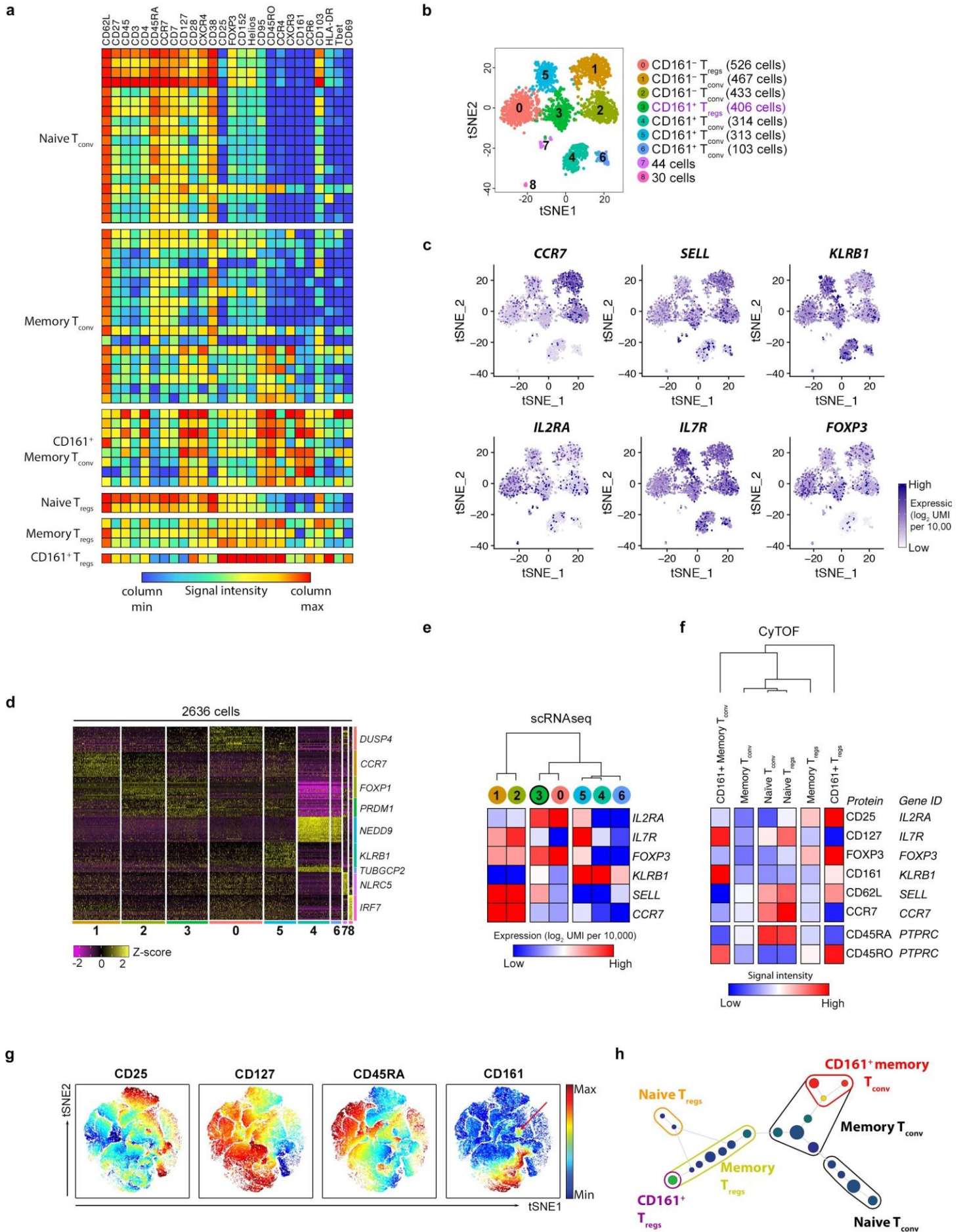


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Human retinoic acid-regulated CD161⁺ regulatory T cells support wound repair in intestinal mucosa

