

Supplementary Figures and Table

The role of Atg8 in the regulation of vacuolar membrane invagination

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Figure S1

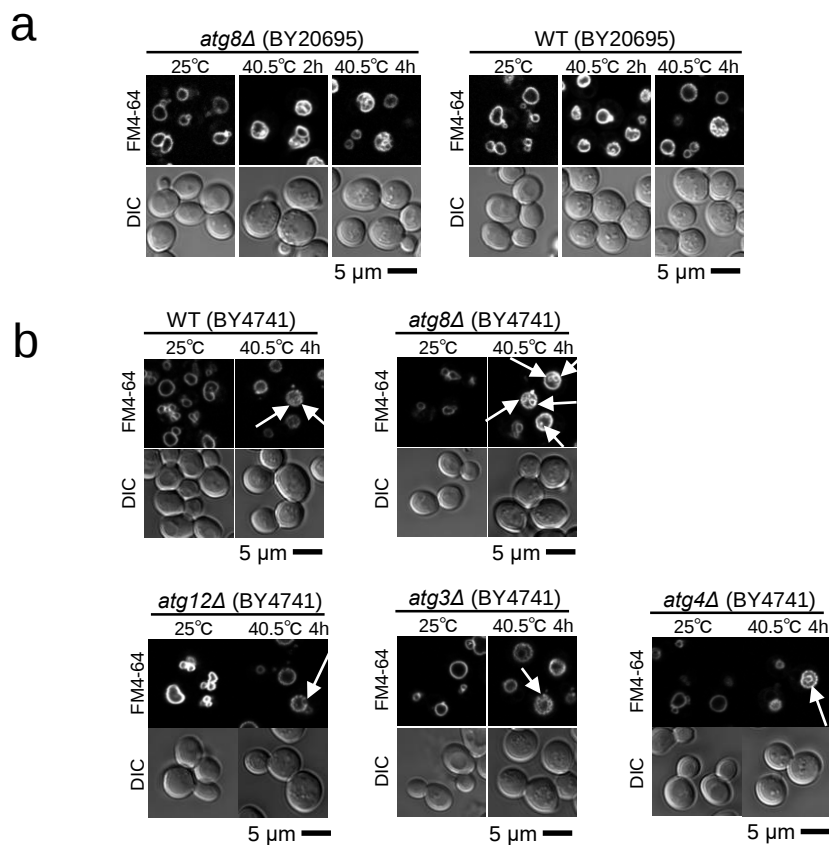


Figure S1. FM4-64 staining.

(a) Strain BY20695 and *atg8Δ* cells.

(b) FM4-64 staining of BY4741, *atg8Δ*, *atg3Δ*, *atg4Δ*, and *atg12Δ* incubated at 40.5° C for 4 h. Arrows indicate vacuolar invaginations

