

Supplementary Materials

Title:

The role of metalloproteases in fertilisation in the ascidian *Ciona robusta*

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Fig. S1

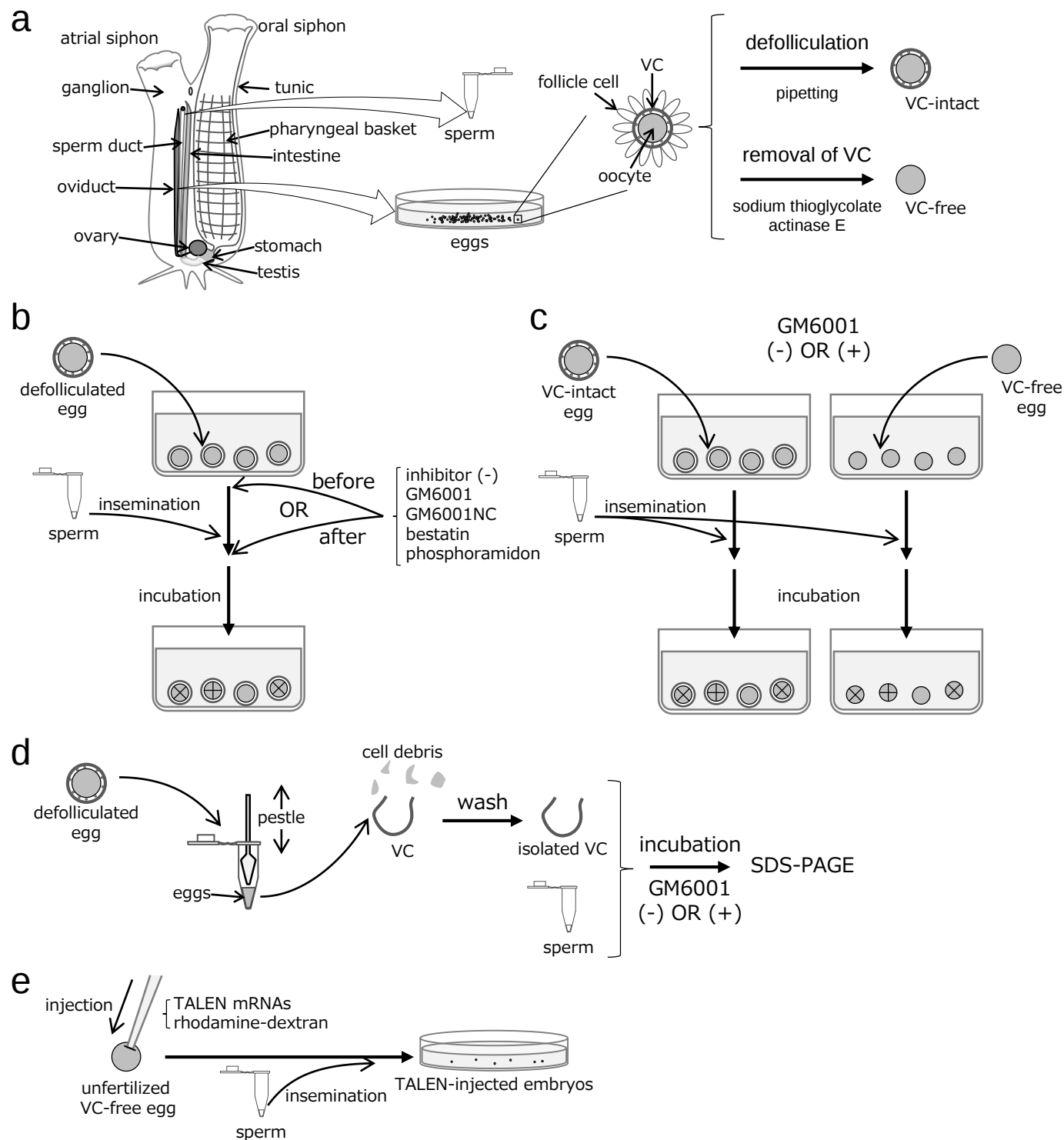


Fig. S1
Brief descriptions of experimental schemes.
a. Surgical harvest of gametes from *Ciona robusta*, defolliculation and VC-removal of the eggs.
b. Fertilization assay of VC-intact eggs in the presence or absence of inhibitors in figure 1 a-c.
c. Fertilization assay of VC-intact or -free eggs in figure 1 d-e.
d. Isolation of the VC and the digestion assay in figure 2. Eggs were mechanically crushed.
e. TALEN-mediated knockout in figure 5. Unfertilized defolliculated eggs were injected with TALEN mRNAs and then inseminated.

