

Enrichment of gut-derived *Fusobacterium* is associated with suboptimal immune recovery in HIV-infected individuals

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Supplemental Methods

Gut microbiota bioinformatics analysis. The paired end raw sequencing reads were joined using fastq-join function from EA-utils¹ under the default parameters of QIIME. Then, the sequences were demultiplexed and quality filtered using the default values of the split_libraries_fastq.py script. Subsequently, demultiplexed reads were clustered in 97% using the default UCLUST algorithm² open-reference OTU picking (pick_open_reference_otus.py) workflow based on Greengenes 13_8 database and PyNAST³ for alignment. Taxonomy was assigned to sequences with the UCLUST consensus taxonomy assigner. In addition, chimera sequences were identified and removed using ChimeraSlayer algorithm within QIIME⁴. Then, phylogenetic tree was constructed using FastTree⁵ alignment algorithm for downstream analysis. In this study, reads were successfully clustered into 14,362 operational taxonomic units (OTUs) with 97% identity.

Downstream analysis including taxa summary plot, alpha diversity and beta diversity were performed using QIIME. The relative abundance of all OTUs in the samples were averaged across the HIV-infected (i.e. HIV optimal and HIV suboptimal) and uninfected samples. Alpha rarefaction curves were computed by calculating the metrics observed OTUs, Phylogenetic distance⁶ and Shannon index⁷ using alpha_rarefaction.py. The nonparametric t-test was done using compare_alpha_diversity.py script and the results were corrected by bonferroni test. Beta diversity was calculated using beta_diversity_through_plots.py. UniFrac⁸ weighted and unweighted were then performed to determine the similarity among the microbial communities and visualized using principal coordinate analysis (PCoA) plots. Subsequently, the non-parametric multivariate analysis of variance with permutation (PERMANOVA), which uses the

unweighted and weighted UniFrac metric to measure phylogenetic distance, was used to test whether between group gut microbiota composition (HIV-infected vs. non-infected & oIR vs. sIR) differed significantly from within group composition.

References

- 1 Aronesty, E. *ea-utils*. " Command-line tools for processing biological sequencing data". (2011).
- 2 Edgar, R. C. Search and clustering orders of magnitude faster than BLAST. *Bioinformatics* **26**, 2460-2461, doi:10.1093/bioinformatics/btq461 (2010).
- 3 Caporaso, J. G. *et al*. PyNAST: a flexible tool for aligning sequences to a template alignment. *Bioinformatics* **26**, 266-267, doi:10.1093/bioinformatics/btp636 (2010).
- 4 Haas, B. J. *et al*. Chimeric 16S rRNA sequence formation and detection in Sanger and 454-pyrosequenced PCR amplicons. *Genome Res.* **21**, 494-504, doi:10.1101/gr.112730.110 (2011).
- 5 Price, M. N., Dehal, P. S. & Arkin, A. P. FastTree: computing large minimum evolution trees with profiles instead of a distance matrix. *Mol. Biol. Evol.* **26**, 1641-1650, doi:10.1093/molbev/msp077 (2009).
- 6 Faith, D. P. & Baker, A. M. Phylogenetic diversity (PD) and biodiversity conservation: some bioinformatics challenges. *Evol Bioinform Online* **2**, 121-128 (2007).
- 7 Shannon, C. E. The mathematical theory of communication. 1963. *MD Comput.* **14**, 306-317 (1997).

- 8 Lozupone, C. & Knight, R. UniFrac: a new phylogenetic method for comparing microbial communities. *Appl. Environ. Microbiol.* **71**, 8228-8235, doi:10.1128/AEM.71.12.8228-8235.2005 (2005).

Figure Legends

Supplementary Fig. S1. Flow cytometry gating strategy. Panel A: Following gating to exclude doublets, CD3⁺ T-cells were sequentially gated for CD4⁺ and CD8⁺ cells. In each of these subsets, the proportion of maturational subsets were identified as follows; naïve (CD45RA⁺CCR7⁺), central memory, CM (CD45RA⁻CCR7⁺), effector memory, EM (CD45RA⁻CCR7⁻) and terminally-differentiated effector memory, TdEM (CD45RA⁺CCR7⁻). Panel B: CD3⁺ cells were sequentially gated for CD4⁺ and CD8⁺ T-cells and subsequently markers of activation (CD38⁺HLA-DR⁺) and senescence (CD57⁺CD28⁻). T-regulatory cells were reported as the proportion of CD3⁺CD4⁺ T-cells expressing CD25⁺Foxp3⁺.

