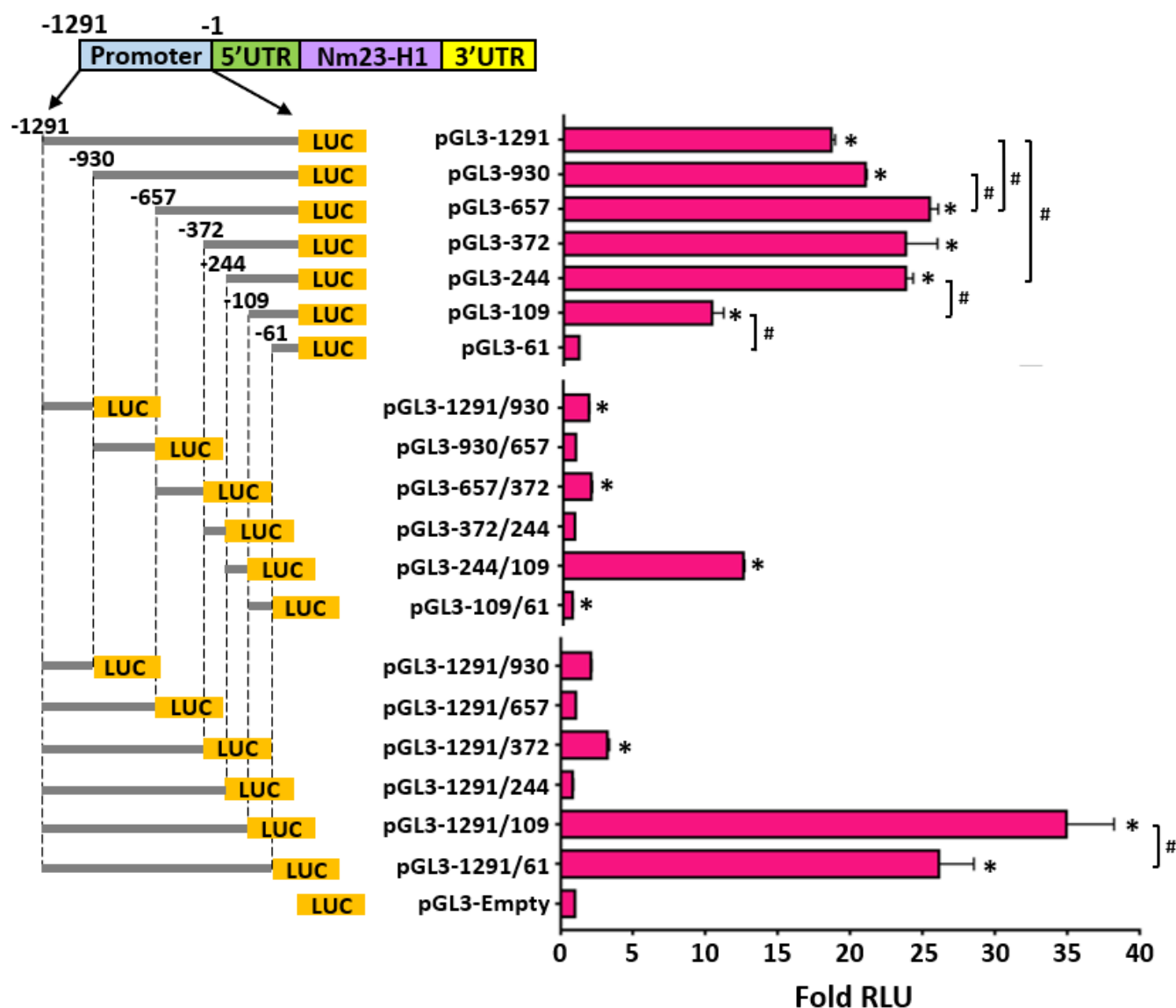


CTCF and EGR1 suppress breast cancer cell migration through transcriptional control of Nm23-H1

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Supplementary Figure S1. Analysis of Nm23-H1 promoter constructs in HEK293 cells. HEK293 cells were transiently transfected with Nm23-H1 promoter constructs and pRL-TK control vector and lysed 48 hours after transfection. Luciferase activities were measured using the Dual-Luciferase Reporter Assay System (Promega). RLU values of reference construct pGL3-Empty: *Firefly* luciferase ($1,128,152 \pm 13,073$), *Renilla* luciferase ($3,427,559 \pm 51,849$). Data represent the mean and standard deviation of three independent trials. *, significance with pGL3-Empty, $p < 0.05$; #, $p < 0.05$.

