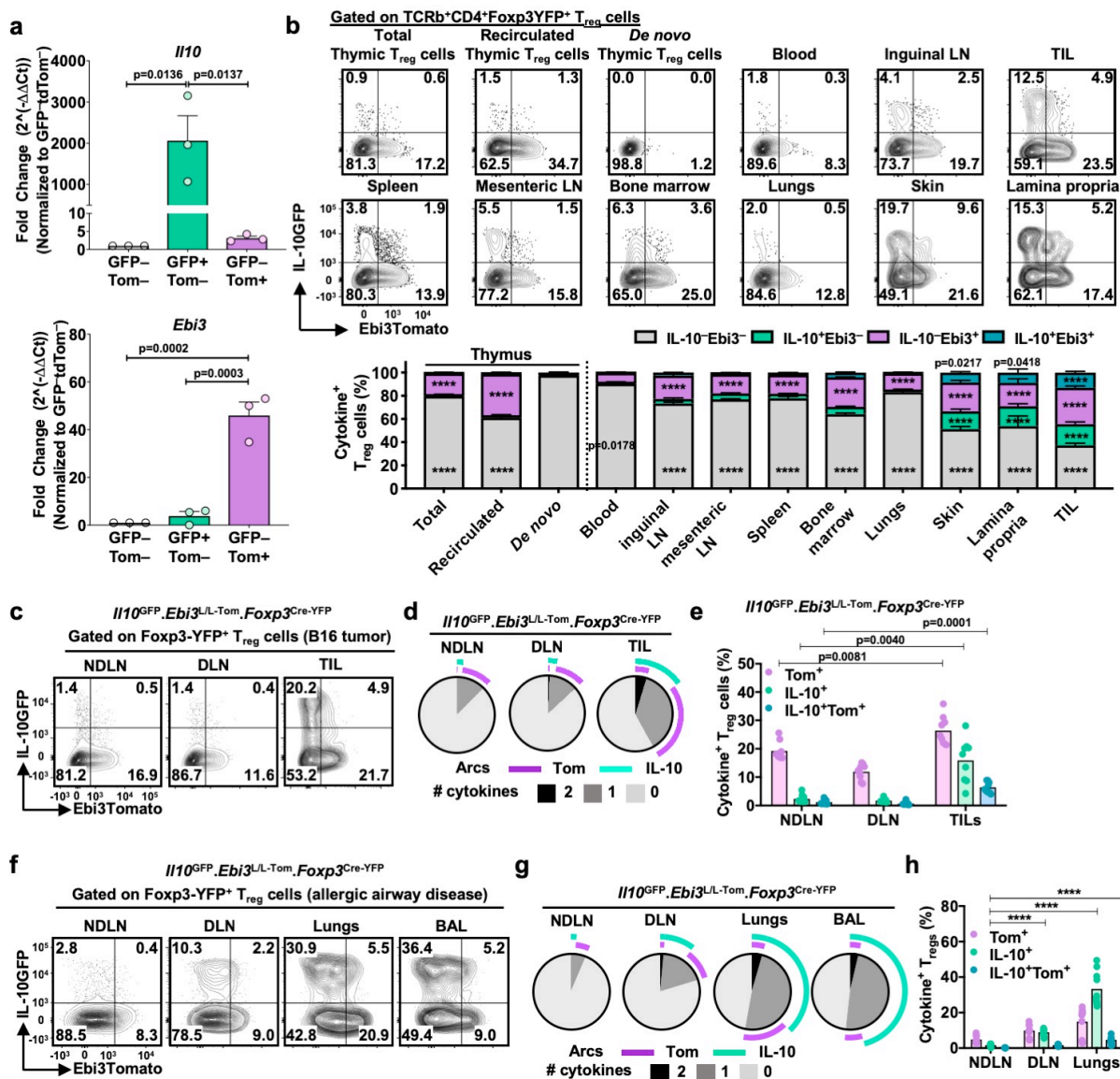


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Adaptive plasticity of IL-10⁺ and IL-35⁺ T_{reg} cells cooperatively promotes tumor T cell exhaustion

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Distribution and characterization of IL-10- and IL-35-expressing T_{reg} cells in inflammatory environments

a, Validation of fluorescent reporters by real-time RT-PCR. Splenic T_{reg} cell subpopulations were sorted from *Il10*^{GFP}.*Ebi3*^{Tom}.*Foxp3*^{Cre-YFP} mice, and *Il10* and *Ebi3* transcripts were analyzed. Relative fold change expression was determined by normalizing to GFP⁺Tom⁺ double-negative T_{reg} cells. Data averaged from 3 independent experiments. Statistical significance was determined by One-way ANOVA with Holm-Sidak multiple comparisons (*P* values as indicated).

b, Representative flow plots depicting the expression of IL-10 and *Ebi3* in T_{reg} cells isolated from the indicated tissues of 8wks old naive *Il10*^{GFP}.*Ebi3*^{Tom}.*Foxp3*^{Cre-YFP} mice (*top*). Stacked-bar graph depicting the percent distribution of subpopulations (*bottom*). Recirculating (CD62L⁺Nrp1⁺) and *de novo* (CD62L⁺Nrp1⁻) T_{reg} cells for Thymus. Data were averaged from 2 independent experiments with *n*=5 mice per tissue, except for thymus, blood, and lamina propria (*n*=3). Error bars represent s.e.m. Statistical significance was determined by comparing tissues to *de novo* Thymus with Two-way ANOVA with Holm-Sidak multiple comparisons (*****p*<0.0001 and other *P* values as indicated).

c, Representative flow plots depicting expression of IL-10⁺ and Tom⁺ cells within Foxp3-YFP⁺ T_{reg} cells isolated from day 14 B16 tumor-bearing *Il10*^{GFP}.*Ebi3*^{L/L-Tom}.*Foxp3*^{Cre-YFP} mice. CD4⁺YFP⁺ T_{reg} cells were assessed for the expression of IL-10 and Tom in NDLN, DLN and TIL.

d, SPICE plots depicting co-expression pattern of IL-10 and Tom on TIL T_{reg} cells from B16 tumor-bearing *Il10*^{GFP}.*Ebi3*^{L/L-Tom}.*Foxp3*^{Cre-YFP} mice as in (**c**). Data averaged from 3 independent experiments with *n*=9 mice total.

