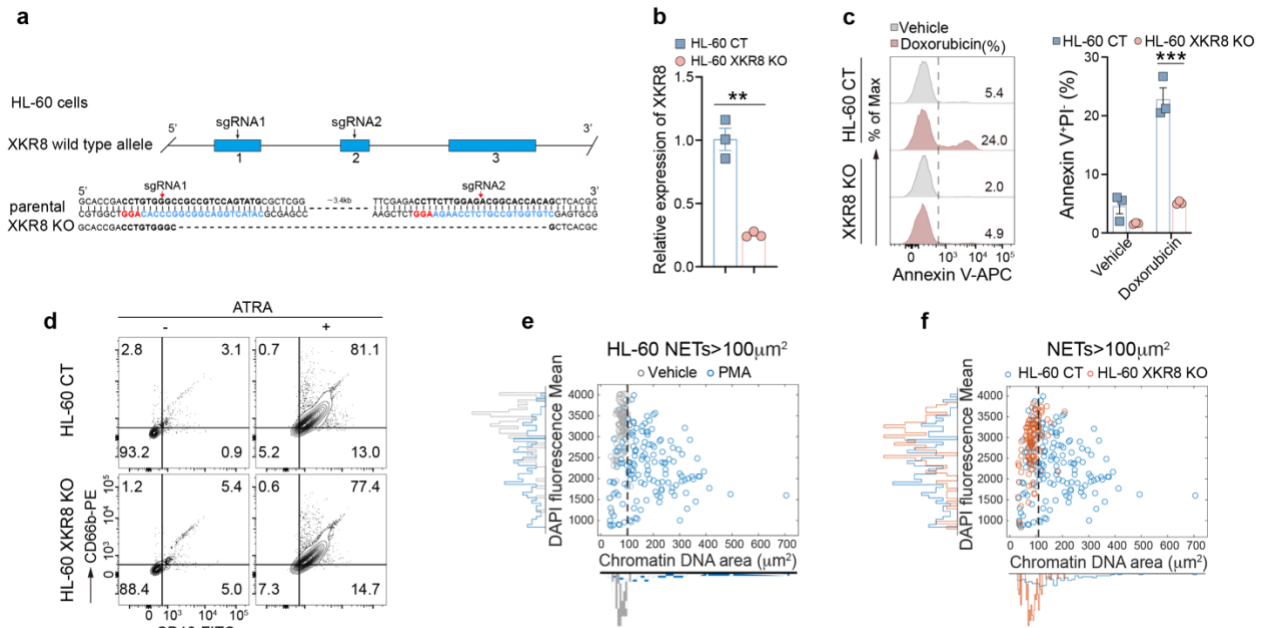
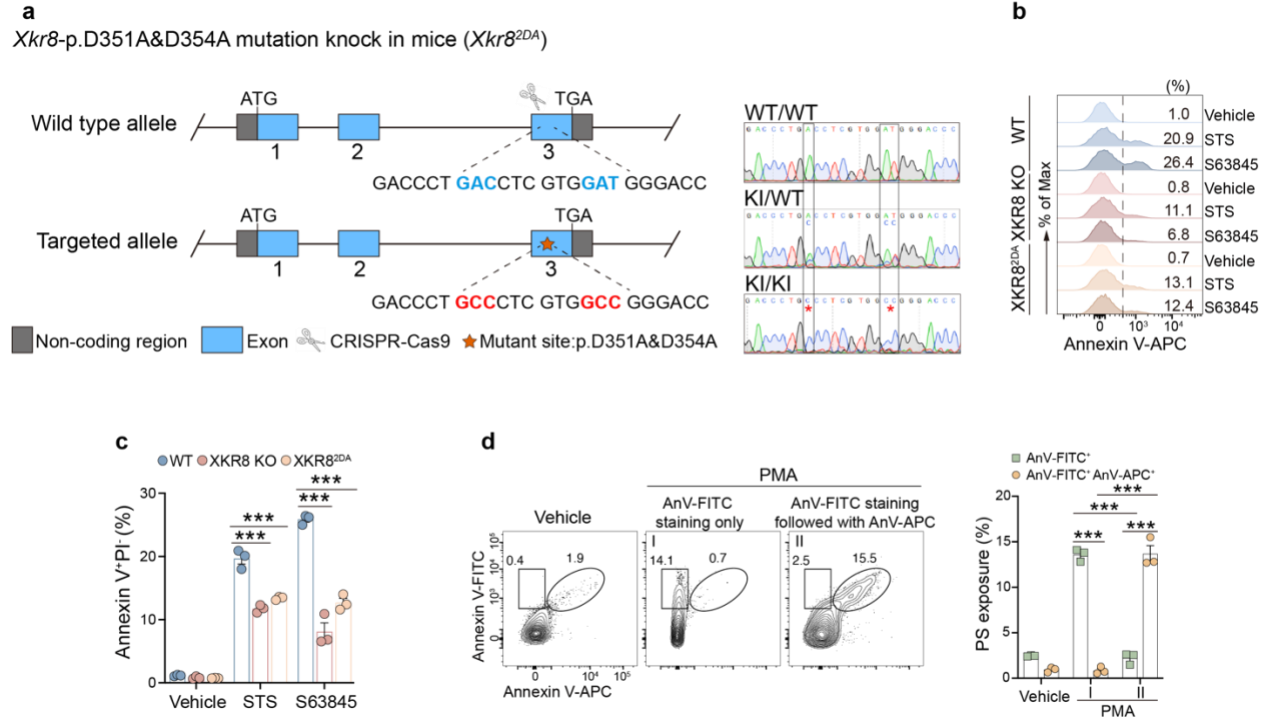