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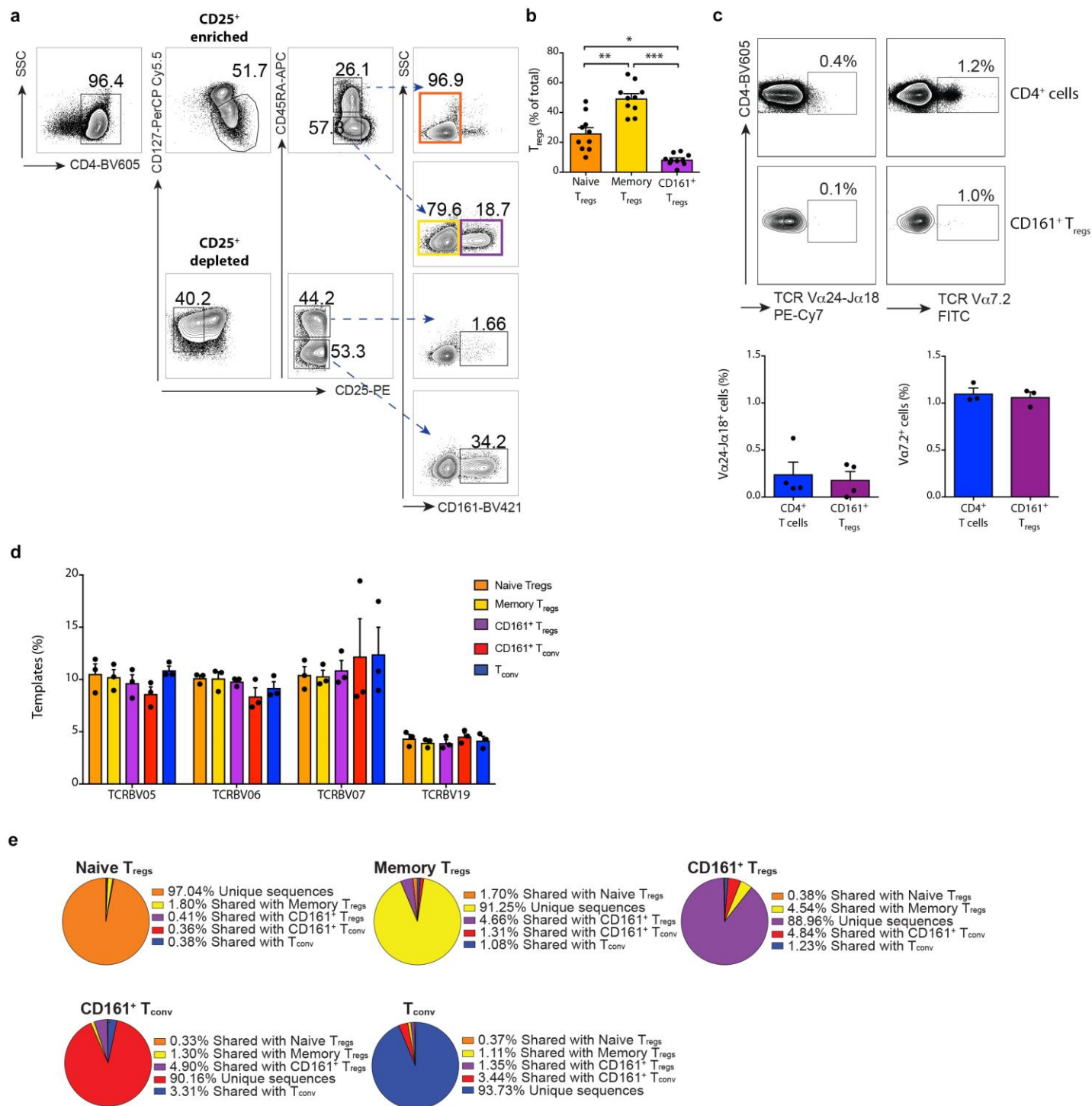
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Supplementary Figure 1

CD161⁺ T_{reg} cells are a discrete population of memory T_{reg} cells.