Supplementary Fig. S2. The top relative abundance of **a)** phyla and **b)** genus across the 46 subjects from the uninfected, HIV optimal and HIV suboptimal groups. The HIV positive samples were arranged according to increasing of CD4 T-cell counts. Histograms are based on the proportion of OTUs per subject.

Supplementary Fig. S3. PCoA plots assessing the beta diversity of microbial communities in i) HIV-infected and uninfected using **a)** unweighted and **b)** weighted UniFrac distance and ii) HIV suboptimal and HIV optimal using **c)** unweighted and **d)** weighted UniFrac distance. PERMANOVA was performed to determine the statistical significance. $p < 0.05$ is considered significant.

Supplementary Fig. S4. Rarefaction curves assessing alpha diversity was plotted for **a)** Observed OTUs, **b)** Phylogenetic distance and **c)** Shannon index. There was no significant difference between the groups tested when using T test, $p > 0.05$ for all comparisons.

Supplementary Fig. S5. Differences in the abundance of bacterial communities between the HIV-infected and uninfected groups. **a)** The uninfected group-enriched taxa are displayed with a positive LDA score (green) while the HIV-infected-enriched taxa are shown with a negative LDA score (red). Only taxa meeting an LDA score threshold >3 are listed. **b)** Bacterial taxa that were differentially abundant in the gut microbiota profiles between the HIV-infected and uninfected subjects visualized using a cladogram generated from LEfSe analysis. **c)** Scatter plot illustrating the association between the relative abundance of *Fusobacterium* and CD4 T-cell counts among the HIV-infected subjects.

Supplementary Table S1. Bacteria taxa which were different in abundance between HIV-infected and uninfected, after Benjamini-Hochberg correction.

Taxonomic Hierarchy	Enriched in Group	Log LDA score	p-value	q-value
Bacteria p__Fusobacteria	HIV positive	4.505	0.0022	0.0154
Bacteria p__Fusobacteria c__Fusobacteriia	HIV positive	4.505	0.0022	0.0154
Bacteria p__Fusobacteria c__Fusobacteriia o__Fusobacteriales	HIV positive	4.505	0.0022	0.0154
Bacteria p__Fusobacteria c__Fusobacteriia o__Fusobacteriales f__Fusobacteriaceae	HIV positive	4.501	0.0032	0.0154
Bacteria p__Fusobacteria c__Fusobacteriia o__Fusobacteriales f__Fusobacteriaceae g__ <i>Fusobacterium</i>	HIV positive	4.500	0.0032	0.0154
Bacteria p__Firmicutes c__Clostridia o__Clostridiales f__Veillonellaceae	HIV positive	4.211	0.0157	0.0326
Bacteria p__Firmicutes c__Clostridia o__Clostridiales f__Veillonellaceae g__ <i>Megamonas</i>	HIV positive	4.063	0.0024	0.0154
Bacteria p__Proteobacteria c__Gammaproteobacteria o__Aeromonadales f__Succinivibrionaceae	HIV positive	3.952	0.0248	0.0421
Bacteria p__Proteobacteria c__Gammaproteobacteria o__Aeromonadales f__Succinivibrionaceae g__ <i>Succinivibrio</i>	HIV positive	3.941	0.0355	0.0450
Bacteria p__Proteobacteria c__Gammaproteobacteria o__Aeromonadales	HIV positive	3.940	0.0144	0.0326
Bacteria p__Tenericutes c__Mollicutes o__Mycoplasmatales	HIV positive	3.300	0.0403	0.0450
Bacteria p__Tenericutes c__Mollicutes o__Mycoplasmatales f__Mycoplasmataceae	HIV positive	3.275	0.0403	0.0450
Bacteria p__Actinobacteria c__Actinobacteria o__Actinomycetales f__Actinomycetaceae g__ <i>Mobiluncus</i>	HIV positive	3.037	0.0253	0.0421