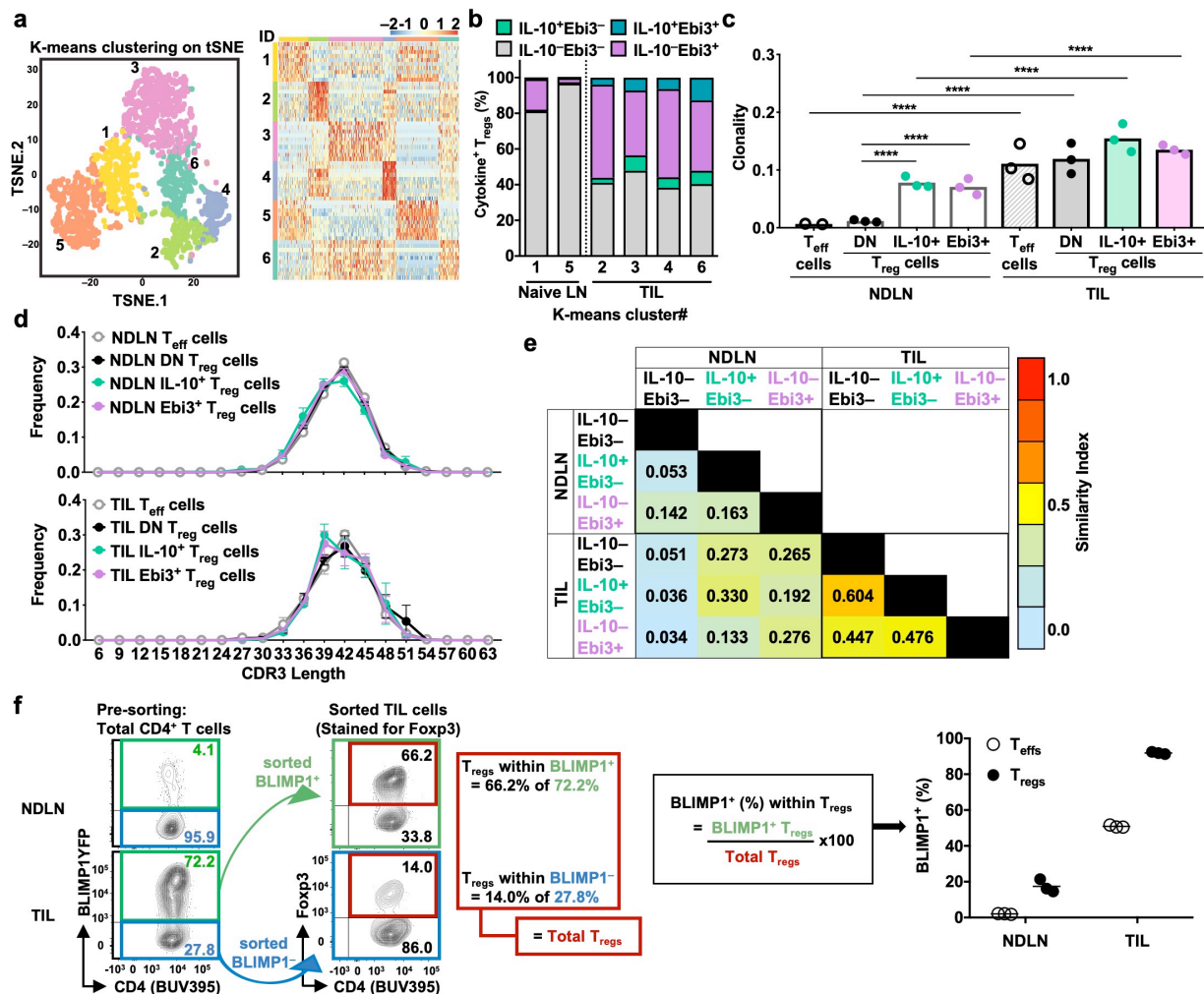
e, Scatter-bar plot depicting percent distribution of T_{reg} cell subpopulations as in (**c**). Bars represent mean values.

f, Representative flow plots depicting the distribution of IL-10⁺ and Tom⁺ cells within Foxp3-YFP⁺ T_{reg} cells isolated from d14 allergic inflammation induced *Il10*^{GFP}.*Ebi3*^{L/L-Tom}.*Foxp3*^{Cre-YFP} mice.

g, SPICE plots depicting co-expression pattern of IL-10 and Tom on T_{reg} cells as described in (**f**). Data averaged from 3 independent experiments with *n*=10 mice total.

h, Scatter-bar plot depicting percent distribution of T_{reg} cell subpopulations as in (**f**). Bars represent mean values.

(**e** and **h**) Statistical significance was determined by Two-way ANOVA with Holm-Sidak multiple comparisons (*****p*<0.0001 and other *P* values as indicated).



Supplementary Figure 2

Substantial TCR clonal overlap among intratumoral T_{reg} cell subpopulations, consistent with lack of distinct transcriptomic signatures

a, scRNAseq tSNE plots of bulk T_{reg} cells from naive LNs and day 14 B16 Tumors (TIL T_{reg} cells) from *Foxp3*^{Cre-YFP} mice, with unsupervised K-means clustering (left) and a heat map depicting the top 10 differentially expressed genes for each of the 6 clusters identified (right).

b, Stacked-bar graph depicting the percent distribution of T_{reg} cell subpopulations within each cluster identified as in (a).