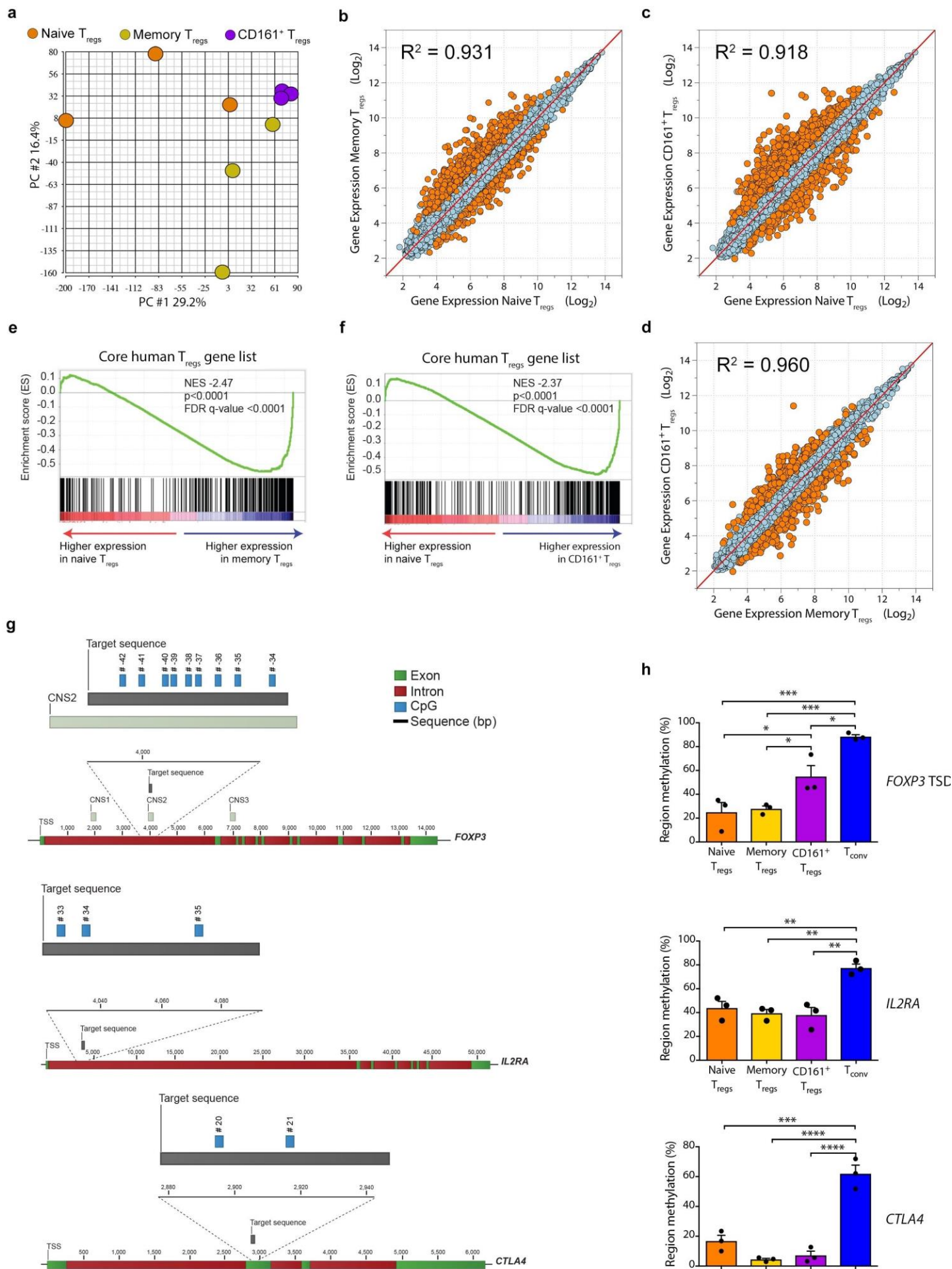
(a) Heat map showing signal intensity of selected markers for each node of the populations shown in Fig. 1c. (b–e) Clustering of populations of T_{conv} and T_{reg} cells by single-cell RNA-seq (scRNAseq) showing the tSNE plot of 2,636 cells from $n = 3$ independent donors separated into six major (clusters 0–6) and two minor (clusters 7 and 8) clusters (b), feature plots showing expression of three T_{reg} markers (*IL2RA* encoding CD25, *IL7R* encoding CD127 and *FOXP3* encoding FOXP3), *KLRB1* (encoding CD161) and two naive/memory markers (*SELL* encoding CD62L and *CCR7* encoding CCR7) (c), a heat map showing the top 20 discriminatory genes per cluster with representative genes of each cluster indicated (d) and population clustering (e) based on expression of the transcripts shown in c. Clusters in b are color-coded and labeled according to expression of the markers shown in e. (f) Clustering of T_{conv} and T_{reg} subpopulations using protein expression data sourced from CyTOF in Fig 1a incorporating the same markers as in e and also including CD45RA and CD45RO. Clustering of subpopulations in f is similar to that from scRNAseq (e). (g,h) viSNE plots (g) and 2D minimum spanning tree (h) of CD4⁺ T cells clustered following staining with anti-CD4, anti-CD25, anti-CD127, anti-CD45RA and anti-CD161 for flow cytometry. Shown in g are heat maps for expression of the indicated markers, with an arrow indicating CD161 expression in T_{reg} cells. Node size in h represents cell number and color CD161 median intensity. Grouped together are naive (circled in orange), memory (circled in yellow) and CD161⁺ (circled in purple) T_{reg} cells, as well as populations of naive (circled in black), memory (circled in black) and CD161⁺ (circled in red) T_{conv} cells. The data in g and h are representative of $n = 4$ experiments. * $P < 0.01$, ** $P < 0.001$, *** $P < 0.0001$ by one-way ANOVA.