Fig. S2

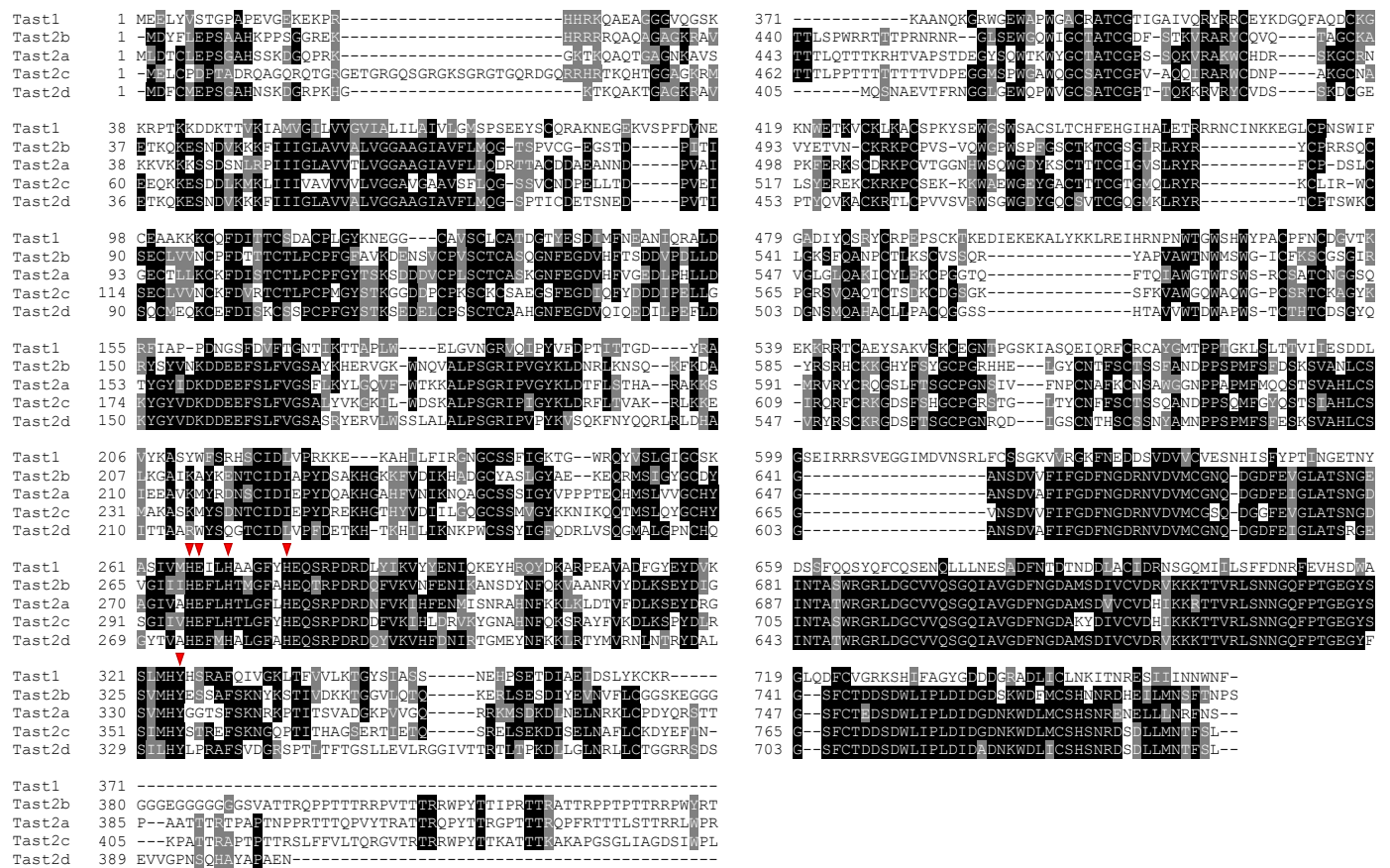


Fig. S2
Multiple sequence alignment of the refined sequences of the predicted products of *Tast* genes. Alignment was conducted with a CLUSTALW program (<http://www.genome.jp/tools-bin/clustalw>) at the default settings with a sorting option -outorder=input. Arrowheads indicate the five catalytic residues of astacins (NCBI Conserved Domains cd04280).

Fig. S3

a

>C9.806_Tast1
MEELYVSTGPAPEVGEKEKPRHHRKQAEAGGGVQGSKKRPTKKDDKTTV 50
KIAMVGILVVGVIALLILAIVLGMSPSEEYSCQRAKNEGEK**VSPFDVNEC** 100
EAAKKKCQCFDITTCSDACPLGYKNEGGCAVSCLCATDGTYESDIMFNEA 150
NIQRALDRFIAPPDNGSFDVFTGNTIK**TTAPLWELGVNGRVQIPYVFD** 200
TTTTGDYRAVYK**ASYWFSRHSCIDLVR**KKKAHILFIR**GNGCSSFIGK** 250
TGW**RQYVSLGIGCSK**ASIVMHEILHAAGFYHEQSRPDRDLYIKVYENI 300
QKEYHRQYDK**ARPEAVADFGYEYDVK**SLMHYHSR**AFQIVGK**LTFVVL**KT** 350
GYSIASSNEHPSETDIAEIDSLYKCKRKAANQKGRWGEWAPWGACR**ATC** 400
GTIGAIVQRYRRCEYKDGQFAQDCKGKNWETKVCKLKACSPKYSEWGSW 450
SACSLTCHFEGHIALETRRRNCINKK**EGLCPNSWIFGADIYQSR**YCR**P** 500
EPSCKTKEDEIEKEKALYKKLEIHRNPNWTGWSHWYPACPFNCDGVTKE 550
KKRRTCAEYSAKVSKEGNTPGSK**IASQEIQR**FCRCAYGMTPTTGKLSL 600
TTVIIESDDLGEIIRRSVEGGIMDVNSRLFCSSGKVVRGKFNEDDSDV 650
VVCVESNHISFYPTINGETNYDSSSQSYQFCQSENQLLLNESADFNTD 700
TNDDLACIDRNSGQMIILSFFDNRFEVHSDWAGLQDFCVGRKSHIFAGY 750
GDDDGRADLICLNKITNRESIIINNWNF