Figure S2

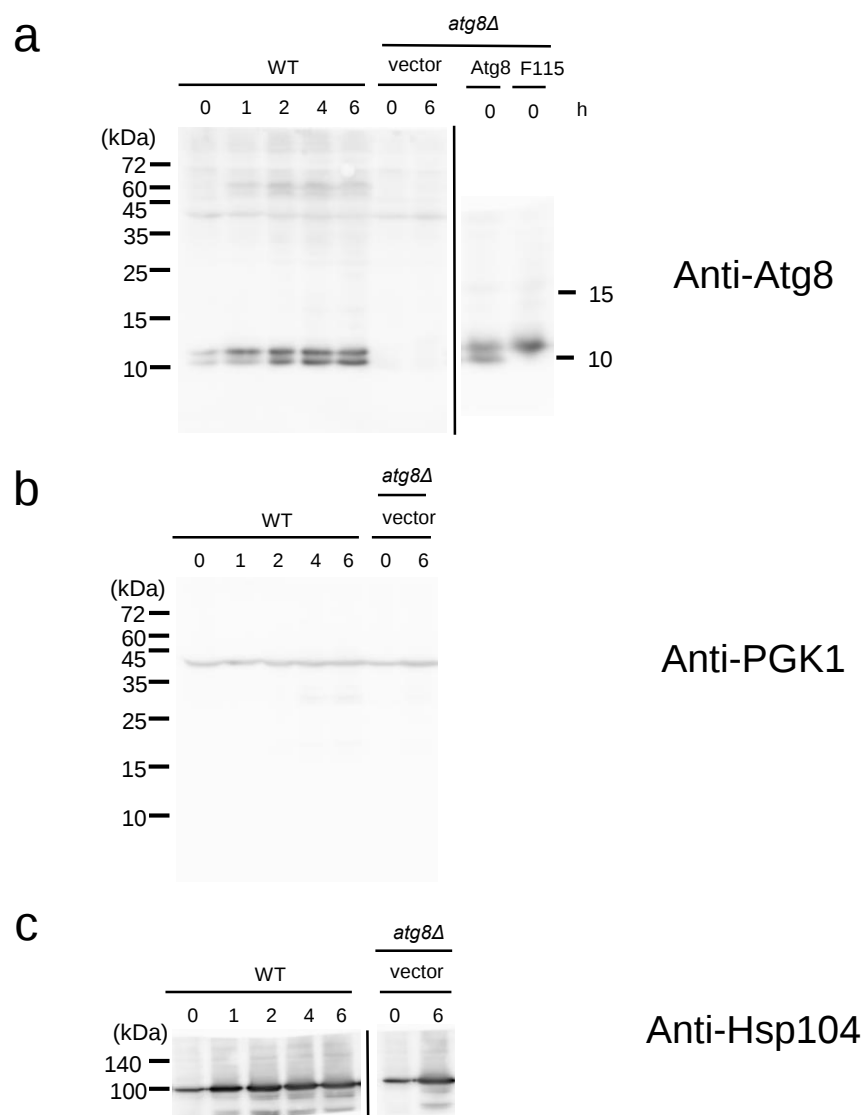


Figure S2. Uncropped images of the blots shown in Fig.2.
(a) Anti-Atg8 antibody.
(b) Anti-PGK1 antibody.
(c) Anti-Hsp104 antibody.

