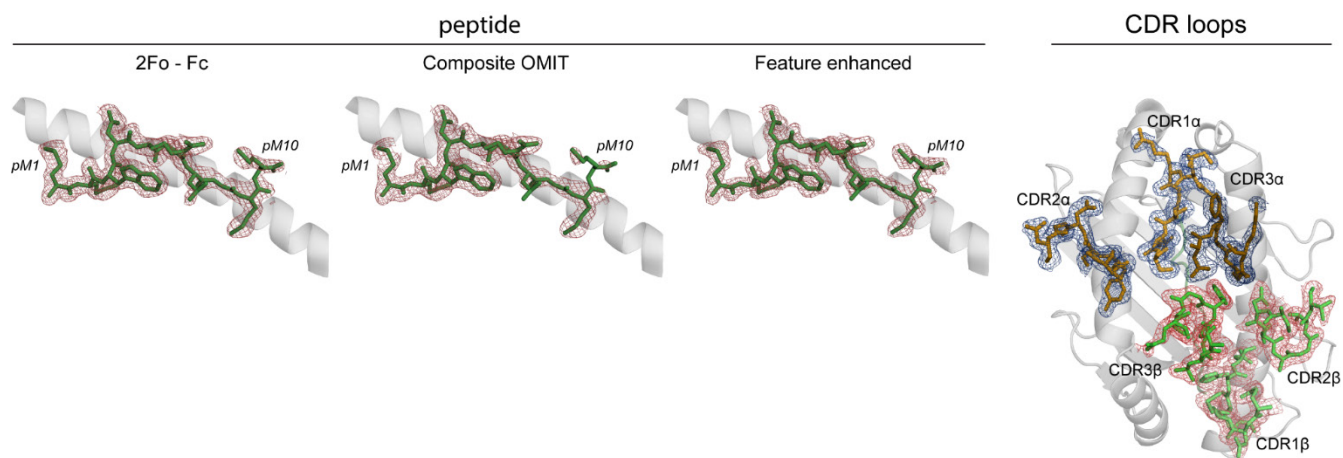


Supplementary Figure 1A. Electron density images for the MMWDRGLGMM/HLA-A2 structure.