c

>C1.533_Tast2a
MLDTCLEPSGAHSSKDQGRPKGKTKQAQTGAGNAKVSKKVKKKSSDSNL 50
RPIIIGLAVVTLVGGAGIAVFLQDRTTACDDAEANNDPVAIGECTLL 100
KCK**FDISTCTTLP**CFGY**TSK**SDDDVCLPSCTCASKNGFEGDVHVFVEDL 150
PHLLDTYGYIDKDDEEFLSFLVGSFLK**YLGQVFWTK**KALPSGRIPVGYK**L** 200
DTFLSTHARAKKSIEEAVKMYR**DNSCIDIEPYDQAKHGAHFVNI**KNQAG 250
CSSSIGYVPPPTQHMSLVVGCHYAGIVAHEFLHTLGLFHEQSRPDRDN 300
FVKIHFENMISNRAHNFKKLKLDTVFDLKSEYDRGSVMHYGGTSFSKNR 350
KPTITSVADGKPVVGQRKMSDKDLNELNR**KLCPDYQR**STTPAATTTRT 400
PAPTNPRTTTTQPVYTRATTQPYTTRGPTTTRQPFRTTTTLSTTRRLWP 450
RTTTLQTTTKRHTVAPSTDEGYSQWTWKYGCATATCGPSSQKVRAKWCHD 500
RSKGCNRNPKFERKSCDRKPCVTGGNHSWQWGDYKSCSTTTCGIGVSLRYR 550
FCPDSLVCVGLGLQAKICYLEKCPGGTQFTQIAWGTWTSWSRCSATCNGG 600
SQMRVRYCRQGSFLTSGCPGNSIVFNPCNAFKCNSAWGNPPAPMFMQ**Q** 650
STSVAHLCSGANSDVAFIFGDFNGDRNVDVMCGNQDGFIEGLATSNGD 700
INTATWRGRLDGCVVQSGQIAVGDfNGDAMSdVVCVDHIKKRTTVRLSN 750
NQGFPTGEGYSGSFCTEDSDWLIPLDIDGDNKWDLMCshsn**RENELLN** 800
RfNS

e

>C1.332_Tast2d
MDFCMEPSGAHNSKDRPKHGKTKQAKTGAGKRAVETKQKESNDVKKKF 50
IIIGLAVVALVGGAGIAVFLMQGSPTICDETSNEDPVTISQCMQKCE 100
FDISKSSPCPFGYSTKSEDELCPSSCTCAAHGNFEGDVQIQEDILPEF 150
LDKYGYVDKDDEEFLSFLVGSASRYERVLWSSLALALPSGRIPVPYKVSQ 200
KFNYQQRLR**LDHAITTAAR**WYSQGTCLDLVPFDETKHTKHILIKNKPWC 250
SSYIGFQDRLVSQGMALGPNCHQGYTVAHEFMHALGFAHEQSRPDRDQY 300
VK**VHFDNI**RTGMeyNFKKLRTYmVRNLNTR**YDAL**SILH**YLPR**AFSVdGR 350
SPTLTFTGSLLEVLRGGIVTTRTLTPKDLLGLNRLCTGGRSDSEVVG 400
PNSQHAYAPaENMQSNAEVTFRNGGLGEWQPWWGCSATCGPTTQKKRVR 450
YCVDSKDCGEPTYQVK**ACKRTLCPVVSVR**WSGWDGYGQCSVTGCGGMK 500
LRYRTCPTSWKCDGNSMQAHACLPLACQGGSSHTAVVTDWAPWSTCTH 550
TCDSGLQVRYRSCK**RGDSFTSGCPG**NRQDIGSCNTHSCSSNYAMNPSP 600
MFSFESKVAHLCSGANSDVAFIFGDFNGDRNVDVMCGNQDGFIEGLA 650
TSR**GEINTATWR**GRLDGCVVQSGQIAVGDfNGDAMSdIVCvDRVKKKTT 700
VRLSNNGQFPTGEGYfGSFCTDDSDWLIPLDIDADNKWDLcshsnRDS 750
DLLMNTFSL