Lipid asymmetry disruption by XKR8 orchestrates neutrophil extracellular trap formation and inhibits fungal infection

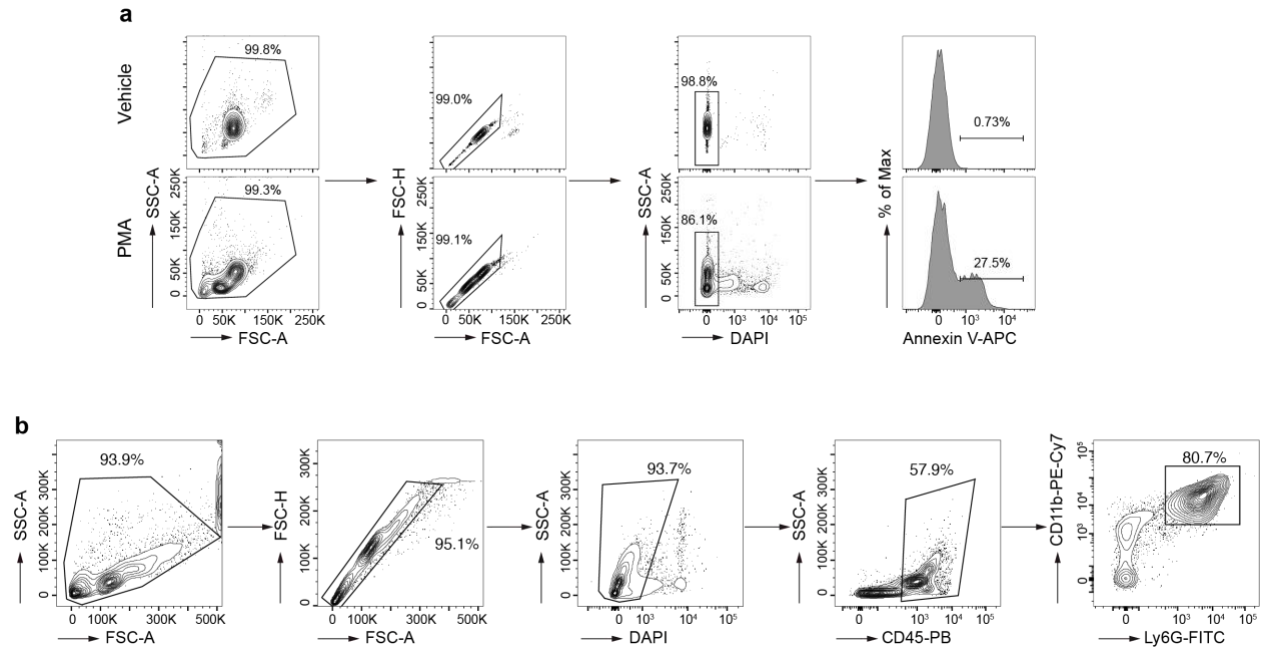
In the format provided by the
authors and unedited