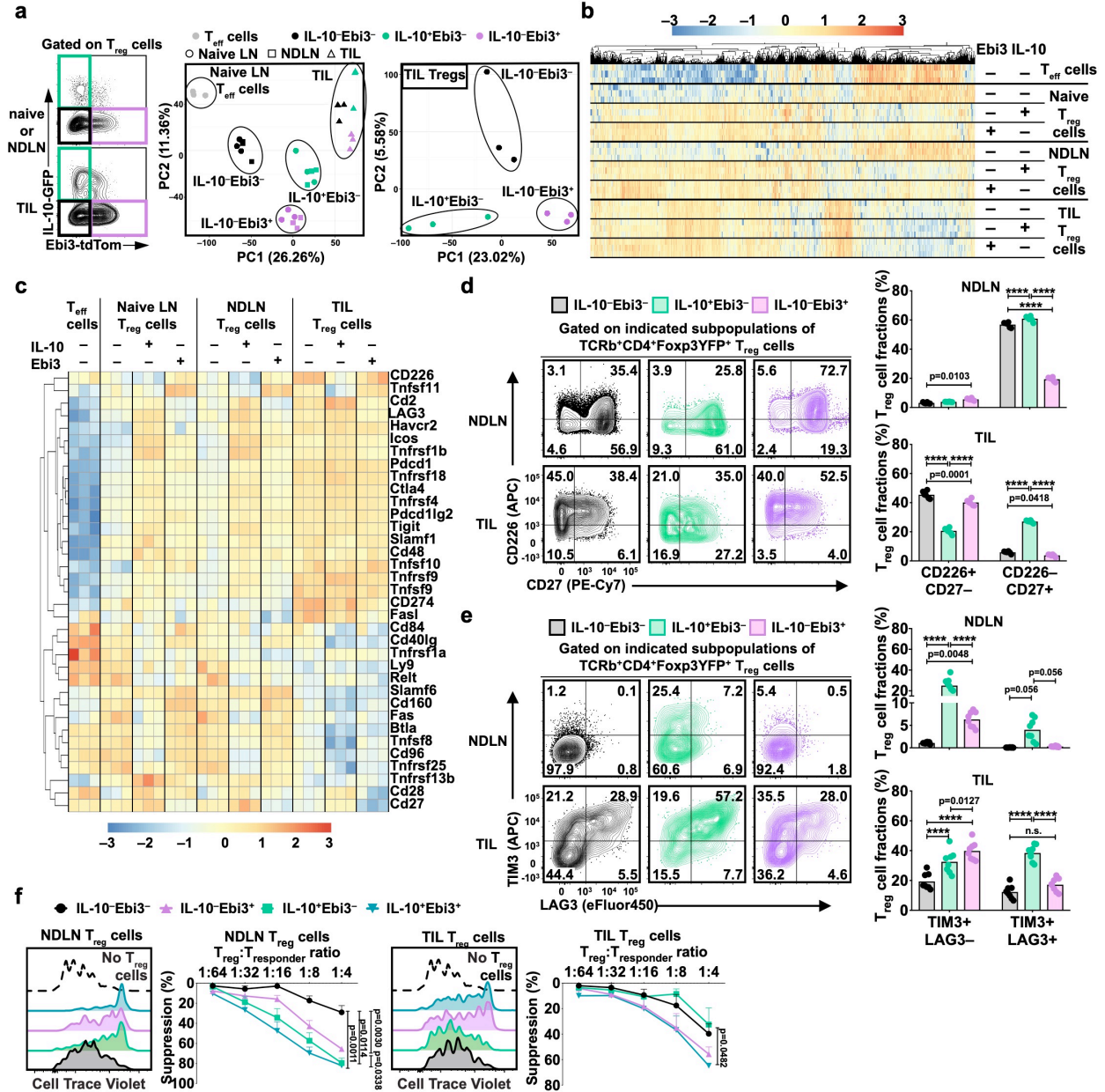
c, Clonality was determined by examining the degree of TCR diversity within each population by TCRseq. Productive CDR3 sequences were used to define the TCR diversity. The indicated T_{eff} control and T_{reg} cell subpopulations were FACS sorted from day 14 B16 tumor-bearing *Il10*^{GFP}.*Ebi3*^{Tom}.*Foxp3*^{Cre-YFP} mice for genomic DNA purification. Bars represent mean values. Statistical significance was determined by One-way ANOVA with with Holm-Sidak multiple comparisons (****p<0.0001).

d, Line graphs representing the mean length of productive CDR3 locus sequence comparing the T_{reg} cells subpopulations and effector T cells. Error bars represent s.e.m.

e, TCR clonal similarity was determined by Morisita-Horn similarity index cross-examining the productive CDR3 sequences of Teff control and T_{reg} cell subpopulations. Representative of three independent mouse samples is shown.

(c-e) Data combined from TCR-sequencing data from three separate, individual *Il10*^{GFP}.*Ebi3*^{Tom}.*Foxp3*^{Cre-YFP} mice. Error bars represent s.e.m.

f, Representative flow plots depicting the BLIMP1YFP expression level plotted against CD4. *(left)*: BLIMP1YFP⁻ and BLIMP1YFP⁺ fractions of CD4⁺ TILs were purified by FACS sorting from d14 B16 tumor-bearing Prdm1^{YFP} transgenic mice, followed by intracellular Foxp3 staining of the sorted cells. Data averaged from three sets of tumors with three tumors per set pooled for sorting. *(right)*: Scatter plot depicting the percent distribution of BLIMP1YFP⁺ T_{effs} and T_{reg} cells.



Supplementary Figure 3

Global transcriptomic analysis of intratumoral T_{reg} cell subpopulations

a, (left) Representative gating strategy for T_{reg} cell subpopulation sorting for RNAseq. Each population was double-sorted. (right) PCA plot demonstrating the segregation of naive LN, NDLN, and B16 tumor derived TIL T_{reg} cell subpopulations isolated from B16 tumor-bearing *Il10^{GFP}.Ebi3^{Tom}.Foxp3^{Cre-YFP}* mice defined by their IL-10 and IL-35 expression pattern (IL-10⁻Ebi3⁻, IL-10⁺Ebi3⁻, and IL-10⁻Ebi3⁺) as well as naive LN effector T cell control (left PCA). A separate PCA plot was generated depicting only TIL T_{reg} cell populations (right PCA). Data generated from three independent sequencing runs. Samples pooled from 4 mice per experiment to sort T_{reg} cell subpopulations. Each dot in PCA plots represent individual sequencing run. For statistical analysis q-value FDR of 0.05 was used to

determine the confidence of adjusted P values.

b, Heat map of top 5,000 differentially expressed genes representing cross-examination of Treg cell subpopulations as in **(a)**.