b

>C1.618_Tast2b
MDYFLEPSAAHKPPSGGREKHRRRRQAQAGAGKRAVETKQKESNDVKKK 50
FIIIGLAVVALVGGAGIAVFLMQGTSPVCGEGSTDPITISECLVVNCP 100
FDTTTCTLPCPFGFfAVKDENSVCVSVCTCASQGNFEGDVHFTSDDVPDL 150
LDRYSYVNKDDEEFLSFLVGSAYKHHERVGK**WNQVALPSGR**IPVGYKLDNR 200
LK**NSQKFK**DALKGAIKAY**KENTCIDIA**PYDSAKHGKK**FVDIKHADG**CYA 250
SLGYAEEQRMSIGYGCDYVGIIIEHFLHTMGFAHEQTRPDRDQFVKVN 300
FENIK**ANS**DY**YNFQK**VAANRVYDLKSEYDIGSVMHYESSAFSKNYKSTIV 350
DKKTGGVLQTQKER**LS**ESDIYEVN**VFLCGGSK**EGGGGGGEGGGGGGGSV 400
ATTTRQPPTTTRRPVTTTTRRPYTTIPRTRATTTRPPTPTTRRPWYRTTT 450
LSPWRRTTTPRNRRLSEWGQWIGCTATCGDFSTKVRAR**YCQVQTAGC** 500
KAVYETVNCKRRKPCPVSVQWGPWSPFGSCTKTCGSGRLRLRYRCPRRSQ 550
CLGK**SFOANPCTLK**SCVSSQRYAPVAWTNWSWGICFKSCSGSIRYRSR 600
HCK**KGHYFSYGCPGR**HHELGYCNTFSCSTSSFANDPPSPMFSFDSKSVAN 650
LCSGANSdVVFIFGDFNGDRNVDVMCGNQDGFfEVLATSNgeINTASW 700
RGRLDGCVVQSGQIAVGDfNGDAMSdIVCvDRVKKKTTVRLSNNGQFPT 750
GEGYSGSFCTDDSDWLIPLDIDGSKWDFMCshnNRDHEILMNSfTNPfS 800

d

>C1.493_Tast2c
MELCPDPTADRQAGRQTGRGETGR**QSGRGKSGR**GTGQRDGQRRHRTK 50
QHTGGAGKRMEEQKKESDCLKMKLIIIVAVVVVLVGGAVGAASFLQGS 100
SVCNDPELLTDPVEISECLVVNCKFDVTRCTLPCPMGYSTKGGDDPCPK 150
SCKCSAEGSFEGDIQFYDDDIPELLGKYGYVDKDDEEFLSFLVGSALYVK 200
GKILWDSKALPSGR**IPIGYK**LDRFLTVAKRLKKEMAKASKMYSdNTCID 250
IEPYDREKHGTHYVDIILQGCSSMVGYKKNIKQQTMSLQYGCHYSgII 300
VHEFLHTLGfYHEQSRPDRDDFVKIHLDRVKYGNahNFQksRAYfVKDL 350
KSPYDLRSIMHYSTREFSK**NGOPTITHAG**SERTIETQsRELSEK**DISEL** 400
NAFLCKDYEFTNKPATTRAPTPTTRSLFFVLTQRGVTRTRRPYTTKAT 450
TTKAKAPGSLIAGDSIWPLTTTLPPTTTTTTTTTVDPEGGMSPWGAWQG 500
CSATCGPVAQQIRARWCDNPAKGCNALSyEREKCKRKPCSEKKKWAEWG 550
EYGACTTTCGTGMQLRYRKCLIRWCPGRSVQ**QAQ**TCtSDKCDGSGKSfKV 600
AWGQWAQWGPCSRTCKAGYKIRQRfCRKGDSfSHGCPGRSTGLTYCNFF 650
SCTSSQANDPPSQMFGYQSTsIAHLCSGVNSdVVFIFGDFNGDRNVDVM 700
CGSQDGGfFVGLATSNgDINTASWRGRLDGCVVQSGQIAVGDfNGDAK**Y** 750
DIVCVDHIKKKTTVRLSNNGQFPTGEGYSGSFCTDDSDWLIPLDIDGDN 800
KWDLMCshsnRDSdLLMNTfSL

f

sum of protein scores (Mascot ver 2.4.1)

	Tast1	Tast2b	Tast2a	Tast2c	Tast2d
ionomycin-	1875	832	324	318	323
ionomycin+	2506	707	581	321	425

g

 m/z 992.82, $z = +2$ 

m/z 1009.02, $z = +2$

 m/z 580.76, $z = +2$ 

m/z 655.30, $z = +2$

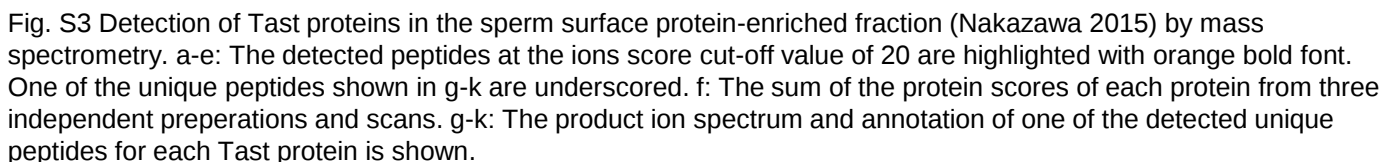
 m/z 817.43, $z = +2$ 

Fig. S4

a

xxxxxx . . . : intron/non-transcriptional region
xxxxxx . . . : exon
ATG: first ATG

