

Supplementary Information

SARS-CoV-2 M^{pro} inhibitors and activity-based probes for patient-sample imaging

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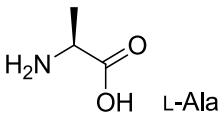
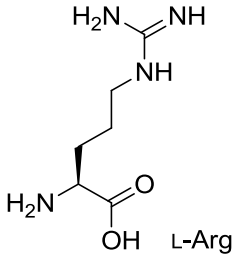
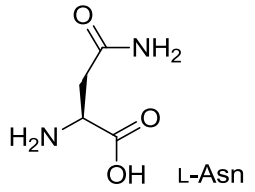
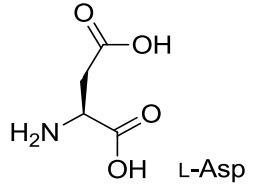
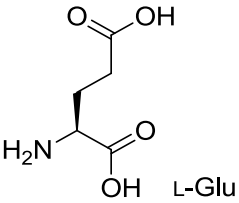
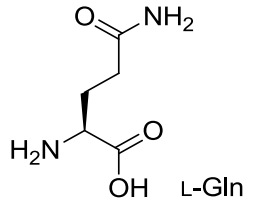
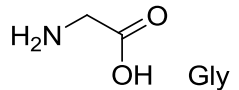
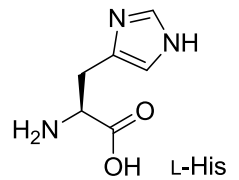
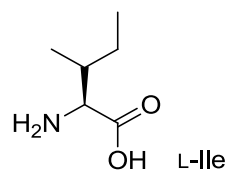
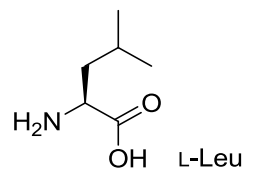
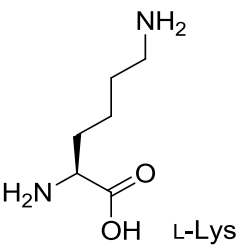
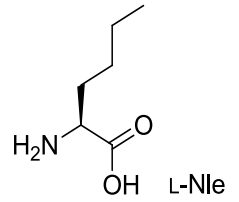
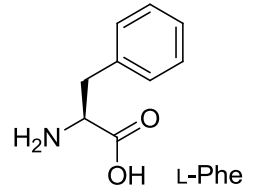
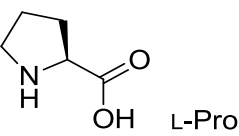
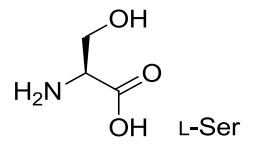
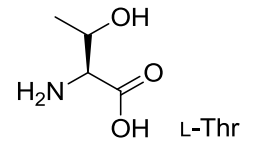
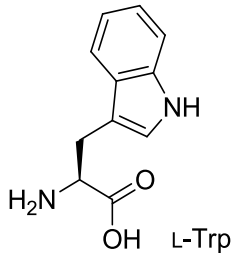
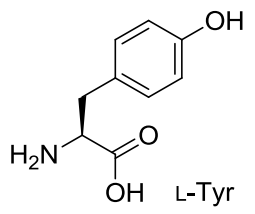
⁹Equal contribution

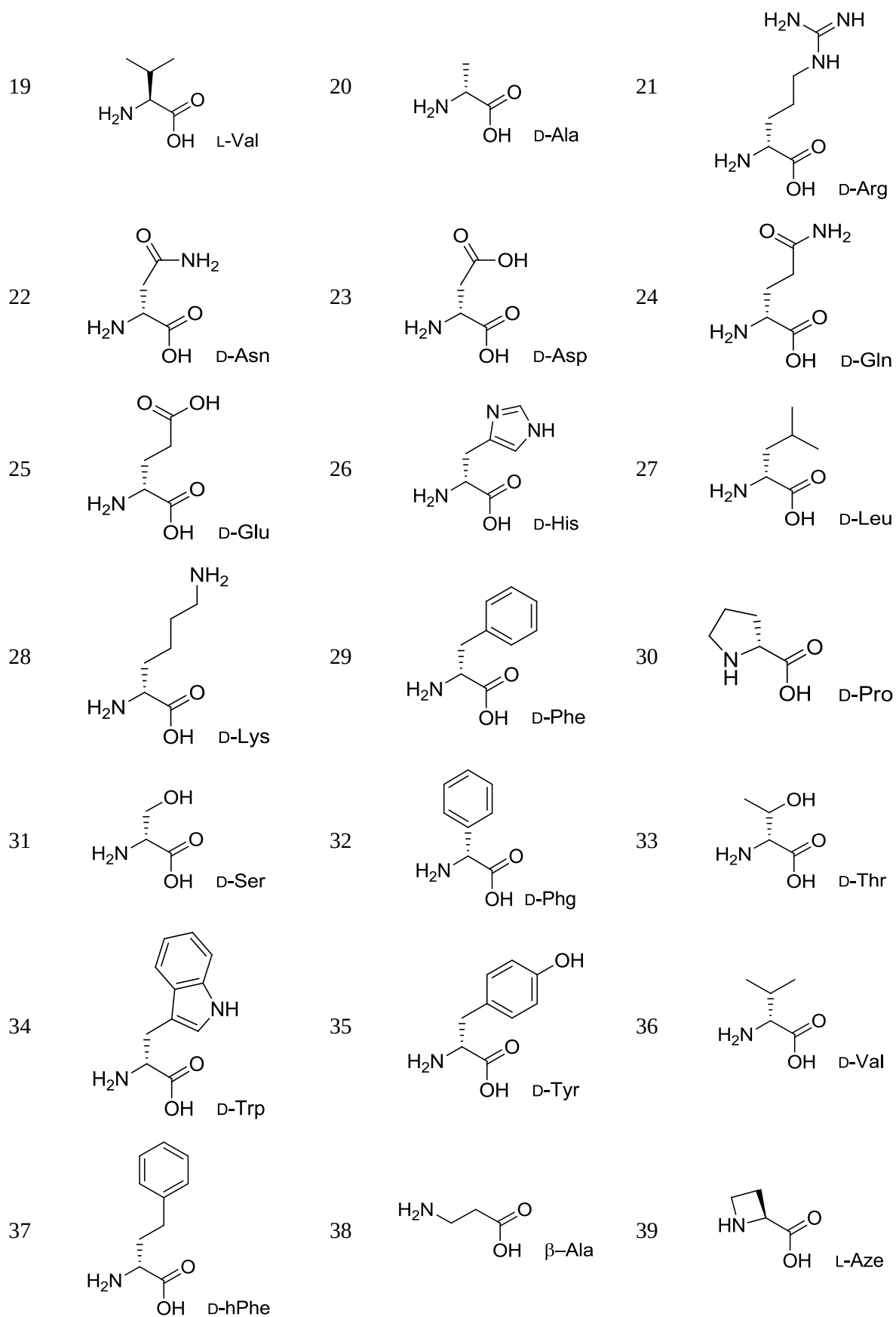
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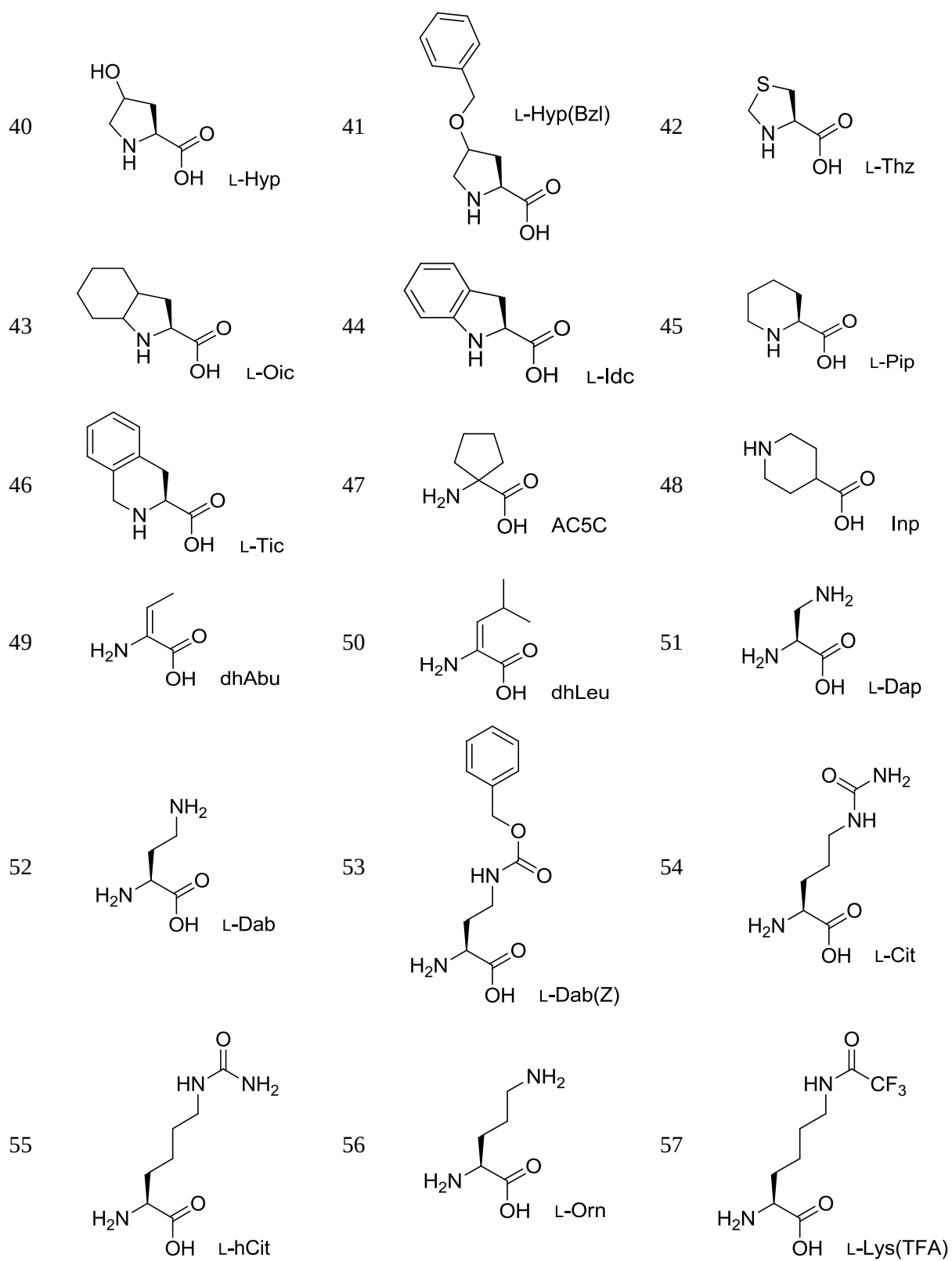
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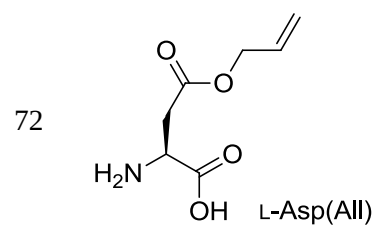
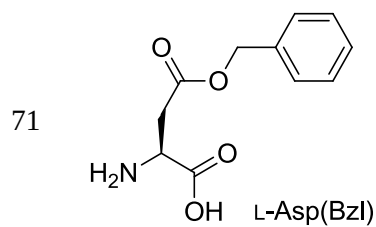
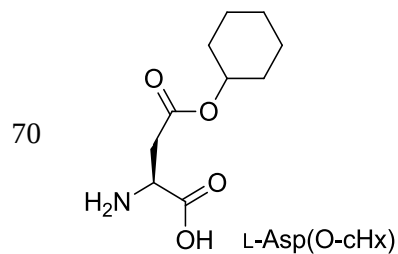
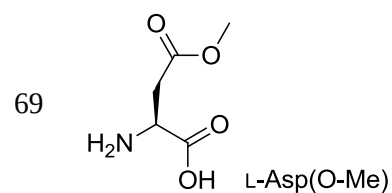
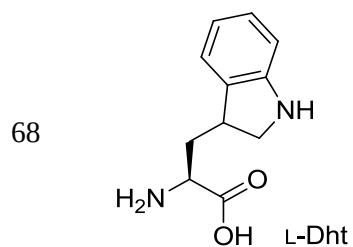
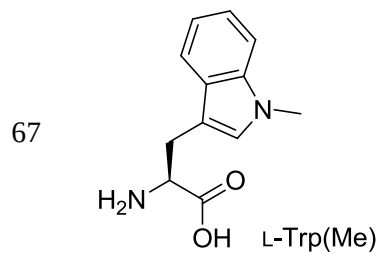
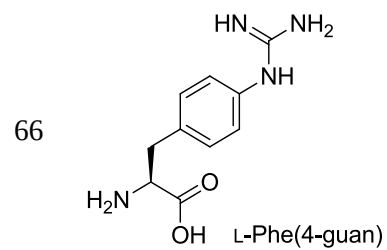
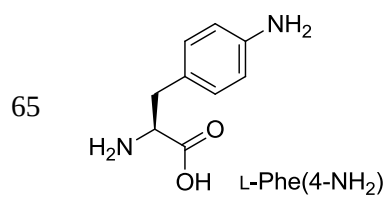
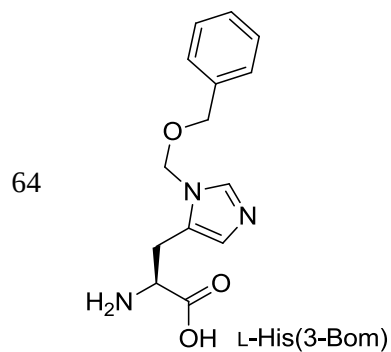
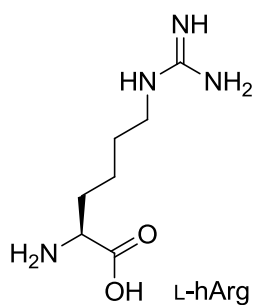
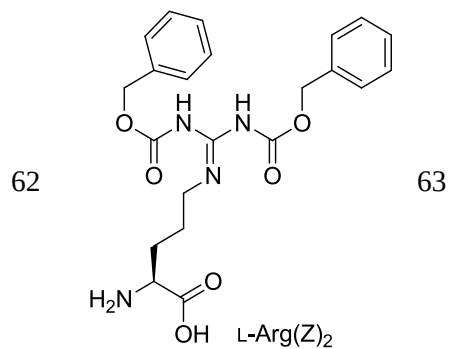
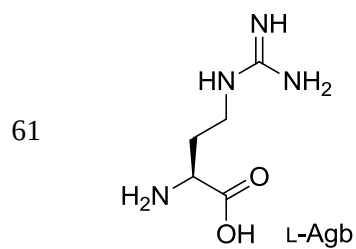
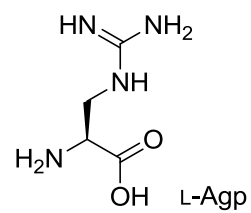
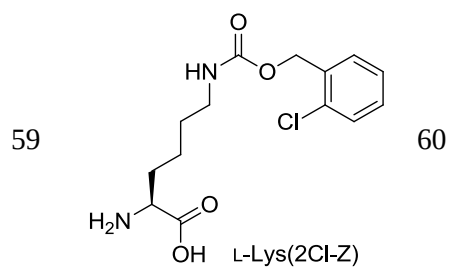
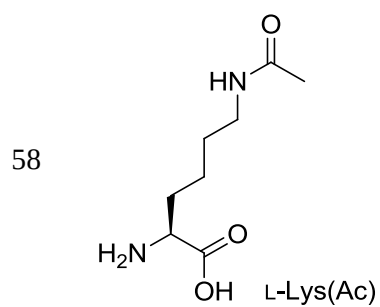
1. Supplementary Tables.....	2-11
2. Supplementary Note.....	11-28