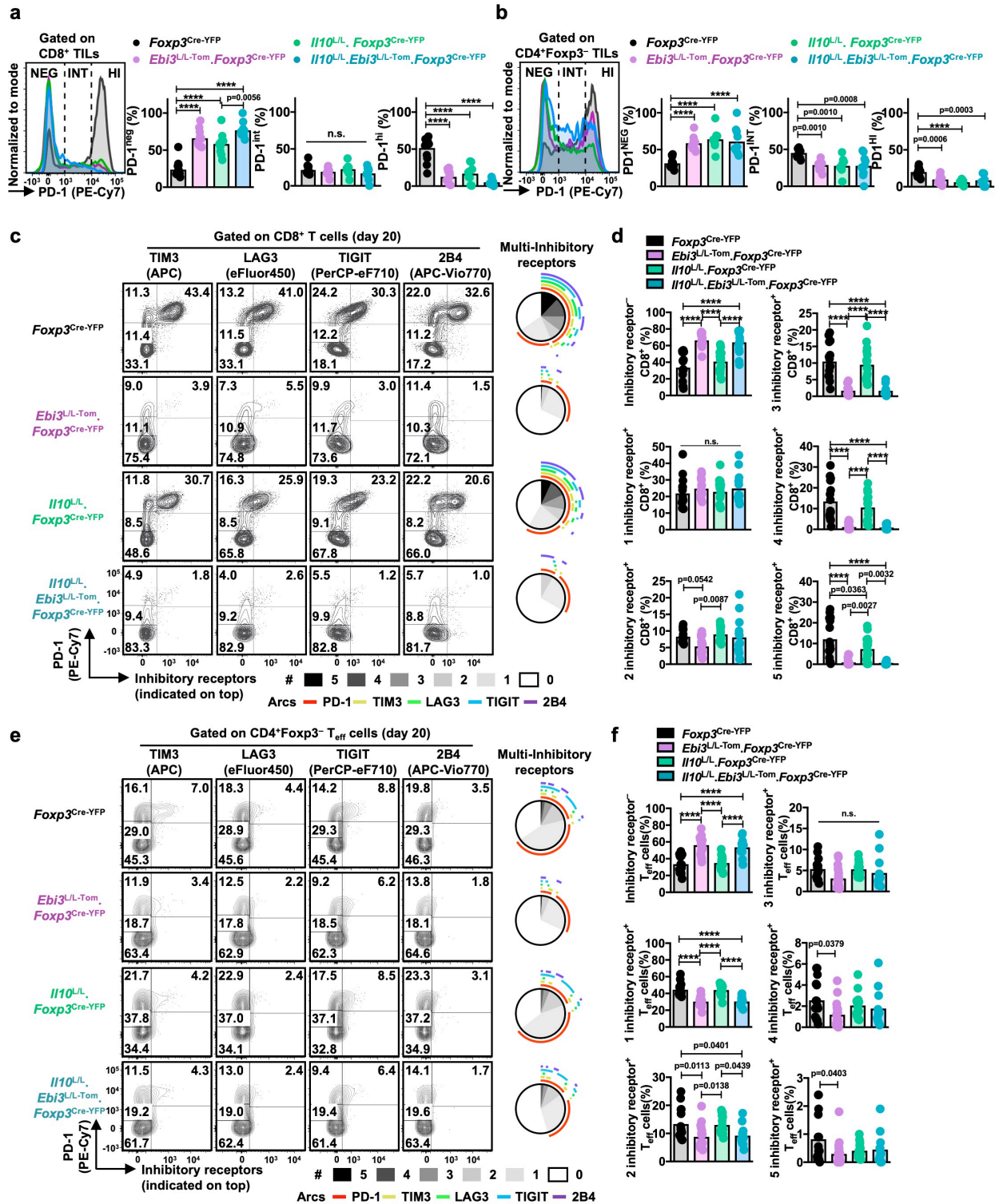
c, Heat map of differentially expressed co-signaling molecules contrasting naive LN T_{eff} cells and T_{reg} cell subpopulations. Only genes that showed significant overall changes are listed.

d, Representative flow plots depicting the expression of CD226 and CD27. Scatter-bar graphs demonstrating T_{reg} cells differentially distributed based on the reciprocal expression of CD226 and CD27.

e, Representative flow plots depicting the expression of TIM3 and LAG3. Scatter-bar graphs demonstrating T_{reg} cells differentially distributed between TIM3 single positive and TIM3⁺LAG3⁺ expression pattern.

(d-e) Data averaged from 2 independent experiments with n=7 mice. Statistical significance was determined by Two-way ANOVA with Holm-Sidak multiple comparisons (**** $p < 0.0001$ and other P values indicated).

f, Microsuppression assay comparing the suppressive function of T_{reg} cell subpopulations (*top*: ND LN Treg cells, *bottom*: TIL T_{reg} cells). Representative half-offset histograms show the proliferation of CD4⁺Foxp3⁻ Teff responder (T_{responder}) cells assessed by the decay of Cell Trace Violet proliferation dye at 1:4 T_{reg}:T_{responder} ratio. Line graphs depicting the summary of 3 independent experiments. Two-way ANOVA with Holm-Sidak multiple comparisons (P values indicated in the graphs).



Supplementary Figure 4

Modulation of inhibitory receptor expression in CD4⁺ and CD8⁺ TILs by T_{reg} cell-derived IL-10 and IL-35

a, Representative histograms depicting PD-1 expression on B16 tumor-infiltrating CD8⁺ TILs from *Foxp3*^{Cre-YFP} (n=13), *Il10*^{L/L}.*Foxp3*^{Cre-}

YFP (n=9), $Ebi3^{L/L-Tom}.Foxp3^{Cre-YFP}$ (n=11), and $Il10^{L/L}.Ebi3^{L/L-Tom}.Foxp3^{Cre-YFP}$ (n=10) mice at day 14 post tumor inoculation. Scatter-bar graphs depicting percent distribution of PD-1^{neg}, PD-1^{int}, and PD-1^{hi} CD8⁺ TILs.

b, Representative histograms depicting the PD-1 expression on CD4⁺Foxp3⁻ T cells isolated from B16 tumor-bearing mice as in **(a)**. Scatter-bar graphs depicting percent distribution of PD-1^{neg}, PD-1^{int}, and PD-1^{hi} CD4⁺Foxp3⁻ T_{eff} cells.