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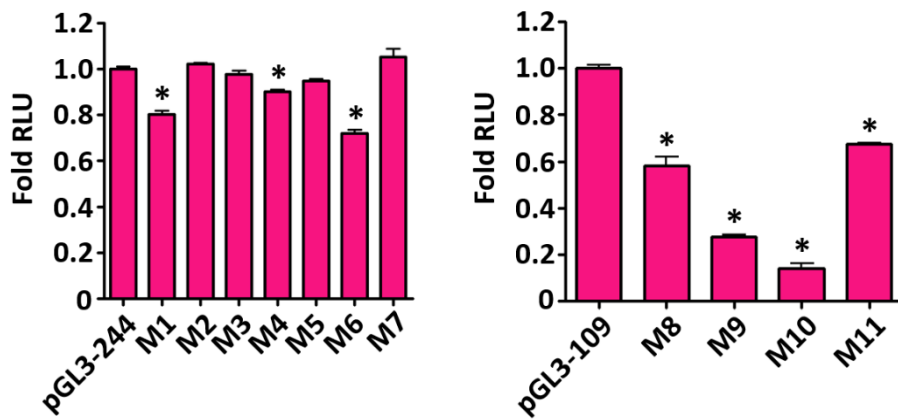
Minimal promoter region (-109 bp to -1 bp)

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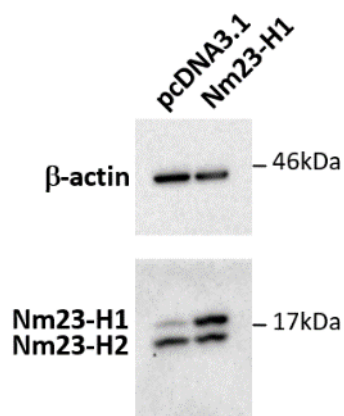
human	AAGCGTTCCGTG
macaque	AAGCGTTCCGTG
mouse	AAAGGTTCCGTG

Supplementary Figure S2. Cross-species alignment of *NME1* promoter region. The proximal and minimal promoter region of *NME1* were aligned to the macaque and mouse sequence using the ECR Browser [55] and ClustalW2 alignment tool [56]. Conserved bases in all three species are indicated by an asterisk.

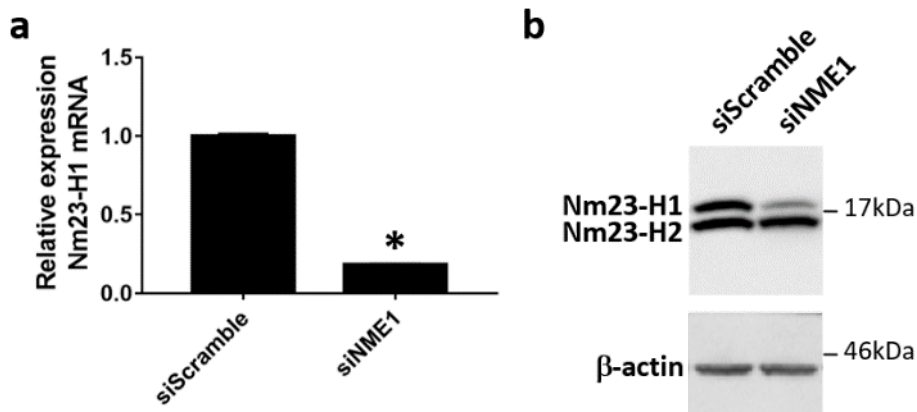


Supplementary Figure S3. Site-directed mutagenesis of transcription factor binding sites.