Supplementary Figure 2

CD161⁺ T_{reg} cells are distinct from other CD161-expressing T cells.

(a) Representative example of the FACS sorting strategy for naive (orange), memory (yellow) and CD161⁺ (purple) T_{reg} cells. (b) Proportions of each T_{reg} subpopulation in healthy human donor peripheral blood (cumulative data from $n = 10$ donors); bars show mean + s.e.m. (c) Representative (top) and cumulative (bottom) expression of TCR V α 24-J α 18 and V α 7.2 in total CD4⁺ T cells and CD161⁺ T_{reg} cells. Both cell types show minimal expression of these invariant TCR chains. Data are from $n = 3$ independent experiments. (d) Contribution of the three highest (BV05, BV06 and BV07) and one of the lowest (BV19) TCRBV families to the overall TCR repertoire in naive, memory and CD161⁺ T_{reg} cells as well as CD161⁺ and total T_{conv} cells. Data are from $n = 3$ independent experiments; bars show mean + s.e.m. (e) Average percentage of TCR sequences either unique or shared among naive, memory and CD161⁺ T_{reg} cells, CD161⁺ and total T_{conv} cells (cumulative data from $n = 3$ experiments). * $P < 0.01$, ** $P < 0.001$, *** $P < 0.0001$ by one-way ANOVA.

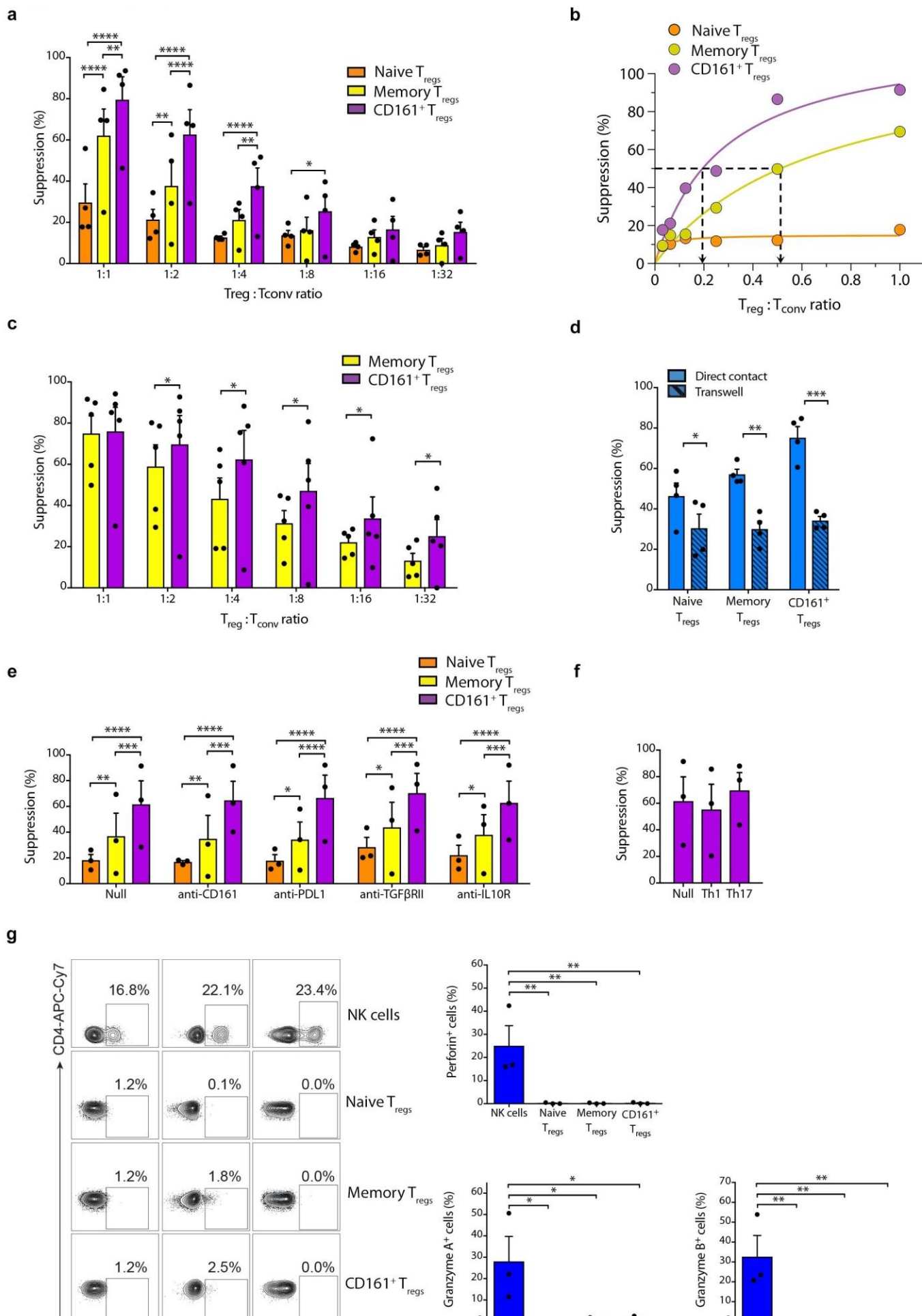


Supplementary Figure 3

Transcriptome and methylome of CD161⁺ T_{reg} cells.