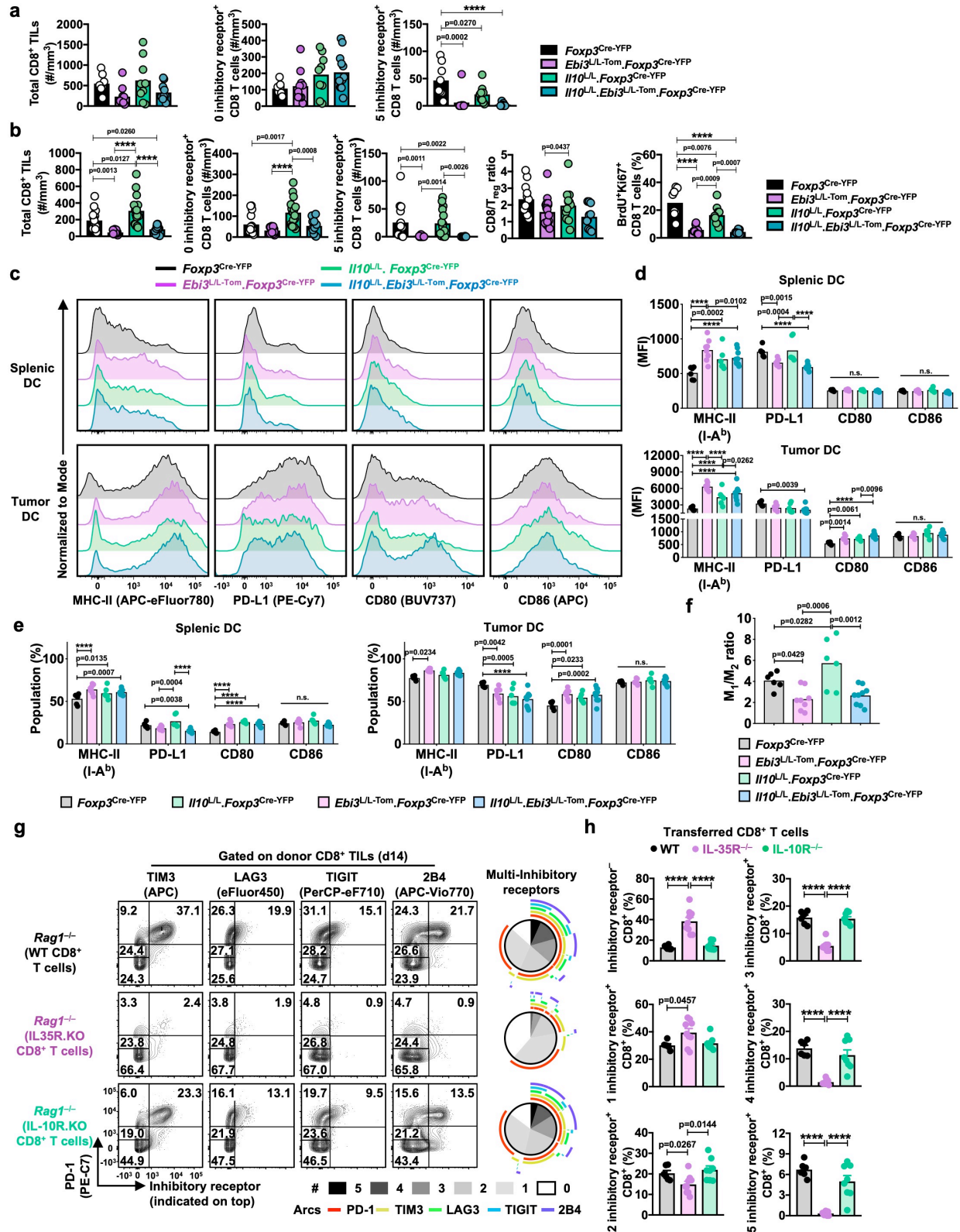
(a-b) Data averaged from 3 independent experiments. Statistical significance was determined by One-way ANOVA with Holm-Sidak multiple comparison (****p<0.0001 and other *P* values as indicated).

c, Representative flow plots depicting the expression of inhibitory receptors (PD-1, TIM3, LAG3, TIGIT, and 2B4) on CD8⁺ T cells from day 20 B16 tumor-bearing $Foxp3^{Cre-YFP}$ (n=15), $Il10^{L/L}.Foxp3^{Cre-YFP}$ (n=20), $Ebi3^{L/L-Tom}.Foxp3^{Cre-YFP}$ (n=17), and $Il10^{L/L}.Ebi3^{L/L-Tom}.Foxp3^{Cre-YFP}$ (n=13) mice. SPICE plots depicting co-expression pattern of multiple inhibitory receptors.

d, Scatter-bar graphs representing the percent distribution of inhibitory receptor-negative (0 & 1 inhibitory receptor-expressing effector-like) and multi-inhibitory receptor⁺ (3-5 inhibitory receptor-expressing exhausted) CD8⁺ T cells as in **(c)**.

e, Representative flow plots depicting expression of inhibitory receptors (PD-1, TIM3, LAG3, TIGIT, and 2B4) on CD4⁺Foxp3⁻ T_{eff} cells from day 20 B16 tumor-bearing mice as in **(c)**. SPICE plots depicting co-expression pattern of multiple inhibitory receptors.

f, Scatter-bar graphs representing the percent distribution of inhibitory receptor-negative (0 & 1 inhibitory receptor-expressing effector-like) and multi-inhibitory receptor⁺ (3—5 inhibitory receptor-expressing exhausted) CD4⁺Foxp3⁻ TILs as in **(e)**. Statistical significance was determined by One-way ANOVA with Holm-Sidak multiple comparison (****p<0.0001 and other *P* values as indicated).