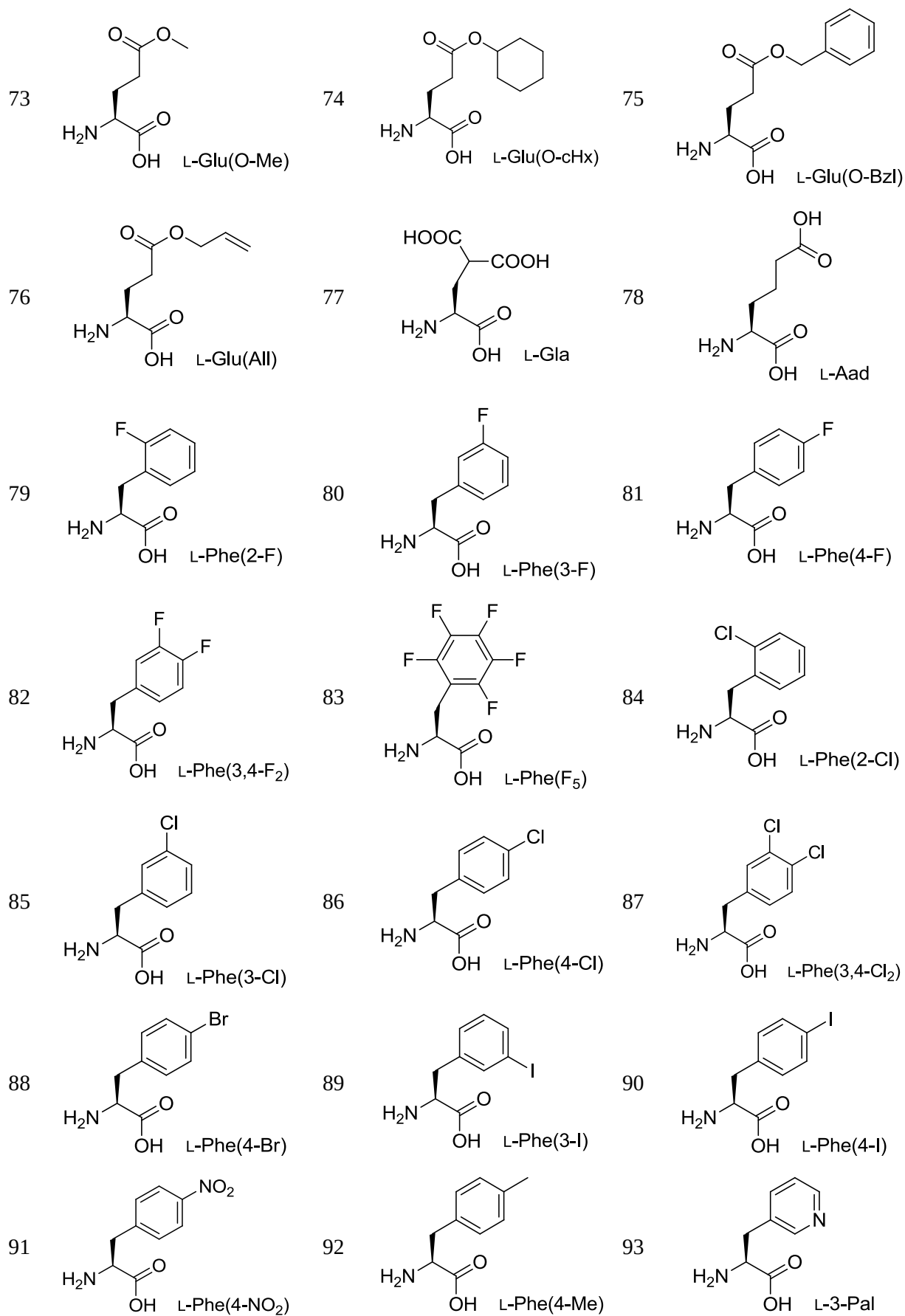
Supplementary Table 1. Structures of fixed natural and unnatural amino acids used in combinatorial library.

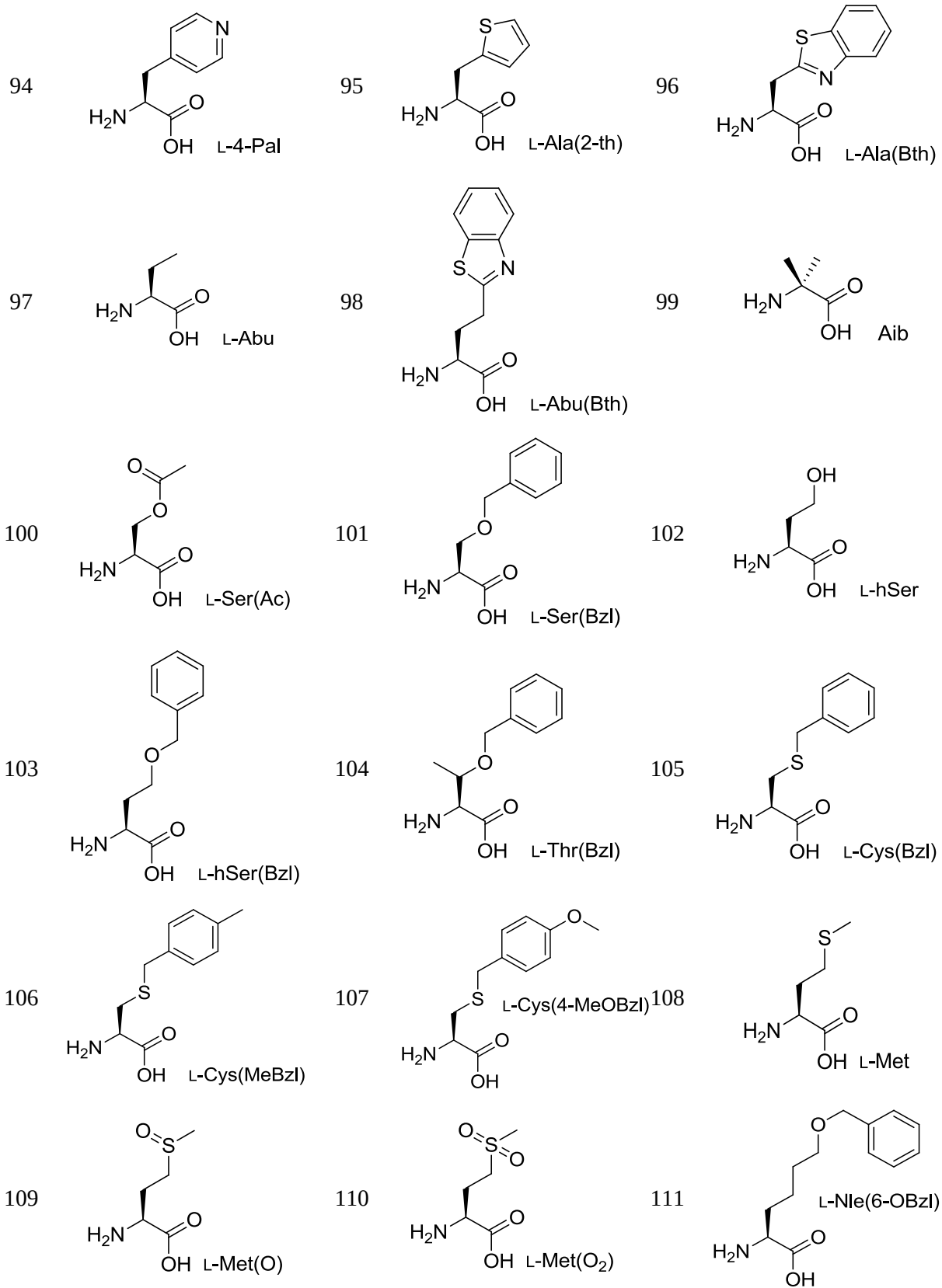
No	Structure + code	No	Structure + code	No	Structure + code
1	 L-Ala	2	 L-Arg	3	 L-Asn
4	 L-Asp	5	 L-Glu	6	 L-Gln
7	 Gly	8	 L-His	9	 L-Ile
10	 L-Leu	11	 L-Lys	12	 L-Nle
13	 L-Phe	14	 L-Pro	15	 L-Ser
16	 L-Thr	17	 L-Trp	18	 L-Tyr