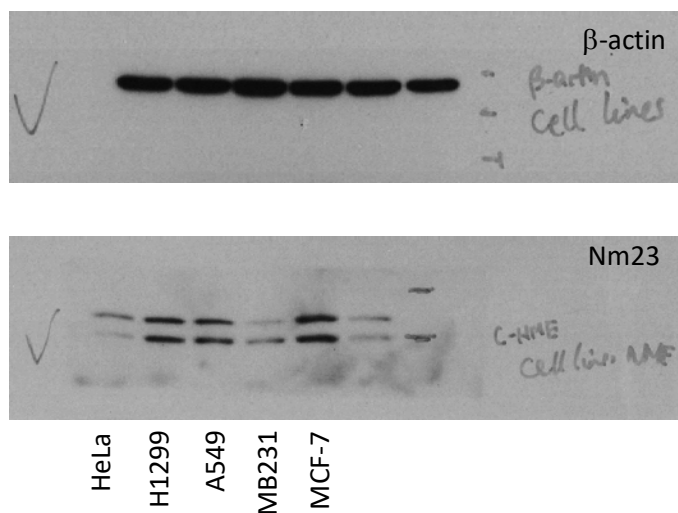
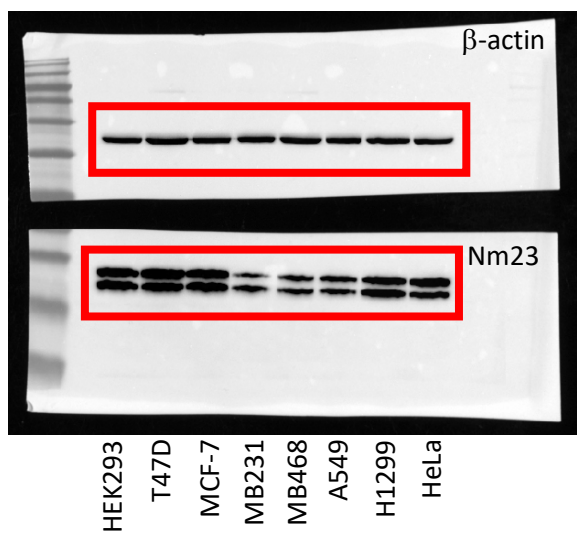
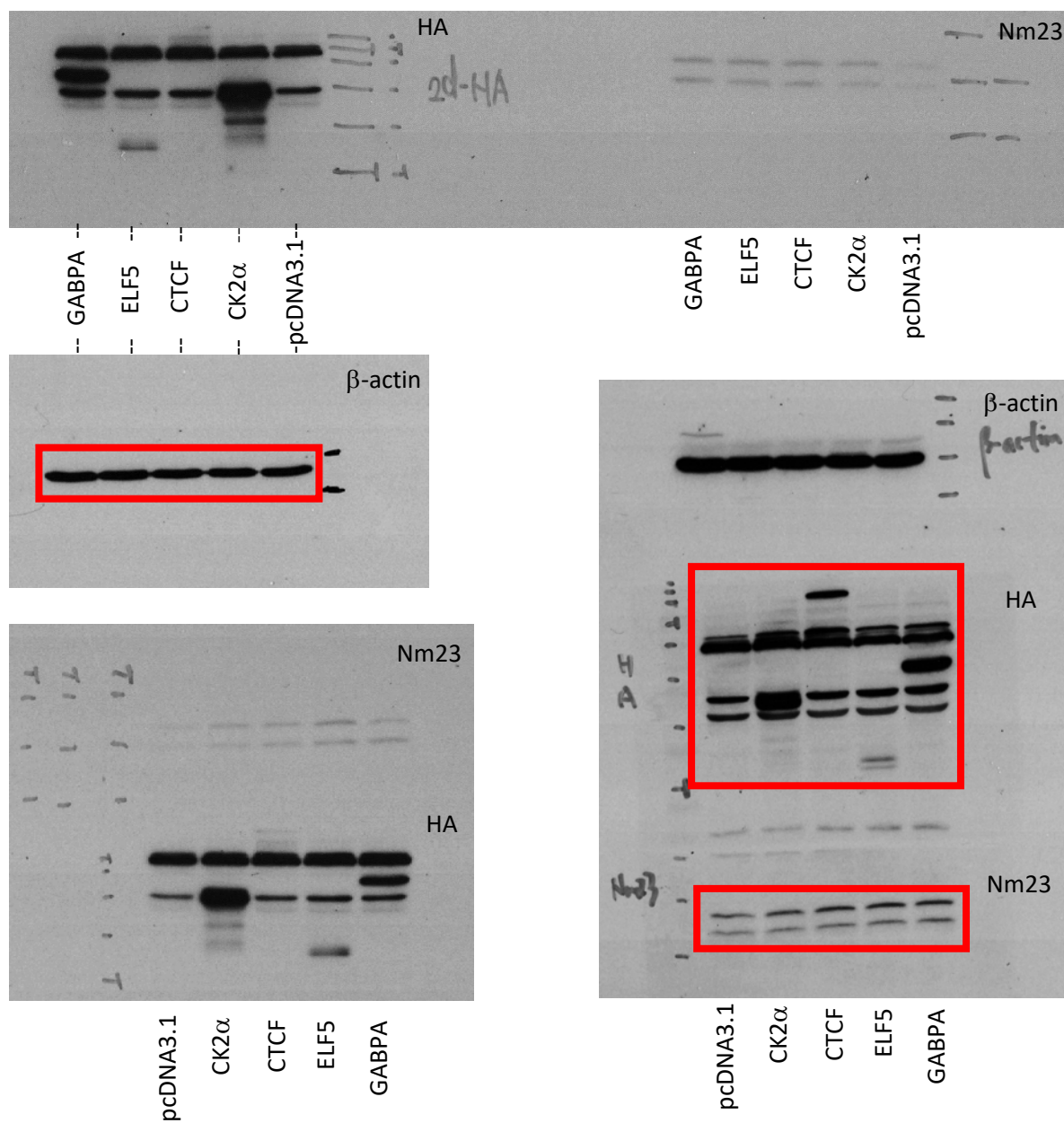
Mutants of the proximal promoter (M1-M7) and minimal promoter (M8-M11) were transiently transfected in HEK293 cells and lysed 48 h after transfection. Luciferase assays were performed using the Dual-Luciferase Reporter Assay System (Promega). RLU values of reference constructs: pGL3-244, *Firefly* luciferase ($31,948,369 \pm 403,518$), *Renilla* luciferase ($3,908,541 \pm 434,200$); pGL3-109, *Firefly* luciferase ($5,316,166 \pm 29,785$), *Renilla* luciferase ($1,329,707 \pm 91,446$). Data represent the mean and standard deviation of three independent trials. *, significance with wild-type promoter, $p < 0.05$.