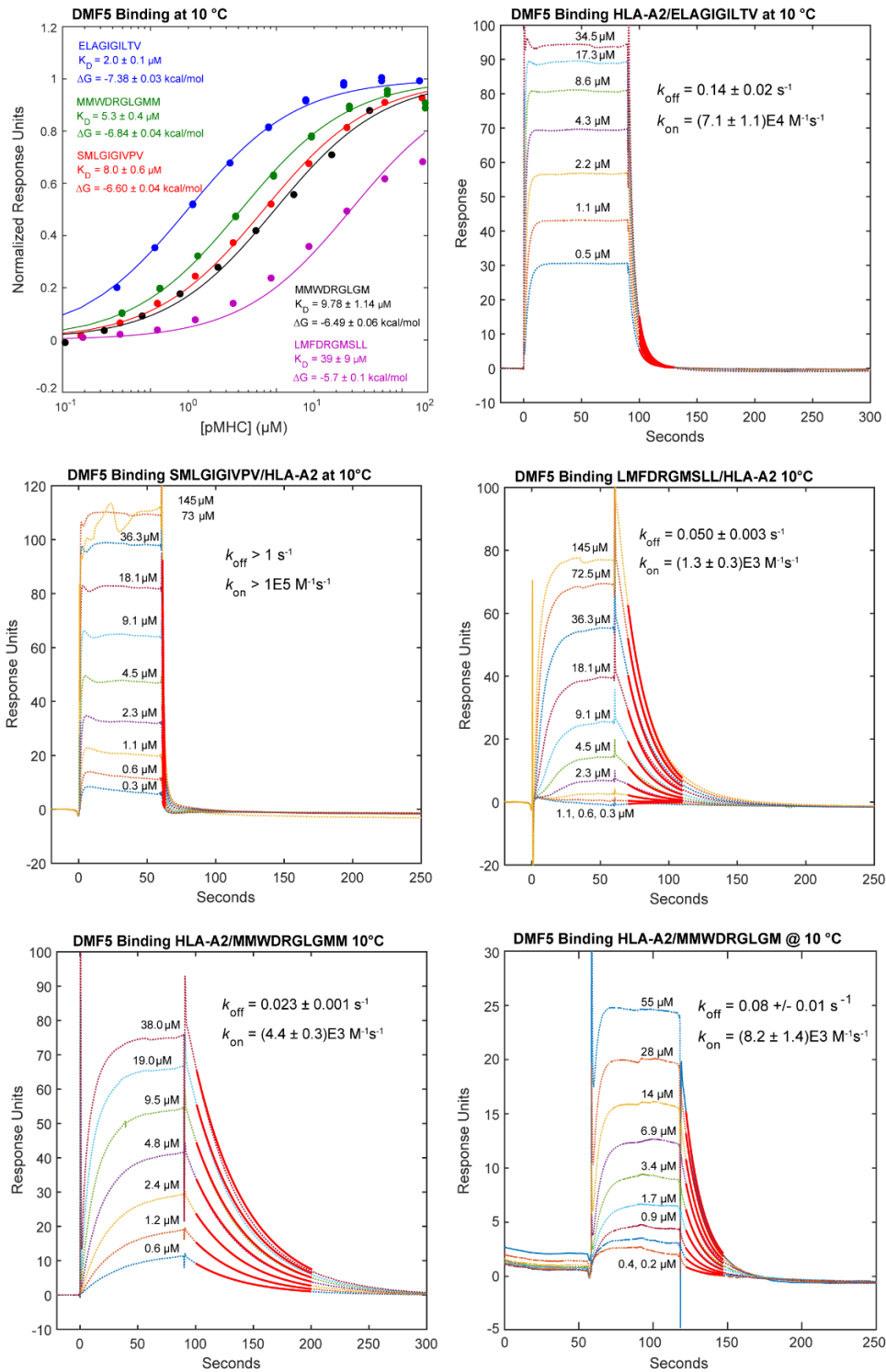
Density contoured at 1σ is shown for 2F_o-F_c, composite omit, and feature enhanced maps for molecule 1 (top) and molecule 2 (bottom). All three maps are shown for the peptide on the left. Only the 2F_o-F_c map is shown for the HLA-A2 $\alpha 2$ helix on the right.