Figure S3

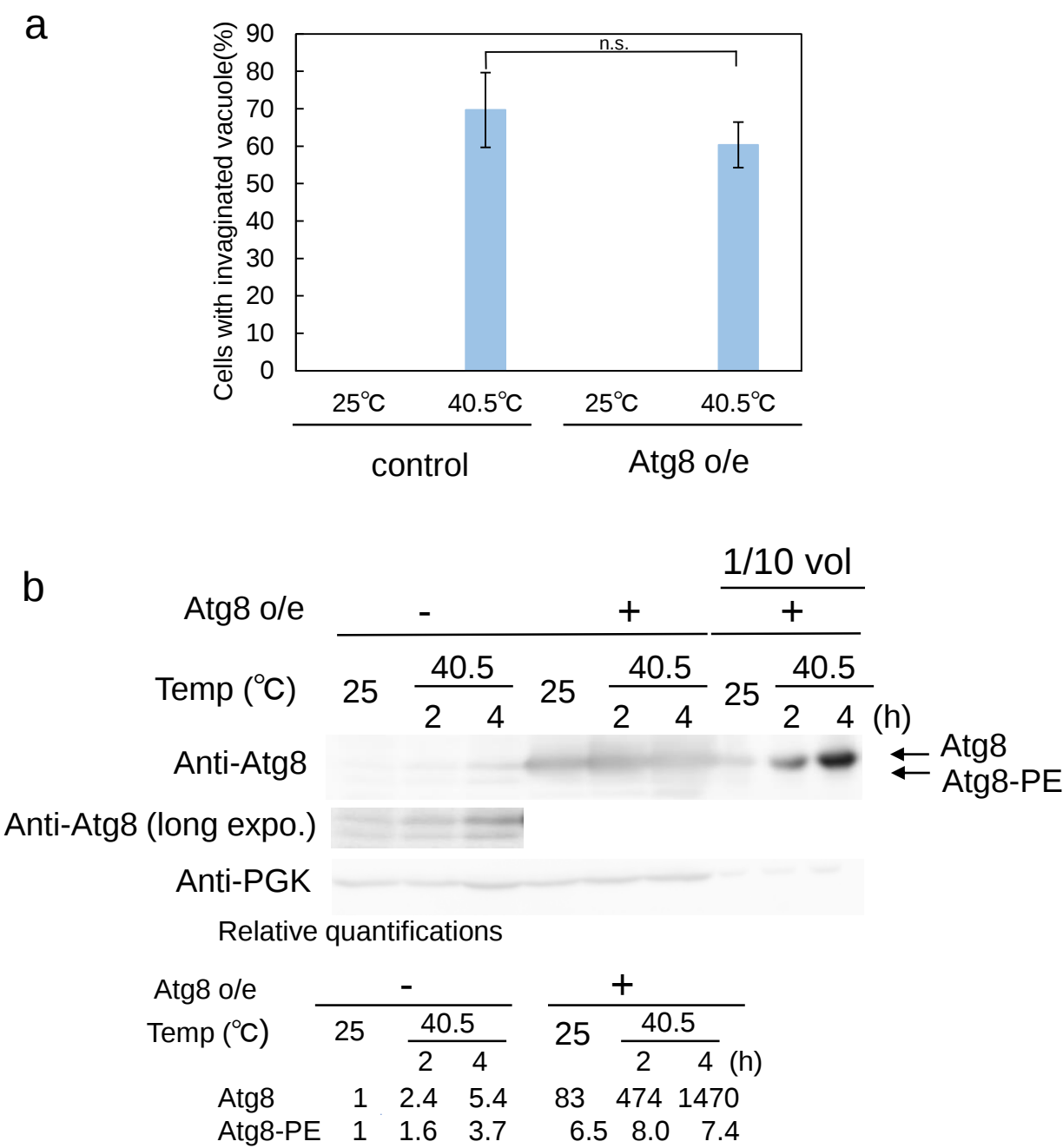


Figure S3. Overexpression of Atg8

(a) Quantifications of vacuolar invaginations of wild-type cells harbouring a control plasmid or a plasmid overexpressing Atg8 at 25 ° C and at 40.5° C for 4 h. Cells were grown in SC-ura media to log phase and heat shocked for 4 h. The mean values of the ratio and standard errors (SE) are shown. $p>0.05$ for t-test.

(b) Western blotting with anti-Atg8 and anti-PGK1. One-tenth vol of lysates of wild-type cells overexpressing Atg8 were also examined. An image with a longer exposure time was added. Relative quantifications were shown at the bottom. Quantifications of cells overexpressing Atg8 were calculated from the results of 1/10 vol of lysates.

