

Supplementary Information for:

Hydrogen cyanide produced by the soil bacterium *Chromobacterium* sp. Panama contributes to mortality in *Anopheles gambiae* mosquito larvae

Sarah M. Short, Sarah van Tol, Hannah J. MacLeod, George Dimopoulos

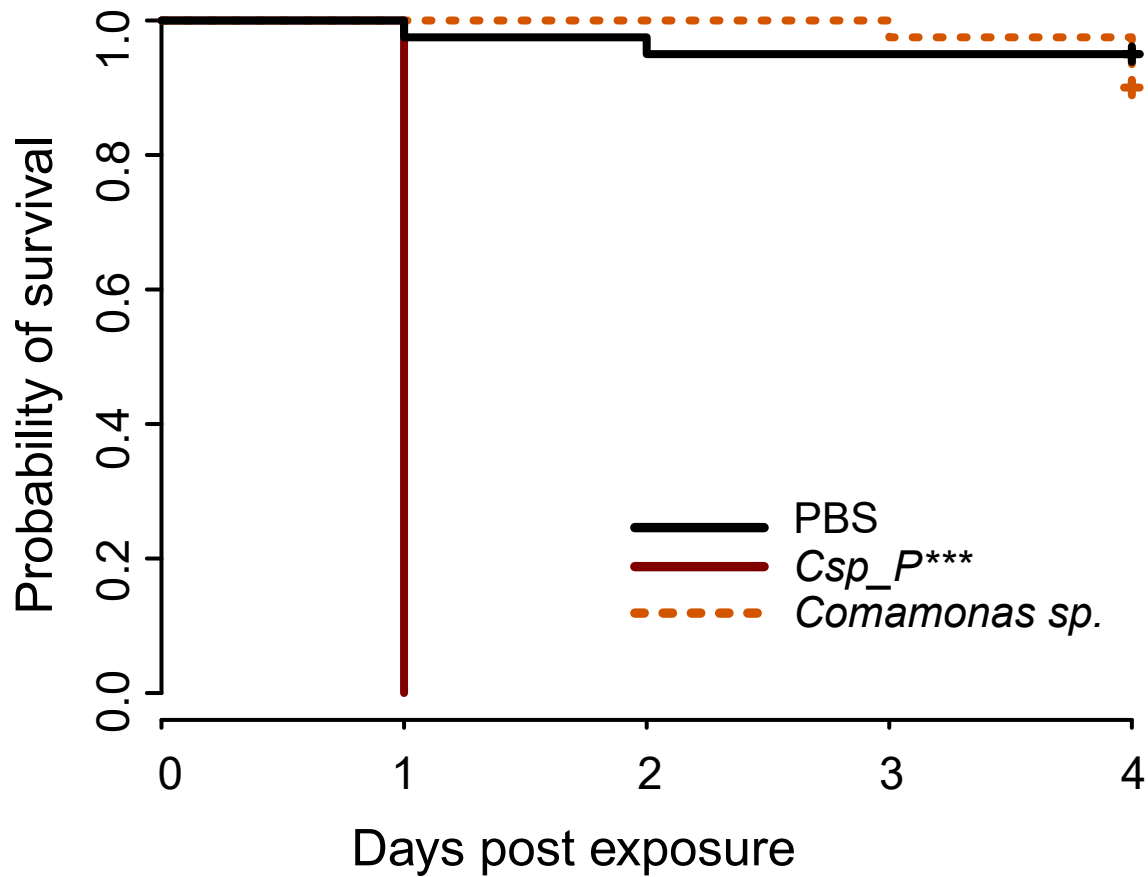


Figure S1: Exposure to *Comamonas sp.*, a common mosquito midgut bacterium does not induce larval mortality. First and second instar *An. gambiae* larvae were treated in groups of ten in 5mL of water containing 10mg of ground larval food. *C.sp_P* or *Comamonas sp.* overnight cultures were diluted to $1.0 (\pm 0.1)$ OD₆₀₀ and 100μL of each culture was added to the larval breeding water. 100μL of 1X PBS was added as a control and survival of larvae was monitored for four days. Compared to the 1X PBS control, *C.sp_P* treated larvae had significantly lower survival ($p = 2.14 \times 10^{-7}$) while *Comamonas sp.* treated larvae did not ($p = 0.435$). The experiment was repeated two independent times and for each experimental replicate two pools of ten larvae were measured per treatment. Data were analyzed using a Cox proportional hazards model. Total sample sizes are as follows: PBS = 40, *C.sp_P* = 40, *Comamonas sp.* = 40.