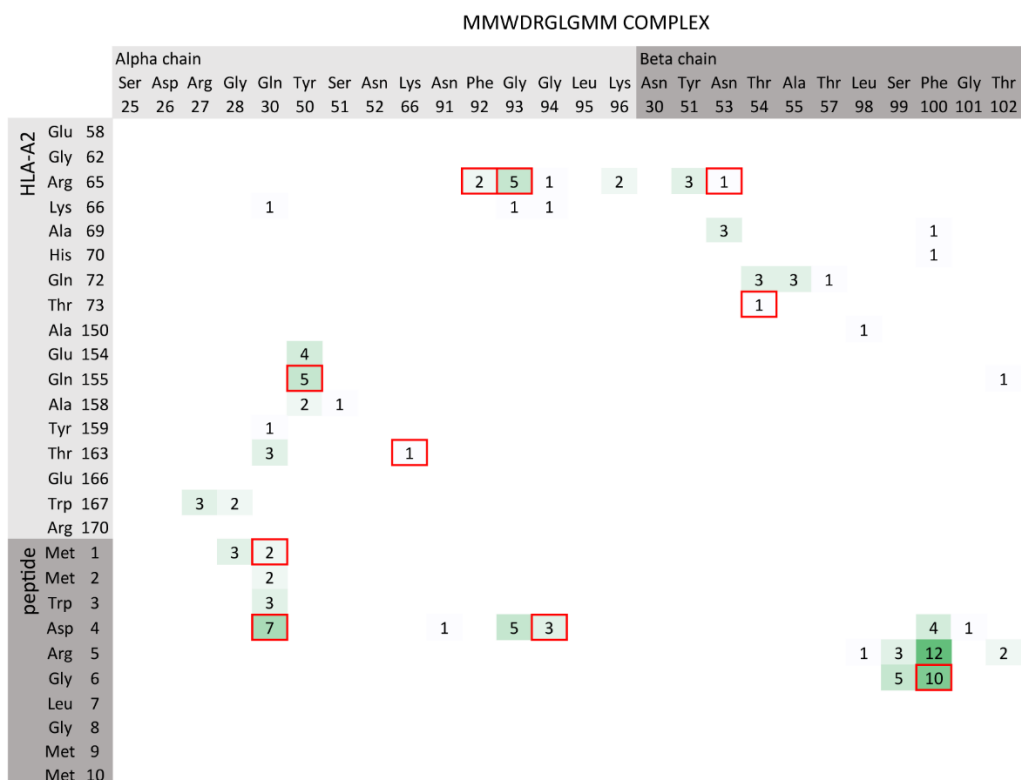
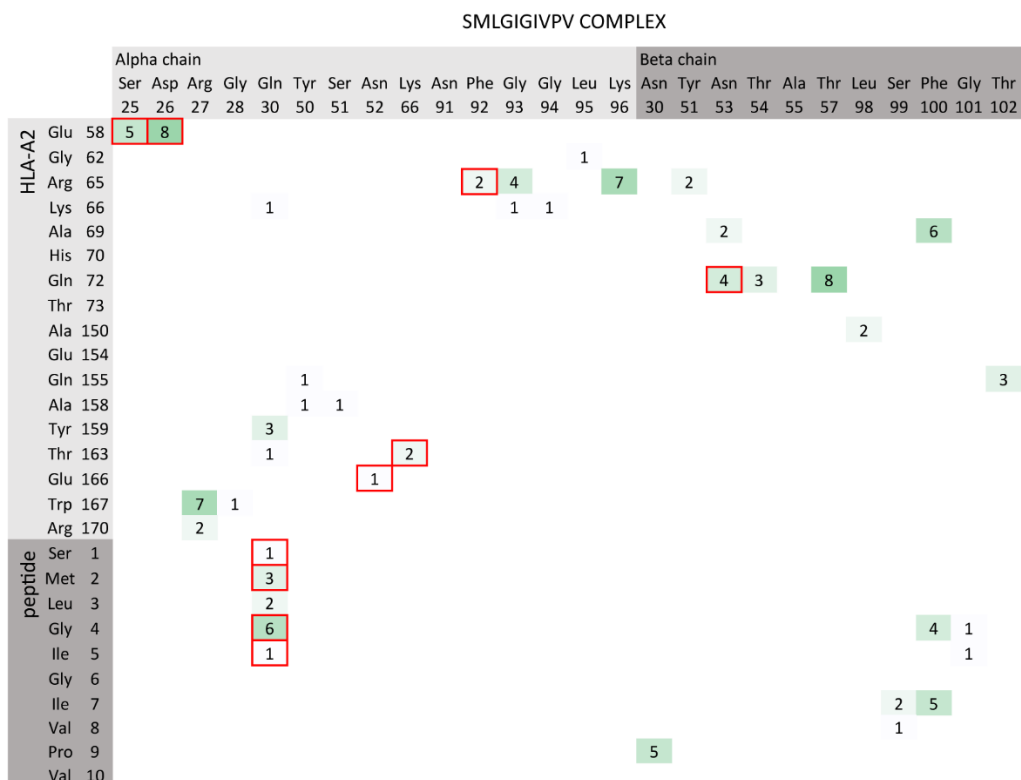
Supplementary Figure 1B. Electron density images for the DMF5-SMLGIGIVPV/HLA-A2 structure. Density contoured at 1σ is shown for 2F_o-F_c and composite omit maps. Both maps are shown for the peptide on the left. Only the 2F_o-F_c map is shown for the CDR loops on the right.