Bacteria p__Proteobacteria c__Betaproteobacteria o__Neisseriales	Uninfected			
f__Neisseriaceae		3.688	0.0324	0.0448
Bacteria p__Proteobacteria c__Betaproteobacteria o__Neisseriales	Uninfected	3.660	0.0324	0.0448
Bacteria p__Firmicutes c__Erysipelotrichi o__Erysipelotrichales f__	Uninfected			
__Erysipelotrichaceae g__ <i>Eubacterium</i> _		3.283	0.0376	0.0450
Bacteria p__Bacteroidetes c__Bacteroidia o__Bacteroidales f__B	Uninfected			
arnesiellaceae_		3.238	0.0395	0.0450
Bacteria p__Firmicutes c__Clostridia o__Clostridiales f__Rumino	Uninfected			
coccaceae g__ <i>Oscillospira</i>		3.180	0.0261	0.0421
Bacteria p__Synergistetes c__Synergistia o__Synergistales f__Det	Uninfected			
hiosulfovibrionaceae g__ <i>Jonquetella</i>		3.145	0.0435	0.0451
Bacteria p__Firmicutes c__Clostridia o__Clostridiales f__Lachnos	Uninfected			
piraceae g__ <i>Moryella</i>		3.082	0.0435	0.0451

Supplementary Table S2. Bacteria taxa which were different in abundance between HIV optimal (oIR) and HIV Suboptimal (sIR) arms, after Benjamini-Hochberg correction.

Taxonomic Hierarchy	Enriched in Group	Log LDA score	p-value	q-value
Bacteria p__Firmicutes c__Bacilli o__Lactobacillales	HIV optimal	4.432	0.0307	0.0208
Bacteria p__Actinobacteria c__Actinobacteria o__Actinomycetales f__Corynebacteriaceae g__Corynebacterium	HIV optimal	4.340	0.0452	0.0292
Bacteria p__Actinobacteria c__Actinobacteria o__Actinomycetales f__Corynebacteriaceae	HIV optimal	4.340	0.0452	0.0313
Bacteria p__Fusobacteria c__Fusobacteriia o__Fusobacteriales f__Fusobacteriaceae	HIV suboptimal	4.695	0.0424	0.0229
Bacteria p__Fusobacteria c__Fusobacteriia o__Fusobacteriales f__Fusobacteriaceae g__Fusobacterium	HIV suboptimal	4.695	0.0424	0.0250
Bacteria p__Fusobacteria	HIV suboptimal	4.688	0.0481	0.0458
Bacteria p__Fusobacteria c__Fusobacteriia	HIV suboptimal	4.688	0.0481	0.0479
Bacteria p__Firmicutes c__Clostridia o__Clostridiales f__Tissierellaceae g__Gallicola	HIV suboptimal	3.158	0.0073	0.0042
Bacteria p__Proteobacteria c__Deltaproteobacteria o__Desulfovibrionales	HIV suboptimal	3.093	0.0479	0.0375
Bacteria p__Proteobacteria c__Deltaproteobacteria	HIV suboptimal	3.093	0.0479	0.0396
Bacteria p__Proteobacteria c__Deltaproteobacteria o__Desulfovibrionales f__Desulfovibrionaceae	HIV suboptimal	3.093	0.0479	0.0417
Bacteria p__Proteobacteria c__Deltaproteobacteria o__Desulfovibrionales f__Desulfovibrionaceae g__Bilophila	HIV suboptimal	3.075	0.0479	0.0438

Supplementary Table S3. Results of CCA showing the associations between CD4 and CD8 T-cell counts and selected bacterial taxa found most abundantly different in LEfSe analysis in optimal and suboptimal immune responders.

Total Samples (N=26)	
Canonical Variate 1	
Variables	Canonical Loadings
Immune Cells	
CD4 count	0.846
CD8 count	-0.051
Gut microbiota	
<i>g_Bilophila</i>	-0.113
<i>g_Gallicola</i>	-0.103
<i>g_Corynebacterium</i>	0.437
<i>g_Fusobacterium</i>	-0.691
<i>o_Lactobacillales</i>	0.582
Aggregate Redundancy	
Coefficients	
Immune cells Gut microbiota	0.300
Gut microbiota Immune cells	0.162

Supplementary Table S4. Results of CCA showing the associations between CD4 T-cell maturational subsets and selected bacterial taxa found significantly associated with CD4 T-cell counts in HIV-infected participants.

Total Samples (N=26)	
Canonical Variate 1	
Variables	Canonical Loadings
Immune Cells	
Naïve CD4	0.509
CD4 Activation	-0.923
CD4 Tregs	-0.639
Gut microbiota	
<i>g_Corynebacterium</i>	-0.199
<i>g_Fusobacterium</i>	-0.951
<i>o_Lactobacillales</i>	0.361
Aggregate Redundancy	
Coefficients	
Immune cells Gut	0.096
microbiota	
Gut microbiota Immune	0.092
cells	