Supplementary Fig. 1 Compromised NETs formation in ATRA-differentiated neutrophil-like HL-60 cells. **a,b**, Strategy for the generation of XKR8 KO HL-60 cells (**a**) and Quantitative real-time PCR (qPCR) analysis of *Xkr8* expression in WT and XKR8 KO cells (**b**), *18S* rRNA as the internal reference. **c**, PS exposure in HL-60 WT and XKR8 KO cells treated with doxorubicin (10 μM). Representative histogram (left) and quantification (right) of PS exposure in living cells (Annexin V⁺ PI⁻) are shown ($n = 3$). **d**, Flow cytometry analysis of CD16 and CD66b in HL-60 cells with or without ATRA treatment, confirming the neutrophil-like differentiation. **e,f**, Definition of NETs by DAPI fluorescence intensity and nuclear area. Cells with a nuclear area greater than 100 μm^2 were classified as NETs in dHL-60 cells (**e**). NETs formation in HL-60 WT and XKR8 KO cells treated with PMA (100 nM) (**f**). Each dot represents a biological replicate (**b,c**). Data are represented as mean $\pm$ SEM from one representative experiment of three (**b,c**) independent experiments. Statistical analysis was performed using two-way ANOVA (**c**) or two-tailed Mann-Whitney test (**b**). ns, not significant, ** $P < 0.01$, *** $P < 0.001$.



Supplementary Fig. 2 Generation and characterization of XKR8^{2DA} knock-in mice. **a**, Strategy for the generation of *Xkr8*^{2DA} knock-in mice carrying D351A and D354A mutations. The genetic modification was confirmed by genotyping, and sequencing, which verified the presence of both mutations in the XKR8^{2DA} allele. **b,c**, PS exposure in live BMNs from WT, XKR8 KO and XKR8 2DA mutant mice treated with S63845 (10 μ M) or STS (10 μ M) ($n = 3$), assessed by flow cytometry. Representative histograms (**b**) and quantification of Annexin V⁺ PI⁻ cells (**c**). **d**, Sustained PS externalization revealed by sequential Annexin V staining. Neutrophils were stained with Annexin V-FITC at 2 hours post-PMA stimulation, washed with 1 x binding buffer, and rested for 15 minutes at 37°C. Cells were then subjected to a second round of Annexin V-APC staining. Representative flow cytometry plot (left) and quantification of Annexin V⁺ DAPI⁻ cells (right). Each dot represents a biological replicate (**c,d**). Data are represented as mean $\pm$ SEM from one representative experiment of three (**c,d**) independent experiments. Statistical analysis was performed using two-way ANOVA (**c,d**). *** $P < 0.001$.



Supplementary Fig. 3 Gating strategies of flow cytometry. a, Gating strategies of PS exposure in purified neutrophils. **b**, Gating strategies of neutrophils in lung and BALF upon LPS or *C. alb* intranasal challenge.

Supplementary Table 1. Immune repertoire in WT and XKR8 KO mice.

Organs	Immune cells		%	Numbers ($\times 10^6$)
Thymus	CD4	WT	10.01 $\pm$ 0.95	14.06 $\pm$ 3.52
		XKR8 KO	10.76 $\pm$ 0.34	11.79 $\pm$ 5.35
	CD8	WT	3.39 $\pm$ 0.43	4.77 $\pm$ 1.32
		XKR8 KO	4.37 $\pm$ 0.18	4.75 $\pm$ 2.01
	Double positive cells	WT	83.03 $\pm$ 1.68	116.76 $\pm$ 25.85
		XKR8 KO	81.12 $\pm$ 0.32	88.27 $\pm$ 38.19
	Double negative cells	WT	3.59 $\pm$ 0.30	5.03 $\pm$ 1.18
		XKR8 KO	3.74 $\pm$ 0.38	3.97 $\pm$ 1.34
Spleen	CD4	WT	20.74 $\pm$ 2.03	13.69 $\pm$ 2.74
		XKR8 KO	20.00 $\pm$ 1.63	12.55 $\pm$ 2.27
	CD8	WT	11.48 $\pm$ 0.98	11.26 $\pm$ 1.14
		XKR8 KO	11.08 $\pm$ 0.29	11.71 $\pm$ 3.22
	B cells	WT	58.92 $\pm$ 2.61	61.03 $\pm$ 6.63
		XKR8 KO	60.00 $\pm$ 2.13	60.42 $\pm$ 17.64
	NK cells	WT	1.79 $\pm$ 0.64	2.50 $\pm$ 0.24
		XKR8 KO	2.47 $\pm$ 0.23	1.92 $\pm$ 1.05
	Neutrophils	WT	0.53 $\pm$ 0.10	0.30 $\pm$ 0.09
		XKR8 KO	0.30 $\pm$ 0.10	0.54 $\pm$ 0.18
	Monocytes	WT	1.20 $\pm$ 0.25	0.46 $\pm$ 0.05
		XKR8 KO	0.68 $\pm$ 0.11	0.74 $\pm$ 0.23
	Macrophages	WT	0.86 $\pm$ 0.06	0.40 $\pm$ 0.05
		XKR8 KO	0.59 $\pm$ 0.03	0.52 $\pm$ 0.09
Peripheral blood	Neutrophils	WT	7.95 $\pm$ 3.13	0.39 $\pm$ 0.22/mL
		XKR8 KO	8.42 $\pm$ 2.68	0.34 $\pm$ 0.28/mL