Supplementary Figure 5

Differential regulation of the tumor microenvironment by T_{reg} cell-derived IL-10 and IL-35

a, Scatter-bar graphs depicting the absolute number of CD8⁺ TILs normalized to tumor volumes (either total, 0inhibitory receptor⁺, or 5inhibitory receptors⁺) isolated from day 14 B16 tumor-bearing *Foxp3*^{Cre-YFP} (n=8), *Il10*^{L/L}.*Foxp3*^{Cre-YFP} (n=9), *Ebi3*^{L/L-Tom}.*Foxp3*^{Cre-YFP}, (n=10) and *Il10*^{L/L}.*Ebi3*^{L/L-Tom}.*Foxp3*^{Cre-YFP} (n=12) mice.

b, Scatter-bar graphs depicting absolute number of CD8⁺ TILs (either total CD8⁺ TILs, 0inhibitory receptor⁺, or 5inhibitory receptors⁺) normalized to tumor volume, CD8/T_{reg} cell ratio, and Brdu⁺Ki67⁺ proliferating CD8⁺ TILs from day 20 B16 tumor-bearing *Foxp3*^{Cre-YFP} (n=15), *Il10*^{L/L}.*Foxp3*^{Cre-YFP} (n=20), *Ebi3*^{L/L-Tom}.*Foxp3*^{Cre-YFP} (n=17), and *Il10*^{L/L}.*Ebi3*^{L/L-Tom}.*Foxp3*^{Cre-YFP} (n=13) mice.

(a-b) Data averaged from 3 independent experiments. Bars represent mean values. Statistical significance was determined by One-way ANOVA with Holm-Sidak multiple comparisons (****p<0.0001 and other *P* values as indicated).

c, Representative histograms depicting expression of indicated markers on splenic and tumor-infiltrating DCs from B16 tumor-bearing *Foxp3*^{Cre-YFP} (n=6), *Il10*^{L/L}.*Foxp3*^{Cre-YFP} (n=8), *Ebi3*^{L/L-Tom}.*Foxp3*^{Cre-YFP} (n=6), and *Il10*^{L/L}.*Ebi3*^{L/L-Tom}.*Foxp3*^{Cre-YFP} (n=9) mice.

d, Scatter-bar graphs depicting the Mean Fluorescent Intensity (MFI) of the markers examined as in **(c)**.

e, Scatter-bar graphs depicting the percent distribution of indicated markers on splenic and tumor-infiltrating DCs examined as in **(c)**.

f, Scatter-bar graphs demonstrating the M₁ vs M₂ subpopulation ratio within the tumor associated macrophage (TAM) population. Statistical significance was determined by One-way ANOVA with Holm-Sidak multiple comparisons (*P* values as indicated).

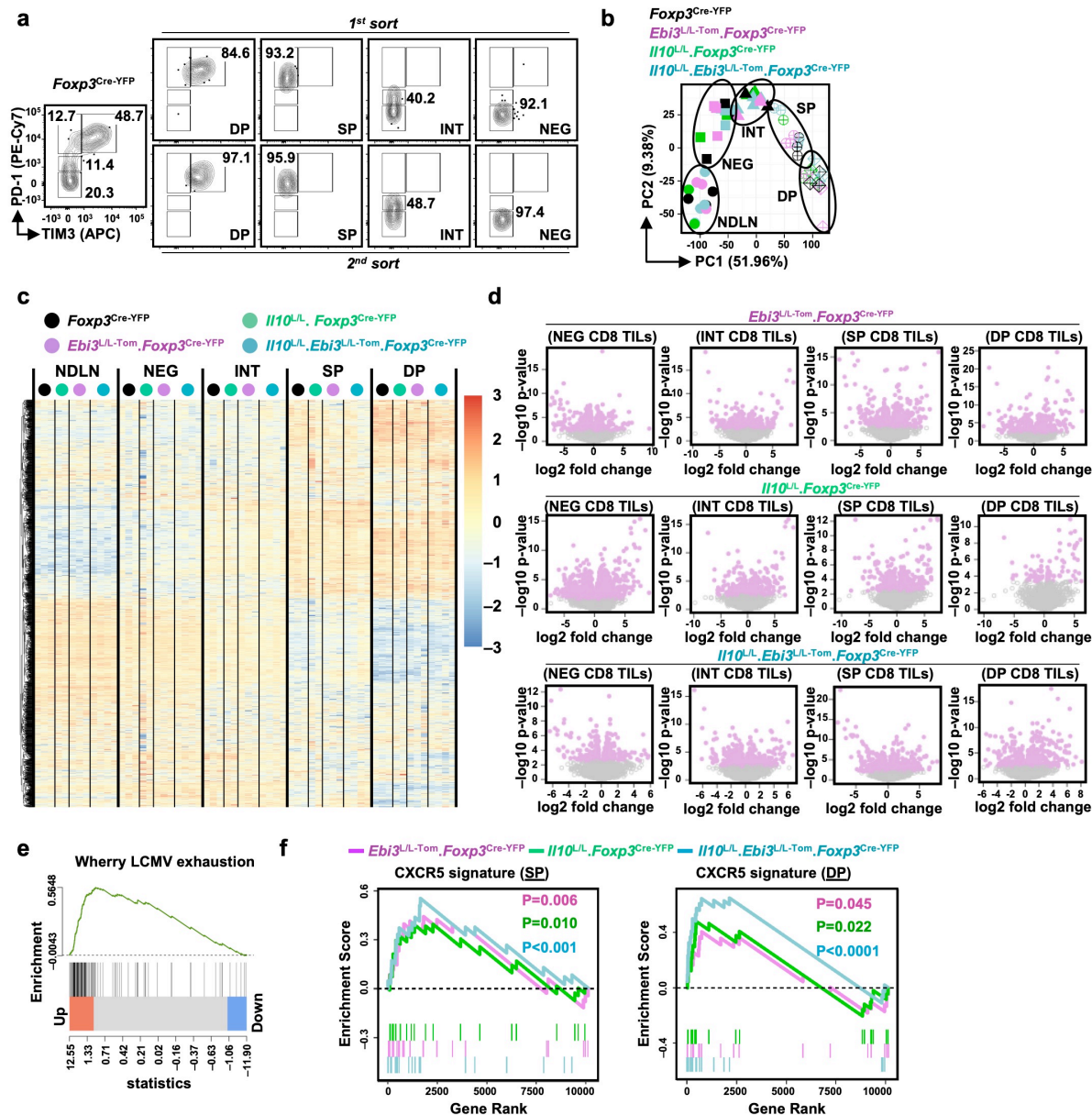
(c-f) Data averaged from 2 independent experiments. Bars represent mean values.