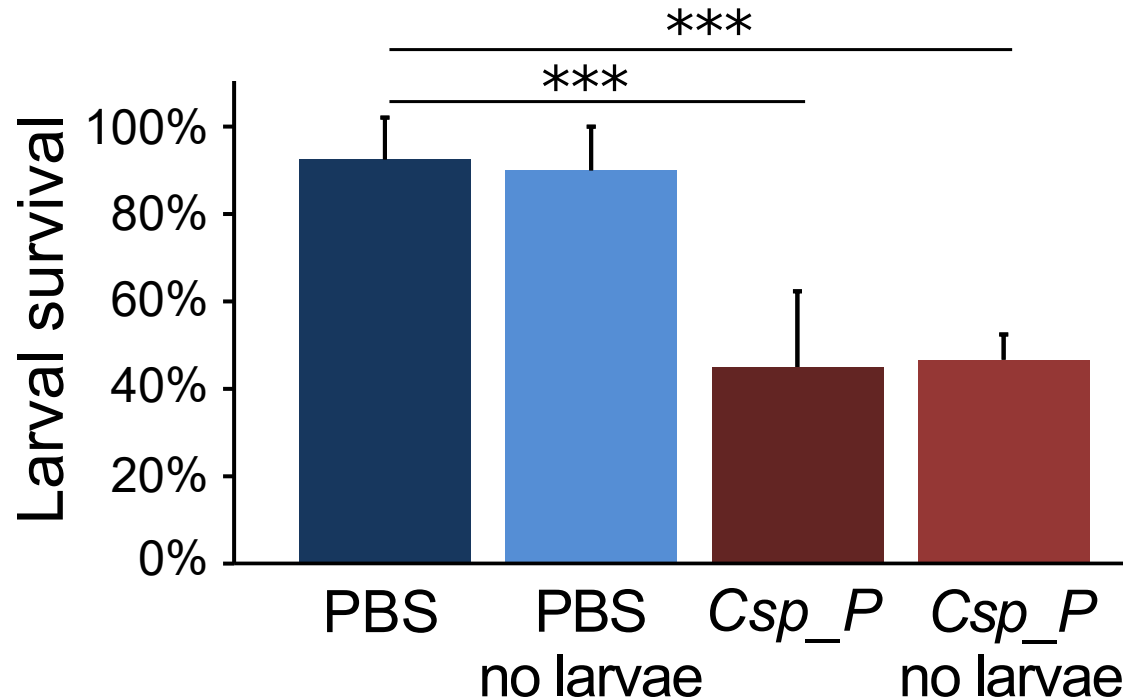


Figure S3: Larvicidal activity not dependent on the presence of larvae. Larval breeding water was treated with either 1X PBS or *C.sp_P*, and incubated for 6-8 hours until all larvae in the *C.sp_P* treatment were dead. In half the treatments, no larvae were added to the larval breeding water. Larval water was then collected and filtered using a 0.2µm filter to remove all live bacterial cells. This filtered water was then used to treat new pools of five *An. gambiae* larvae and survival was recorded after 16-17 hours exposure to the filtered water. The “PBS” and “*C.sp_P*” data presented here are the same as those presented in Figure 3. Larvae exposed to *C.sp_P* treated water suffered significant mortality regardless of whether larvae were co-incubated with *C.sp_P* bacteria before filtration (with larvae, $p = 0.0011$, without larvae, $p = 0.00097$). The graph shows average percent survival across replicate experiments and error bars represent one standard deviation. For each experiment two pools of five larvae were measured per treatment and the experiment was repeated three independent times. For all treatments, total sample size was $n = 30$.