C9.806 (*Tast1*) exons 1 and 2

left

spacer

right

: target

gttgctacaacgtatcac

cagacatgtatacgggtaac

cacaatttttaccagaacctgtc

→ C9.806exon1 primer

caatttttatttatgtttttcaacattttgcattaagtttaatttttttttttaattta

acataatttatgttcacaaatttttggcggggaaattttggatcatcgtttaaaattcagac

exon 1

ATAATTTTTTACGATTGTAAAA**ATG**GAGGAACTGTATGTTAGCACTGGCCCTGCTCCGGA

C9.806-1 C9.806-2

AGTTGGGGAAAAGGAAAAACCTCGTCATCATCGCAAGCAGGCAGAAGCTGGCGGGGGGTGG

C9.806-3

TAAGTCTGTGTTACGTAATATATTTGATTAgaaacaaaaacaacaaccaccaacaaagt

aacatacttgaaaactcctaaactggcacgaggtgtatatgaagtattgaagtgttatac

actgttgtttttcagccacagagataaagttgcattttattcattcaattttatgtttatg

← C9.806e1r primer

tttagcctcaaacactaacagctgagcaaatacacacaaactagttaacaataagtgtgt

exon 2

ataattttttatttggtgtttactattttataccacattcaatttttttagTTCAAGGATCT

AAAAAACGTCCAACCTAAGAAAGATGATAAACTACTGTTAAATAGCAATGGTTGGAATA

C9.806-4

CTTGTTGTTGGAGTAATAGCTCTCATTCTGGCTATTGTATTAGgtgagtggttttaaacgt

taaagtgtacgttatattatcatgtaaatggatgaaagcagttaataatggaaaaattgg

cacagtaatttttagaggatattgatagtccttgatagaagacatgggtaaaaggggggcttg

ggtccaattttataaacatgggactatacaagaagacgg

← C9.806e2r primer

cggtaattatttaaatataaaaaatacggaaaaacgaacaaaacagcactacaaagtgtta

ataagtgtaaaacaaattaattaataataaaaaatacctttctaaatacataaaacacta

Fig. S4 Continued

b **C1.533 (*Tast2a*) exon 1**
aataacagcagcggcgagccctgaacctagattagaggcgcgccaaccactctgccacca
aacttaacaagatgcaaagttttattgattctcttttcttttatcaagttgtataccatt
C1.533exon1 primer →
exon 1
GACAAACTAGATTTGTAACAACAACCGCTTTTCAAATTGCATAAACTTTAAAATCTCTT
TATCACATTTTACATA**ATG**TTGGATACTTGTCTGGAACCATCTGGTGCACATAGCTCAAA
C1.533-1 C1.533-2
AGATGGCCAGCCAAGAAAAGGAAAGACAAAACAAGCCCAAACTGGAGgtagtgtactaag
ttcttgctatTTaatttttataaagtaatatatgcaaacatttaaagctataaatataag
ggtgtacagtttttttaccctaaacaccaaagtagggcgatggaaaattattgatgttga
← C1.533e1r primer
gttatgtgtatTTtttgaaaagtgtacattagtgtataacagactatagcactacttatgt
ttaatttgtaccaacaacttaaattgcactacagcaggtatTTaaaccaagagagtt

c **C1.332 (*Tast2d*) exon 1**
ttagtttttaaacacgtgatttgaacgttgtacatctggtctttgcgttcttattctgtg
C1.332e1r primer →
aacttattttgtaaaaaaatttaattttgttttgtctgaaacgtgttattttgttgcag
exon 1
AAGGTCTATAGTATTAGGCTAGTATATTGTGACAAAATATGTATGCAAAGTAATCTTAAT
TGAATTAACAAAATGGATTTTTGT**ATG**GAACCTTCTGGTGCACACAACCTCTAAAGATGGT
C1.332-1 C1.332-2
CGTCCCAAACATGGAAAAACAAAACAAGCCAAAACTGGAGgtgatggttttataattgtc
tcatattatgtaatatgttatatagaaaattttgtgatataattataatgttccttatgt
agtttttgtgttaatgagaacatcctactggcaccgcggaagtggtagcgccacatgc
ctctaaccagcggtaattggttcaaggcttgtcactgctaccattgtggacgttgggca
← C1.332exon1 primer
agacacaaattgtacaaattagcatccacacataaatattaaaaaaattccccaaaaata