Figure S4

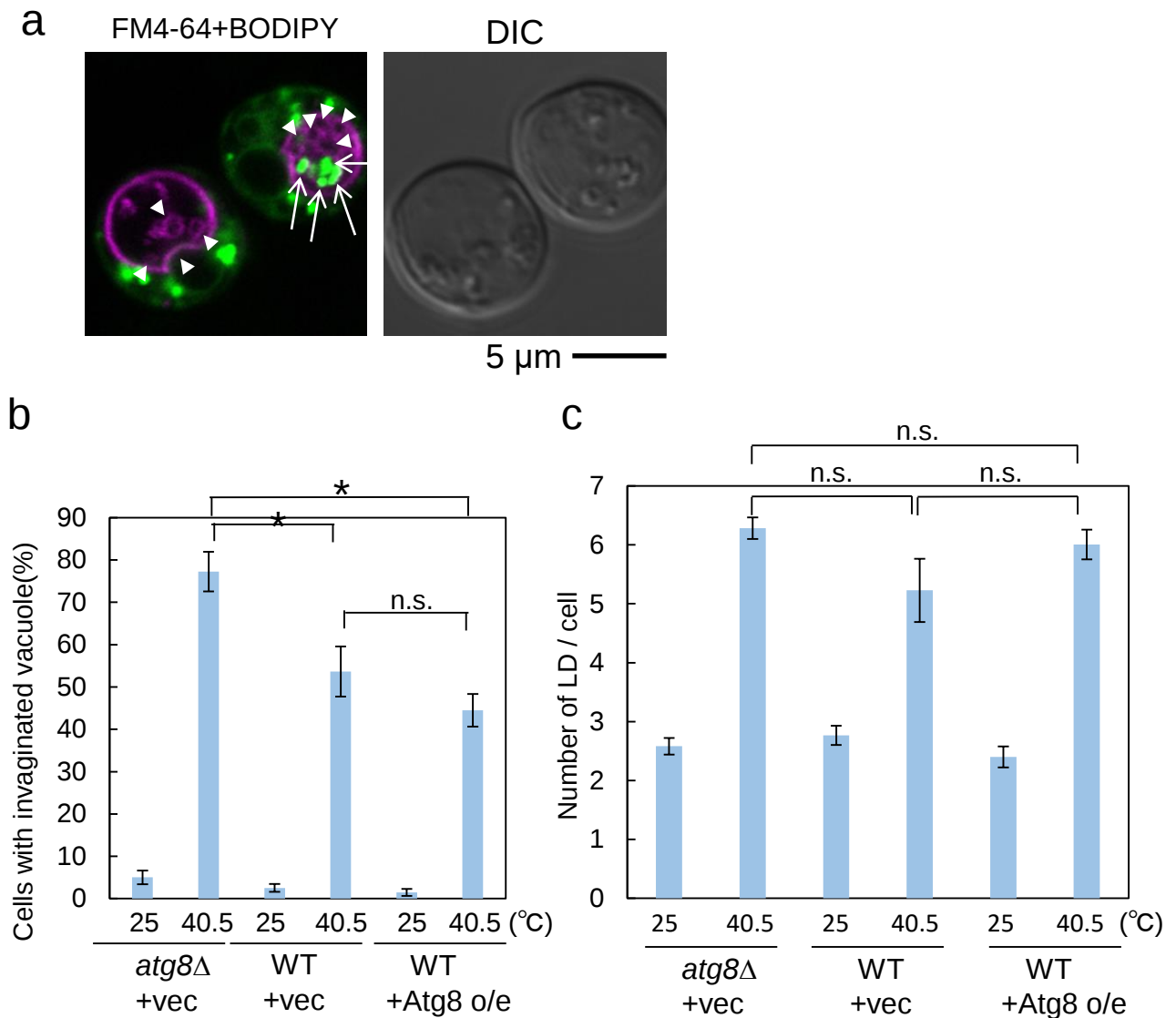


Figure S4. Relationship with LDs

(a) BODIPY493/503 and FM4-64 fluorescence in *atg8Δ* cells incubated at 40.5° C for 4 h. Arrows and arrowheads indicate vacuolar invaginations with LDs and vacuolar invaginations without LDs, respectively.

(b) Quantification of vacuolar invaginations by *atg8Δ* deletion or Atg8 overexpression. Cells harbouring a vector or 2μm-based Atg8-expressing plasmid were grown in SC-Ura media. Cells were centrifuged and suspended in YPD and heat shocked for 4 h. The mean values of the ratio and standard errors (SE) are shown. Experiments were repeated for 5 times. Statistical significance : * indicates $p < 0.05$, and n.s. indicates $p > 0.05$ for t-test.

(c) Number of LDs by *atg8Δ* deletion or Atg8 overexpression. The mean values of the ratio and standard errors (SE) are shown. Experiments were repeated for 3 times. Statistical significance : n.s. indicates $p > 0.05$ for t-test.