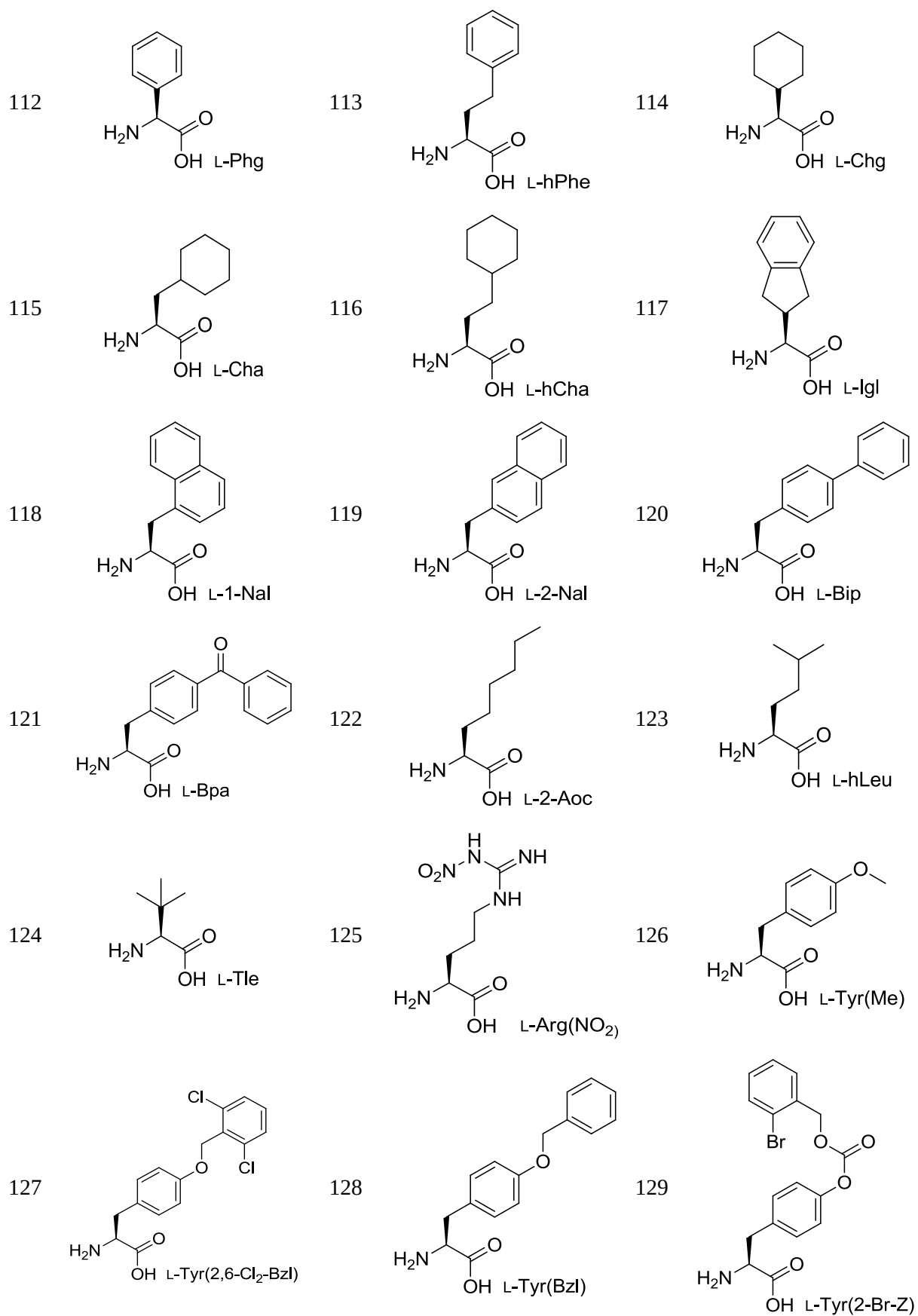


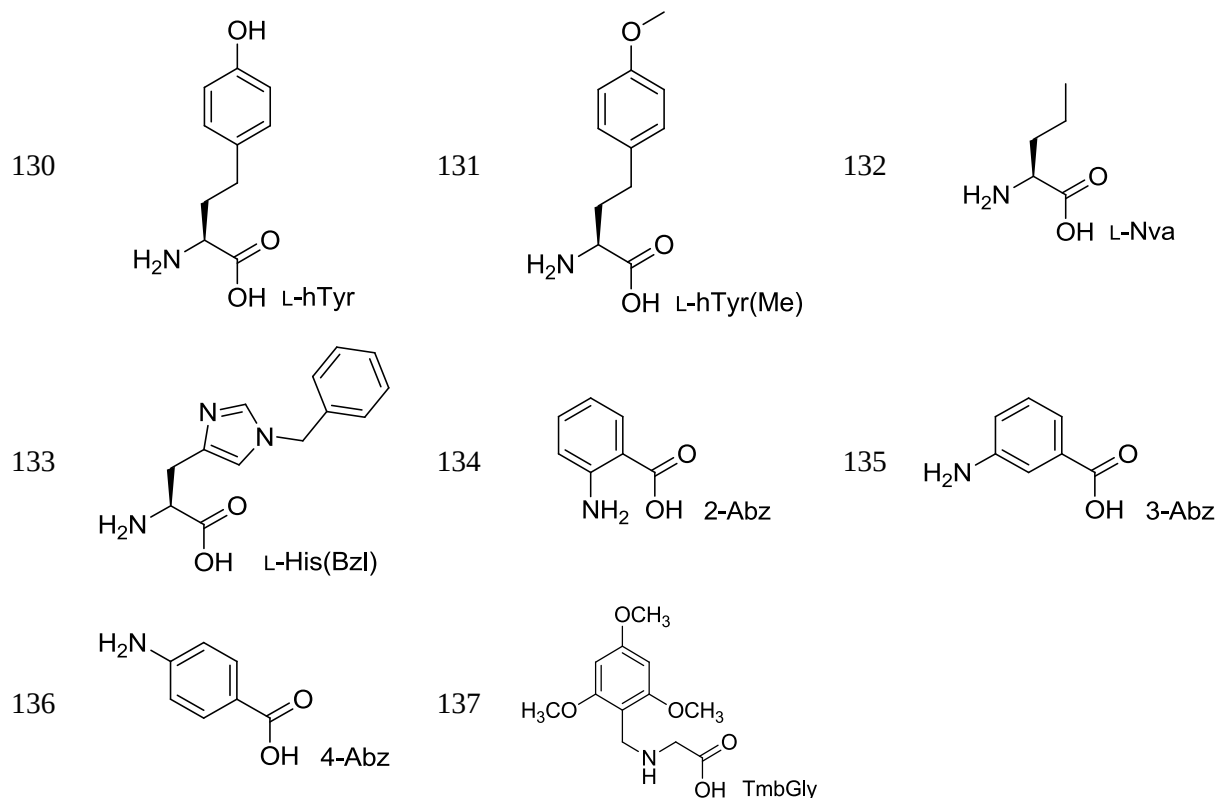




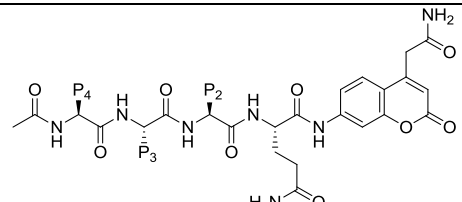
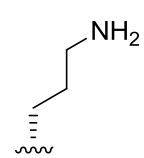
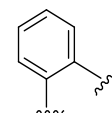
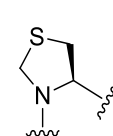
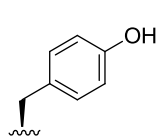
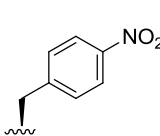








Supplementary Table 2. Structures of the most recognizable amino acids at each tested position by SARS-CoV-2 M^{pro}.

					
P4		P3		P2	
structure	Relative activity, %	structure	Relative activity, %	structure	Relative activity, %
	100		100		100
	77.5		83		54
	77		82		50

Protein	34
Ligand/ion	57
Water	43
R.m.s. deviations	
Bond lengths (Å)	0.01
Bond angles (°)	1.8

*Values in parentheses are for highest-resolution shell.

Supplementary Note

Synthetic procedures

Individual substrate synthesis

ACC-labeled substrates were synthesized on the solid support according to the solid phase peptide synthesis method described elsewhere¹. In brief, Fmoc-ACC-OH (2.5 eq) was attached to a Rink-amide resin using HOBt (2.5 eq) and DICl (2.5 eq) in DMF as coupling reagents. Then, the Fmoc protecting group was removed using 20% piperidine in DMF (three cycles: 5, 5, and 25 min). Fmoc-Gln(Trt)-OH (2.5 eq) was coupled to the H₂N-ACC-resin using HATU (2.5 eq) and 2,4,6-collidine (2.5 eq) in DMF. After Fmoc group removal, the Fmoc-P2-OH (2.5 eq) amino acid was attached (HOBt and DICl (2.5 eq) in DMF). Amino acids in P3 and P4 positions were coupled in the same manner. The free *N*-terminus was acetylated using HBTU, AcOH and DIPEA in DMF (5 eq of each reagent). Then, the resin was washed five times with DMF, three times with DCM and three times with MeOH, and dried over P₂O₅. Substrates were removed from the resin with a mixture of TFA/TIPS/H₂O (% v/v/v, 95:2.5:2.5), precipitated in Et₂O, purified on HPLC and lyophilized. The purity of each substrate was confirmed using analytical HPLC. Each substrate was dissolved in DMSO at a final concentration of 10 mM and stored at -80°C until use.

Internally quenched fluorescence substrates were synthesized on the solid support in the same manner as described previously².

Activity-based probe and inhibitor synthesis

The synthesis of the biotinylated activity-based probe involved three sequential steps. In the first step, the Biotin-PEG(4)-Abu-Tle-Leu-OH fragment was synthesized using a 2-chlorotrityl chloride resin. Fmoc-Leu-OH (2.5 eq) was dissolved in anhydrous DCM, pre-activated with DIPEA (3 eq) and added to the cartridge with resin (1 eq). After 3h, the mixture was filtered, washed 3 times with DCM and 3 times with DMF, and the Fmoc group was removed using 20% piperidine in DMF. Fmoc-Tle-OH, Fmoc-Abu-OH and Fmoc-PEG(4)-OH were attached

to the resin using HATU (2.5 eq) and 2,4,6-collidine (2.5 eq) in DMF as coupling reagents. The biotin tag was coupled to H₂N-PEG(4)-Abu-Tle-Leu-resin using 2.5 eq HBTU and 2.5 eq DIPEA in a DMF:DMSO mixture (1:1, v/v). After 3 h, the resin was washed 3 times with DMF, 3 times with DCM, and 3 times with MeOH and dried over P₂O₅. Finally, the peptide fragment was removed from the resin with a mixture of DCM/TFE/AcOH (v/v/v, 8:1:1). The solution was filtered and concentrated. The obtained crude peptide was dissolved in ACN:H₂O (v/v, 3:1), lyophilized, and then used without further purification (purity > 95%). In the second step, Fmoc-Gln(Trt)-VS was synthesized in the same manner as described elsewhere³. In the third step, the Fmoc group was removed from Fmoc-Gln(Trt)-VS using a mixture of diethylamine:ACN (1:1, v/v). Biotin-PEG(4)-Abu-Tle-Leu-OH was pre-activated with HATU (1.2 eq) and 2,4,6-collidine (3 eq) in DMF and attached to the H₂N-Gln(Trt)-VS (1 eq). The reaction was agitated for 2 h and the product was purified on HPLC. Finally, Biotin-PEG(4)-Abu-Tle-Leu-Gln(Trt)-VS was treated with a mixture of TFA/DCM/TIPS (% v/v/v/ 70:27:3) to remove the Trt group. After 40 min, solvents were evaporated and the probe was purified on HPLC.

Ac-Abu-Tle-Leu-Gln-VS, Ac-Abu-DTyr-Leu-Gln-VS, Bodipy-PEG(4)-Abu-DTyr-Leu-Gln-VS, and Cy5-PEG(4)-Abu-Tle-Leu-Gln-VS were synthesized in the same manner, but Boc-PEG(4)-Abu-Tle-Leu-OH, Boc-PEG(4)-Abu-DTyr-Leu-OH, Ac-Abu-DTyr-Leu-OH, and Ac-Abu-Tle-Leu-OH were synthesized on the 2-chlorotrityl chloride resin.

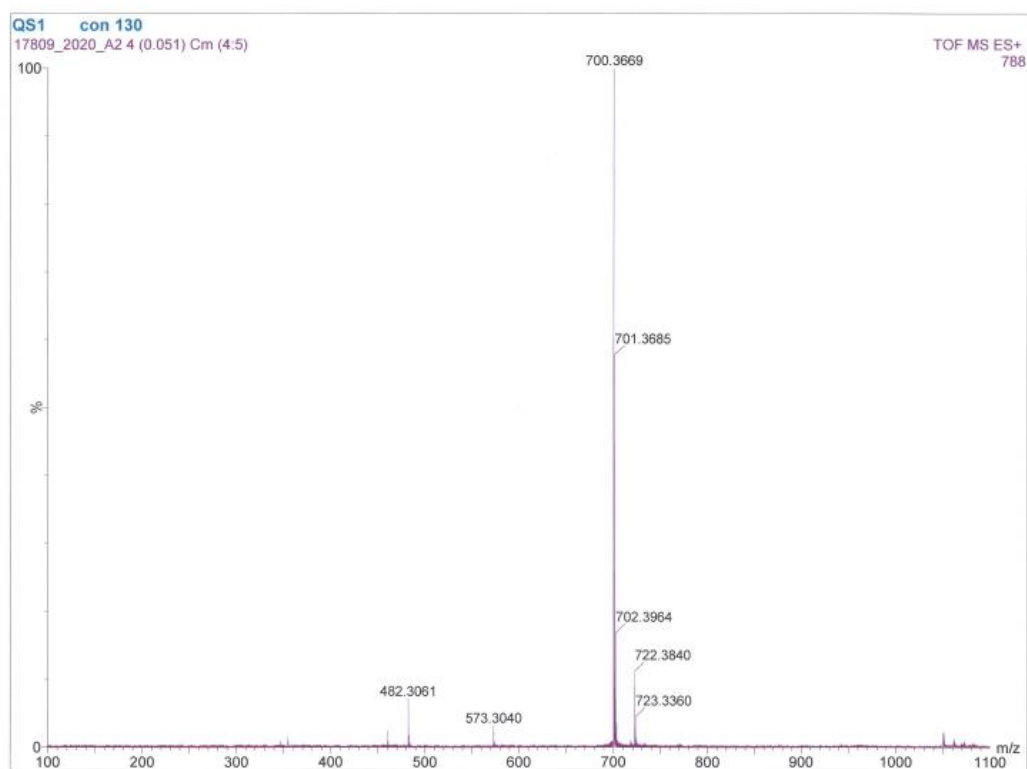
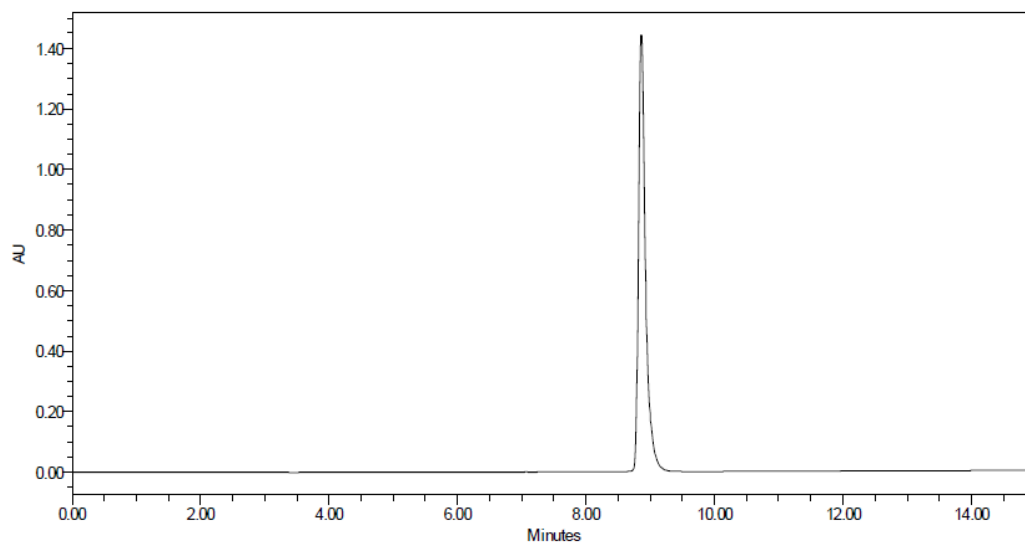
References

1. Poreba, M., Salvesen, G.S. & Drag, M. Synthesis of a HyCoSuL peptide substrate library to dissect protease substrate specificity. *Nature Protocols* **12**, 2189-2214 (2017).
2. Poreba, M. et al. Highly sensitive and adaptable fluorescence-quenched pair discloses the substrate specificity profiles in diverse protease families. *Scientific Reports* **7** DOI: 10.1038/srep43135 (2017).
3. Rut, W., Poreba, M., Kasperkiewicz, P., Snipas, S.J. & Drag, M. Selective Substrates and Activity-Based Probes for Imaging of the Human Constitutive 20S Proteasome in Cells and Blood Samples. *Journal of Medicinal Chemistry* **61**, 5222-5234 (2018).