Organs	Immune cells		%	Numbers ($\times 10^6$) *
Bone marrow	CD4	WT	1.51 $\pm$ 0.28	0.61 $\pm$ 0.10
		XKR8 KO	1.38 $\pm$ 0.22	0.58 $\pm$ 0.14
	CD8	WT	0.64 $\pm$ 0.26	0.25 $\pm$ 0.09
		XKR8 KO	0.67 $\pm$ 0.25	0.29 $\pm$ 0.12
	B cells	WT	34.20 $\pm$ 5.05	14.11 $\pm$ 3.12
		XKR8 KO	35.94 $\pm$ 2.82	14.98 $\pm$ 1.79
	NK cells	WT	0.82 $\pm$ 0.21	0.33 $\pm$ 0.07
		XKR8 KO	0.65 $\pm$ 0.10	0.27 $\pm$ 0.05
	Neutrophils	WT	33.50 $\pm$ 4.35	13.74 $\pm$ 2.28
		XKR8 KO	34.38 $\pm$ 2.21	14.33 $\pm$ 1.60
	Monocytes	WT	8.06 $\pm$ 1.12	3.30 $\pm$ 0.49
		XKR8 KO	7.45 $\pm$ 0.96	3.08 $\pm$ 0.30
	Macrophages	WT	11.48 $\pm$ 1.60	4.68 $\pm$ 0.59
		XKR8 KO	10.63 $\pm$ 0.58	4.43 $\pm$ 0.47

	Spleen weight (mg)	Thymus weight (mg)
WT	69.92 $\pm$ 5.64	65.10 $\pm$ 11.29
XKR8 KO	65.26 $\pm$ 10.56	58.60 $\pm$ 14.64

* Counts of immune cell subsets in the femur and tibia of the mouse's two hind legs.

Data are presented as mean $\pm$ SD.

Supplementary Table 2. gRNA, crRNA, and PCR sequences used in this study

Name	Sequence (5' to 3')	Note
<i>XKR8</i> -crRNA-1	CATACTGGACGGCGGCCAC	
<i>XKR8</i> -crRNA-2	CTGTGGTGCCGTCTCCAAGA	
<i>XKR8</i> -sgRNA-1	GCTCGGCGGCCGCTACCTGT	
<i>XKR8</i> -sgRNA-2	GTGAGTGCTTCGCCCCGGA	
<i>TRPC3</i> -crRNA-1	GTCAACTGCGTGGACTACAT	
<i>TRPC3</i> -crRNA-2	TCGAGGATCAATGCCTACAA	
<i>TRPV2</i> -crRNA-1	AGTCAACCTCAACTACCGAA	
<i>TRPV2</i> -crRNA-2	CTCTTCAATGCGGTCTCCCG	
<i>TRPV4</i> -crRNA-1	TCGATAGTAGATGTCACGGA	
<i>TRPV4</i> -crRNA-2	CACCCTATATGAGTCCTCGG	
<i>PIEZO1</i> -crRNA-1	CAACGCTGGCCCCTACACGG	
<i>PIEZO1</i> -crRNA-2	AGGACATCCCCAACGCCATC	
<i>Xkr8</i> -1-F	GAATGCATCAAGGGCTGTCC	RT-PCR primers for WT and XKR8 KO mice
<i>Xkr8</i> -1-R	ACTGGTAGTATTCCGCCTGG	
<i>Xkr8</i> -2-F	GGCCGTTGTCCAGTACGTG	RT-PCR primers for <i>Xkr8^{fl/fl}</i> and <i>Xkr8^{fl/fl} Ly6G^{Cre}</i> mice
<i>Xkr8</i> -2-R	GCAAACACCTATACAGGTAGCC	
<i>18s</i> -F	CTGCCGTCTGAGTGATCGC	<i>18s</i> as the internal reference for murine cells
<i>18s</i> -R	GCTGGGGCTGAGGAAAGTG	
<i>XKR8</i> -F	CCTGGGTGACTCATAGCTCCT	RT-PCR primers for HL-60 WT and XKR8 KO cell lines
<i>XKR8</i> -R	GCCAGTGGTAGTACACAAGCC	
<i>18S</i> -F	GTAACCCGTTGAACCCATT	<i>18S</i> as the internal reference for HL-60 cell lines
<i>18S</i> -R	CCATCCAATCGGTAGTAGCG	