Figure S5

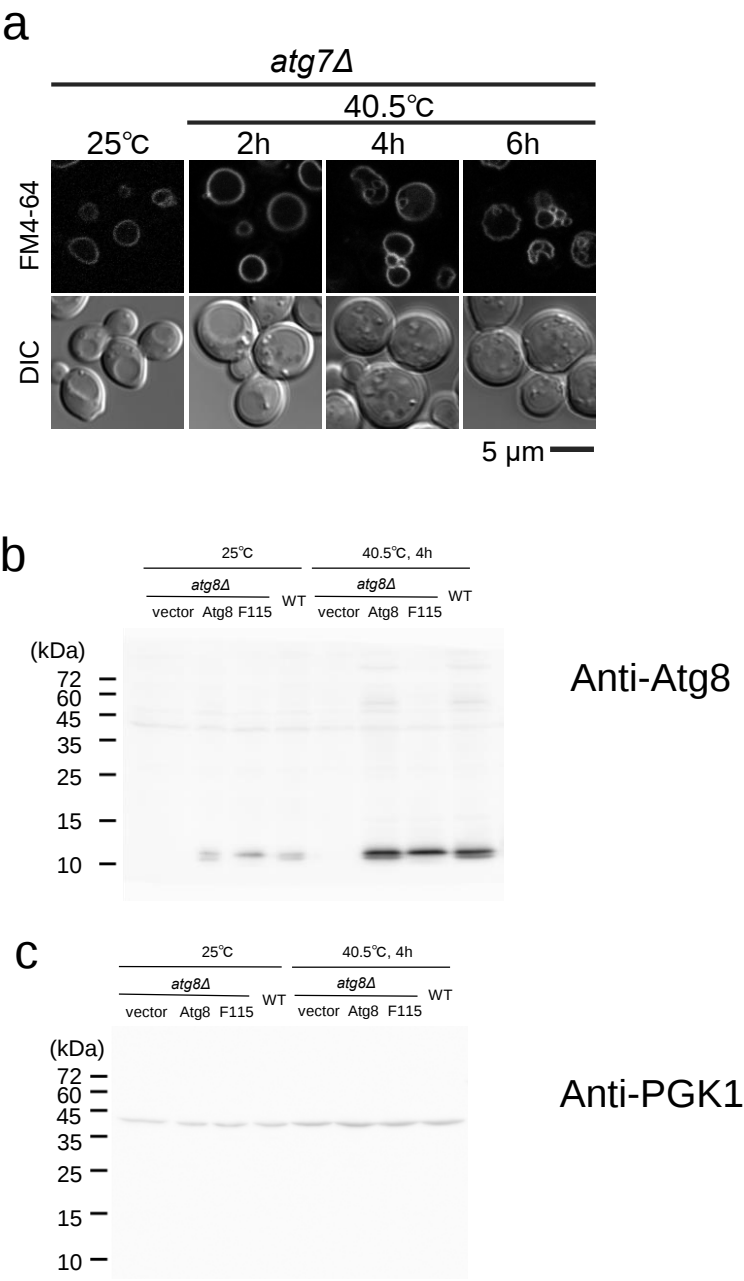


Figure S5. Lipidation-independent function of Atg8

(a) FM4-64 stained cells with equal degree of contrast adjustments of images between the cells grown at 25° C and heat stressed cells in Figure 3a.

(b) and (c) Uncropped images of the blots shown in Fig. 3d.

Figure S6

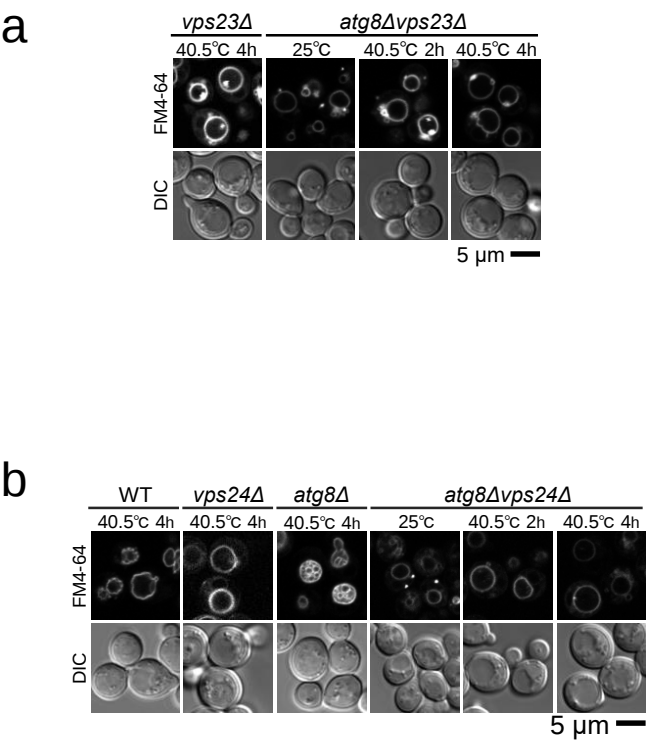


Figure S6. FM4-64 staining.

(a) FM4-64 staining of *vps23Δ* and *atg8Δvps23Δ* cells. Scale bar, 5 μm.

(b) FM4-64-stained cells with the same image contrast adjustments between cells grown at 25° C and the heat-stressed cells in Fig. 4a.