(a) Principal-component analysis (PCA) 2D mapping of variance in expression of transcripts in freshly isolated naive, memory and CD161⁺ T_{reg} cells ($n = 3$). (b–d) Scatterplots showing correlation between mean gene expression of freshly isolated T_{reg} subpopulations ($n = 3$). (e,f) GSEA plots for core human T_{reg} signature genes comparing freshly isolated naive to memory T_{reg} cells (e) and to CD161⁺ T_{regs} cells (f) ($n = 3$ per group). NES, normalized enrichment score; empirical P and multiple-test adjusted q values from GSEA are shown. (g) Schematic representation of *FOXP3*, *IL2RA* and *CTLA4* gene loci showing the target sequence and conserved CpGs assayed for methylation analysis in Fig. 2e. (h) Mean region methylation percentage of *FOXP3* TSDR, *IL2RA* and *CTLA4* loci from three male donors. Data are from $n = 3$ independent experiments; bars show mean + s.e.m. * $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$; **** $P < 0.0001$ by one-way ANOVA.

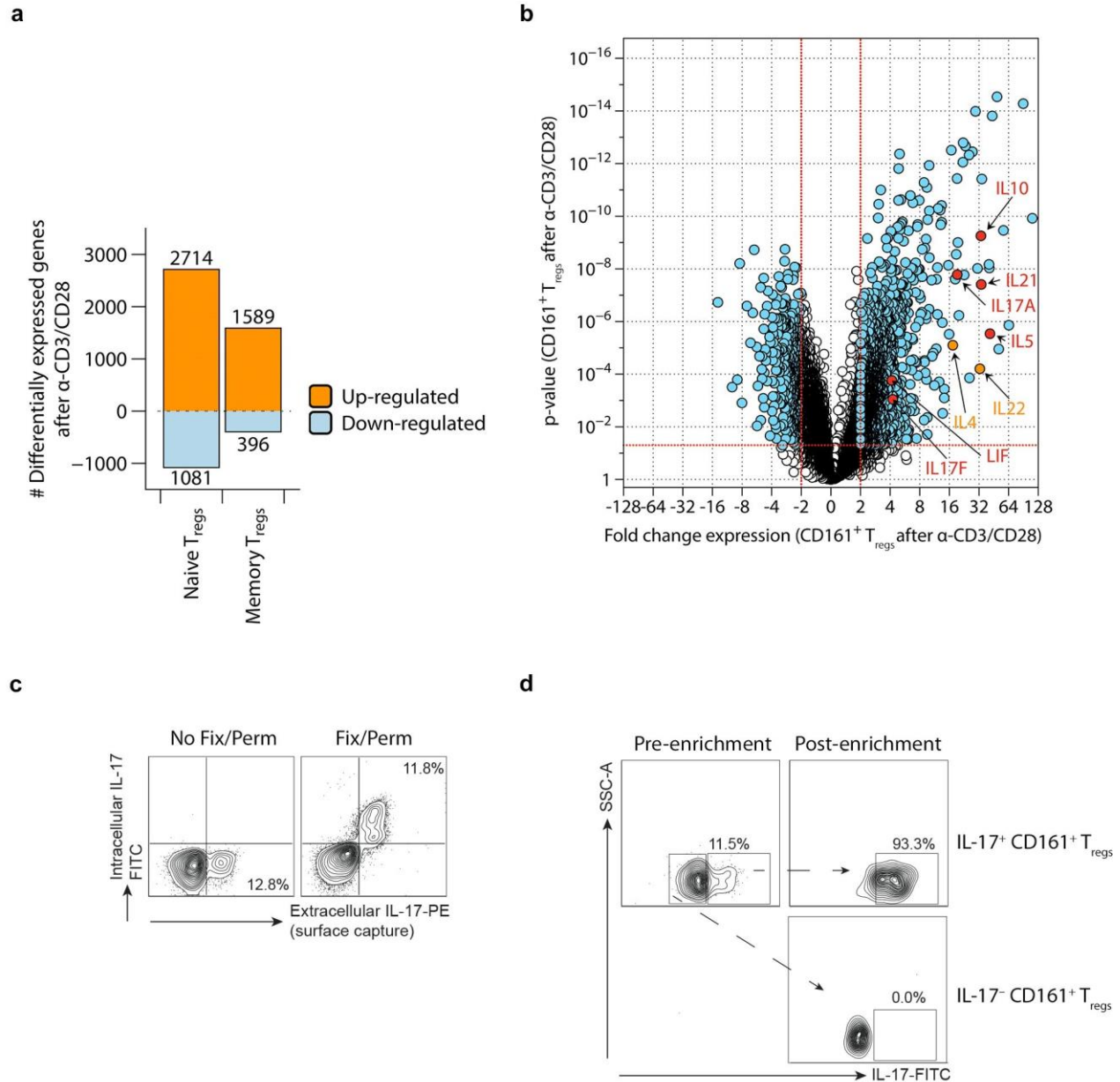
surface protein expression on memory and CD161⁺ T_{reg} cells; shown are representative flow cytometry plots (left) and cumulative data from $n = 3$ independent experiments (right). (c) CD161 expression on T_{reg} cells co-cultured for 5 d with DCs with and without BMS493 at increasing concentrations. Shown are representative plots (left) and cumulative data (right) from $n = 3$ independent experiments. (d,e) Sequence logo for RARA DNA motif (d) and schematic representation of the *KLRB1* and *CCR9* gene loci (± 5 kb) showing the predicted binding sites of RARA (e); the red arrowhead indicates the binding site for *CCR9* selected for analysis in CHIP-qPCR. Bars show mean + s.e.m. throughout the figure. * $P < 0.05$ by paired t -test.