(c-e) Statistical significance was determined by Two-way ANOVA with Holm-Sidak multiple comparisons (****p<0.0001 and other *p*-values as indicated).

g, Representative flow plots depicting the expression of inhibitory receptors and SPICE plots showing the multi-inhibitory receptor co-expression pattern on the donor CD8⁺ T cells infiltrating B16 tumor-bearing reconstituted *Rag1*^{-/-} mice 14 days post-tumor inoculation as described in **Fig. 5a**.

h, Scatter-bar graphs representing the percent distribution of donor CD8⁺ TILs based on the number of inhibitory receptors expressed. (WT: n=10, IL-10R^{-/-}: n=6, IL-35R^{-/-}: n=7). Bars represent mean values. Statistical significance was determined by One-way ANOVA with Holm-Sidak multiple comparisons (****p<0.0001 and other *P* values indicated).

(g-h) Data averaged from 2 independent experiments.



Supplementary Figure 6

Transcriptional analysis of CD8⁺ TIL subsets from IL-10/IL-35 T_{reg} cell-deficient tumor microenvironments

a, Double-sorting scheme used to fractionate the CD8⁺ TIL subsets from the control *Fcpx3*^{Cre-YFP} and T_{reg} cell-cytokine deficient environments into the NEG, INT, SP and DP fractions. Total CD8⁺ T cells were sorted from the NDLN for each of the 4 genotypes. Data represent at least 5 independent RNAseq experiments.

b, PCA depicting the NDLN and CD8⁺ TILs subsets (NEG, INT, SP and DP) isolated from day 14 B16 tumor-bearing *Fcpx3*^{Cre-YFP} (n=3), *Il10*^{L/L}.*Fcpx3*^{Cre-YFP} (n=2), *Ebi3*^{L/L-Tom}.*Fcpx3*^{Cre-YFP} (n=3), and *Il10*^{L/L}.*Ebi3*^{L/L-Tom}.*Fcpx3*^{Cre-YFP} (n=4) mice. Each of symbol represent an independent RNAseq run replicate.

c, Heat map of top 5,000 differentially expressed genes representing CD8⁺ TILs from NDLN and NEG, INT, SP, and DP subsets. Genes were considered differentially expressed if their q-value FDR was below 0.05.

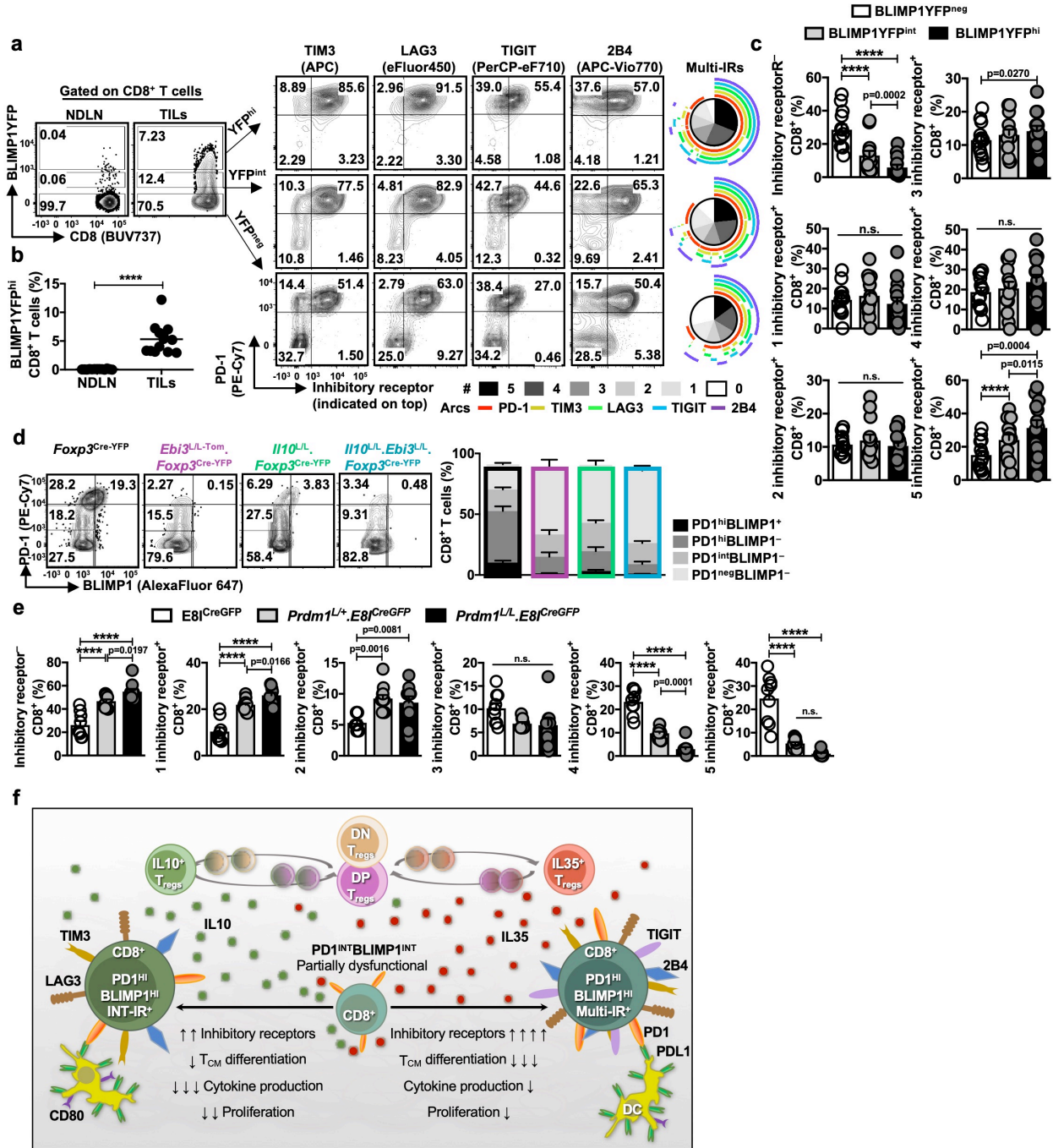
d, Volcano plots contrasting CD8⁺ TIL subpopulations (NEG = PD-1^{neg}, INT = PD-1^{int}, SP = PD-1^{hi}TIM3⁻, DP = PD-1^{hi}TIM3⁺) isolated from *Il10*^{L/L}.*Foxp3*^{Cre-YFP}, *Ebi3*^{L/L-Tom}.*Foxp3*^{Cre-YFP}, and *Il10*^{L/L}.*Ebi3*^{L/L-Tom}.*Foxp3*^{Cre-YFP} mice against the corresponding populations from *Foxp3*^{Cre-YFP} control mice. Colored dots represent genes with q-value false discovery rate of <0.05.