Figure S7

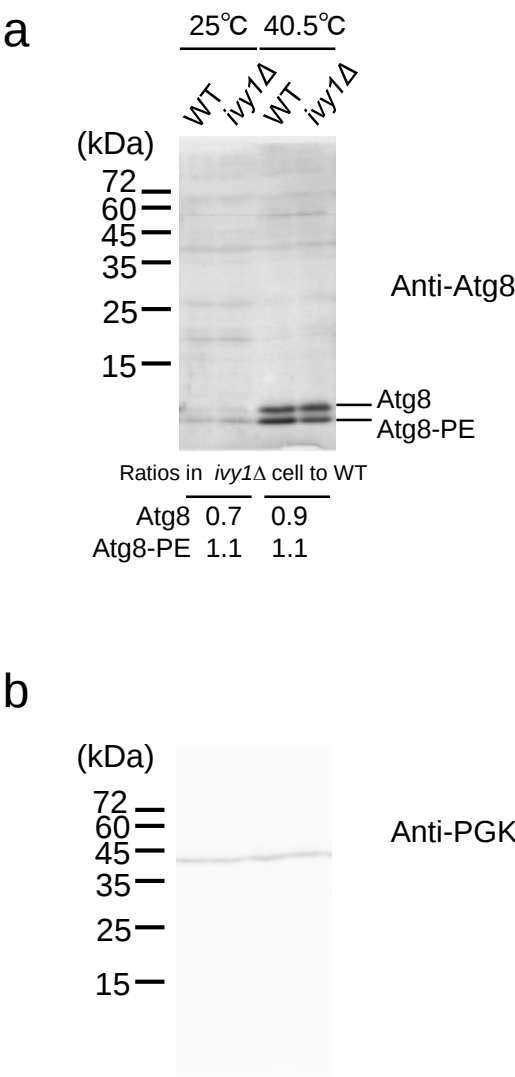


Figure S7. Atg8 expression in wild-type and *ivy1Δ* cells.
(a and b) Western blotting of anti-Atg8 and anti-PGK1 in wild-type and *ivy1Δ* cells at 25° C and 40.5° C for 6 h. Relative ratios of the level of Atg8 and Atg8-PE in *ivy1Δ* with respect to wild-type for each temperature are shown.