HRMS and analytical chromatograms of synthesized substrates

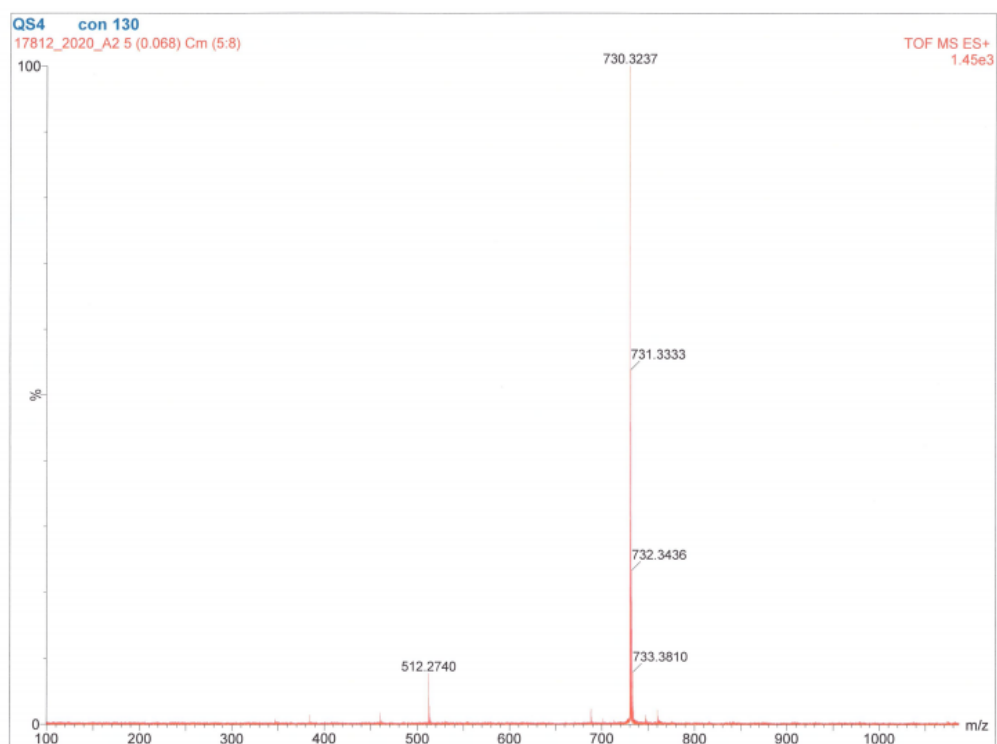
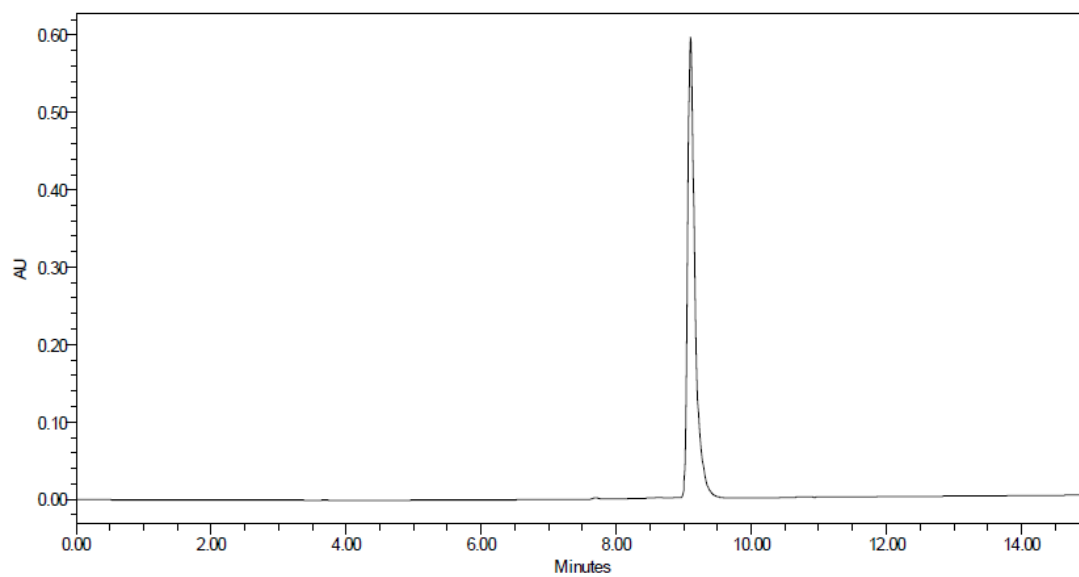
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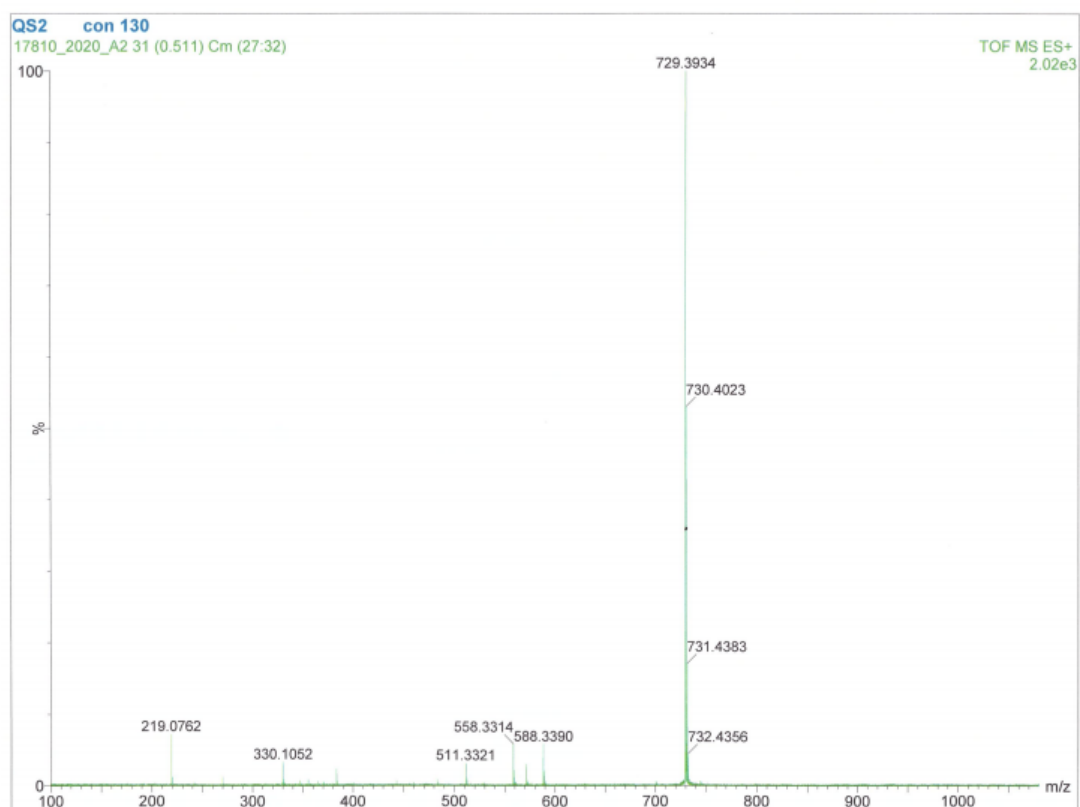
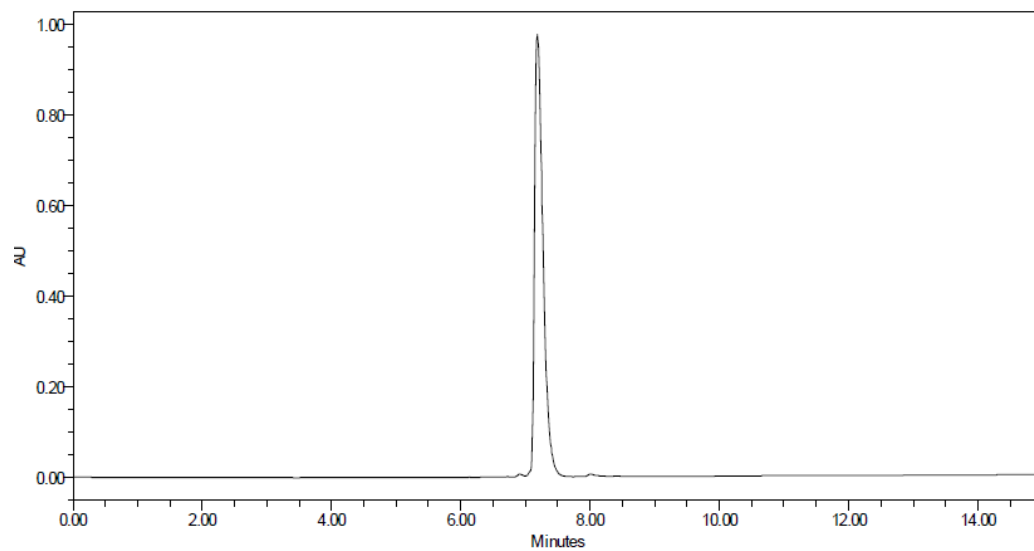
2 Ac-Thz-Tle-Leu-Gln-ACC

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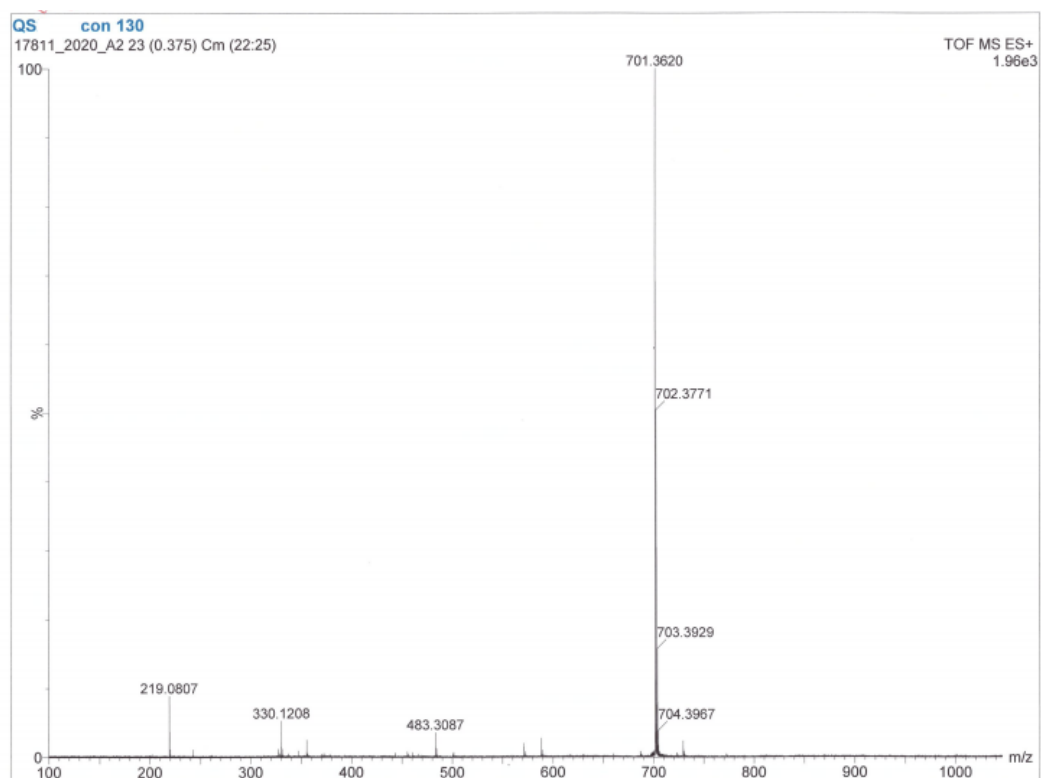
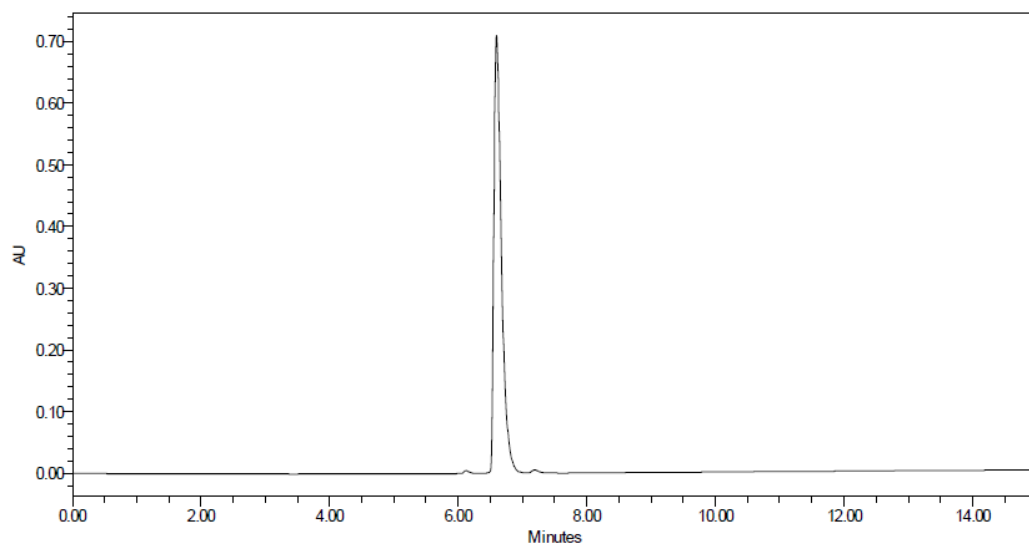
3 Ac-Val-Lys-Leu-Gln-ACC

m/z: $[M+H]^+$ for $C_{35}H_{52}N_8O_9$ ($m/z_{\text{calcd}} = 729.3931$; $m/z_{\text{found}} = 729.3934$)