Fig. S4 Continued

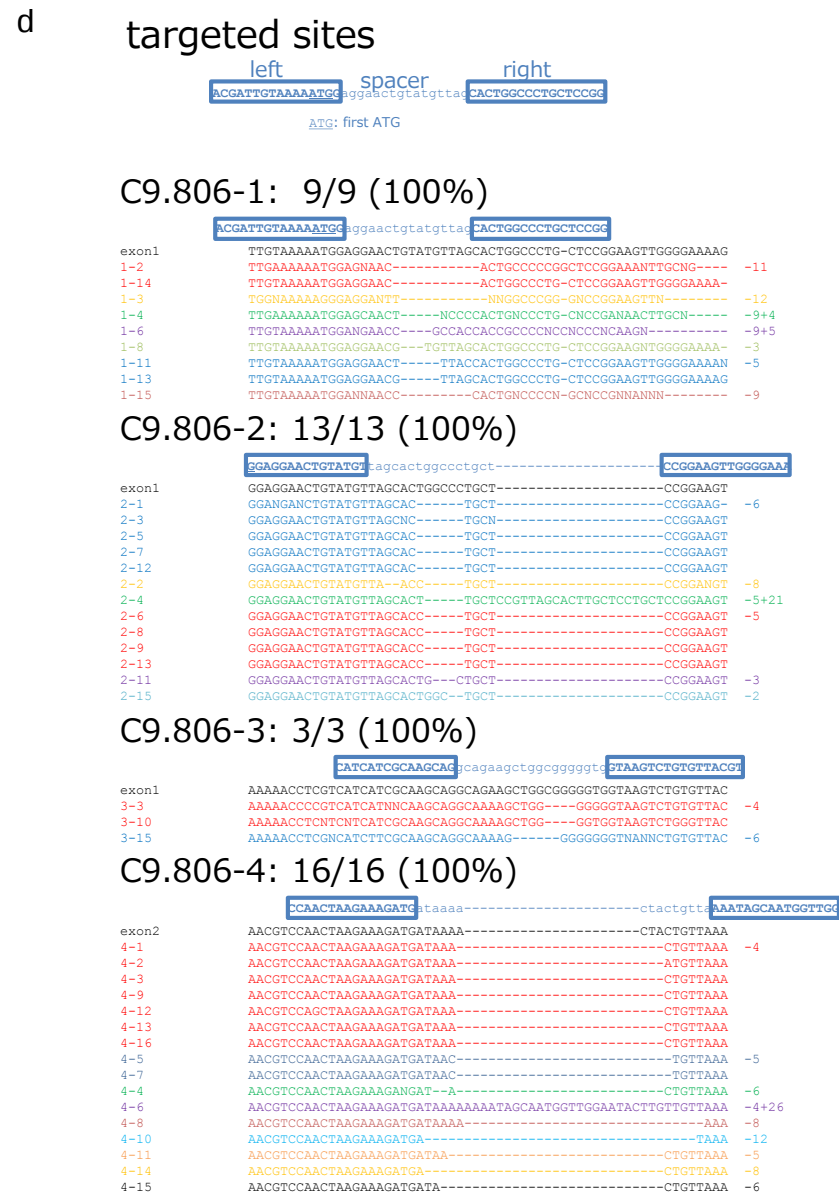
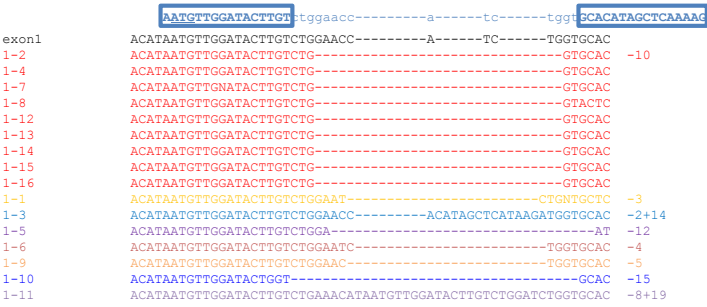


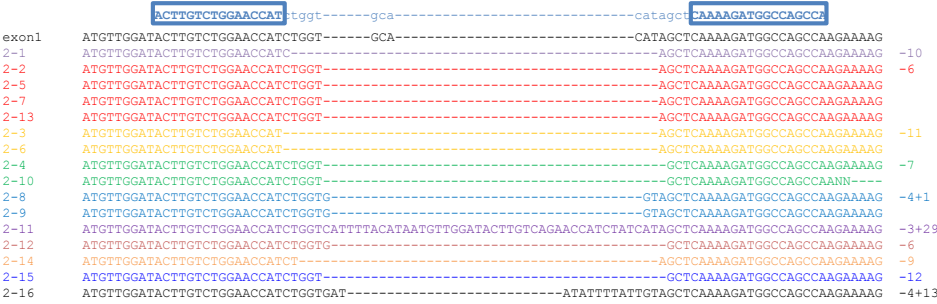
Fig. S4 Continued

e

C1.533-1: 16/16 (100%)