Figure S8

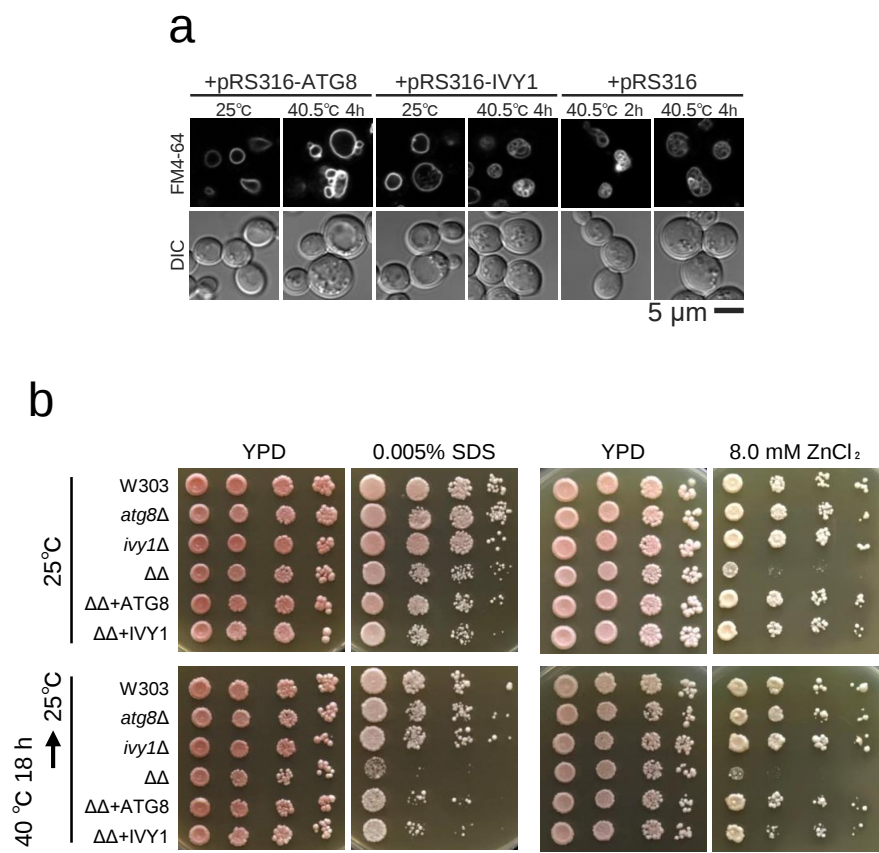


Figure S8. Suppression of invagination and growth sensitivities of *atg8Δivy1Δ* cells at 25° C by expression of either Atg8 or Ivy1.

(a) Vacuolar morphology. FM4-64 staining of *atg8Δivy1Δ* cells harboring a plasmid expressing Atg8 or Ivy1 or a vector (pRS316) are shown.

(b) Growth phenotype. Wild-type, *atg8Δ*, *ivy1Δ* *atg8Δivy1Δ* with a vector, *atg8Δivy1Δ* with a plasmid expressing Atg8, *atg8Δivy1Δ* with a plasmid expressing Ivy1 were diluted by ten-fold and spotted on YPD, YPD+0.005% SDS or YPD+8.0 mM ZnCl₂. Cells were placed at 25° C or at 40 ° C for 18 h followed by the incubation at 25° C. Cells were incubated at 25° C for 3 days for YPAD, 4 days for YPD+ 0.005 % SDS, and 8 days for YPD +8.0 mM ZnCl₂ plates, respectively.

Table S1. Strains used in this study

Name	Genotype	Source/Reference
W303	MATa <i>ade2-1 can1-100 his3-12,16 leu2-3,112 trp1-1 ura3-1</i>	Rothstein
Y1373	W303a, <i>atg8Δ::KanMX</i>	This study
Y1408	W303a, <i>atg8Δ::KanMX</i>	This study
Y1390	W303a <i>PGK1-GFP::HIS3</i>	This study
Y1412	W303a <i>PGK1-GFP::HIS3, atg8Δ::KanMX</i>	This study
Y1444	W303a, <i>atg8Δ::KanMX, pRS306::URA3</i>	This study
Y1445	W303a, <i>atg8Δ::KanMX, pRS306-ATG8::URA3</i>	This study
Y1446	W303a, <i>atg8Δ::KanMX, pRS306-ATG8-F115::URA3</i>	This study
Y995	W303a, <i>vps23Δ::KanMX</i>	Ishii et al, 2018
Y1287	W303a, <i>vps24Δ::KanMX</i>	Ishii et al, 2018
Y1406	W303a, <i>atg8Δ::KanMX, vps24Δ::KanMX</i>	This study
Y1545	W303a, <i>atg8Δ::KanMX, vps23Δ::KanMX</i>	This study
Y1375	W303a, <i>ivy1Δ::KanMX</i>	Ishii et al, 2018
Y1539	W303a, <i>atg8Δ::KanMX, ivy1Δ::KanMX</i>	This study
Y1542	W303a, <i>atg8Δ::KanMX, ivy1Δ::KanMX, PGK1-GFP::HIS3</i>	This study
Y1416	W303a, <i>atg7Δ::KanMX</i>	This study
BY4741	MATa <i>his3Δ1 leu2Δ0 ura3Δ0 met15Δ0</i>	Euroscarf
atg8	BY4741, <i>atg8Δ::KanMX</i>	Euroscarf
atg3	BY4741, <i>atg3Δ::KanMX</i>	Euroscarf
atg4	BY4741, <i>atg4Δ::KanMX</i>	Euroscarf
atg12	BY4741, <i>atg12Δ::KanMX</i>	Euroscarf
Y1410	W303, <i>IVY1-GFP::HIS3</i>	This study
Y1589	W303, <i>IVY1-GFP::HIS3, atg8Δ::KanMX</i>	This study
Y1590	W303, <i>IVY1-GFP::HIS3, vps24Δ::KanMX</i>	This study
BY20695	MATα, <i>his3-Δ1, leu2Δ0, met15&Delta0, ura3Δ0</i>	NBRP
Y1570	BY20695, <i>atg8Δ::KanMX</i>	This study