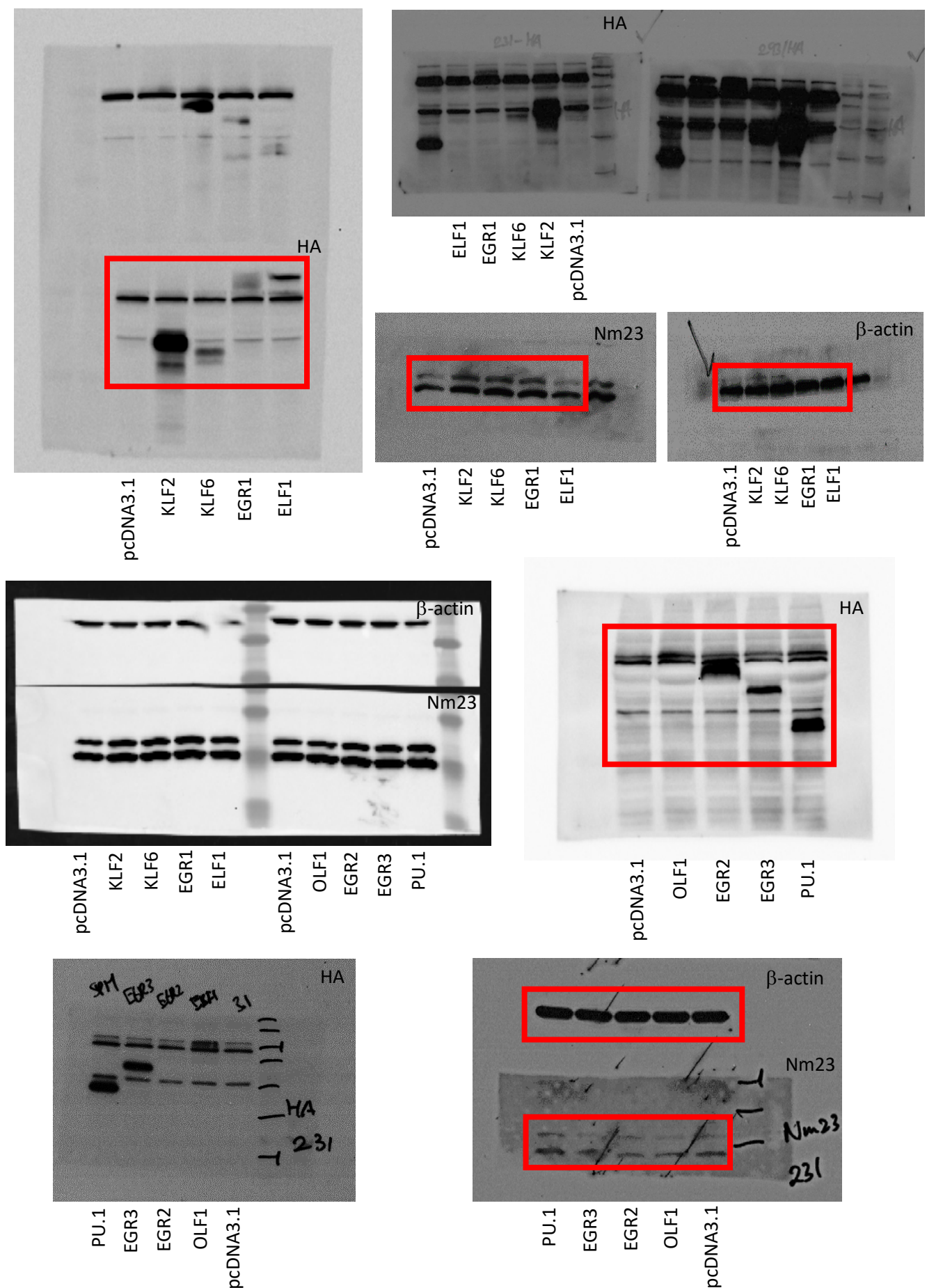


Supplementary Figure S4. Transient expression of Nm23-H1 in MDA-MB-231 cells. MDA-MB-231 cells were transiently transfected with Nm23-H1 or pcDNA3.1 control vector. Proteins were separated on 15% acrylamide gels and immunoblots were probed with corresponding antibodies. Data are shown as a representative experiment from three independent trials.