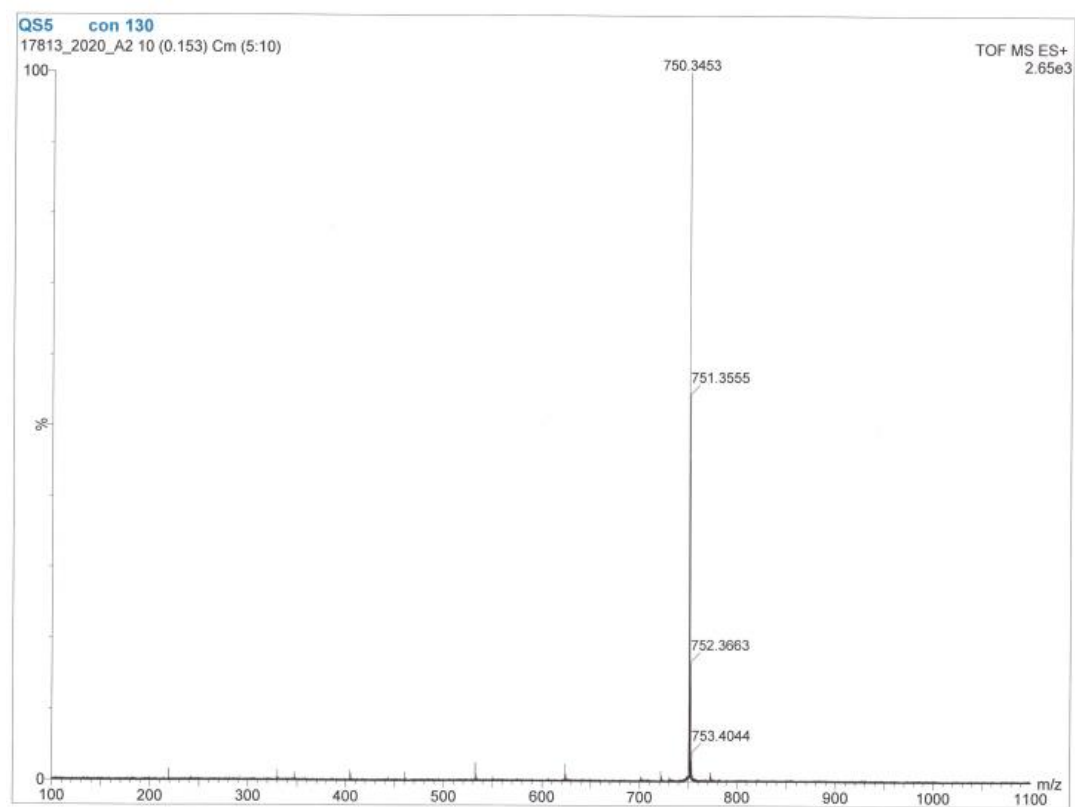
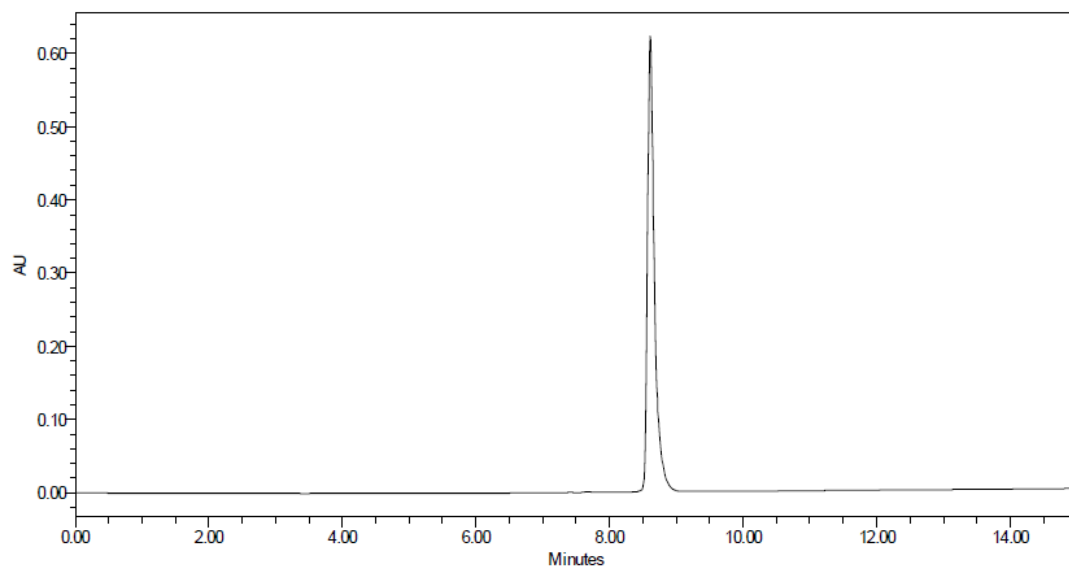
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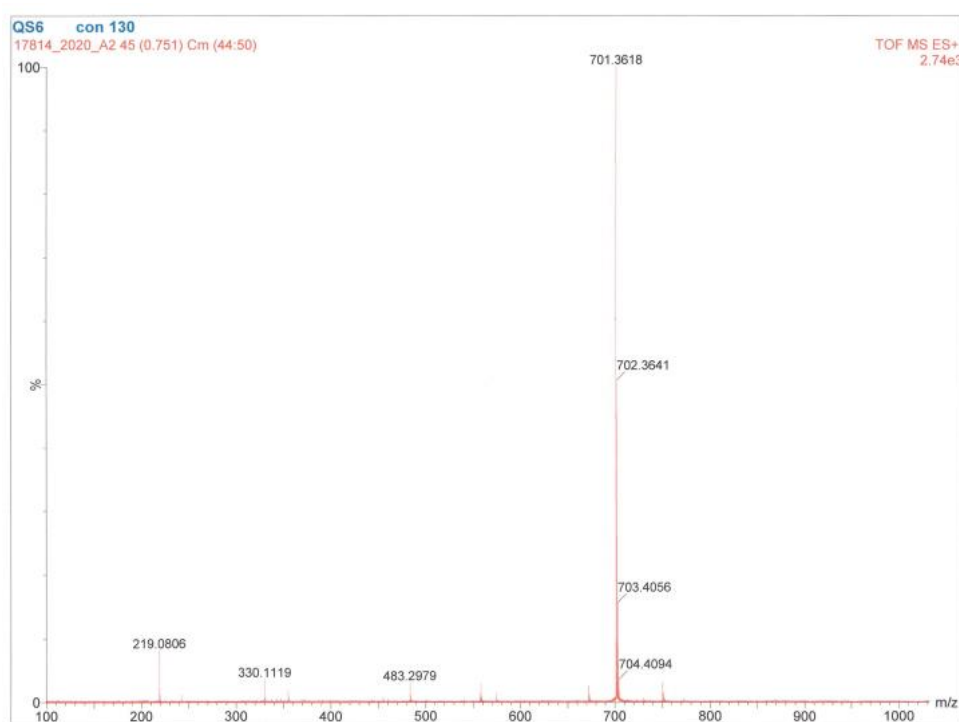
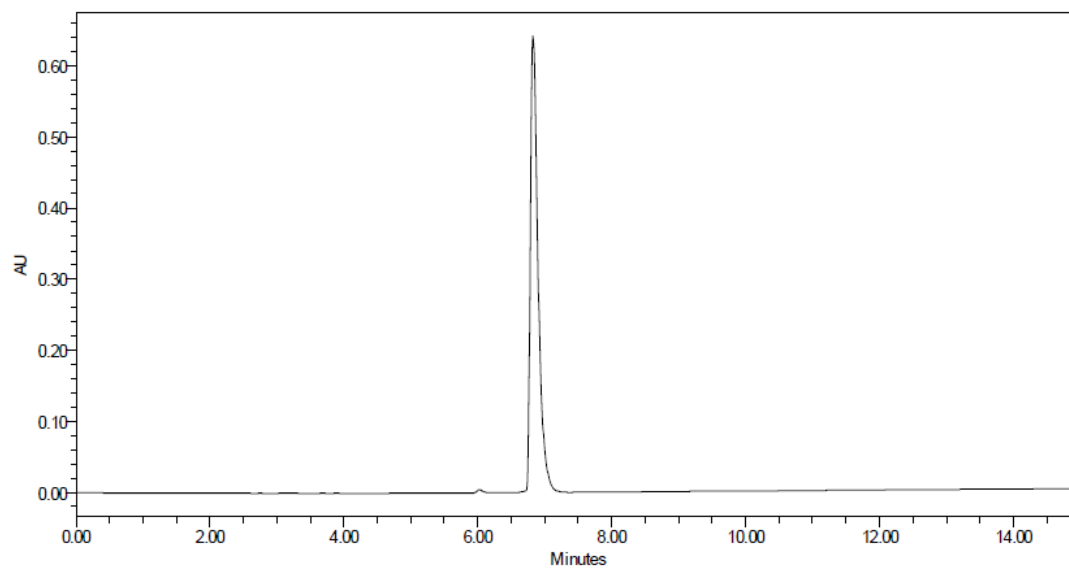
5 Ac-Abu-DTyr-Leu-Gln-ACC

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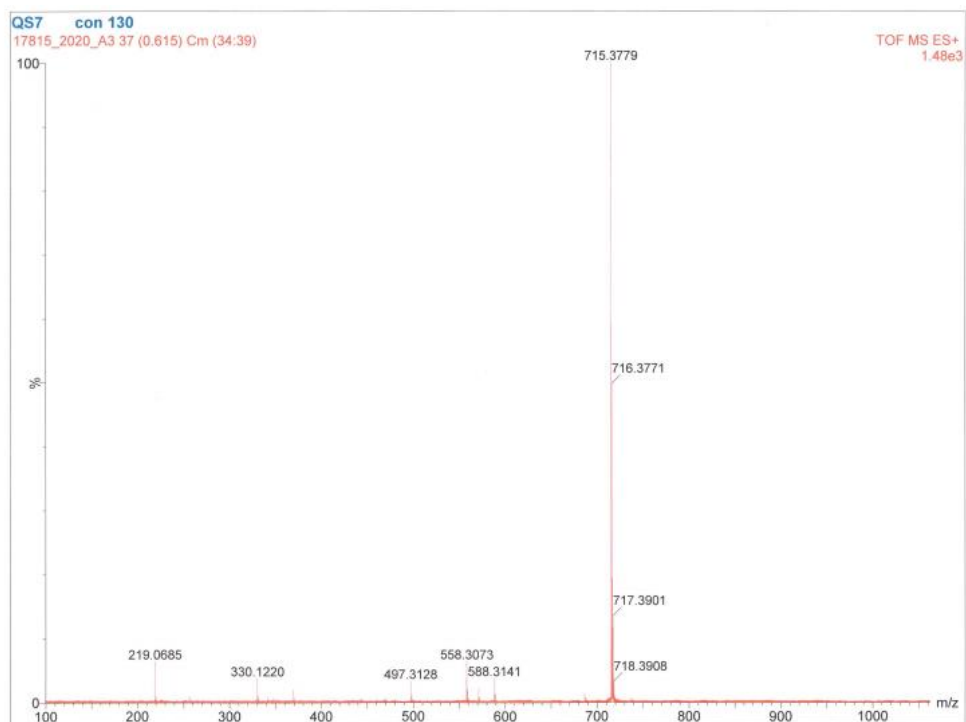
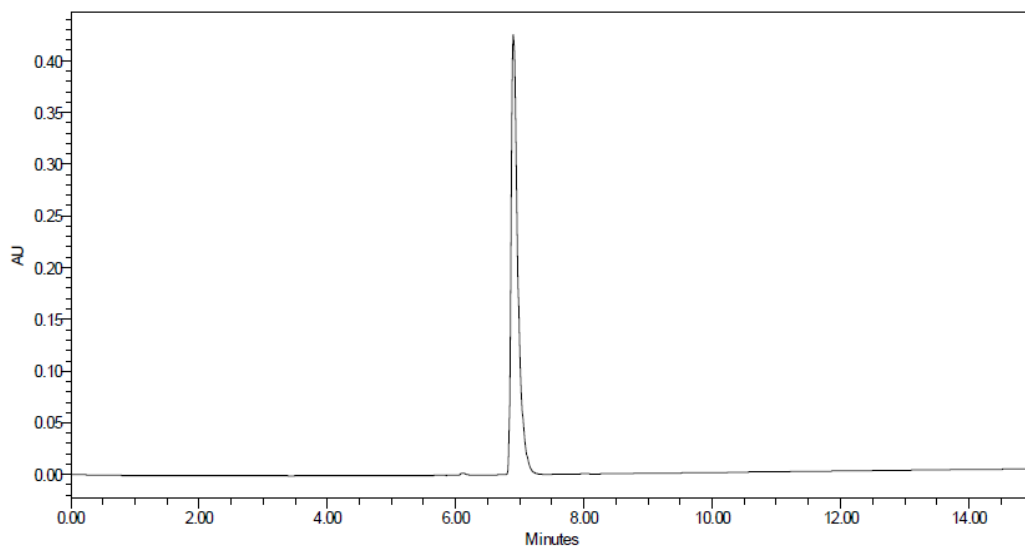
6 Ac-Abu-Orn-Leu-Gln-ACC

m/z: $[M+H]^+$ for $C_{33}H_{48}N_8O_9$ ($m/z_{\text{calcd}} = 701.3618$; $m/z_{\text{found}} = 701.3618$)