e, GSEA plot depicting the comparison of the exhaustion program in chronic LCMV infection versus the tumor exhaustion signature (derived by PD-1^{hi} to PD-1^{neg} CD8⁺ T cell comparison), with *P* values as indicated.

f, GSEA plot depicting the gene signature of PD-1 checkpoint blockade responsive TCF1⁺CXCR5⁺ CD8⁺ T cell subset in SP CD8⁺ TILs (*left*) and DP CD8⁺ TILs (*right*) from cytokine deficient-T_{reg} cell environments (*Il10*^{L/L}.*Foxp3*^{Cre-YFP}, *Ebi3*^{L/L-Tom}.*Foxp3*^{Cre-YFP}, and *Il10*^{L/L}.*Ebi3*^{L/L-Tom}.*Foxp3*^{Cre-YFP}); *P* values as indicated.

(e-f) Statistical significance was determined by the rankSumTestWithCorrelation test implemented in the limma package with correction for loss in degrees of freedom due to correlation among genes. All *P* values are two sided. No correction for multiple hypothesis was performed.

(a-f) Data are averaged from at least 5 independent RNASeq experiments with genotype-specific CD8⁺ T cell replicates as indicated - *Foxp3*^{Cre-YFP} (n=3), *Il10*^{L/L}.*Foxp3*^{Cre-YFP} (n=2), *Ebi3*^{L/L-Tom}.*Foxp3*^{Cre-YFP} (n=3), and *Il10*^{L/L}.*Ebi3*^{L/L-Tom}.*Foxp3*^{Cre-YFP} CD8s (n=4).



Supplementary Figure 7

IL-10⁺ and IL-35⁺ T_{reg} cells drive BLIMP1 induction on TILs *in vivo*

a, Representative flow plots depicting the expression of BLIMP1YFP expression levels in NDLN and TIL CD8⁺ T cells from B16 tumor-bearing *Prdm1*^{YFP} transgenic reporter mice (n=13). CD8⁺ TILs representative flow and SPICE plots demonstrating the expression of inhibitory receptors on the BLIMP1YFP^{hi}, YFP^{int} and YFP^{neg} CD8⁺ T cell fractions.

b, Scatter plots showing the percent distribution of YFP^{hi} subpopulation among CD8⁺ T cells as in (a). Statistical significance was

determined by paired two-tailed Student's t-test (**** $p < 0.0001$).

c, Scatter-bar graphs depicting the percent distribution of CD8⁺ TILs based on the number of inhibitory receptors expressed as in **(a)**. Bars represent mean values. Statistical significance was determined by One-way ANOVA with Holm-Sidak multiple comparison (**** $p < 0.0001$ and other P values as indicated).

(a-c) Data averaged from 3 independent experiments, with $n=13$ mice/group.

d, Representative flow plots depicting the protein level expression pattern of PD-1 and BLIMP1 on CD8⁺ T cells from B16 tumor-bearing *Foxp3*^{Cre-YFP} ($n=14$), *Il10*^{L/L}.*Foxp3*^{Cre-YFP} ($n=11$), *Ebi3*^{L/L-Tom}.*Foxp3*^{Cre-YFP} ($n=12$), and *Il10*^{L/L}.*Ebi3*^{L/L-Tom}.*Foxp3*^{Cre-YFP} ($n=14$) mice, with stacked bar graph depicting percent distribution of PD-1^{hi}, PD-1^{int}, and PD-1^{neg} CD8⁺ T cells co-expressing PD-1 and BLIMP1. Data averaged from 3 independent experiments.

e, Scatter-bar graphs representing the percent distribution of CD8⁺ T cells expressing 0-5 inhibitory receptors from B16 tumor-bearing *E8l*^{Cre-GFP} ($n=9$), *Prdm1*^{L/+}.*E8l*^{Cre-GFP} ($n=12$), and *Prdm1*^{L/L}.*E8l*^{Cre-GFP} ($n=10$) mice as in **Fig. 7d**. Data averaged from 3 independent experiments. Bars represent mean values. Statistical significance was determined by One-way ANOVA with Holm-Sidak multiple comparison (**** $p < 0.0001$ and other P values as indicated).

g, Graphical summary of the regulatory functions of T_{reg}-derived IL-10 and IL35 depicted by their common and non-redundant effects on TILs. T_{reg}-derived IL35 predominantly regulated IR-expression and T_{CM} differentiation while T_{reg}-derived IL-10 played a greater role in limiting cytokine production and proliferation of CD8⁺ TILs.