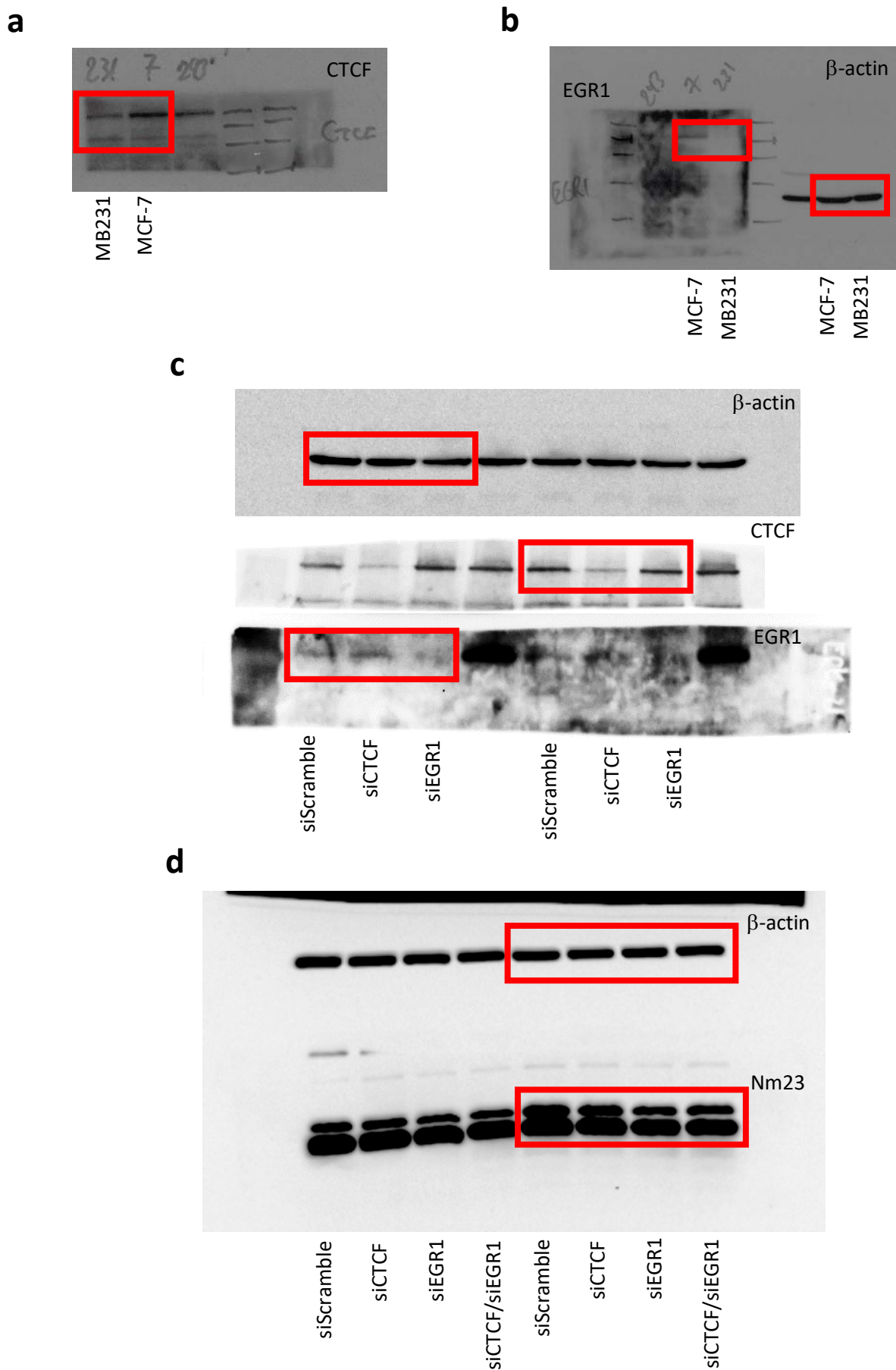


Supplementary Figure S5. Knockdown of Nm23-H1 by siRNA in MCF-7 cells. (a) MCF-7 cells were transiently transfected with siNME1 or siScramble control. RNA extraction was performed with TRIzol, followed by cDNA synthesis and qPCR. *, significance with siScramble, $p < 0.05$. Data represent the mean and standard deviation of three independent trials. (b) Proteins were separated on 15% acrylamide gels and immunoblots were probed with corresponding antibodies.

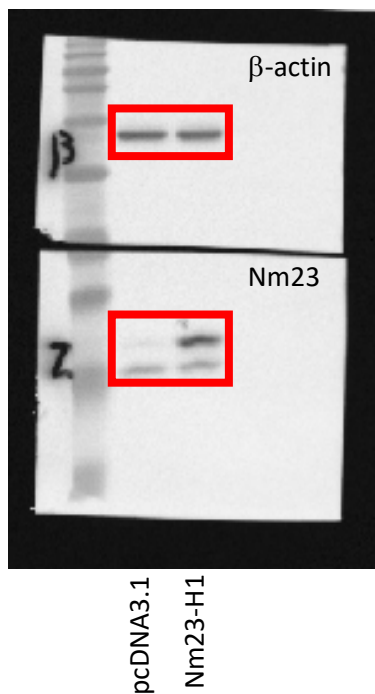
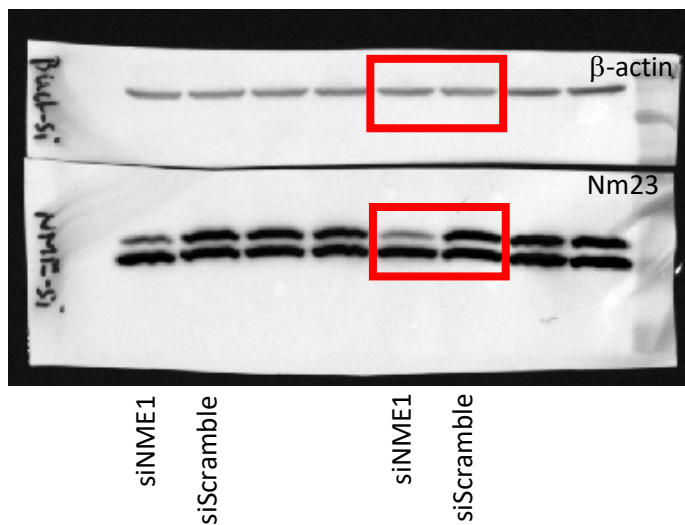
a**b**