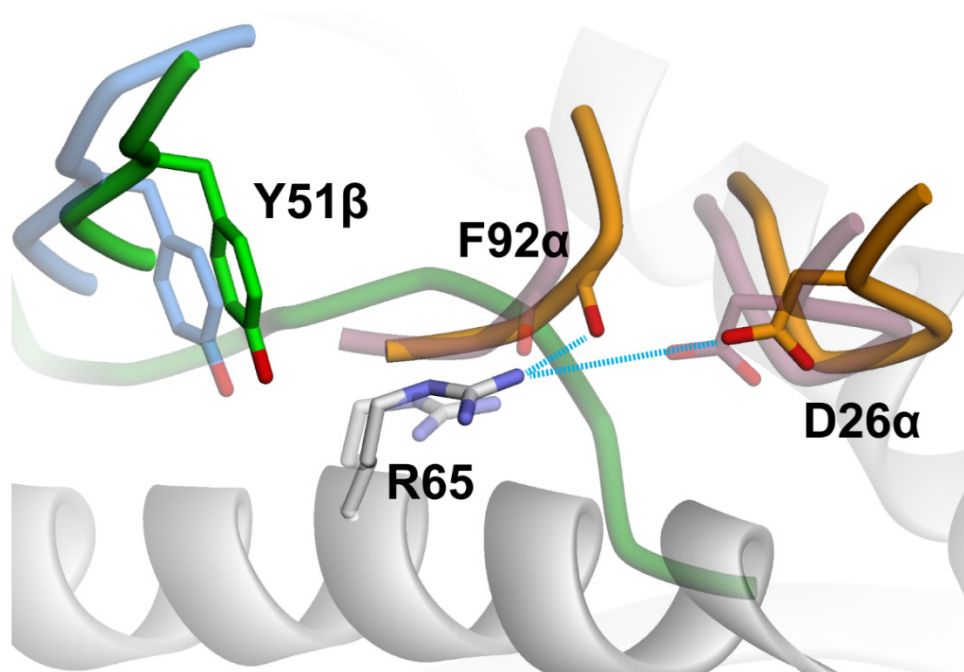
Supplementary Figure 1C. Electron density images for the DMF5-MMWDRGLGMM/HLA-A2 structure. Density contoured at 1σ is shown for 2F_o-F_c, composite omit, and feature enhanced maps. All maps are shown for the peptide on the left. Only the 2F_o-F_c map is shown for the CDR loops on the right.



Supplementary Figure 2. SPR data for assessing the kinetics of DMF5 binding GIG, DRG, and the C-terminally truncated MMWDRGLGM peptides presented by HLA-A2. The top panel shows steady state binding data used to determine K_D values at 10 °C. The bottom four panels show kinetic data at 10 °C used to determine k_{off} rates. Association rates were determined from K_D and k_{off} values ($k_{\text{on}} = K_D/k_{\text{off}}$). Errors are standard fitting errors for global analysis (K_D and k_{off}) or propagated from errors in K_D and k_{off} (k_{on}). These experiments indicate that DMF5 binds the GIG peptides with faster kinetics than the DRG peptides, supporting the conclusion that TCR binding the DRG peptides occurs with a register shift. The truncated MMWDRGLGM nonamer is recognized with faster kinetics than the parental decamer, including a more rapid association rate.



Supplementary Figure 3. Contact maps of the TCR-pMHC complexes with the SMLGIGIVPV (top) and MMWDRGLGMM (bottom) peptides. Contacts are defined as interatomic distances ≤ 4 Å. Numbers indicate the number of contacts, with cells colored according to a white (min) or green (max) color scale. Elements with residues participating in interfacial hydrogen bonds are boxed in red. Amino acid numbering is adjusted from the PDB files to facilitate a direct comparison.



Supplementary Figure 4. TCR electrostatic interactions with Arg65. Interactions with Arg65 that help drive recognition of HLA-A2 and restrict its orientation over peptide/HLA-A2 complexes (ref. 34 in main text) are conserved in the MMWDRGLGMM and GIG interfaces. Coloring is as defined in Figure 6 in the main manuscript.

Supplementary Video 1. Animation shown the transition of the MMWDRGLGMM/HLA-A2 complex from TCR-free to TCR-bound. The video highlights the peptide's register shift and resulting C-terminal extension upon binding of the DMF5 TCR. The animation was generated via interpolation paired with a refinement step to minimize VDW clashes.