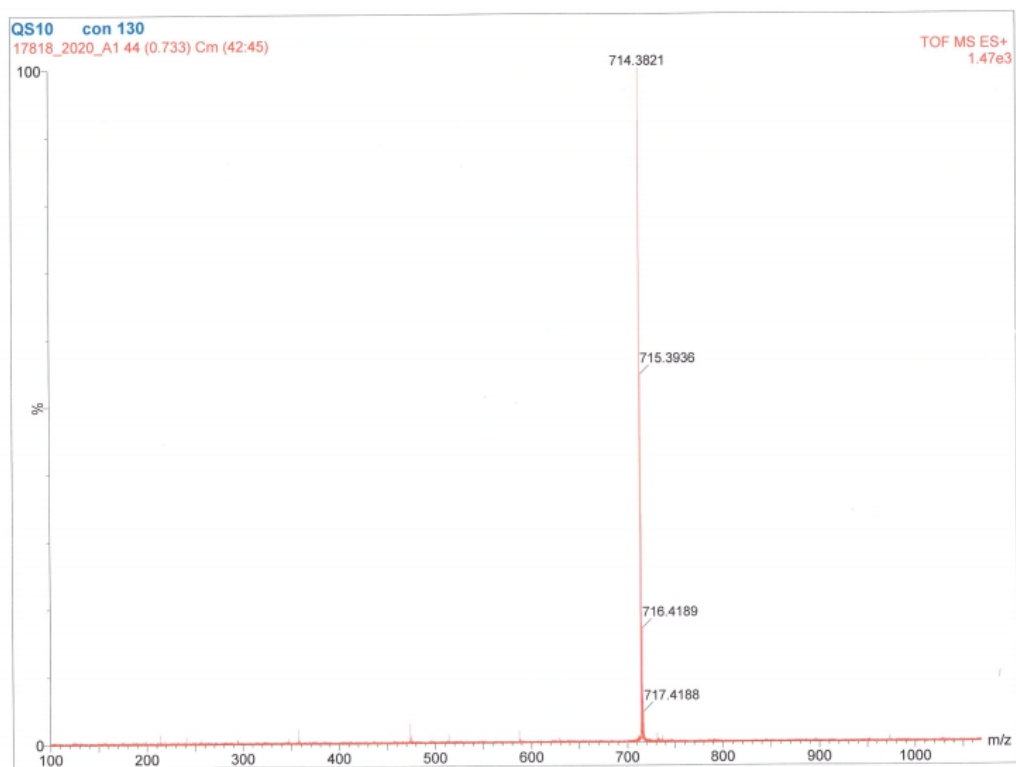
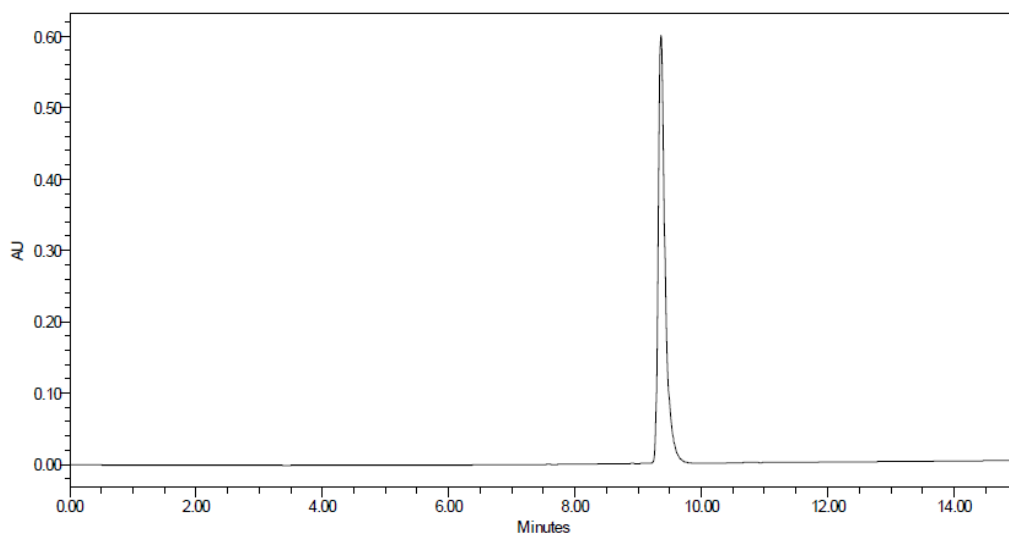
7 Ac-Abu-Lys-Leu-Gln-ACC

m/z: $[M+H]^+$ for $C_{34}H_{50}N_8O_9$ ($m/z_{\text{calcd}} = 715.3774$; $m/z_{\text{found}} = 715.3779$)



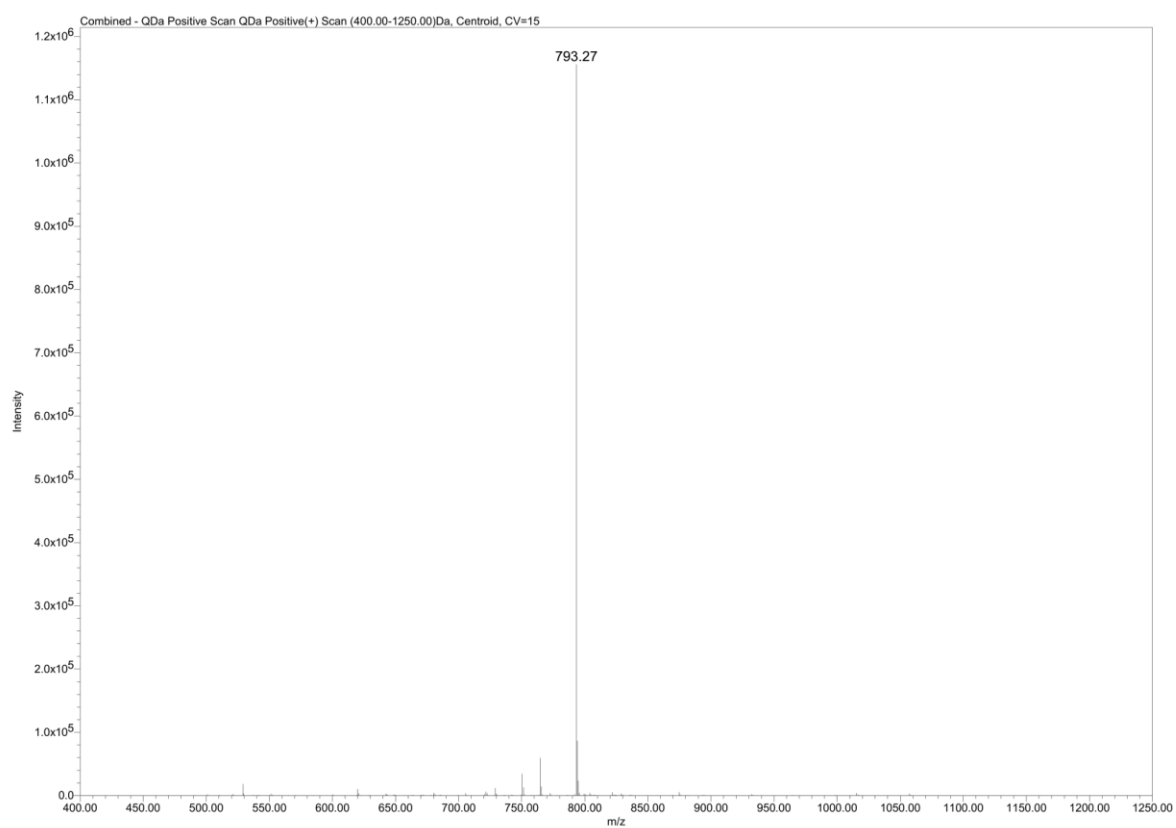
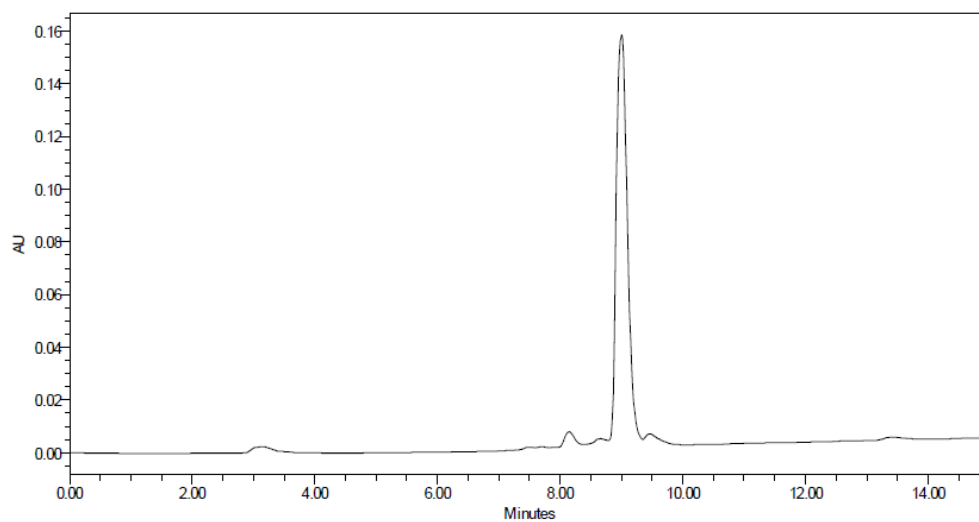
8 Ac-Abu-Tle-hLeu-Gln-ACC

m/z: $[M+H]^+$ for $C_{35}H_{51}N_7O_9$ ($m/z_{\text{calcd}} = 714.3822$; $m/z_{\text{found}} = 714.3821$)



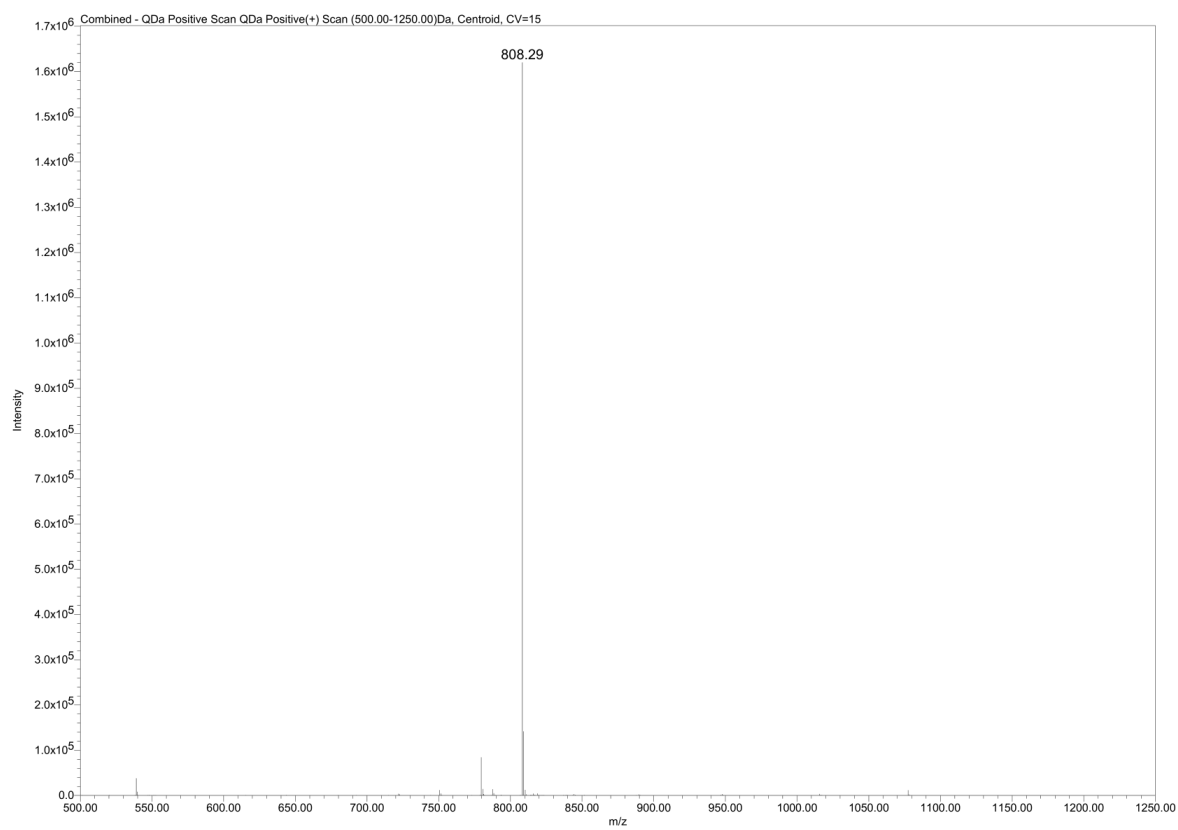
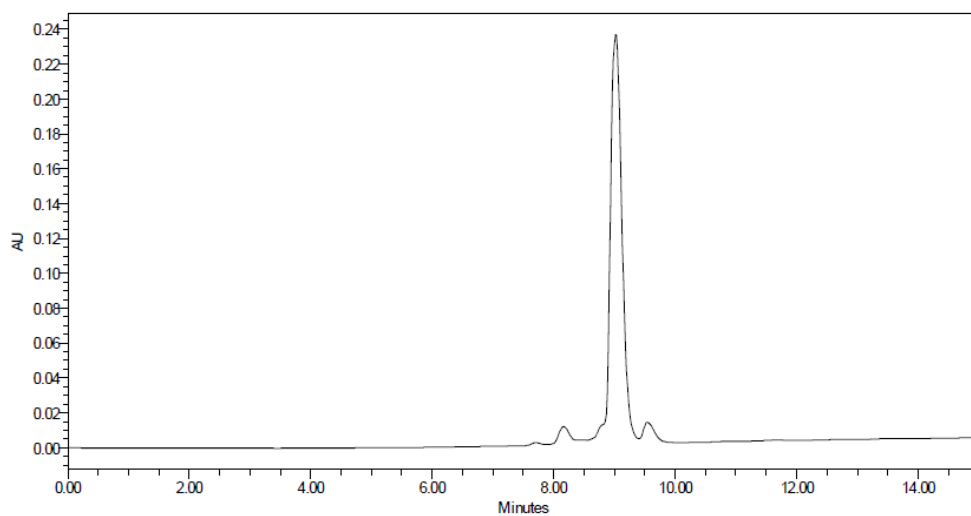
9 ACC-Gly-Abu-Tle-Leu-Gln-Ser-Gly-Phe-Arg-Lys(dnp)-Lys-NH₂

m/z: [M+H]²⁺ for C₇₂H₁₀₅N₂₁O₂₀ (m/z_{calcd} = 792.89; m/z_{found} = 793.27)