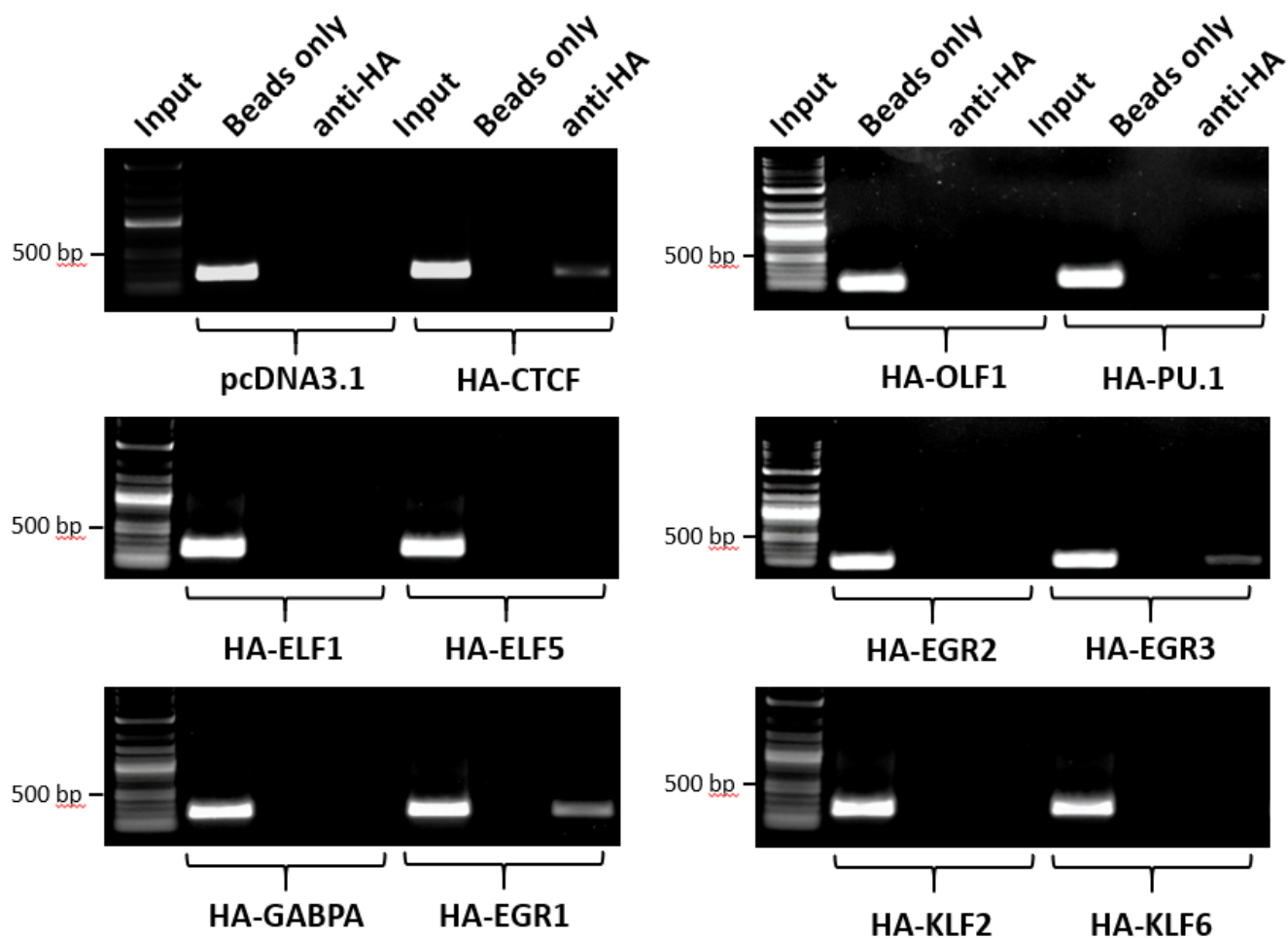
Supplementary Figure S6. Unprocessed original version of Western blots (1). (a) Analysis of Nm23-H1 protein expression in breast cancer cell lines, **Fig. 1a**. (b) Transcription factors regulating Nm23-H1 protein levels in MDA-MB-231 cells, **Fig. 3c**.



Supplementary Figure S7. Uncropped original version of Western blots (2). (a,b) Endogenous expression of CTCF and EGR1 in breast cancer cell lines, **Fig. 4a**. (c) siRNA-knockdown of CTCF and EGR1 in MCF-7 cells, **Fig. 7a**. (d) Nm23-H1 expression upon CTCF and EGR1 knockdown in MCF-7 cells, **Fig. 7d**.

a**b**

Supplementary Figure S8. Uncropped original version of Western blots (3). (a) Transient expression of Nm23-H1 in MDA-MB-231 cells, Supplementary Figure S4. (b) Knockdown of Nm23-H1 by siRNA in MCF-7 cells, Supplementary Figure S5.



Supplementary Figure S9. Unprocessed original version of agarose gels. Transcription factors binding to Nm23-H1 promoter in MDA-MB-231 cells, **Fig. 3d**.