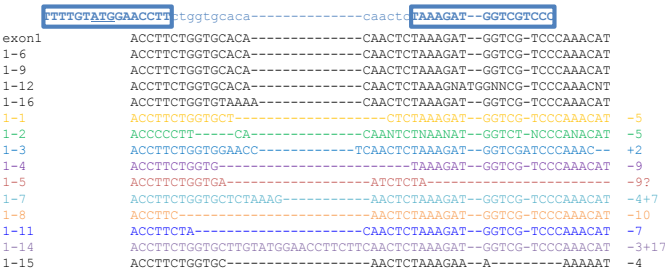


C1.533-2: 16/16 (100%)



f

C1.332-1: 10/14 (71.4%)



C1.332-2: 14/14 (100%)



Fig. S4
Designs of TALEN pairs targeting Tact genes.
a-c. The targets of the left and right TALENs and the primers for genotyping are shown in the genome sequences.
d-f. The genotypes observed in the TALEN-injected tailbud larvae and the efficiency of mutagenesis caused by TALEN pairs.

Fig. S5

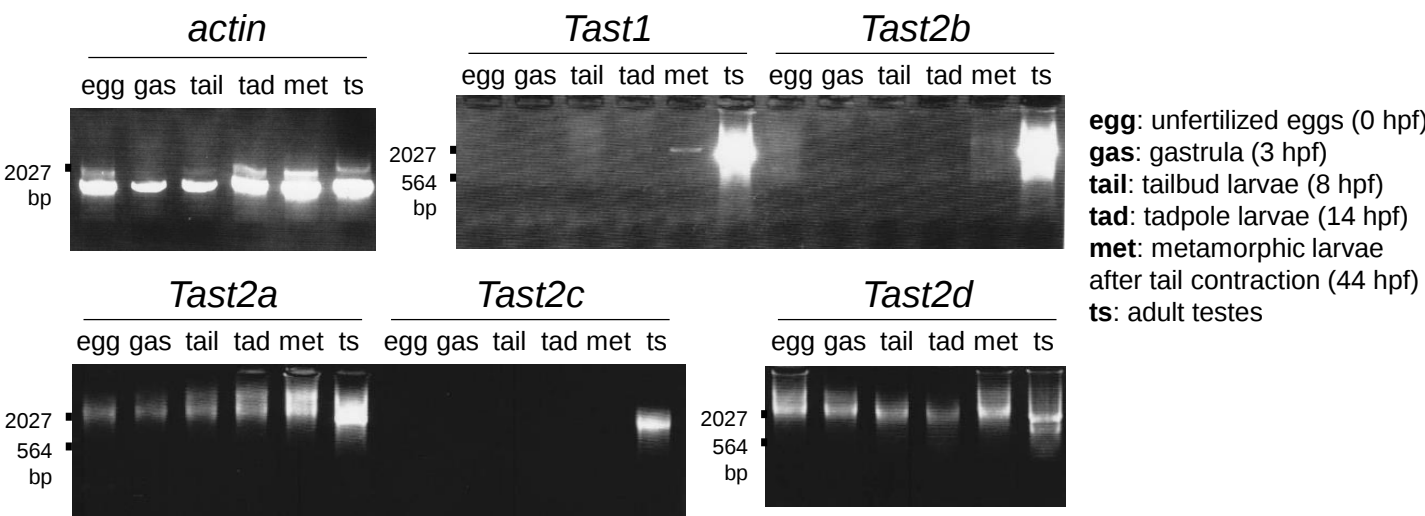


Fig. S5 At least three *Tast* genes were expressed at the developmental stages.