Table S1: Model outputs for Statistical Analyses**Figure 1A:**

```
coxph(formula = Surv(day, status, type = c("right")) ~ treatment + rep + pool)
```

n= 355, number of events= 97

	coef	exp(coef)	se(coef)	z	Pr(> z)
CspP 10 ⁵	0.69003	1.99378	0.41689	1.655	0.0979 .
CspP 10 ³	-0.45264	0.63595	0.52716	-0.859	0.3905 .
CspP 10 ¹	-0.65412	0.51990	0.55782	-1.173	0.2409 .
CspP 0.1	-1.58830	0.20427	0.78183	-2.032	0.0422 *
CspP 10 ¹⁷	3.95136	52.00587	0.42587	9.278	<2e-16 ***
rep2	0.47306	1.60490	0.26506	1.785	0.0743 .
rep3	0.26788	1.30719	0.27574	0.971	0.3313 .
pool2	0.05954	1.06135	0.20336	0.293	0.7697 .

Figure 1B:

```
coxph(formula = Surv(day, status, type = c("right")) ~ treatment + rep + pool)
```

n= 240, number of events= 102

	coef	exp(coef)	se(coef)	z	Pr(> z)
CspP 10 ⁵	-0.33149	0.71785	0.39705	-0.835	0.404 .
CspP 10 ³	0.36408	1.43919	0.34165	1.066	0.287 .
CspP 10 ¹	0.15112	1.16314	0.35436	0.426	0.670 .
CspP 0.1	-0.02647	0.97388	0.37166	-0.071	0.943 .
CspP 10 ¹⁷	1.35025	3.85838	0.32948	4.098	4.17e-05 ***
rep2	-0.03130	0.96918	0.20013	-0.156	0.876 .
pool2	-0.02145	0.97878	0.20064	-0.107	0.915 .

Figure 2:

```
coxph(formula = Surv(day, status, type = c("right")) ~ treatment + rep)
```

n= 260, number of events= 141

	coef	exp(coef)	se(coef)	z	Pr(> z)
C. aquaticum	1.73922	5.69291	0.31997	5.436	5.46e-08 ***
CspP	3.01887	20.46817	0.31796	9.494	< 2e-16 ***
C. subtsugae	-0.12424	0.88317	0.64027	-0.194	0.846 .
C. vaccinii	2.32464	10.22296	0.31987	7.268	3.66e-13 ***
C. violaceum	1.36668	3.92232	0.33550	4.074	4.63e-05 ***
rep2	0.04868	1.04988	0.23295	0.209	0.834 .
rep3	0.09991	1.10507	0.23457	0.426	0.670 .
rep4	-0.45661	0.63343	0.25110	-1.818	0.069 .

Figure 3:

```
glm(formula = status ~ rep + treat/pool, family = binomial(link = logit))
```

	Estimate	Std. Error	z value	Pr(> z)
(Intercept)	-2.9892	0.9428	-3.170	0.00152 **
rep2	0.8844	0.8837	1.001	0.31697 .
rep3	-0.1025	0.8416	-0.122	0.90303 .
treatCspP	3.6174	0.8678	4.169	3.06e-05 ***
treatPBS:pool2	-17.0511	2364.3872	-0.007	0.99425 .
treatCspP:pool2	2.0318	1.1475	1.771	0.07662 .