	Gene	Primer sequence
<i>NME1</i> promoter constructs	(1291/1)	F: 5'-TTACTCGAGAGTCTCGCTCTGTCACCCAGGCT-3' R: 5'-AGTAAGCTTGCACGGAACGCTTCTGCG-3'
	(930/1)	F: 5'-TTACTCGAGCTGGCCCTAGGTCCTGAGGATT-3' R: 5'-AGTAAGCTTGCACGGAACGCTTCTGCG-3'
	(657/1)	F: 5'-CGGCTCGAGCTGTATTAGCATCTCACAACATAAGATTTGGC-3' R: 5'-AGTAAGCTTGCACGGAACGCTTCTGCG-3'
	(372/1)	F: 5'-TGA CT CGAGCTAGAATAATGCGTCCACGTAGTAGGT-3' R: 5'-AGTAAGCTTGCACGGAACGCTTCTGCG-3'
	(244/1)	F: 5'-ATTCTCGAGGTGTGAGCGCCACCTCTC-3' R: 5'-AGTAAGCTTGCACGGAACGCTTCTGCG-3'
	(109/1)	F: 5'-TGA CT CGAGAGAAAGTTGTTCCCTAGAATGACTGCCTACT-3' R: 5'-AGTAAGCTTGCACGGAACGCTTCTGCG-3'
	(61/1)	F: 5'-TTACTCGAGGCGAGCGGTGTTCTGCAAAATG-3' R: 5'-AGTAAGCTTGCACGGAACGCTTCTGCG-3'
	(1291/930)	F: 5'-TTACTCGAGAGTCTCGCTCTGTCACCCAGGCT-3' R: 5'-CGCAAGCTTGCAAATAGTTTTAGACGACACCAAAAGACC-3'
	(1291/657)	F: 5'-TTACTCGAGAGTCTCGCTCTGTCACCCAGGCT-3' R: 5'-CGCAAGCTTATTTTGCACAAGGCATGACATCATCAC-3'
	(1291/372)	F: 5'-TTACTCGAGAGTCTCGCTCTGTCACCCAGGCT-3' R: 5'-CGCAAGCTTGCCCTAGGGATAAAACAGTATGCTC-3'
	(1291/244)	F: 5'-TTACTCGAGAGTCTCGCTCTGTCACCCAGGCT-3' R: 5'-AGCAAGCTTGTTGCGTGAAGTCCAAAGAGTGC-3'
	(1291/109)	F: 5'-TTACTCGAGAGTCTCGCTCTGTCACCCAGGCT-3' R: 5'-AGTAAGCTTGCAGTTAACTTCCGGCGCTAG-3'
	(1291/61)	F: 5'-TTACTCGAGAGTCTCGCTCTGTCACCCAGGCT-3' R: 5'-GCTAAGCTTCCACGCTTCTCTTGGGAGT-3'
	(930/657)	F: 5'-TTACTCGAGCTGGCCCTAGGTCCTGAGGATT-3' R: 5'-CGCAAGCTTATTTTGCACAAGGCATGACATCATCAC-3'
	(657/372)	F: 5'-CGGCTCGAGCTGTATTAGCATCTCACAACATAAGATTTGGC-3' R: 5'-CGCAAGCTTGCCCTAGGGATAAAACAGTATGCTC-3'
	(372/244)	F: 5'-TGA CT CGAGCTAGAATAATGCGTCCACGTAGTAGGT-3' R: 5'-AGCAAGCTTGTTGCGTGAAGTCCAAAGAGTGC-3'
	(244/109)	F: 5'-ATTCTCGAGGTGTGAGCGCCACCTCTC-3' R: 5'-AGTAAGCTTGCAGTTAACTTCCGGCGCTAG-3'
	(109/61)	F: 5'-TGA CT CGAGAGAAAGTTGTTCCCTAGAATGACTGCCTACT-3' R: 5'-GCTAAGCTTCCACGCTTCTCTTGGGAGT-3'
Others	ELF1 cDNA	F: 5'- ATGGCTGCTGTTGTCCAACAGAACGACCTA-3' R: 5'-AAAAGAGTTGGGTTCCAGCAGTTCGTTTTGTTTCA-3'
	OLF1 cDNA	F: 5'-ATGTTTGGGATTGAGGAAAGCATCCAAC-3' R: 5'-TCACATAGGAGGAACAATCATGCCAGATATC-3'

Supplementary Table S1. Primers for the cloning of *NME1* promoter regions and cDNAs. *NME1* promoter construct (1291/1) indicates that the region from -1,291 bp to -1 bp upstream of TSS of *NME1* is cloned into pGL3-Basic. Other cDNAs are cloned into pcDNA3.1. F, forward primer. R, reverse primer.

Gene	Primer sequence	Amplicon size
NME1	F: 5'-AAGGAGATCGGCTTGTGGTTT-3'	60
	R: 5'-CTGAGCACAGCTCGTGTAAATC-3'	
GAPDH	F: 5'-AAGTTGTCATGGATGACCTTGGC-3'	206
	R: 5'-GGCGTCTTCACCACCATGGAG-3'	

Supplementary Table S2. Primers for amplification of cDNA in qPCR reactions. F, forward primer. R, reverse primer.