Supplementary Figure 5

CD161⁺ T_{reg} cells are highly regulatory.

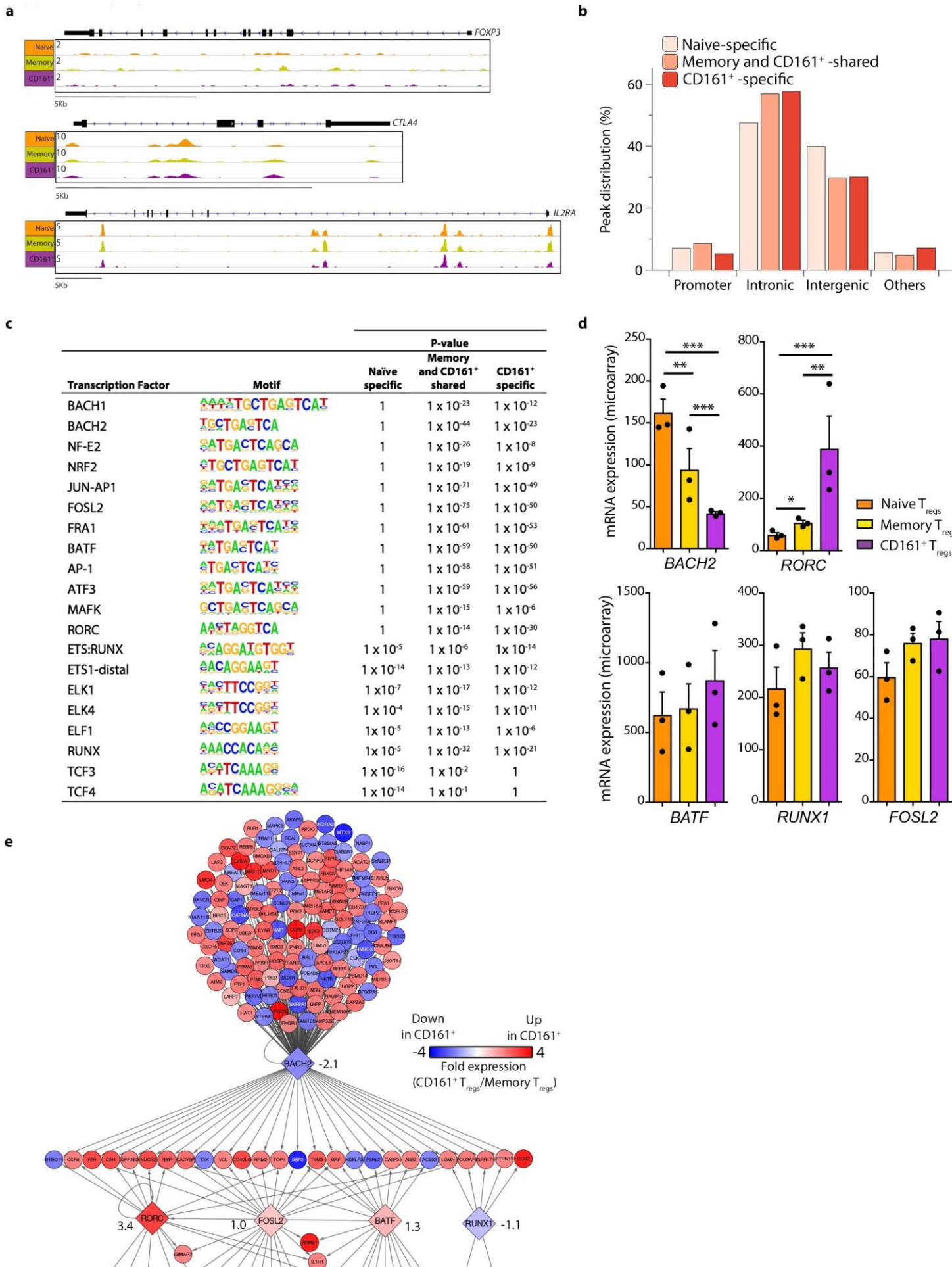
(a,b) Cumulative mean percentage suppressive function of freshly isolated naive, memory and CD161⁺ T_{reg} cells from Fig. 4a ($n = 4$ experiments) (a) and representative IC₅₀ value calculation (b; dashed arrows show the IC₅₀ for CD161⁺ and memory T_{reg} cells). (c) Cumulative mean percentage suppressive function of memory and CD161⁺ T_{reg} cells after in vitro expansion for 2 weeks ($n = 4$ experiments). (d) Suppression of T_{conv} cell proliferation when in direct contact with T_{reg} cells or when separated by a Transwell at a T_{reg}:T_{conv} ratio of 1:1 (cumulative data from $n = 4$ independent experiments). (e) Suppressive function of T_{reg} cells under neutral (null) conditions or in the presence of blocking antibodies directed against CD161, PDL1, TGFβRII or IL10R, all at a T_{reg}:T_{conv} ratio of 1:2 (cumulative data from $n = 3$ independent experiments). (f) Suppressive function of CD161⁺ T_{reg} cells under neutral (null) conditions or in the presence of T_H1 and T_H17 skewing conditions, all at a T_{reg}:T_{conv} ratio of 1:2 (cumulative data from $n = 3$ independent experiments). (g) Expression of perforin, granzyme A and B in subpopulations of T_{reg} cells, with NK cells as a positive control; shown are representative flow cytometry plots for each marker (left) and cumulative data from $n = 3$ independent experiments (right). Bar charts show mean + s.e.m. throughout the figure. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$, **** $P < 0.0001$ by two-way ANOVA (a–e) and one-way ANOVA (g).