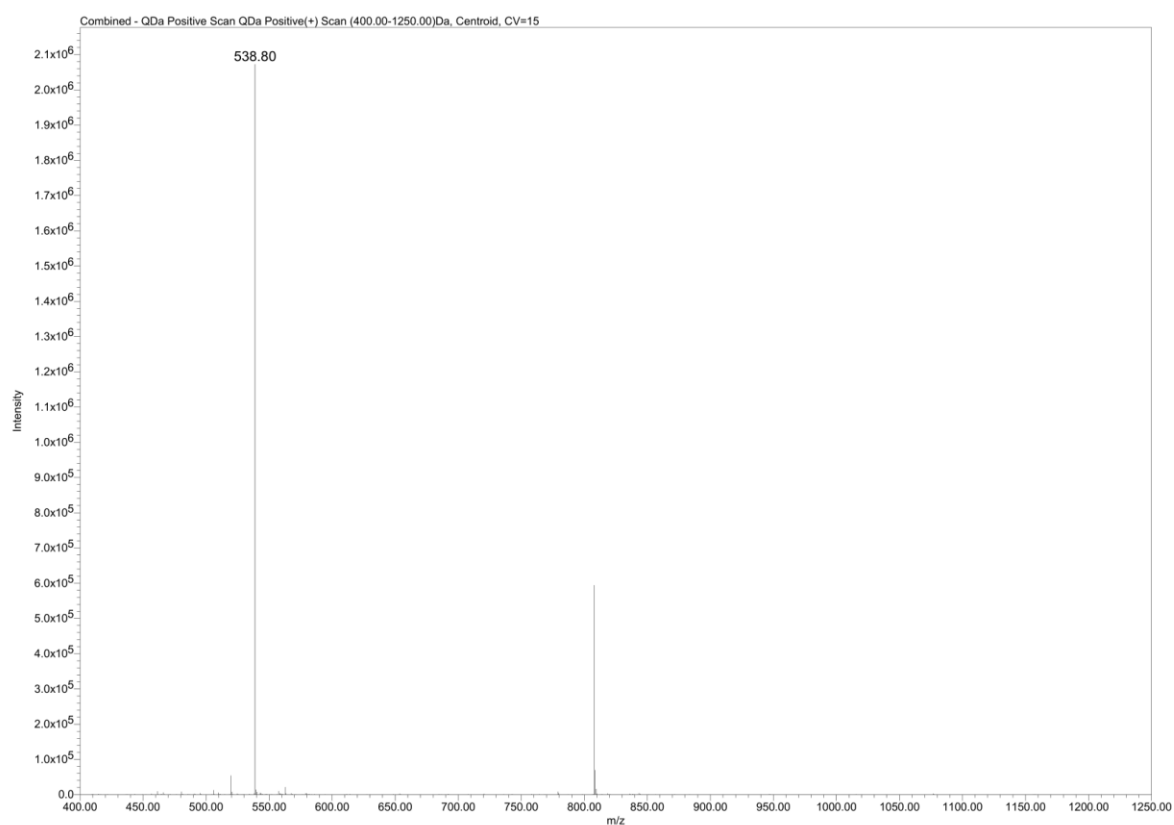
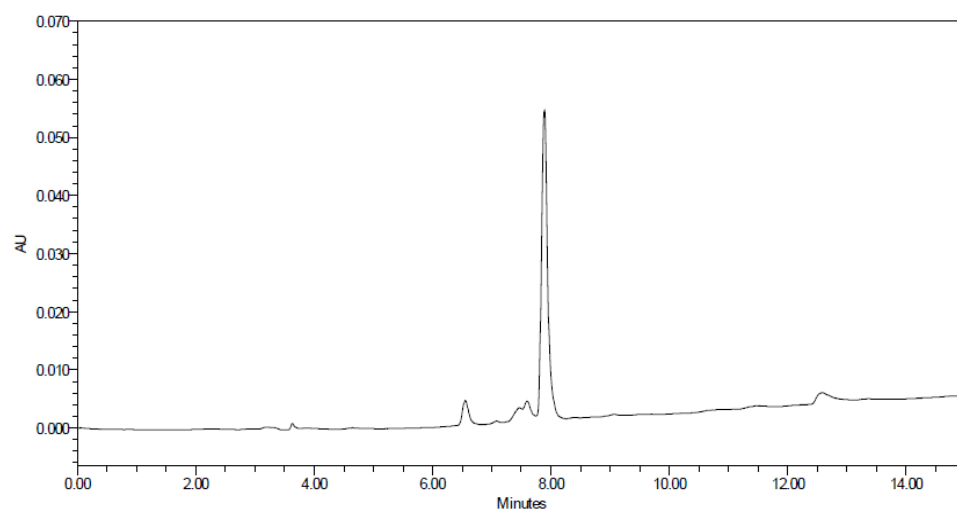
10 ACC-Gly-Thz-Tle-Leu-Gln-Ser-Gly-Phe-Arg-Lys(dnp)-Lys-NH₂

m/z: [M+H]²⁺ for C₇₂H₁₀₃N₂₁O₂₀S (m/z_{calcd} = 807.87; m/z_{found} = 808.29)



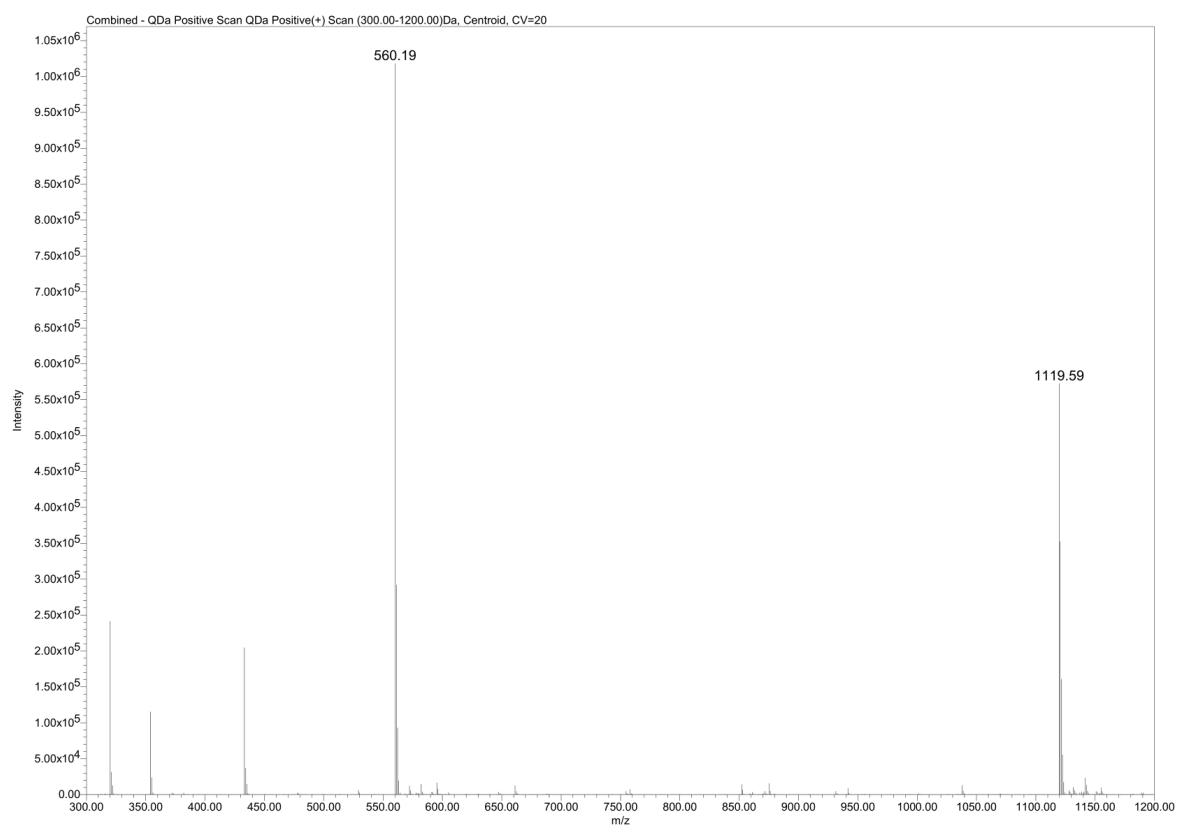
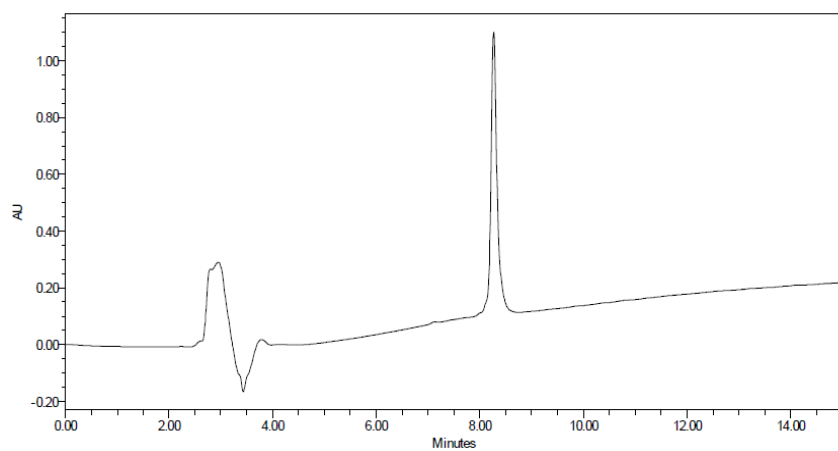
11 ACC-Gly-Val-Lys-Leu-Gln-Ser-Gly-Phe-Arg-Lys(dnp)-Lys-NH₂

m/z: [M+H]³⁺ for C₇₃H₁₀₈N₂₂O₂₀S (m/z_{calcd} = 538.60; m/z_{found} = 538.80)