Fig. S6

lane No.	1	2	3	4	5	6
VC	+	+	+	+	-	-
sperm	-	-	+	+	+	+
GM6001	-	+	-	+	-	+

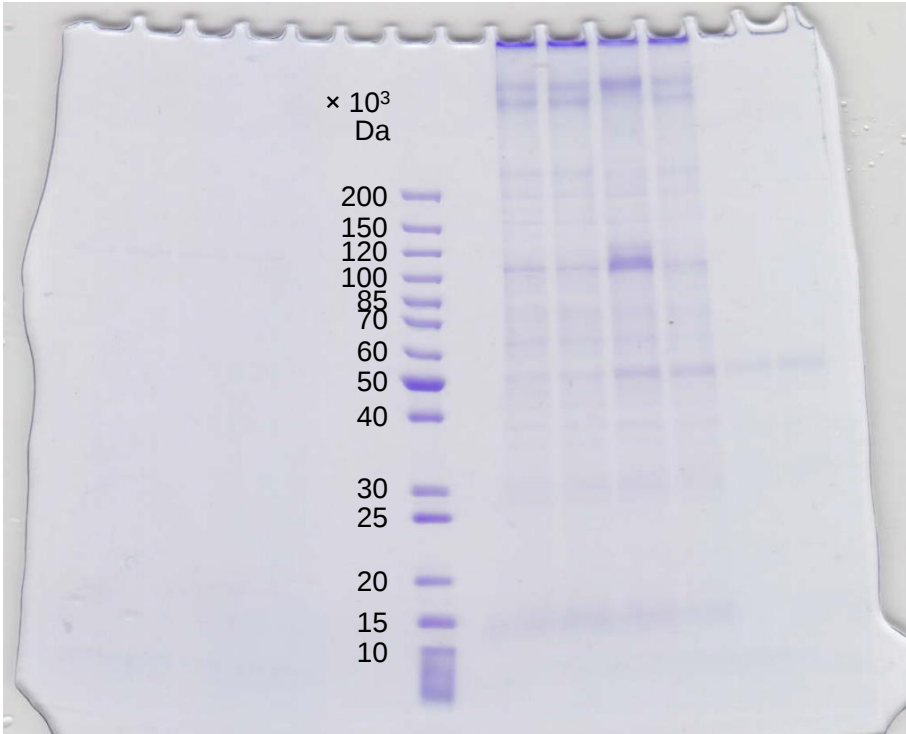


Fig. S6 The whole image of the gel shown in Figure 2.

Supplementary Table 1 The primers for amplification of the full coding sequences of *Tast* genes.

Target	Primer name	Sequence	Restriction site
<i>Tast1</i>	C9.806for	TTGGTACCATTGTAAAAATGGAGGAAC	<i>KpnI</i>
<i>Tast1</i>	C9.806rev	TGTGGATCCACAAGATAACAATAAGC	<i>BamHI</i>
<i>Tast2b</i>	C1.618for	TAGGTACCAAAATAAAATCTGAAATGG	<i>KpnI</i>
<i>Tast2b</i>	C1.618rev	AATGGATCCAATGAAATGTCTTTTATG	<i>BamHI</i>
<i>Tast2a</i>	C1.533for	TCGGTACCACATTTTACATAATGTTG	<i>KpnI</i>
<i>Tast2a</i>	C1.533rev	CTGGGATCCTAGATGTAAACTGGTT	<i>BamHI</i>
<i>Tast2c</i>	C1.493for2	AAAggtacCGAATTATTCAAGTGGCC	<i>KpnI</i>
<i>Tast2c</i>	C1.493rev	TTGGGATCCATTTTGTGTGATCAC	<i>BamHI</i>
<i>Tast2d</i>	C1.332for	GAGATATCAAATGGATTTTGTATGG	<i>EcoRV</i>
<i>Tast2d</i>	C1.332rev	AACGGATCCAATTCATTAATATAAC	<i>BamHI</i>

Supplementary Table 2 The primers designed for 5'-RACE of *Tast2c*.

Target	Primer name	Sequence	Restriction site
BglII site + oligo (dC)	BglII-(dC)10	AAGATCTCCCCCCCCC	<i>BglII</i>
C1.493 GSP for 5'-RACE	C1.493RACE1	ctcggatCCTTTAACATAGAGAGCAC	<i>BamHI</i>

Supplementary Table 3. The primers used in confirmation of the sequences of cloned *Tast* genes.

Target	Primer name	Sequence	Restriction site
pBluescript universal primer-binding site	M13-20	CGACGTTGTAAAACGACGGCCAGT	-
pBluescript universal primer-binding site	T3 promoter	ATTAACCCCTCACTAAAGGGAA	-
<i>Tast1</i>	C9.806seq1	GGGATTGGTTGTTCAAAAGC	-
<i>Tast1</i>	C9.806seq2	TGTGTTCCCTTCACATTGTC	-
<i>Tast2b</i>	C1.618seq1	TATGGGTGCGATTATGTCGG	-
<i>Tast2b</i>	C1.618seq2	ACTGTGACCTTCTAGGACAG	-
<i>Tast2a</i>	C1.533seq1	GCTGTTCTTCTTCCATCGGG	-
<i>Tast2a</i>	C1.533seq2	TGTCCCCAAGCAATTTGAG	-
<i>Tast2c</i>	C1.493seq1	AAGATATTTCTGAGCTGAATGC	-
<i>Tast2c</i>	C1.493seq2	CCACATGTAGTTGTGCATGC	-
<i>Tast2d</i>	C1.332seq1	GAATTTCTTGACAAATACGGG	-
<i>Tast2d</i>	C1.332seq2	GCCATCACACTTCCATGATG	-

Supplementary Table 4. The primers for PCR amplification and genotyping of TALEN-targeted sequences.

Target	Primer name	Sequence	Restriction site
<i>Tast1</i> exon 1	C9.806exon1	cagacatgtatacggttaac	-
<i>Tast1</i> exons 1~2	C9.806e2r	ccgtcttctttgtatagtc	-
<i>Tast2a</i> exon 1	C1.533exon1	ctgccaccaacttaacaag	-
<i>Tast2a</i> exon 1	C1.533e1r	catcaataattttccatcgcc	-
<i>Tast2d</i> exon 1	C1.332e1f	ctttgcgttcttattctgtg	-
<i>Tast2d</i> exon 1	C1.331exon1	acaagccttgaaccaattac	-