Figure 4A:

```
glm(formula = status ~ rep + treat, family = binomial(link = logit))
```

	Estimate	Std. Error	z value	Pr(> z)
(Intercept)	-2.7030	0.6940	-3.895	9.82e-05 ***
rep2	0.4258	0.5328	0.799	0.424 .
rep3	0.1479	0.5382	0.275	0.784 .
rep4	0.1479	0.5382	0.275	0.784 .
treatcsp	2.7246	0.6808	4.002	6.27e-05 ***
treatcsp10k	-16.0520	1029.4223	-0.016	0.988 .
treatcsp3-10k	-1.1525	1.1778	-0.979	0.328 .
treatcsp3k	2.7246	0.6808	4.002	6.27e-05 ***

treatpbs10k	-0.4328	0.9423	-0.459	0.646
treatpbs3-10k	-0.4328	0.9423	-0.459	0.646
treatpbs3k	-1.1736	1.1776	-0.997	0.319

Figure 4B:

Heat treatment experiment:

Room temperature:

```
glm(status~rep+treatment/well,data=rt, family=binomial(link=logit))
```

	Df	Deviance	Resid. Df	Resid. Dev	Pr(>Chi)
NULL			59	74.920	
rep	2	2.9435	57	71.976	0.2295194
treatment	1	14.8347	56	57.142	0.0001174 ***
treatment:well	2	0.9822	54	56.159	0.6119638

37 degrees:

```
glm(status~rep+treatment/pool,data=ts, family=binomial(link=logit))
```

	Df	Deviance	Resid. Df	Resid. Dev	Pr(>Chi)
NULL			59	73.304	
rep	2	1.4919	57	71.812	0.474286
treatment	1	8.5075	56	63.304	0.003537 **
treatment:pool	2	3.4841	54	59.820	0.175158

55 degrees:

```
glm(status~rep+treatment/pool,data=ff, family=binomial(link=logit))
```

	Df	Deviance	Resid. Df	Resid. Dev	Pr(>Chi)
NULL			59	73.304	
rep	2	1.3964	57	71.907	0.49747
treatment	1	24.2913	56	47.616	8.281e-07 ***
treatment:pool	2	5.1472	54	42.469	0.07626 .

70 degrees:

```
glm(status~rep+treatment/pool,data=s, family=binomial(link=logit))
```

	Df	Deviance	Resid. Df	Resid. Dev	Pr(>Chi)
NULL			59	81.503	
rep	2	0.9590	57	80.544	0.6191014
treatment	1	12.2854	56	68.259	0.0004565 ***
treatment:pool	2	2.1969	54	66.062	0.3333882

90 degrees:

```
glm(status~rep+treatment/pool,data=n, family=binomial(link=logit))
```

	Df	Deviance	Resid. Df	Resid. Dev	Pr(>Chi)
NULL			59	73.304	
rep	2	1.933	57	71.371	0.38049
treatment	1	34.443	56	36.929	4.39e-09 ***
treatment:pool	2	5.951	54	30.977	0.05101 .

Figure 4C:

Initial model:

```
glm(status~rep+waterharvest+treat*vac_cent/pool,data=og, family=binomial(link=logit))
```

Final model:

```
glm(status~rep+treat*vac_cent,data=og, family=binomial(link=logit))
```

Anova:

	Df	Deviance	Resid. Df	Resid. Dev	Pr(>Chi)
NULL			238	313.42	
rep	1	11.087	237	302.33	0.0008692 ***
treat	1	23.458	236	278.88	1.277e-06 ***
vac_cent	1	15.764	235	263.11	7.174e-05 ***
treat:vac_cent	1	27.467	234	235.65	1.598e-07 ***

Figure 6:

Original model:

```
glm(status~waterharvest+exptrep+bact*antidote/pool, family=binomial(link=logit))
```

Final model:

```
glm(status~bact*antidote, family=binomial(link=logit))
```

	Df	Deviance	Resid. Df	Resid. Dev	Pr(>Chi)	
NULL			159	154.43		
bact	1	4.1680	158	150.26	0.0411951	*
antidote	1	14.4453	157	135.81	0.0001443	***
bact:antidote	1	5.0559	156	130.76	0.0245421	*

Figure 7A:

```
survdifff(formula = Surv(day, status) ~ treatment)
```

	N	Observed	Expected	(O-E)^2/E	(O-E)^2/V
treatment=csp	262	140	159	2.38	5.78
treatment=pbs	476	377	358	1.06	5.78

Chisq= 5.8 on 1 degrees of freedom, p= 0.0162