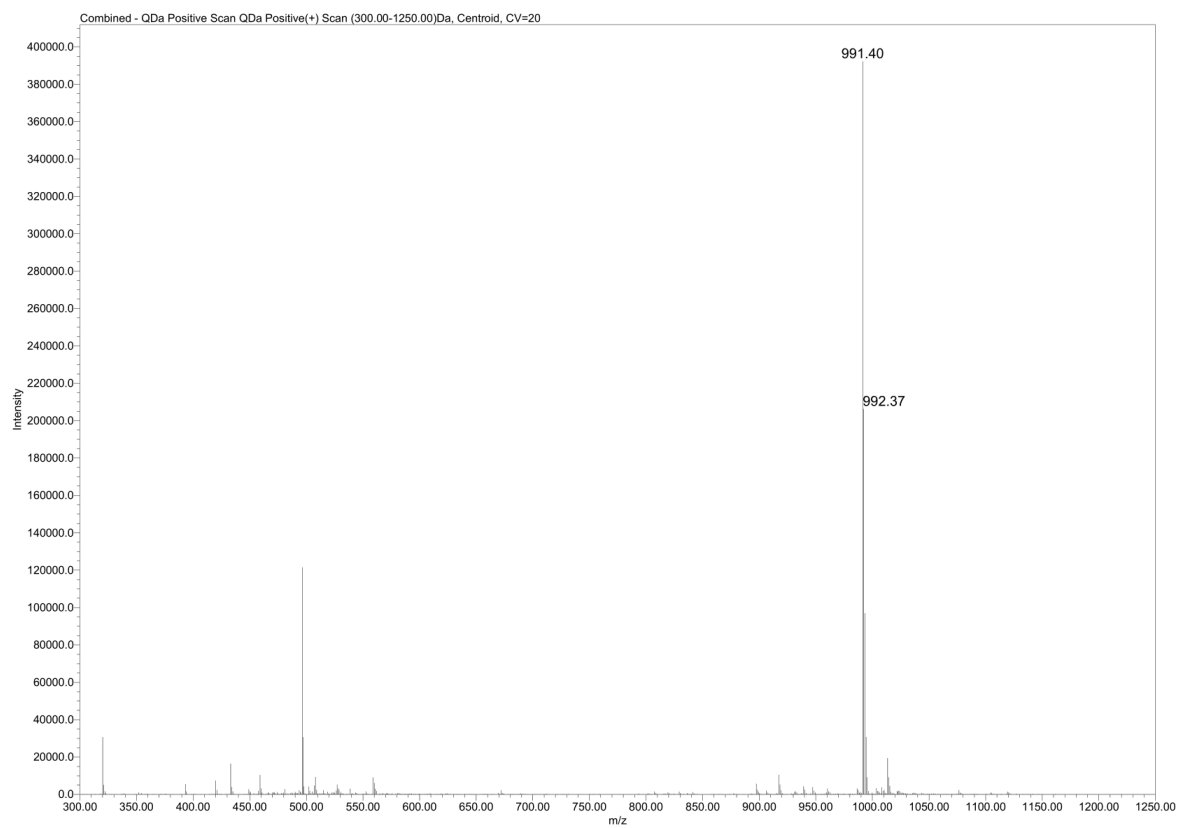
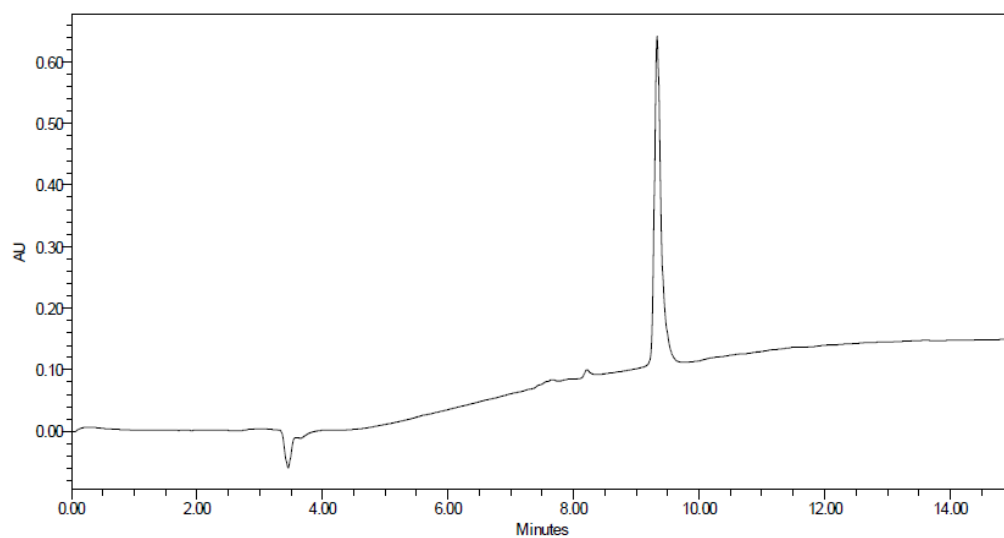
12 Ac-Abu-Tle-Leu-Gln-VS

m/z: $[M+H]^+$ for $C_{25}H_{45}N_5O_7S$ (m/z calcd = 560.31; m/z found = 560.19)



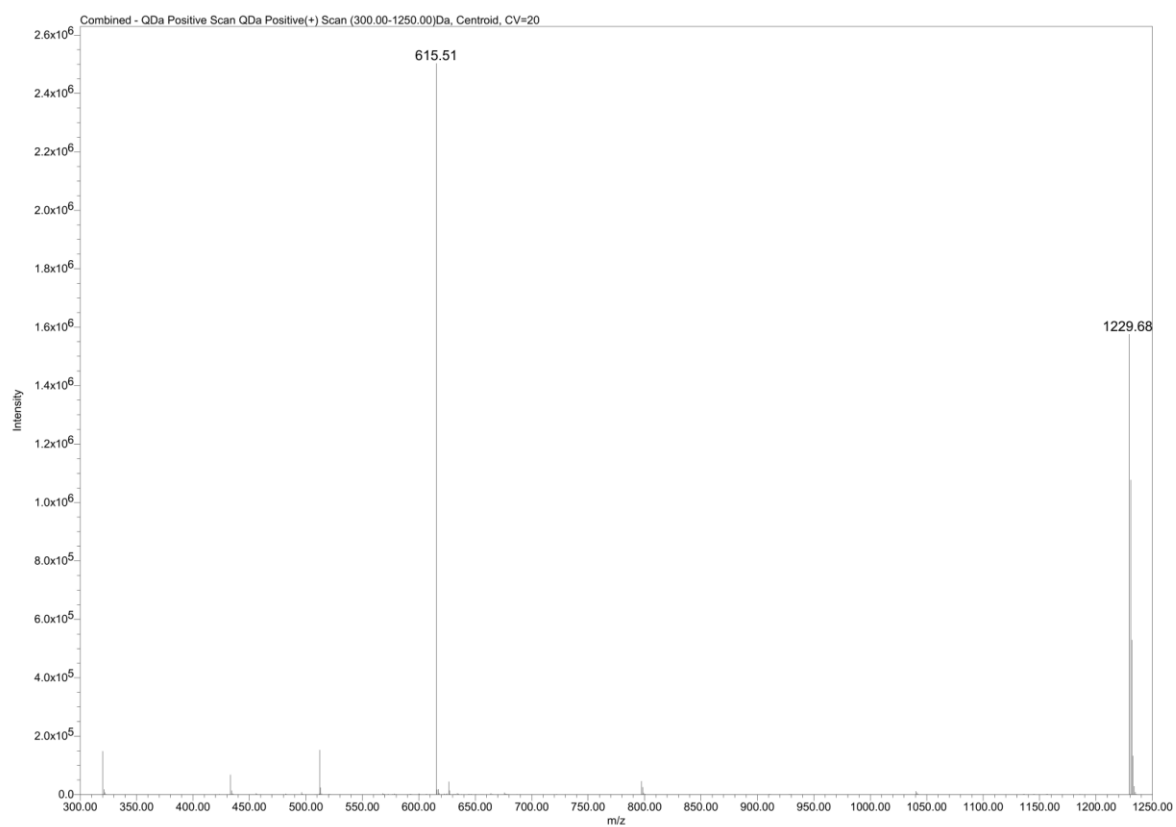
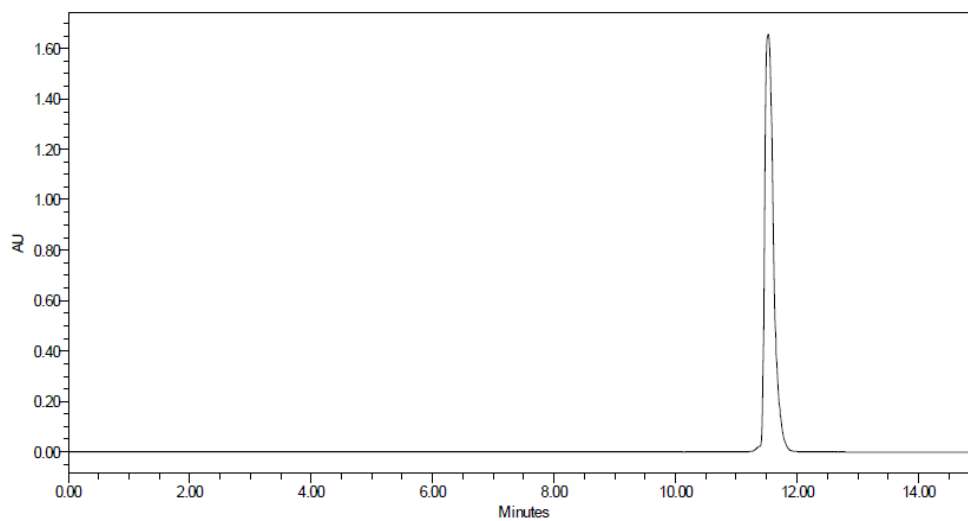
13 Biotin-PEG(4)-Abu-Tle-Leu-Gln-VS

m/z: $[M+H]^+$ for $C_{44}H_{78}N_8O_{13}S_2$ (m/z $_{\text{calcd}}$ = 991.52; m/z $_{\text{found}}$ = 991.40)



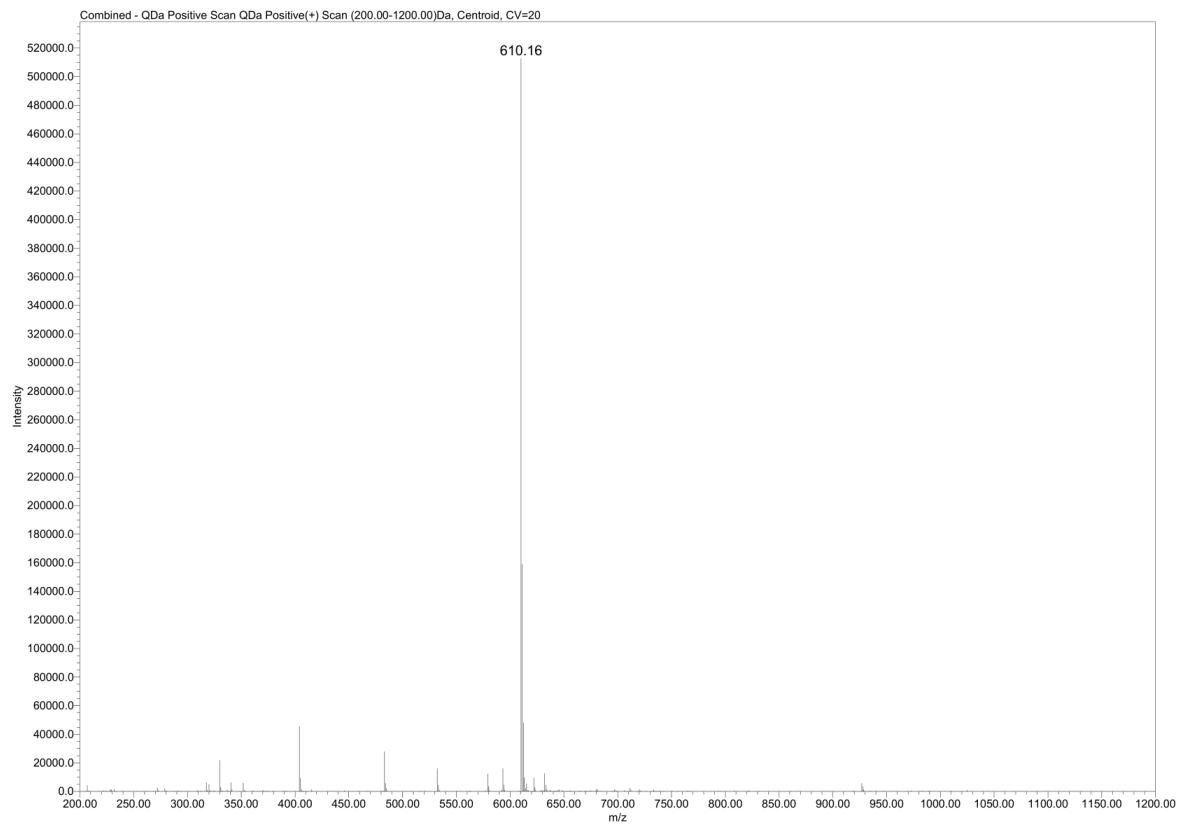
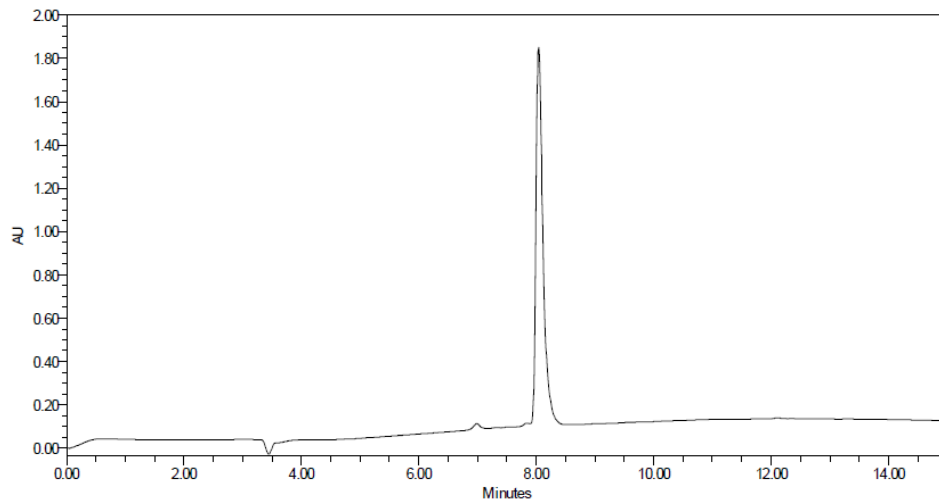
14 Cy5-PEG(4)-Abu-Tle-Leu-Gln-VS

m/z: $[M+H]^+$ for $C_{66}H_{101}N_8O_{12}S$ (m/z_{calcd} = 1229.73; m/z_{found} = 1229.68)



15 Ac-Abu-DTyr-Leu-Gln-VS, Ac-QS5-VS

m/z: $[M+H]^+$ for $C_{28}H_{43}N_5O_8S$ (m/z $_{calcd}$ = 610.29; m/z $_{found}$ = 610.16)



16 Bodipy-PEG(4)-Abu-DTyr-Leu-Gln-VS, Bodipy-QS5-VS

m/z: $[M+H]^+$ for $C_{51}H_{75}BF_2N_8O_{13}S$ (m/z_{calcd} = 1089.52; m/z_{found} = 1089.59)