Supplementary Figure 6

CD161⁺ T_{reg} cells produce multiple cytokines upon activation.

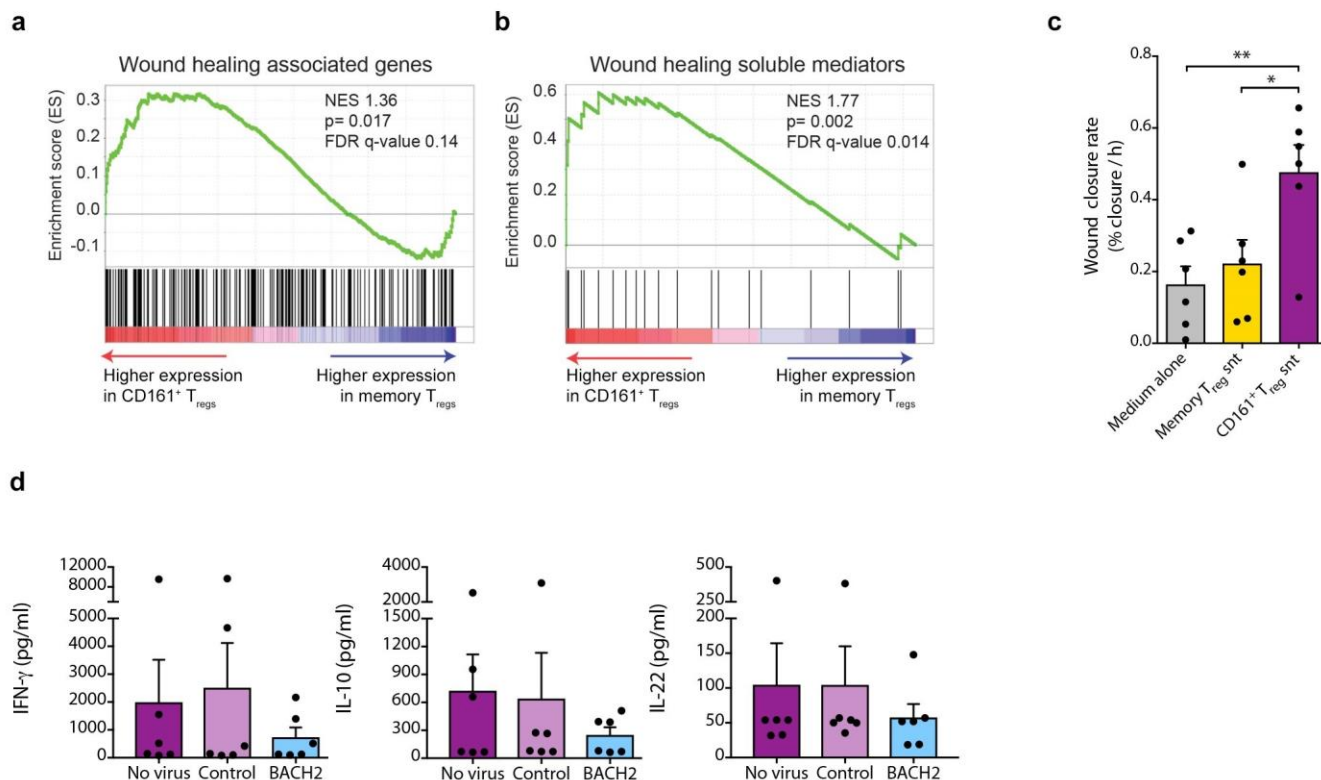
(a) Differentially expressed genes (at least twofold change at $P < 0.05$) following 4 h of stimulation of memory and naive T_{reg} cells with anti-CD3/CD28. (b) Volcano plot showing significant genes differentially expressed in CD161⁺ T_{reg} cells after 4 h of activation with anti-CD3/CD28. Differentially expressed genes are shown in blue; red indicates significantly upregulated cytokine genes that are differentially expressed compared to memory T_{reg} cells (see Fig. 5c); orange indicates other cytokine genes of interest upregulated in activated CD161⁺ T_{reg} cells. The data in a are b are from three independent experiments. (c,d) Cell sorting of IL-17⁺ and IL-17⁻ CD161⁺ T_{reg} cells by surface IL-17 capture. Shown are representative flow cytometry plots with double staining for intracellular and extracellular (captured on the surface) IL-17 with (right) and without (left) fixation and permeabilization (c) and the sorting strategy using IL-17 surface capture and post-sorting purity for the IL-17⁺ and IL-17⁻ fractions of CD161⁺ T_{reg} cells (d). Shown in c and d are representative examples from $n = 3$ and $n = 4$ independent experiments, respectively.



Supplementary Figure 7

Global analysis of T_{reg} regulomes.

(a) Open chromatin regions (OCRs) at prototypical T_{reg} gene loci from Fig. 6a. (b) Genomic distribution of ATAC peaks (promoter, intragenic or intergenic regions) in the three groups shown in Fig. 6a. (c) *P* values (cumulative hypergeometric test *P* values calculated by Homer) for transcription factor (TF) motifs shown in each cluster in Fig. 6c. Data in **a–c** are from *n* = 3 experiments. (d) Expression of key TFs participating in gene regulation in CD161⁺ T_{reg} cells. Shown is the normalized signal intensity (mean + s.e.m.) from *n* = 3 microarrays for each T_{reg} population. (e) Integrated TF network showing the contribution of each TF to DEGs and overlap between them (fold difference in expression of each TF is also indicated next to the TF). **P* < 0.05, ***P* < 0.01, ****P* < 0.001 (Partek analysis of microarrays).



Supplementary Figure 8

CD161⁺ T_{reg} cells accelerate wound healing.

(a,b) GSEA plots for wound healing–associated genes (a) and wound healing–associated soluble mediators (b) comparing activated CD161⁺ to memory T_{reg} cells. Data are from $n = 3$ independent experiments. NES, normalized enrichment score; empirical P and multiple-test-adjusted q values from GSEA are indicated. (c) Wound closure rate of Caco-2 cells cultured in the presence of either medium alone or medium supplemented with culture supernatants (snt) from activated memory or CD161⁺ T_{reg} cells. Shown are cumulative data from $n = 6$ independent experiments. (d) Concentration (pg/ml) of stated cytokines in supernatants of CD161⁺ T_{reg} cells transduced, or not, with control lentivirus or lentivirus encoding BACH2 (cumulative data from $n = 6$ experiments). Bar charts show mean + s.e.m.; * $P < 0.05$, ** $P < 0.01$ by one-way ANOVA.